Phenobarbital-Induced Alterations in Vitamin D Metabolism

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ABSTRACT The metabolic fate of intravenously injected vitamin D$_3$-1,2-$^3$H (D$_3$-$^3$H) was studied in two normal individuals on chronic phenobarbital therapy. Silicic acid column chromatography of lipid-soluble plasma extracts obtained serially for 96 hr after D$_3$-$^3$H injection demonstrated a decreased plasma D$_3$-$^3$H half-life and increased conversion to more polar metabolites. The polar metabolites formed included several with chromatographic mobility identical to 25-hydroxycholecalciferol. Disappearance of this latter material was also accelerated. A child with rickets and a normal volunteer studied before and after a 2 wk course of phenobarbital therapy demonstrated similar alterations in D$_3$-$^3$H metabolism. When liver microsomes from 3-wk-old Sprague-Dawley rats treated with phenobarbital were incubated with D$_3$-$^3$H, polar metabolites were produced with chromatographic mobility similar to the plasma D$_3$-$^3$H metabolites from phenobarbital-treated humans. Similar incubations employing 25-hydroxycholecalciferol-26,27-$^3$H as the substrate also demonstrated an increased conversion to polar metabolites. The data suggest that the reported increased incidence of osteomalacia observed in patients on chronic anticonvulsant therapy may be the result of an accelerated conversion of vitamin D and its active metabolite, 25-hydroxycholecalciferol, to polar metabolites by drug-induced liver microsomal enzymes.

INTRODUCTION

An increased incidence of rickets in children on long-term anticonvulsant therapy was first reported in 1968 by Kruse (1). More recently, Richens and Rowe (2) have observed a striking incidence of hypocalcemia and elevations of serum alkaline phosphatase in an institutionalized adult epileptic population, the degree of hypocalcemia correlating positively with the total dosage of anticonvulsant drugs received. Simultaneously, Dent, Richens, Rowe, and Stamp (3) demonstrated that the osteomalic bone changes in adults on long-term anticonvulsant therapy respond rapidly to vitamin D supplementation.

In a preliminary report (4) we have observed that human subjects on chronic phenobarbital therapy manifest an increased rate of disappearance of vitamin D$_3$-$^3$H (D$_3$-$^3$H)$^1$ from the plasma, and that liver microsomes from phenobarbital-treated animals are capable of rapidly converting D$_3$-$^3$H to more polar metabolites in vitro. We now report that (a) patients on chronic phenobarbital therapy rapidly convert injected D$_3$-$^3$H to more polar metabolites, some of which are metabolically inactive; (b) this increased conversion can be rapidly induced in individuals receiving a conventional dose of phenobarbital; and (c) administration of phenobarbital to rats induces increased in vitro hepatic conversion of D$_3$-$^3$H and 25-hydroxycholecalciferol-$^3$H (25-OH-D$_3$-$^3$H) to more polar metabolites, the increased conversion being associated with increased microsomal hydroxylase activity.

METHODS

Two normal adult females age 45 and 43 yr (E. D. and G. D.) who had been taking phenobarbital (90-120 mg/day) orally in divided doses for more than 1 yr, an 11 month old male infant with previously untreated rickets (C. C.) and a 30 yr old male normal volunteer (J. H.) were used for these studies. All subjects had previously been on adequate diets containing approximately 800 IU of vitamin D and were studied during hospitalization after 7-10 days of adaptation to diets containing 800-1000 IU of vitamin D per day. C. C. and J. H. were restudied after a 2 wk course of phenobarbital, 2 mg/kg per day and 1.5 mg/kg per day, respectively, in divided oral doses. 7-9 μCi

$^1$Abbreviations used in this paper: D$_3$-$^3$H, vitamin D$_3$-1,2-$^3$H; DPH, diphenylhydantoin; 25-OH-D$_3$, 25-hydroxycholecalciferol; 25-OH-D$_3$-$^3$H, 25-hydroxycholecalciferol-$^3$H.

