Thermal Environment and Acid-Base Homeostasis in Human Infants during the First Few Hours of Life *

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It has recently been shown that labor and delivery result in variable degrees of respiratory and metabolic acidosis in the human infant (1). With the prompt establishment of effective ventilation, the healthy newborn achieves a relatively normal acid-base state in the first few hours of life (2). The optimal thermal environment for this recovery is unknown.

It is generally accepted that the newborn infant is a true homeotherm from the time of birth. Consequently, it is reasonable to suppose that the increased metabolism in a cool environment might lead to an accumulation of lactic, pyruvic, and other organic acids, thereby aggravating the existing acidosis. This would be particularly true if metabolic needs were to exceed oxygen availability to the tissues. Conversely, it could be proposed that the cool environment might favorably influence the acid-base homeostasis if the formation of acid metabolites were reduced at a lower body temperature.

With these possibilities in mind a study was designed to explore the relationship between acid-base status and thermal environment in the first hours of life. It was hoped that such information would contribute to our understanding of the optimal environmental temperature for the newborn.

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Methods

In an initial series of observations, 24 healthy full-term infants from the obstetric service of the Sloane Hospital were studied. The infants were divided into two groups, "warm" and "cool." Each of these two groups consisted of six infants delivered by elective Cesarean section and six delivered vaginally. The two groups were considered comparable in regard to premedication, anesthesia, birth weight, and Apgar score (3) at 1 minute (see Table I). All vaginal deliveries were uncomplicated, being either spontaneous or by low forceps.

The environment to which a particular infant was assigned was determined before delivery by a random permutation of numbers in groups of four. The period of observation was continued for approximately 2 hours from the time of birth. In the warm group deep rectal temperature was maintained at approximately 37.0°C by placing the infant immediately after birth under a thermostatically controlled radiant heating device. They were left exposed to the radiant heat except when draping was necessary for obtaining blood samples. Infants in the "cool" group were left uncovered at room temperature (mean, 25.0°C; range, 22.5 to 26.5°C). Temperatures in all infants were monitored continuously by thermistor probes and a multichannel polygraph. Thermistors were placed in the rectum (8 cm from anus), on the skin of the anterior abdominal wall, and in the room air. In the warm group the skin thermistor was shielded from the direct heating of the infrared lamp.

1 Apgar scoring system. This provides objective means of evaluating the over-all condition of a newborn infant. It consists of assigning a score of 0, 1, or 2 to each of five criteria, namely heart rate, respiratory performance, reflex irritability, muscle tone, and color. The score is recorded routinely at 1 minute of age. Under this system, infants who score 7 or higher are classed as being in good condition, those who score between 4 and 6 are moderately depressed, and those whose score falls between 0 and 3 are severely depressed.

