**Edg-1, the G protein–coupled receptor for sphingosine-1-phosphate, is essential for vascular maturation**

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Received for publication July 28, 2000, and accepted in revised form September 12, 2000.

Sphingolipid signaling pathways have been implicated in many critical cellular events. Sphingosine-1-phosphate (SPP), a sphingolipid metabolite found in high concentrations in platelets and blood, stimulates members of the endothelial differentiation gene (Edg) family of G protein–coupled receptors and triggers diverse effects, including cell growth, survival, migration, and morphogenesis. To determine the in vivo functions of the SPP/Edg signaling pathway, we disrupted the Edg1 gene in mice. Edg1−/− mice exhibited embryonic hemorrhage leading to intrauterine death between E12.5 and E14.5. Vasculogenesis and angiogenesis appeared normal in the mutant embryos. However, vascular maturation was incomplete due to a deficiency of vascular smooth muscle cells/pericytes. We also show that Edg-1 mediates an SPP-induced migration response that is defective in mutant cells due to an inability to activate the small GTPase, Rac. Our data reveal Edg-1 to be the first G protein–coupled receptor required for blood vessel formation and show that sphingolipid signaling is essential during mammalian development.


**Introduction**

Sphingolipids have emerged as important signaling molecules in a variety of biologic processes (1–3). SPP in particular has come to the fore as a mediator of an extracellular signaling pathway through its interaction with the family of G protein–coupled receptors known by the acronym, Edg (endothelial differentiation gene) (4). Edg-1, the first of these receptors described, was identified as a gene induced during human endothelial cell differentiation (5). Activation of the Edg receptors triggers diverse effects including proliferation, survival, migration, morphogenesis, adhesion molecule expression, and cytoskeletal changes and has led to the view that the Edg receptor signaling pathways may have important roles in many physiological and pathological events (reviewed in refs. 6–10).

The Edg family can be subdivided into either receptors for SPP or for lysophosphatidic acid. The Edg receptors for SPP activate different and sometimes overlapping G protein–mediated intracellular signaling pathways. For instance, Edg-1 couples directly to the Gi pathway, whereas Edg-3 and -5 stimulate G13 and G12/13 pathways with differing degrees of potency (4, 11–14). Moreover, the expression pattern of individual Edg receptors changes during development and differentiation, leading to different combinations on cells and tissues (15–18). The diverse receptor expression and activation of divergent signaling pathways may explain the pleiotropic responses to SPP but have made functional analysis difficult.

To determine the functions of the SPP/Edg-1 signaling pathway, we have disrupted Edg1 in mice. Homozygous Edg1 mutant mice die in utero due to massive embryonic hemorrhage. They undergo normal vasculogenesis and angiogenesis but are severely impaired in vessel maturation due to a defect in the recruitment of mural cells to vessel walls. The results reveal the SPP receptor Edg-1 as mediating a novel G protein–coupled signaling pathway required for blood vessel development.

**Methods**

Generation of Edg1 mutant mice. To generate the Edg1 knockout mice, we cloned a 10-kb genomic DNA fragment containing the entire Edg1 gene from a 129/Sv
library. As shown in Figure 1a, the Edg1 gene is composed of two exons and an intron (16). The second large exon contains a 5' UTR region, the entire open reading frame region, and approximately 1.8 kb of the 3' UTR region. For knocking-in the LacZ reporter gene and targeted inactivation of the Edg1 gene, a NeoI site in the beginning of the Edg1 open reading frame was used to insert a LacZ-neo (neomycin-resistant gene) cassette (19). In the construct used for disruption of the Edg1 gene, the LacZ coding region is preceded by an internal ribosomal entry sequence (20). Therefore, targeted insertion generates a bi-cistronic transcription unit in which the expression of the β-galactosidase reporter protein is under the control of Edg1 transcriptional regulatory elements. The herp simplex virus thymidine kinase (TK) gene was located outside the homologous sequence to prevent random integration.