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of a D$_3$-$^3$H preparation with a specific activity of 152 mCi/mmole (New England Nuclear Corp., Boston, Mass.) were administered i.v. in 0.7-0.8 ml of absolute ethanol over 10-15 sec to each subject in the fasting state. 15-20 ml samples of heparinized blood were collected at 5, 15, and 30 min and at 1, 2, 4, 8, 12, and 24 hr after the D$_3$-$^3$H injection. For the subsequent 4 days, plasma was obtained at 12 hr intervals.

Plasma samples were subjected to combustion for determination of total $^3$H and extracted with methanol-chloroform for determination of total lipid soluble radioactivity (5). Chloroform extracts of 24-, 48-, 72-, and 96-hr plasma samples were also dried in a flash evaporator under nitrogen, the residue dissolved in Skellysolve B (a petroleum ether-diethyl ether-methanol elution procedure as described by Ponchon and DeLuca (6). 10-ml fractions were collected with a flow rate of 1 ml/min and radioactivity monitored as previously described (5).

The interval between initial and repeat studies in C. C. was 5 consecutive days. The peak I-V region in J. H. was rechromatographed on a fiberglass-backed silica gel thin-layer medium (Gelman type SG; Gelman Instrument Company, Ann Arbor, Mich.) and separated into 25-OH-D$_3$-$^3$H and more polar metabolites with a solvent system of hexane/acetonitrile (V:V, 2:1) and the mixture homogenized in a Potter-Elvehjem homogenizer fitted with a Teflon pestle. The aqueous- and lipid-soluble phases were separated and portions of each phase subjected to lipid scintillation counting as previously described (5). The remainder of the lipid phase was evaporated to dryness under a stream of nitrogen and redissolved in Skellysolve B. The products of the D$_3$-$^3$H incubation were applied to silicic acid columns as described above. The products of the 25-OH-D$_3$-$^3$H incubation were partitioned onto fiber glass-backed silica gel thin-layer medium.

Definitive chromatography was confirmed in each subject by calculation of D$_3$-$^3$H half-life determined by thin-layer chromatography of plasma samples taken at the more frequent intervals noted above (5). The peak I-V fraction eluted by silicic acid chromatography of 72-hr plasma samples from the two patients on chronic phenobarbital therapy (E. D. and G. D.) was rechromatographed on a Celite partition column (Johns-Manville, N. Y.) developed with the mobile phase (20% chloroform-80% Skellysolve B equilibrated with 90% methanol-10% water) with 5-ml fractions collected according to the technique of Suda et al. (7). When unlabeled 25-hydroxycholecalciferol (25-OH-D$_3$) (generously supplied by the Upjohn Co., Kalamazoo, Mich.) was added to the original silicic acid column for spectrophotometric identification of material migrating as 25-OH-D$_3$ fractions were collected as usual, evaporated under nitrogen, made up to 5 ml with absolute alcohol and the optical density read at 264 m$_u$ in a Beckman DU spectrophotometer (Beckman Instruments, Inc., Fullerton, Calif.).

In the animal experiments, phenobarbital (100 mg/kg per day) dissolved in 0.50 ml of normal saline was administered i.p. to 50 g immature male Sprague-Dawley rats for 5 consecutive days. The animals were anesthetized under ether and the livers perfused in vivo with ice-cold normal saline, removed, blotted dry, weighed, and homogenized in two vol of 0.25 M sucrose at 4°C. Separation of liver homogenates into nuclear, mitochondrial, microsomal, and cytoplasmic fractions was accomplished by differential centrifugation according to the technique of Schneider and Hogeboom (8). The protein content of the various cell fractions obtained was determined by the method of Lowry, Rosebrough, Farr, and Randall (9). For incubation, samples of various cell fractions from treated and control animal livers were taken so as to contain equivalent amounts of protein nitrogen (equivalent to that obtained from 400 mg of control liver).