2 Experimental model made by Airshields, Inc., Hatboro, Pa.
TABLE I

Summary of clinical data in 24 infants*

<table>
<thead>
<tr>
<th>Group</th>
<th>Birth wt</th>
<th>Apgar score</th>
<th>Pregnancy</th>
<th>Medication</th>
<th>Anesthesia</th>
<th>Delivery</th>
<th>Complication</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>kg</td>
<td>I min</td>
<td>mg</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Warm VV</td>
<td>3.3</td>
<td>5</td>
<td>Normal</td>
<td>Demerol 75</td>
<td>N₂O</td>
<td>LF</td>
<td>Nuchal cord</td>
</tr>
<tr>
<td></td>
<td>3.2</td>
<td>7</td>
<td>Normal</td>
<td>0</td>
<td>Caudal and N₂O</td>
<td>LF</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>3.1</td>
<td>7</td>
<td>Normal</td>
<td>Demerol 25</td>
<td>Caudal and N₂O</td>
<td>LF</td>
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</tr>
<tr>
<td></td>
<td>2.7</td>
<td>8</td>
<td>Normal</td>
<td>Demerol 50</td>
<td>N₂O</td>
<td>NSD</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>3.4</td>
<td>7</td>
<td>Normal</td>
<td>0</td>
<td>Saddle block</td>
<td>NSD</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>2.6</td>
<td>6</td>
<td>Normal</td>
<td>Demerol 75</td>
<td>Caudal and N₂O</td>
<td>NSD</td>
<td>None</td>
</tr>
<tr>
<td>CS</td>
<td>3.0</td>
<td>8</td>
<td>Normal</td>
<td>Atropine 0.4</td>
<td>Spinal</td>
<td>Elective CS</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>4.7</td>
<td>7</td>
<td>Normal</td>
<td>Atropine 0.4</td>
<td>Spinal</td>
<td>Elective CS</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>3.5</td>
<td>8</td>
<td>Mild</td>
<td>Atropine 0.4</td>
<td>Spinal</td>
<td>CS</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>2.7</td>
<td>8</td>
<td>Normal</td>
<td>Atropine 0.4</td>
<td>Spinal</td>
<td>Elective CS</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>3.0</td>
<td>8</td>
<td>Normal</td>
<td>Atropine 0.4</td>
<td>Spinal</td>
<td>Elective CS</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>3.3</td>
<td>9</td>
<td>Normal</td>
<td>Atropine 0.4</td>
<td>Spinal</td>
<td>Elective CS</td>
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</tr>
<tr>
<td>Cool VV</td>
<td>3.0</td>
<td>9</td>
<td>Normal</td>
<td>Seconal 100</td>
<td>N₂O</td>
<td>LF</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>2.5</td>
<td>8</td>
<td>Normal</td>
<td>Demerol 75</td>
<td>N₂O</td>
<td>LF</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>3.3</td>
<td>6</td>
<td>Normal</td>
<td>Demerol 75</td>
<td>Epidural</td>
<td>LF</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>3.2</td>
<td>8</td>
<td>Normal</td>
<td>0</td>
<td>Caudal</td>
<td>NSD</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>3.2</td>
<td>8</td>
<td>Normal</td>
<td>0</td>
<td>Caudal</td>
<td>NSD</td>
<td>None</td>
</tr>
<tr>
<td>CS</td>
<td>3.0</td>
<td>7</td>
<td>Normal</td>
<td>Atropine 0.4</td>
<td>Thioptental, Succinylcholine, N₂O</td>
<td>Elective CS</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>3.6</td>
<td>8</td>
<td>Normal</td>
<td>Atropine 0.4</td>
<td>Thioptental, Succinylcholine, N₂O</td>
<td>Elective CS</td>
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</tr>
<tr>
<td></td>
<td>2.7</td>
<td>9</td>
<td>Normal</td>
<td>Atropine 0.4</td>
<td>Thioptental, Succinylcholine, N₂O</td>
<td>Elective CS</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>3.0</td>
<td>7</td>
<td>Normal</td>
<td>Atropine 0.4</td>
<td>Thioptental, Succinylcholine, N₂O</td>
<td>Elective CS</td>
<td>None</td>
</tr>
<tr>
<td></td>
<td>2.4</td>
<td>8</td>
<td>Normal</td>
<td>Atropine 0.4</td>
<td>Spinal</td>
<td>Elective CS</td>
<td>None</td>
</tr>
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<td></td>
<td>3.2</td>
<td>7</td>
<td>Normal</td>
<td>Atropine 0.4</td>
<td>Spinal</td>
<td>Elective CS</td>
<td>None</td>
</tr>
</tbody>
</table>

* VV = vaginal vertex; CS = Cesarean section; LF = low forceps; NSD = normal spontaneous delivery.