Gene targeting in TC1 embryonic stem (ES) cells and generation of chimeric and heterozygous mice were as described previously (21). One targeted ES clone was used to establish chimeric mice, which were crossed with C57BL/6 mice to obtain Edg1 heterozygotes. All mice analyzed were obtained from intercrosses of the Edg1 heterozygotes. Edg1 genotypes were determined by Southern blot and PCR analyses of genomic DNA isolated from ES cells, yolk sacs and tail biopsies. For genotyping by PCR, the primers were: 5'TAGCAGCTATGTTGCCCATCTA3 (Primer 1), 5'GATCCGTGACGATAGATGCG3' (Primer 2), 5'TGGAGGTGACGAGTTTACCTGGA3' (Primer 3), and 5'TCAACACCGCAGATGAGTT3' (Primer 4).

Primers 1 and 2 detected the wild-type Edg1 allele and amplified an approximately 630-bp fragment. Primers 3 and 4 detected the Edg1mut allele and amplified an approximately 350-bp fragment. Forty-five cycles of 94°C (1 minute), 55°C (1 minute), and 72°C (3 minutes) were used.

Histological analysis. Embryos at embryonic days (E) 9.5–16.5 were removed from the mother after heterozygous mating. Then the embryos were fixed and processed to be embedded in paraffin. Serial sections (5-μm thick) were made at 15-μm intervals and stained with hematoxylin and eosin (H&E).

Paraffin sections were deparaffinized and rehydrated. Antigen retrieval was accomplished by 30-minute incubation at 95°C in Target Retrieval Solution (DAKO Corp., Carpenteria, California, USA). Endogenous peroxidase activity was quenched by incubation with 5% hydrogen peroxide in methanol for 5 minutes. Specimens were incubated with anti–smooth muscle actin (EPOS anti-SM α actin, HRP; no. U7033; DAKO Corp.) for 1 hour at room temperature. After washing with PBS, peroxidase reaction was visualized with diaminobenzidine/hydrogen (DAB/hydrogen peroxide).

To define the developmental and tissue-specific expression patterns of Edg1 through X-Gal staining, embryos dissected out from the decidua at various developmental stages were fixed in 2% formaldehyde/2% glutaraldehyde in PBS for 10 minutes. They were washed in PBS and then incubated in PBS containing 5 mM K3Fe(CN)6, 2 mM MgCl2, and 1 mg/ml X-Gal at 37°C overnight. Reactions were stopped by rinsing embryos with PBS, followed by further fixation in 4% paraformaldehyde.

Whole-mount embryo immunostaining. Embryos were dissected out and fixed in 4% paraformaldehyde in PBS at 4°C overnight. They were then dehydrated through a methanol series and stored in 100% methanol at –20°C. The embryos were bleached in 6% hydrogen peroxide/methanol for 1 hour at room temperature and rehydrated through a methanol series to PBS + 0.1% Tween 20 (PBST). They were incubated in a blocking solution (4% BSA in PBST) twice, for 1 hour each time. The embryos were incubated with rat mAb’s (anti–PECAM-1: no.1951D; anti-CD34: no. 09431D; anti–VE-cadherin: no. 28091D; PharMingen, San Diego, California, USA), diluted 1:200 in 10% goat serum and 4% BSA in PBST at 4°C overnight. Embryos were washed with 4% BSA in PBST at room temperature and then incubated with peroxidase-conjugated goat anti-rat Ig in 10% goat serum and 4% BSA in PBST at 4°C overnight. Peroxidase reaction was visualized with DAB/hydrogen peroxide.

RT-PCR and immunoblotting. Total RNA was isolated from E12.5 mouse embryos and cultured cells using Trizol (Life Technologies Inc., Gaithersburg, Maryland, USA) and treated with DNaseI (Life Technologies). Total RNA (5 μg) was reverse transcribed using Superscript Preamplification System (Life Technologies) according to the manufacturer’s instructions. PCR was performed on 2 μl of the RT reaction in a volume of 50 μl using AmpliTaq Gold polymerase (Perkin-Elmer Corp., Norwalk, Connecticut, USA). The PCR conditions were as follows: initial denaturation at 95°C for 10 minutes followed by up to 35 cycles of denaturation at 95°C (1 minute), annealing at 55°C (1 minute), and extension at 72°C (1 min). Amplified PCR products were analyzed by electrophoresis on a 2% agarose gel. PCR primer pairs were as follows:

Flt-1: 5'TGTTGGAGAAACACTTGTTGACCT3', 5'TGGAGAACAGCAGACTCTT3'
Flk-1: 5'TCTGTGTTGCTGCTGGAGA3'; 5'GTATCATTTCCAACCACCT3'
Tie-1: 5'TCCTTTGGCTGCTCCACACTCT3'; 5'ACACACACATTGCCATCAT3'
Tie-2: 5'CCCTCTACACTGGTCATTTA3'; 5'CCACACACTTTGTTTCA3'
Ang-1: 5'AGGGAGGGAAAAAAGAGAAGAG3', 5'GTAGCATGAGAGGGCATTTG3'
Ang-2: 5'TGCGCTACACTCCAGAAGAC3', 5'TATTTACTGCTGAACCTCC3'
PECAM-1: 5'GTCACTGGGACATGGCTGAG3'; 5'CCCTCTGCGACCTTTGCTGAA3'
VE-cadherin: 5'GGATCAGAGGCTCAGACAG3', 5'CTGGCGTTCACGTTTGC3'
Smad5: 5'CTTTTCAACCACCCCACAC3', 5'TCATAGCGGAGGCCTGAAC3'
endoglin: 5'TACTCATGTCCCTGTACCCAGCC3',
For immunoblotting, detergent extracts of E12.5 embryos were analyzed for protein expression using antibodies against the following proteins: VE-cadherin (catalog no. 28091D; PharMingen); N-cadherin (catalog no. SC-7939; Santa Cruz Biotechnology Inc., Santa Cruz, California, USA); and P-cadherin (catalog no. SC-7893), PDGF-B (catalog no. SC-7878), and VEGF (catalog no. SC-507; all from Santa Cruz Biotechnology). A peroxidase-conjugated secondary antibody was used, and the reaction was visualized with the ECL + Plus Western blotting system (Amersham Pharmacia Biotech, Piscataway, New Jersey, USA).

In vitro assays. Chemotactic migration of cells was measured in a modified Boyden chamber as described previously using polycarbonate filters (25 × 80 mm; 12 μm pore size) coated with collagen type I (50 μg/ml in 5% acetic acid), which promotes uniform attachment to and migration across the filter without formation of a barrier (22). SPP or medium without serum was placed in the lower chamber as chemoattractants. Mouse embryonic fibroblasts were harvested and added to the upper chamber at 5 × 10⁴ cells per well. Each data point was the average number of cells in four random fields, each counted twice. Each determination represents the average ± SD of three individual wells. For the detection of GTP-bound activated Rac, embryonic fibroblasts were serum-starved for 24 hours before they were treated with SPP (200 nM) for 5 minutes. The cell lysates were used for affinity precipitation with the PAK-1 p21 binding domain–conjugated (PDB-conjugated) agarose beads (Upstate Biotechnology Inc., Lake Placid, New York, USA) as described elsewhere (23), or were used without fractionation to determine total Rac levels. Rac was visualized by immunoblotting with an mAb.

Results
Generation of Edg1 knockout mice with an inserted β-galactosidase (LacZ) reporter gene. Edg1 consists of two exons (16), with the entire coding region in the second exon (Figure 1a). To disrupt Edg1 in mouse ES cells, we constructed a replacement-type targeting vector in which the Edg1 coding region, containing 382 amino acids, was disrupted after the first 42 amino acids by a Neo cassette containing a internal ribosome entry sequence (19). This targeting strategy should both create a disrupted Edg1 allele and enable analysis of Edg1 expression in mice by creation of an Edg1-LacZ hybrid transcript driven by the endogenous Edg1 promoter elements. The targeting vector was linearized and electroporated into the TC-1 ES cell line. Genomic DNA from G418- and ganciclovir-resistant clones was analyzed by Southern blotting. Of the 120 clones examined, about 60% contained the 2.5-kb BamHI band diagnostic of a homologous recombina-
revealed inheritance of the Edg1 mutant allele at the expected mendelian frequency (Table 1). Up to E11.5, Edg1−/− embryos appeared phenotypically normal. At E12.5, the Edg1−/− embryos could be identified by their abnormal yolk sacs, which were edematous, with less blood in the otherwise normal looking, highly branched, vasculature (Figure 2a, arrows). After removing the yolk sac, intraembryonic bleeding was evident in the Edg1−/− embryos (Figure 2b). The pericardial cavity of mutant embryos was enlarged and filled with fluid. The limbs of mutant embryos were underdeveloped and rounded with areas of bleeding. In comparison, age-matched wild-type embryos had more developed, fan-shaped limbs. Much less blood was found in the yolk sac blood vessels of E13.5 Edg1−/− embryos compared with those of the mutant embryos obtained 1 day earlier (Figure 2c, arrows). At E13.5, massive intraembryonic bleeding could be observed through the yolk sac. In addition to widespread hemorrhage, severe edema was observed.