Cell fraction samples were incubated with 3 IU of D$_3$-$^3$H (519 mCi/mmole) or 3 IU of 25-OH-D$_3$-$^3$H (315 mCi/mmole) added in 0.20 ml of methanol to achieve a final volume of 5.6 ml containing 3.8 ml of 0.1 M potassium phosphate buffer (pH 7.4) with 1.60 ml of 0.1 M Tris buffer (pH 7.4), 0.5 m mole MgCl$_2$, NADPH (3 mg), glucose-6-phosphate (14 mg) and glucose-phosphate-dehydrogenase (5 KU). Incubation was carried out in a Duboff metabolic shaker under a 95% O$_2$/5% CO$_2$ gas phase according to the technique of Conney and Klutch (10). The reaction was terminated at 60 min by the addition of 5 vol of a solution of methanol-chloroform (V:V, 2:1) and the mixture homogenized in a Potter-Elvehjem homogenizer fitted with a Teflon pestle. The aqueous- and lipid-soluble phases were separated and portions of each phase subjected to lipid scintillation counting as previously described (5).

RESULTS

Disappearance and metabolic fate of vitamin D$_3$ in plasma. The plasma half-life of D$_3$-$^3$H as determined by silicic acid column chromatography in two otherwise normal adults (E. D. and G. D.) on chronic phenobarbital therapy (1.3-1.5 mg/kg per day) was significantly decreased when compared to a group of 12 normal adults (7 males and 5 females, aged 26-48 yr) not on phenobarbital studied under similar conditions (Table I). Moreover, administration of phenobarbital orally for 2 wk to C. C., a child with previously untreated rickets (1.5 mg/kg per day), and J. H., a normal adult volunteer (2 mg/kg per day), resulted in

<table>
<thead>
<tr>
<th>Table I</th>
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<tr>
<td>Effect of Phenobarbital Treatment on Plasma Half-Life of D$_3$-$^3$H</td>
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<tr>
<td>Control</td>
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<tr>
<td>hr</td>
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<tr>
<td>C. C. (rickets)</td>
</tr>
<tr>
<td>J. H. (normal adult)</td>
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<tr>
<td>G. D. (normal adult)</td>
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<tr>
<td>E. D. (normal adult)</td>
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<tr>
<td>Normal adult controls</td>
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</table>

Half-lives were calculated by analysis of semilogarithmic plots of plasma D$_3$-$^3$H (expressed as per cent administered dose) disappearance with time determined by silicic acid column chromatography for the interval beginning at 12 hr and terminating 96 hr after D$_3$-$^3$H administration. Control values are given as mean ± SEM.
a similar decrease in Da-H plasma half-life. Silicic acid column profiles of the plasma lipid soluble radioactivity 72 hr after injection of Da-H are illustrated in Fig. 1.

The material in peak III has been isolated previously from human plasma after Da-H administration and identified as unaltered vitamin D3 with potent in vivo antirachitic activity (5). In the multigradient chromatographic system employed in this study, peak IV ordinarily represents 25-OH-D3, the biologically active metabolite of vitamin D (12). Further confirmation of the chromatographic identity of peak IV as 25-OH-D3 in these studies was obtained by adding 0.50 mg of pure 25-OH-D3 to 72-hr plasma extracts from E. D. and G. D. and applying to silicic acid columns. Fractions were collected as usual, and samples taken for measurements of optical density at 264 μM as well as radioactivity by liquid scintillation counting. The optical density of 25-OH-D3 at 264 μM in each case coincided exactly with the peak IV radioactivity. Additional confirmation was obtained by isolating the peak IV metabolite from silicic acid columns and rechromatographing on Celite partition columns (7). In each instance, a single peak of radioactivity was obtained which was exactly coincident with 25-OH-D3 as determined by optical density measurements.

Normally, 25-OH-D3 is derived from vitamin D3 by hydroxylation in the liver, possibly by a specific mitochondrial hydroxylase (13, 14). Peak V is ordinarily comprised of at least three biologically active vitamin D metabolites, 21, 25-, 25, 26-, and 1,25-dihydroxy cholecalciferol as well as several unidentified components (7, 15-17). The identity of peaks VI and VII is presently unknown but they have been demonstrated to be biologically inactive (12).

The chromatographic profile of Da-H metabolites in the 72-hr control plasma samples from J. H. (normal subject) is typical of the normal column profile (Fig. 1, top). During the second Da-H study performed after 2 wk of phenobarbital therapy, the percentage of radioactivity in J. H.'s 72 hr plasma migrating as Da-H was decreased and the percentage migrating as 25-OH-D3-H (peak IV) and more polar metabolites (peak V, VI, VII) was increased. In the two normal subjects on long-term phenobarbital therapy, the changes were even more striking (Table II).