Initial blood samples were taken from the artery of a doubly clamped segment of the umbilical cord. Subsequent specimens were obtained either by direct needle puncture of the femoral artery or by catheterization of the umbilical artery. The samples were drawn into greased, heparinized syringes and stored in iced water. A certain number of the acid-base determinations were done on "arterialized" capillary blood, since results from this laboratory have shown a good correlation between arterial blood and that obtained from a heel prick, provided the extremity is adequately warmed for 10 minutes and precautions are taken to prevent the loss of CO₂. All determinations were done within 30 minutes of taking the sample.

Hydrogen ion activity of whole blood was determined using the radiometer microglass electrode (4). Buffer base (B.B.), base excess (B.E.), a total CO₂ and CO₂ tension (Pco₂) were calculated from pH measurements.

a Base excess, a term recently introduced by Astrup, Andersen, Jørgensen, and Engel (5), represents the amount of fixed acid or base in milliequivalents per liter required to restore the pH of a blood sample to 7.38 at 38°C and at a Pco₂ of 40 mm Hg. By definition the normal value is zero. Since a negative value for base excess is somewhat confusing, we have elected to use the term base deficit. Thus base excess connotes metabolic alkalosis and base deficit, metabolic acidosis.
of blood equilibrated at two known CO₂ tensions according to the method described by Astrup, Siggaard Andersen, Jørgensen, and Engel (3) and Siggaard Andersen and Engel (6). As all pH determinations were done at 38° C, the obtained values were corrected for differences in temperature by the Rosenthal factor of 0.0147 pH per degree C (7). Similarly, corrections were made for Paco₂ by using the factor of 4.4% per 1° C given by Bradley, Stupfel, and Severinghaus (8). It was arbitrarily decided to use the babies' deep rectal temperature to calculate the above corrections.

The mean pH was computed from individual pH values after converting them into microequivalents of hydrogen ion per liter. Over the rather narrow pH range these means did not differ significantly from the arithmetic means of the numerical pH values.

Buffer base and base excess were determined on all samples at full oxygen saturation; however, only base excess or base deficit values are referred to in the results section, because buffer base, unlike base excess, is dependent upon the hemoglobin concentration (9). This latter determination was not carried out on all samples.

Similar observations were made on eight healthy and four depressed full-term infants, in whom measurements of oxygen consumption as well as acid-base state were obtained under both warm and cool conditions. The healthy infants all had an Apgar score of 7 or higher at 1 minute, whereas the depressed group scored 1, 2, 2, and 6, respectively. All of the latter infants responded promptly to resuscitation, and their subsequent nursery progress was uneventful. These infants were placed as soon as possible after delivery, usually within 15 minutes, into a double-walled box which was in series with a closed circuit for measurement of oxygen consumption. Air was circulated at a constant temperature (approximately 34° C). Radiant heat loss was prevented by circulating water at 37° C through the double wall of the box. In this way the infants' rectal and skin temperatures were maintained at about 37° C. After a period of at least 2 hours under these conditions during which time the healthy infants achieved a relatively normal acid-base status, blood samples were obtained. The temperature in the circuit was then reduced to approximately 23° C. When the circuit temperature had equilibrated, the infant's oxygen consumption was again measured. This mild cold stress was maintained for approximately 60 minutes, after which further blood samples were obtained.

The samples were analyzed for pH, Paco₂, CO₂ content, and base deficit. In most cases, lactate and pyruvate levels were determined by modifications of the methods of Barker and Summerson for lactate (10) and Friedemann and Haugen for pyruvate (11).

Results

These will be presented in two sections, the first dealing with the 24 healthy infants randomly assigned either to a "warm" or "cool" environment and the second dealing with the infants studied under both thermal conditions.