Figure 1
Targeted disruption and embryonic expression of the Edg1 gene. (a) Schematic representation of the Edg1 targeting strategy. The structure of the mouse Edg1 locus is shown at the top, the structure of the Edg1 targeting vector in the middle, and the predicted structure of the homologous recombinant locus on the bottom. RT-5′ and RT-3′, primers for RT-PCR. B, BamHI; BglI, BglII; PBSK, PBSK, pBluescript vector. (b) Genotyping of mouse offspring from the Edg1 heterozygous mating. Wild-type Edg1 locus yielded a 9.5-kb BamHI band. Disrupted Edg1 locus yielded a 2.5-kb BamHI band. No Edg1−/− mice were found born alive. (c) RT-PCR analysis of total RNA from E12.5 mouse embryos by using RT-5′ and RT-3′. Edg1+/+ and Edg1+/− RNA yielded the predicted 630-bp amplification product. No amplification product was detected from Edg1−/− RNA. (d, e) Whole-mount of Edg1−/−E9.5 and E10.5 embryos stained with X-Gal. H, heart; DA, dorsal aorta; ISA, intersomatic arteries; CP, capillaries; TC, telencephalon; ACV, anterior cardinal vein. (f) Longitudinal section of dorsal aorta (DA) from E10.5 Edg1−/− embryo. LacZ staining is seen in arterial ECs (AEC). (g) Longitudinal section of posterior cardinal vein (PCV) from E10.5 Edg1−/− embryo. LacZ staining is seen in arterial endothelial cells (AEC) but not in venous endothelial cells (VEC). (h) Transverse section of dorsal aorta from E12.5 Edg1−/− embryo. Vascular ECs and VSMCs are stained. EC, endothelial cell; VSMC, vascular smooth muscle cell. Scale bars = 50 μm.
throughout the body of Edg1−/− embryos (Figure 3d). No Edg1−/− embryos survived beyond E14.5 (Table 1).

Normal vasculogenesis in Edg1−/− embryos. To define the vascular system in the Edg1−/− embryos, the morphology of the vasculature was characterized by whole-mount immunohistochemical staining using mAb’s against markers for endothelial cells. Antibodies to CD34 (Figure 2, e and f) and platelet endothelial cell adhesion molecule-1 (PECAM-1) (data not shown) revealed a substantially normal arborized vascular network both in the mutants and age-matched control embryos. High magnification views showed capillary sprouts in the head of mutant embryos (Figure 2, g and h, black arrowheads). However, the small vessels in the forebrain of mutant embryos appeared dilated and stained darker than controls with antibodies against PECAM-1 and CD34 (white arrowheads).

Because SPP signaling through Edg-1 was shown to be involved in adherens junction assembly in HUVEC cells in vitro (24), we investigated the expression of VE-cadherin, P-cadherin, E-cadherin, N-cadherin, and PECAM-1 in Edg1−/− embryos by using whole-mount immuno-

The Journal of Clinical Investigation | October 2000 | Volume 106 | Number 8

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and the phase of angiogenesis that entails vessel sprouting and penetration had occurred.

**Vascular smooth muscle defects in Edg1−/− embryos.** After the initial formation of the vascular plexus, vessels mature by the stabilization of the endothelial vascular network through a recruitment and differentiation process that ultimately results in the investment of vessel walls with mural cells (26).