The time course of disappearance of Da-H and appearance of 25-OH-D3-H and more polar metabolites in control and phenobarbital-treated subjects is illustrated in Figs. 2 and 3. The disappearance of Da-H and the appearance of 25-OH-D3-H and peaks V-VII were markedly accelerated in the phenobarbital-treated subjects. The generated 25-OH-D3-H peak increased with time, reaching maximal value by 24 hr (1.8 times the Da-H half-life) in the phenobarbital-treated subjects and then began to fall off rapidly. In contrast, the level of the 25-OH-D3 peak was still rising after 96 hr (3.5 times the Da-H half-life) in control subjects. The relatively earlier onset of decay of the 25-OH-D3-H peak suggests increased metabolism as well as more rapid appearance of this metabolite in phenobarbital-treated subjects.

The portion of plasma radioactivity occurring as aqueous-soluble metabolites was determined by subtracting plasma lipid-soluble radioactivity from total

**TABLE II**

<table>
<thead>
<tr>
<th></th>
<th>III*</th>
<th>IV*</th>
<th>V*</th>
<th>VI*</th>
<th>VII*</th>
</tr>
</thead>
<tbody>
<tr>
<td>J. H. (control)</td>
<td>1.46</td>
<td>1.31</td>
<td>0.04</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>J. H. (after phenobarbital)</td>
<td>1.27</td>
<td>2.28</td>
<td>0.15</td>
<td>0.05</td>
<td>0.02</td>
</tr>
<tr>
<td>E. D. (chronic phenobarbital)</td>
<td>0.13</td>
<td>8.55</td>
<td>0.56</td>
<td>0</td>
<td>0.09</td>
</tr>
<tr>
<td>G. D. (chronic phenobarbital)</td>
<td>0.14</td>
<td>7.27</td>
<td>0.50</td>
<td>0.13</td>
<td>0.35</td>
</tr>
</tbody>
</table>

* Chromatographic fractions as designated by Ponchon and DeLuca (11) and designated in Figs. 1 and 4.

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and chronic phenobarbital therapy.

Figure 2 Time course of plasma vitamin D$_3$ disappearance and peak IV appearance in three normal controls ($\downarrow$, mean ±SEM), J. H. control period (○), J. H. after phenobarbital (●), and two subjects, E. D. (▲) and G. D. (■), on chronic phenobarbital therapy.

Figure 3 Time course of appearance of peaks V, VI, and VII in three normal controls ($\downarrow$, mean ±SEM), J. H. control period (○), J. H. after phenobarbital (●), and two subjects, E. D. (▲) and G. D. (■), on chronic phenobarbital therapy.

plasma radioactivity determined by combustion. The per cent plasma aqueous-soluble metabolites for J. H. and C. C. after phenobarbital and E. D. and G. D. did not vary significantly from the normal control values of 25.3 ±3.7, 19.6 ±5, 22.3 ±3.8, and 24.0 ±2.3 at 24, 48, 72, and 96 hr, respectively.

In vitro metabolism of D$_3$-3H and 25-OH-D$_3$-3H hepatic cell fractions. Subcellular components obtained by differential centrifugation from homogenates of livers from 3-wk old male Sprague-Dawley rats treated with

![Figure 4](image-url)

**Figure 4** Column chromatographic profiles of chloroform extracts of reaction mixtures from incubations of D$_3$-3H with microsomes from control and phenobarbital-treated rats. 3 IU of D$_3$-3H were incubated aerobically for 60 min at 37° with 11 mg of microsomal protein from control and phenobarbital-treated animals in the presence of a NADPH-generating system (9). The dashed lines indicate the chromatographic position of added (26-27)-D$_3$-25-hydroxycholecalciferol. Chromatographic techniques were identical to those described in Fig. 1.