Section I

The results were analyzed in two stages, first by comparing the effect of the mode of delivery within both warm and cool groups and secondly by comparing the effect of environmental temperature. It was established by the t test that

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**Fig. 1.** Mean rectal and skin temperatures in 24 infants at two different environmental temperatures during the first 2 hours of life. Temperature at birth is assumed to be 37.5° C. Measurements were not obtained in all infants. □ = rectal temperature, ▽ = skin temperature—cool; ■ = rectal temperature, ▼ = skin temperature—warm. Vertical bars represent ± 1 SE of the mean.
there were no significant differences between infants delivered by Cesarean section and those born vaginally either at birth or subsequently. Therefore, the values obtained within each temperature group were combined and the "warm" group compared with the "cool" by the t test.

Temperature. In the warm group the mean rectal and skin temperatures were maintained at approximately 37 to 37.5°C (Figure 1). In the cool group there was a pronounced fall in skin as well as deep body temperature after delivery, the fall having the characteristics of an exponential function. In the initial 15 minutes after birth the skin temperature fell almost 4°C. During the ensuing 45 minutes a further drop of about 0.5°C occurred, the temperature finally leveling at approximately 33°C after the first hour. The rectal temperature reached approximately 35°C after 2 hours. From the age of 15 minutes, there was a temperature gradient of 2.5 to 3°C between skin and rectum. The cooler infants were much more active than those in the warm environment. Intermittent shivering was observed, although seldom in the first 15 minutes.

Hydrogen ion activity. The rate of rise of pH in both warm and cool groups was similar, being fairly rapid over the first hour and more gradual over the second hour as normal values were approached (Figure 2). At the end of 2 hours the mean pH value was 7.36 in both warm and cool groups.

CO₂ tension. PCO₂ fell rapidly in both groups (Figure 3). However, at the end of the first hour the value in the warm group was significantly higher (40 mm Hg) compared to the cool group (33 mm Hg) (p < 0.02). At the end of the second hour there was still a significant difference between the two groups (p < 0.01), the mean value in the warm group being 37 mm Hg compared to 31 mm Hg in the cool group.

Total CO₂. The initial values in the two groups were similar, but after birth there was a small but consistent difference (Figure 4). At the end of 2 hours the mean value in the warm group was 22 mEq per L as opposed to 19 mEq
per L in the cool group. The difference was statistically significant (p < 0.01).

**Base deficit.** The mean values at birth were essentially identical in the two groups (Figure 5). Infants kept in the warm environment showed a steady elimination of base deficit, the mean value reaching 1.5 mEq per L at the end of 2 hours. By contrast the base deficit in the cool group after 2 hours averaged 5.5 mEq per L, an insignificant fall during the cold exposure. The difference between the means at 1 and 2 hours was statistically significant (p < 0.01).

**Section II**

Data from eight healthy infants studied under both warm and cool conditions are summarized in Table II. The rectal and skin temperatures were maintained at approximately 37° C for the first 2 hours of life. During this period, the infants showed almost complete recovery from birth asphyxia. The mean pH rose from 7.26 to 7.35; Pco2 fell from 58 to 39 mm Hg, and base deficit fell from 5.6 to 2.1 mEq per L. After being placed in the cool environment (23° C), where they remained for the next hour, the oxygen consumption immediately doubled. Despite this increase in metabolic rate, there was a fall in rectal temperature to a mean value of 34.2° C. The rate of fall in temperature was gradual compared with the initial precipitous fall seen in infants placed in a cool environment at birth. During this time the pH remained unchanged, although there was a significant increase in base deficit (p < 0.05). This was accompanied by a reduction in Pco2 (p < 0.05) and total CO2 (p < 0.01). Blood lactate rose slightly; however, the ratio of lactate to pyruvate remained unchanged in three infants and showed only a small increase in two.