Vascular smooth muscle cells (VSMCs) first appear on the ventral side of the aorta in E10.5 embryos, followed by migration to the dorsum (27). By E11.5, the aorta is completely enveloped by VSMCs (27). To assess this aspect of vessel development in the Edg1 mutant embryos, VSMCs were identified using an antibody to SMαA. In longitudinal sections of E12.5 control embryos stained with anti-SMαA, the dorsal aortae were found to be completely surrounded by VSMCs (Figure 3a). The aortae of Edg1−/− mice were strikingly different. SMαA-positive VSMCs were present on the ventral surface; however, VSMCs were deficient along the entire length of the dorsal surface examined (Figure 3b). Transverse sections of aortae from control embryos showed two to three layers of VSMCs surrounding the
vessel (Figure 3c). In contrast, similar sections from Edg1−/− embryos showed that the aortae were covered only ventrally by poorly organized SMαA-expressing cells (Figure 3d). SMαA-expressing cells were not found on the dorsal side of the mutant aortae. These results suggested initial recruitment and differentiation had taken place to produce VSMCs on the ventral side of the mutant aorta, but that the process leading to the complete envelopment of the vessel was defective. Endothelial cell morphology appeared normal in E11.5 embryos, before the onset of bleeding (Figure 3e). In the E12.5 embryos, after the onset of bleeding, the dorsal aortic surface (uncovered by VSMCs) appeared abnormal and discontinuous (Figure 3, d and f).

In E12.5 control embryos, the majority of medium-sized arteries, identified by association with SMαA-positive VSMCs, were surrounded by a continuous layer of VSMCs (Figure 3g). Only rare vessels were found with an incomplete covering by VSMCs. In contrast, a substantial fraction of intracerebral arteries in the mutant embryos displayed a discontinuous or patchy covering by VSMCs (Figure 3h). Bleeding from these arteries was apparent by the presence of blood cells in the surrounding tissue space (Figure 3h).

The muscular layers in the gastrointestinal tract (Figure 3, i and j) and bronchial tree (data not shown) were well developed in the mutant embryos, indicating that there was not a generalized defect in smooth muscle.

The blood vessel defects in the Edg1−/− embryos were further analyzed by electron microscopy. Small blood vessels from the limb of E12.5 Edg1−/− embryos illustrated a marked reduction of VSMCs/pericytes adjacent to the endothelial cells (Figure 4, a and b). The endothelial cell body was very thin, and in some areas, fragmented (data not shown). Intracerebral capillaries of mutant embryos appeared without associated microvascular pericytes (Figure 4, c and d). The endothelial cell nuclei of the mutant capillaries were abnormally rounded and enlarged. The areas surrounding these vessels were generally much less densely packed with cells. Mutant blood vessels contained normal-appearing, electron-dense interendothelial junctions (Figure 4, e and f), suggesting that endothelial cell-cell junctions formation occurred in the absence of Edg-1. Given that PDGF-B and its receptor β are critical for the investment of capillaries with pericytes (28–30), we determined their expression in mutant embryos. We found that in Edg1−/− embryos PDGF-B expression was normal as determined by Western analysis and that PDGF-receptor β was highly expressed in mesenchymal cells by immunohistochemical analysis (data not shown). Mice deficient in the transcription factor LKLF also show marked reductions in VSMCs and pericytes around vessels (31). RT-PCR of E12.5 Edg1−/− RNA indicated that LKLF expression was similar to control levels (data not shown).

**Figure 4**
Reduced VSMCs and pericytes in the Edg1−/− vessels. (a–d) EM microscopic analyses of representative small blood vessels from the limb (a and b) and brain capillaries (c and d) from E12.5 wild-type and Edg1−/− embryos. Reduced number of VSMCs (bracket in b) and the lack of capillary pericytes (PC) were found in the Edg1−/− embryos. Notice the abnormally rounded EC nucleus in the Edg1−/− capillary (d). (e and f) EC junctions (arrows) in wild-type and mutant embryos. Note the normal EC junction (E) in the Edg1−/− embryo (f). BC, blood cell. ×2,000 (a–d); ×50,000 (e and f).
SPP on Rac activation in wild-type and Edg1−/− fibroblasts (Figure 5c). In wild-type fibroblasts, SPP treatment resulted in a substantial increase in the amount of activated Rac. By contrast, no increase in activated Rac could be detected after SPP treatment of Edg1−/− fibroblasts, demonstrating that Edg-1 is required for the SPP induction of activated Rac.