<table>
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<tr>
<th>TABLE III</th>
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<tr>
<td><strong>Effect of Phenobarbital Treatment on Hepatic Microsomal Metabolism of D$_3$-3H</strong></td>
</tr>
<tr>
<td>Unaltered D$_3$-3H</td>
</tr>
<tr>
<td>% of lipid-soluble radioactivity</td>
</tr>
<tr>
<td>Media control</td>
</tr>
<tr>
<td>Control</td>
</tr>
<tr>
<td>Phenobarbital</td>
</tr>
</tbody>
</table>

Values represent the mean ±SEM of at least three determinations. Incubation conditions were identical to those described in Fig. 4. Media control values were obtained by incubating D$_3$-3H in the absence of microsomes. Water-soluble metabolites were determined by previously described techniques (5). Hexobarbital (1000 nmoles) was added in parallel incubations to flasks containing 11 mg of microsomal protein from control and phenobarbital-treated animals.
Phenobarbital were incubated with D₃-H and the results compared with incubations with equivalent amounts of protein nitrogen from corresponding subcellular fractions of control animals. Whereas microsomes from control animal livers had a minimal effect on D₃-H at the concentrations employed, hepatic microsomes from phenobarbital-treated animals catalyzed an accelerated conversion of D₃-H to lipid-soluble products which were more polar than 25-OH-D₃-H (Fig. 4). Additionally, there was a 250% increase in total water-soluble vitamin D₃ metabolites. These changes were associated with increased microsomal oxidation of hexobarbital (Table III). Similarly, whereas 25-OH-D₃-H was only slightly metabolized by the liver microsomes obtained from control animals, liver microsomes obtained from phenobarbital-treated animals increased conversion of 25-OH-D₃-H to more polar metabolites (Table IV). Incubation of either D₃-H or 25-OH-D₃-H with nuclear, mitochondrial, or cytoplasmic hepatic cell fractions from phenobarbital-treated animals demonstrated no difference from incubations with similar fractions from control animals.

The metabolic products obtained from D₃-³C incubated in vitro with microsomes from phenobarbital-treated rats were cochromatographed on silicic acid columns with the D₃-H metabolites from the plasma of the patients on chronic phenobarbital therapy (Fig. 5). D₃-³C metabolites derived from microsome incubation had a mobility almost identical to the plasma metabolites obtained from the human subjects on phenobarbital.

**DISCUSSION**

The demonstration of accelerated metabolism of vitamin D₃ in subjects on phenobarbital therapy would provide a rational basis for the reported increased incidence of osteomalacia in individuals on chronic anticonvulsant therapy (1, 2). The results of the present experiments suggest that this accelerated metabolism is the result of drug-stimulated increases in hepatic microsomal activity.

**TABLE IV**

<table>
<thead>
<tr>
<th></th>
<th>Unaltered 25-OH-D₃-H</th>
<th>Water-soluble metabolites</th>
<th>Hexobarbital oxidized</th>
<th>% of lipid-soluble radioactivity</th>
<th>% of total radioactivity</th>
<th>nmol/100 mg liver</th>
</tr>
</thead>
<tbody>
<tr>
<td>Medial control</td>
<td>95.6 ±0.6</td>
<td>0.3 ±0.2</td>
<td>—</td>
<td>95.6 ±0.6</td>
<td>0.3 ±0.2</td>
<td>52 ±3</td>
</tr>
<tr>
<td>Control</td>
<td>83.8 ±1.4</td>
<td>2.7 ±0.4</td>
<td>52 ±3</td>
<td>83.8 ±1.4</td>
<td>2.7 ±0.4</td>
<td>52 ±3</td>
</tr>
<tr>
<td>Phenobarbital</td>
<td>61.4 ±1.9</td>
<td>8.9 ±1.1</td>
<td>528 ±21</td>
<td>61.4 ±1.9</td>
<td>8.9 ±1.1</td>
<td>528 ±21</td>
</tr>
</tbody>
</table>

Values represent the mean ±SEM of at least five determinations. Incubation conditions were identical to those described in Table III with the substitution of 25-OH-D₃-H as the substance.

**FIGURE 5** Cochromatography of plasma D₃-³H metabolites from a patient (E. D.) on chronic phenobarbital therapy (closed figures) with metabolic products obtained from in vitro incubation of D₃-³C with microsomes from phenobarbital-treated rats (open figures). The optical density of pure 25-OH-D₃-³C (●) added to the patient plasma extract is indicated on the scale at right. The patient plasma was obtained 72 hr after the i.v. injection of D₃-³H. Microsomal incubation conditions were identical to those described for Fig. 3. Chromatographic techniques were identical to those described for Fig. 1.