Similar measurements made in four depressed infants (Table III) revealed distinct differences in behavior. Exposure to the cool environment was followed in all instances by a fall in blood pH. This was due to a pronounced metabolic acidosis, as shown by a rise in base deficit up

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**TABLE II**

*Mean values ± SE for pH, Pco2, total CO2, base deficit, lactate, pyruvate, O2 consumption, and rectal temperature in eight healthy newborn infants, Apgar score 7 or higher at birth, and under warm and cool conditions*

<table>
<thead>
<tr>
<th></th>
<th>pH</th>
<th>Pco2 mm Hg</th>
<th>Total CO2 mmol/L</th>
<th>Base deficit mEq/L</th>
<th>Lactate* mEq/L</th>
<th>Pyruvate* mEq/L</th>
<th>O2 consumption ml/kg/min</th>
<th>Rectal temp. °C</th>
</tr>
</thead>
<tbody>
<tr>
<td>Birth</td>
<td>7.26 ± 0.020</td>
<td>58 ± 5</td>
<td>27 ± 0.5</td>
<td>5.6 ± 1.4</td>
<td>4.0 ± 0.6</td>
<td>0.27 ± 0.07</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Warm, 34° C, age 2 hrs</td>
<td>7.35 ± 0.006</td>
<td>39 ± 2.1</td>
<td>22 ± 1.0</td>
<td>2.1 ± 1.7</td>
<td>2.8 ± 0.4</td>
<td>0.27 ± 0.04</td>
<td>5.0 ± 0.3</td>
<td>37 ± 0.2</td>
</tr>
<tr>
<td>Cool, 23° C</td>
<td>7.35 ± 0.013</td>
<td>34 ± 2.1</td>
<td>20 ± 0.6</td>
<td>4.7 ± 1.5</td>
<td>3.4 ± 0.5</td>
<td>0.22 ± 0.05</td>
<td>10.8 ± 0.5</td>
<td>34.2 ± 0.4</td>
</tr>
<tr>
<td>Mean change between warm and cool</td>
<td>0</td>
<td>-4.9 ± 1.8</td>
<td>-2.0 ± 0.6</td>
<td>+2.6 ± 0.7</td>
<td>+0.6 ± 0.3</td>
<td>-0.05 ± 0.02</td>
<td>+5.8 ± 1.0</td>
<td>+2.8 ± 0.3</td>
</tr>
</tbody>
</table>

* The mean values for lactate and pyruvate represent five of these eight infants.
to 10 mEq per L. There was a consistent fall in total CO$_2$ and PCO$_2$, although the magnitude of the change varied from infant to infant. The increase in oxygen consumption in the cool environment was generally less. Blood lactate levels rose only slightly; however, there was a considerable increase in the lactate to pyruvate ratio in all infants.

**Discussion**

This study has confirmed earlier observations (12) that there is a fall of body temperature after birth if the infant is kept at room temperature (20 to 23°C). This fall is initially extremely rapid. The two main contributing factors are evaporation of water from the body surface and lungs and a high thermal conductance. The skin temperature falls even more rapidly than the deep body (rectal) temperature. As shown by Brück, Brück, and Lemtis (13), the fall in skin temperature observed upon exposure to cold is partly due to a decrease in skin blood flow.

The fall in body temperature, however, does not imply that the newborn infant is polikilothermic. It has been shown in this and other studies (14) that the human newborn infant, in common with most mammals, is a true homeotherm, since there is an increase in metabolic rate upon exposure to a cool environment. This fall in temperature indicates therefore that in the immediate neonatal period, heat production is insufficient to make up for the heat loss.

For technical reasons it has not been possible to measure metabolic rate during the first minutes of life when the infant's temperature is falling rapidly. It may well be that thermogenesis is impaired at this time. Hypoxia and hypercapnea are known to reduce the metabolic response to cooling (15-17), and chemoreceptor stimulation may abolish shivering (18, 19). Since the infant is both hypoxic and hypercapnic as a result of the delivery process, it is reasonable to suppose that each of these factors could be contributing to a decreased heat production.

By the age of 2 hours thermal stability of the newborn appears to be increased as evidenced by a slower rate of fall in body temperature when the infant is subjected to a comparable cold stress. This could be due either to decreased heat loss or to improved thermogenesis. Although it is difficult to assess the relative contributions made by these two mechanisms in the neonate, from thermodynamic considerations the former should play the more important role.