Discussion
A number of receptor-mediated signaling pathways have been identified that coordinate the stages of blood vessel formation. Disruption of the genes encoding these receptors and ligands in mice has been instrumental in defining their roles (reviewed in refs. 26, 37, 38). Vasculogenesis is dependent on VEGF and its receptor tyrosine kinases, Flk-1 and Flt-1, expressed in endothelial cells (39–42). VEGF, Flk-1, and Flt-1 knockout mice die between E8.5 and E9.5 as a result of defects in the formation of the primitive vasculature. Angiogenesis and vascular remodeling require the Tie-2 receptor tyrosine kinase on endothelial cells and its ligand, angiopoietin-1. Without Tie-2, mice have normal vasculogenesis but defective vessel sprouting, branching, and remodeling; they die at E10.5. Mice devoid of angiopoietin-1 have a similar phenotype (43–45).

Signaling pathways have also been implicated in the recruitment and differentiation of mural cells during vessel maturation. PDGF-B and its receptor-β have been shown to be essential for the recruitment of mesenchymally derived mural cell precursors to vessel walls (30, 46). Disruption of the PDGF-B or PDGF receptor-β genes in mice leads to lethal hemorrhage and edema in the perinatal stage owing to a lack of microvasculature pericytes (28–30). TGF-β1 induces differentiation of VSMCs (47) after their recruitment to endothelial walls (48, 49). The endothelial TGF-β1 binding protein, endoglin, and its downstream signaling molecule, SMAD5, both important in the TGFβ1 signaling pathway, have essential roles in VSMC differentiation (50, 51). Endoglin- and Smad5-deficient mice die between E10.5 and E11.5 with a lack of VSMCs around major vessels.

These studies point to the critical roles played by receptor tyrosine kinases during vascular development. In contrast, the involvement of G-protein–coupled signaling pathways have not been as well characterized during development. Such pathways are important because disruption of the Ga13 gene results in defects in embryonic vasculature formation, presumably due to a migration defect (52). However, upstream receptors involved had not been defined.

Role of Edg-1 during early vascular system development. We found that Edg1 was highly expressed in the cardiovas-
cular system during early embryonic development. Vascular endothelial cells expressed Edg1 at relatively high levels, although expression was almost exclusively found in the endothelial cells of arteries rather than of veins. A low but detectable expression was also found in VSMCs surrounding the aorta. Expression of Edg1 was prominent in cardiomyocytes, although no gross abnormalities were detected in the developing heart. In addition to the vascular system, Edg1 was found highly expressed in the developing central nervous system as has been shown previously (16, 18).

Severe bleeding caused lethality in Edg1–/– embryos between E12.5 and E14.5. However, the mutant embryos showed a substantially normal blood vessel network when stained with antibodies to markers specific for differentiated endothelial cells such as PECAM-1 and CD34. The expression of genes known to be important for vasculogenesis and angiogenesis was not measurably affected in Edg1–/– embryos. These genes included VEGF, Flk-1, Flt-1, Ang-1, and Tie-2. Each phenotype of these knockout mice is quite distinct from that of the Edg1–/– mice, which develop a relatively normal appearing vascular network and die between E12.5 and E14.5. These results indicate that Edg1 is dispensable for vascular endothelial cell differentiation, proliferation, migration, and tube formation during vasculogenesis and for the stage of angiogenesis involving vessel sprouting and branching. The morphology and viability of endothelial cells in the mutants appeared normal until the onset of severe bleeding, suggesting that the morphology changes were secondary to the lack of supporting VSMCs and subsequent disruption of the vessels in the mutant embryos.

Although SPP signaling through Edg-1 has been shown to regulate adherens junction formation in vitro in human endothelial cells, we found no evidence of aberrant endothelial junctions in Edg1 mutant mice during development. Ultrastructural analysis of mutant endothelial cells revealed normal appearing cell-cell junctions. The phenotype of the Edg1 mutant embryos was also not in keeping with significant defects in endothelial cell junctions. Recently, it was shown that VE-cadherin, an important component of adherens junctions, controls an endothelial cell survival pathway through its intracellular interaction with β-catenin (25). Disruption of this pathway in mice resulted in impaired angiogenesis, increased endothelial apoptosis, and embryonic death by E9.5. The Edg1 mutant mice exhibited none of these characteristics. Thus the formation of functional endothelial junctions apparently proceeds normally during early development in the absence of Edg-1. This could indicate that there is functional redundancy among members of the Edg family for this process and that other Edg proteins may substitute for Edg-1. It may also indicate that the SPP-Edg regulation of adherens junction assembly is not required for blood vessel formation during development. However, this would not preclude a role for the pathway during angiogenesis in the adult.