It has been well established that the induction of increased liver microsomal enzyme activity by phenobarbital results in decreased in vivo biologic activity of steroid compounds. Levin, Welch, and Conney demonstrated that pretreating immature female rats with phenobarbital in doses as low as 2 mg/kg per day diminished the uterotrophic response to administered estradiol or estrone, decreased the uterine concentration of injected tritiated estrogens and increased production of polar estrogen metabolites (18, 19). Also, phenobarbital treatment has been demonstrated to decrease the seminal vesicle-stimulating effect of androgens (20) and to diminish the effect of progesterone in association with increased hepatic hydroxylation and decreased total body accumulation of the steroid and its metabolites (21).

Similarly, induction of liver microsomal enzyme activity with diphenylhydantoin (DPH) has been demonstrated to have important clinical effects on steroid metabolism in humans. Werk, Sholitan, and Olinger (22) demonstrated that DPH therapy sharply decreased plasma cortisol levels, increased urinary excretion of 6-OH cortisol and resulted in clinical improvement in patients with Cushing’s syndrome. More recently, Choi, Thrasher, Werk, Sholitan, and Olinger (23) have reported that DPH administration in patients with various disorders resulted in a decreased plasma cortisol half-life.
life, with a compensatory increase in cortisol secretory rate. Additionally, Jubiz and coworkers (24) have reported decreased effectiveness of dexamethasone in suppressing morning plasma cortisol levels in association with increased urinary and biliary excretion of conjugated dexamethasone or its metabolites in individuals on DPH therapy.

Similarities between the drug and steroid hydroxylases in hepatic microsomes have led to the suggestion that drugs and steroids are substrates for the same hepatic P-450 enzyme system (25). Hence, agents which increase the activity of hepatic drug-metabolizing enzymes could be expected to increase hydroxylation of steroid compounds as well. The drug-induced microsomal enzymes hydroxylate steroids predominantly at the 6α, 7α, and 16α positions (26). Thus, the resultant steroid metabolites tend to be more polar than their parent compound. Since vitamin D₃ and 25-OH-D₃ are structurally similar to the steroid hormones and are readily accumulated by the liver (27, 28), these compounds might well be expected to be metabolized to more polar products by the hepatic P-450 system. Although the polar lipid-soluble D₃-'H metabolites occurring in increased proportion in phenobarbital-treated humans are as yet unidentified, their chromatographic similarity to metabolites produced by phenobarbital-induced rat liver microsomal enzymes makes it likely that some of the human plasma metabolites are the result of increased liver microsomal activity. There are, however, quantitative differences between the metabolites produced in the in vitro D₃-'H studies and those noted in the plasma of patients treated with phenobarbital. Whereas the major portion of the radioactivity in human 72-hr plasma samples occurred as 25-OH-D₃, peaks V, VI, and VII were the more prominent D₃-'H metabolites in the microsomal incubations.

The explanation for the observed differences in vivo and in vitro metabolites is not entirely clear. Since it has been reported that the hepatic vitamin D₃ 25-hydroxylase is of mitochondrial origin, it is not surprising that the microsomal incubations in these studies did not produce significant amounts of 25-OH-D₃. On the other hand, mitochondrial fractions from phenobarbital-treated animals were likewise not found to produce increased amounts of 25-OH-D₃. It is possible that differences between patterns of plasma and intra-cellular metabolites or differences between in vivo and in vitro pathways might account for the differences between in vivo and in vitro metabolite patterns. A more likely explanation relates to the fact that, due to accelerated microsomal metabolism, it would be anticipated that body stores of vitamin D₃ and 25-OH-D₃ would gradually become depleted in individuals on chronic phenobarbital therapy. Since it has been demonstrated both in rats (29) and humans (30) that the fraction of an injected tracer dose of D₃-'H converted to 25-OH-D₃-'H varies inversely with body vitamin D pool size, an increased proportion of injected D₃-'H appearing as 25-OH-D₃-'H in phenobarbital-treated subjects would be expected if these individuals have decreased levels of vitamin D₃ and 25-OH-D₃. Our recent observations that plasma levels of 25-OH-D₃ measured by competitive protein-binding assay (31) are depressed in individuals on chronic phenobarbital therapy* support this hypothesis.