Despite a difference in deep body temperature between the two groups of infants in Section I, both the rate of rise of pH and the absolute values achieved during the first 2 hours of life were similar. In addition, the healthy infants exposed to a cool environment at 2 hours of age showed very little, if any, change in pH. Analysis of base deficit data, however, revealed the presence of metabolic acidosis in all infants subjected to cold stress. In addition there was concomitant
reduction in Pco2, probably on a compensatory basis. This respiratory compensation for metabolic acidosis was complete in all high score infants irrespective of whether they were exposed to a cool environment immediately after delivery or at the age of 2 hours. The ratio of lactate to pyruvate in these infants remained relatively constant when they were transferred from a warm to a cool environment, suggesting that the rise in oxygen consumption during cold stress was not associated with a persistent oxygen debt.

The biochemical changes induced by cold stress in the four depressed infants differed from those seen in the healthy ones in several respects, namely, fall in pH, greater increase in base deficit, smaller increase in oxygen consumption, and increased lactate to pyruvate ratio. These observations suggest an inadequate circulatory and respiratory response to cold stress. There was a partial respiratory compensation. In two infants in whom base deficit exceeded 10 mEq per L, Pco2 was lowered to 25 mm Hg. Had alveolar ventilation been impaired by prematurity, aspiration of meconium, or depression of the respiratory center, or had there been disturbances in ventilation perfusion ratio as a result of intrapulmonary or intracardiac shunts, it is unlikely that such a compensatory reduction in Pco2 could have occurred. In this event there would have been an even greater fall in pH during exposure to a cool environment. Even if elimination of CO2 in the depressed newborn were adequate, it is doubtful that cardiac output could be sufficiently increased to satisfy the greater demand for oxygen under conditions of lowered environmental temperature. Although it is obvious that no statistical conclusions can be drawn from the small number of depressed infants, there was a consistent fall in pH in the cool environment. Also, the increase in base deficit in every instance was greater than the mean change in the healthy infants by at least twice the standard error of the mean.

An identical hydrogen ion concentration at two different temperatures does not indicate the same acid-base environment. Because of changes in the dissociation constant of water, the neutrality point shifts approximately 0.05 pH upward when temperature is lowered from 38° C to 35° C. It is not yet known whether homeostatic mechanisms regulate towards a constant pH or a constant alkaline departure from neutrality. The body maintains a constant pH at normal body temperatures which is 0.6 pH on the alkaline side of the neutrality point for water (6.8 at 38° C). If the body temperature is lowered and the pH remains constant, then the degree of shift towards the alkaline side becomes less. In this sense, even the vigorous infants in the present study were less alkaline at lower body temperatures. If blood, in vivo, behaves similarly to that in vitro, then the pH would be expected to rise at lower temperatures.

The blood of hibernating bats, in vivo, does behave in this way, the pH at 8° C being 7.67 as compared with 7.40 at 38° C (20). It might be inferred from this that the normal pH at lower body temperatures is higher than 7.4; however, since hypothermia is an unnatural state for humans, it is at present not possible to define the ideal acid-base environment at lower body temperatures.

From a quantitative point of view these considerations are probably insignificant over the comparatively small temperature range in this study. They might, however, be of importance under conditions of deep hypothermia.

**Summary**

In mature newborn infants, recovery from birth asphyxia was influenced by the cold stress of normal room temperature. Under these conditions vigorous infants were able to achieve and maintain a relatively normal pH. This was accomplished by increasing CO2 elimination to compensate for a developing metabolic acidosis. Infants depressed for even a brief period at birth were unable to maintain their pH and developed a more pronounced metabolic acidosis in the cold environment.

**Acknowledgments**

We wish to express our appreciation to Mrs. L. Grann and Dr. S. S. Daniel for their invaluable assistance.

**References**