**Functions of Edg-1 during vessel maturation.** In dorsal aorta, VSMC investment is initiated on the ventral side with a condensation of SMαA-positive cells. The recruitment process continues dorsally until VSMCs have completely enveloped the endothelial tube (27). We found that the aortae of wild-type and heterozygous embryos were surrounded by several layers of elongated VSMCs. The aortae in the Edg1–/– embryos were strikingly abnormal in VSMC investment. Aortic sections demonstrated the presence of multiple layers of SMαA-positive cells, but only at the ventral surface. These results suggest that the defect in VSMC investment of vessel walls in Edg1 mutant embryos was expressed after the initial VSMC recruitment to the ventral aortic surface has taken place. This vessel abnormality was distinct from that observed in mice deficient in endoglin, the TGF-β binding protein on endothelial cells, and in SMAD5, the TGF-β1 signaling molecule. Both of these proteins are required for VSMC differentiation; the respective knockout mice have severely defective VSMC development with almost no SMαA-positive cells around vessels (50, 51).

In addition to a defect in mural cell recruitment in dorsal aorta, the Edg1–/– mice exhibited defects in the smaller vessels and in the microvasculature. By electron microscopy, we found evidence of a lack of pericytes associated with capillaries. When pericytes are deficient as in PDGF-B–/– mice, dilated microvessels develop that are prone to rupture (53). Similarly, in the Edg1–/– mice, dilation of small cranial vessels and bleeding were evident.

**How does Edg-1 so dramatically influence the recruitment of VSMCs to vessels during development?** With fibroblasts from the Edg1 mutant mice, we found that in the absence of Edg-1, the SPP-induced activation of Rac seen in wild-type cells did not occur. Rac is a key regulator of the actin cytoskeleton and of associated activities such as cell motility (36, 54). Thus, the Edg1–/– fibroblasts, without this signaling pathway, were unable to mount a migration response to SPP. This signaling pathway may also operate in VSMCs, and its disruption could be responsible for the defect in vascular maturation seen in the Edg1–/– mice. Consistent with this hypothesis, we found that Edg-1 mediates migration of normal VSMCs toward SPP (data not shown). SPP is abundantly stored in platelets and secreted after stimulation (55). Other blood cells, including erythrocytes, neutrophils, and mononuclear cells, produce and secrete SPP constitutively (56), resulting in significant SPP levels in blood. During maturation of the dorsal aorta and possibly other vessels, plasma containing SPP could escape from immature, leaky vessels and act as a signal to recruit VSMCs that are differentiating in the proximity of vessel walls (Figure 5d).

Recently, it was shown that the zebra fish gene, *mil*, encodes an SPP-binding, G protein-coupled receptor of the Edg family that directs the migration of heart precursors to the midline during embryonic development (57). Interestingly, *mil* does not function in the
migrating precursor cells, but in the paraxial cells at the midline, presumably by creating a permissive environment for migration. Sequence and functional similarities suggest that mld may be the ortholog of the mammalian Edg5 gene. However, the indirect influence of mld on migrating cells, as well as the high level of expression of Edg1 in arterial endothelial cells, raises the possibility that Edg-1 stimulation on endothelial cells may regulate the recruitment of VSMCs (Figure 5d). This might occur via the upregulation of adhesion molecules on endothelial cells, or by stimulating the secretion of recruitment factors for VSMCs. Further studies using tissue-specific knockouts of Edg1 will be required to address precisely how Edg-1 functions. Nevertheless, our data demonstrate an indispensable role of Edg1 in vascular maturation and, together with the results showing mld is essential for heart organogenesis, indicate that different members of the Edg receptor family regulate distinct aspects of cardiovascular development through sphingolipid signaling pathways.

In summary, we have uncovered a unique and vital role for the G protein–coupled receptor, Edg-1, in blood vessel formation during development. The SPP-Edg signaling pathway may also have important functions in adult vascular biology. SPP stimulation of Edg receptors on endothelial cells in vitro results in proliferation, migration, and tube formation (24, 32, 58), all requisite for angiogenesis. Treatment through sphingolipid signaling pathways.

Acknowledgments
We thank J. Van Broklyn for help in the early stages of this project and A. Howard for producing the figures.