It is also theoretically possible that displacement of D₃-'H or 25-OH-D₃-'H from plasma-binding protein by phenobarbital might account for an increased plasma disappearance rate. This is quite unlikely, however, since phenobarbital in concentrations of 500 µg/ml, approximately 50 times the usual therapeutic plasma levels, does not displace 25-OH-D₃-'H from partially purified 25-OH-D₃ plasma-binding protein in vitro.⁹

Decreased vitamin D effect in individuals receiving agents which induce hepatic microsomal enzymes could result either from a decreased biologic activity of the polar vitamin D metabolites or increased excretion of these more polar compounds. Decreased biologic activity of the vitamin D metabolites might play some role since the appearance of inactive peak VI and VII D₃-'H metabolites was increased both in human subjects on phenobarbital and in vitro incubations of D₃-'H with microsomes from phenobarbital-treated rats. Additionally, it is unlikely that the in vitro peak V metabolite represents any of the known biologically active vitamin D₃ metabolites with peak V mobility, since evidence to date indicates that 1,25-dihydroxycholecalciferol is derived from 25-OH-D₃ exclusively in the kidney (32), while there is no evidence to suggest that either 21,25-, or 25,26-dihydroxycholecalciferol is produced by the liver.⁹ On the other hand, induction of hepatic drug-metabolizing enzymes by DPH is associated with increased urinary and biliary excretion of polar steroid metabolites (24). In normal humans, a fraction of i.v. injected D₃-'H appears rapidly in the bile and urine as water-soluble metabolites, a major portion of which are glucuronides and acid conjugates (5). Hence, increased production of polar vitamin D metabolites due to increased hydroxylation and/or glucuronidation might well result in increased biliary and urinary loss of vitamin D.

It is of interest that clinical osteomalacia can be demonstrated in relatively large proportions of epileptics on chronic anticonvulsant therapy (1, 2), while clinical evidence of deficiencies of other steroids such

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⁹H. F. DeLuca. Personal communication.
as cortisol or estrogen are apparently not commonly observed in this group. This would perhaps be expected, since endogenous levels of cortisol, estrogen, and androgens are known to be regulated through pituitary feedback mechanisms, whereas, to date, there is only fragmentary evidence that production of the active 25-OH-D₃ metabolite may be subject to hepatic feedback regulation (14). The demonstration that anticonvulsant-induced osteomalacia responds rapidly to vitamin D therapy (1, 2) suggests, however, that increasing the endogenous level of vitamin D by exogenous supplementation is effective in correcting the drug-induced deficiency. A corollary of the observed decreased effectiveness of normal dietary vitamin D in individuals on chronic anticonvulsant therapy is the anticipation that increased hepatic drug-metabolizing activity would impart resistance to exogenously administered vitamin D. In this regard, there is preliminary evidence that phenobarbital pretreatment may protect rats from vitamin D–induced hypercalcemia (3). This raises the possibility that treatment with a nonhypnotic barbiturate such as phenobarbital (33) might be effective, relatively nontoxic therapy for vitamin D intoxication. Additionally, the resistance to vitamin D occasionally seen in individuals with postsurgical hypoparathyroidism on phenobarbital therapy may be attributable to induction of hepatic drug-metabolizing enzymes.

The immediate implication of these studies is that care should be exercised in interpreting D₃-¹⁴C kinetic data in persons receiving medications known to induce hepatic drug-metabolizing enzymes. Additionally, since the hepatic drug-metabolizing systems are stimulated by a wide variety of commonly used agents, including barbiturates, methylphenylpyridylhydantoin, DPH, meprobamate, chloridiazepoxide, and chlorpromazine (34), among others, the potential for altered vitamin D metabolism in many clinical situations is apparent. Therefore, patients on high doses of drugs known to stimulate hepatic drug-metabolizing enzymes, especially those individuals with marginal dietary intake or diseases predisposing to vitamin D malabsorption, should be carefully evaluated for evidence of vitamin D deficiency.

ACKNOWLEDGMENTS

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