The Effect of Hyperventilation on Distal Nephron Hydrogen Ion Secretion


From the Department of Medicine, University of Toronto, Toronto M5S 1A8, Canada and Baylor College of Medicine, Houston, Texas 77025

Abstract This study was designed to determine the effect of acute hyperventilation on distal nephron hydrogen ion secretion. The blood PCO₂ declined and stabilized rapidly when bicarbonate loaded rats were hyperventilated. In contrast, the urine PCO₂ declined slowly, resulting in an early increase in the urine minus blood (U-B) PCO₂ which could not be obliterated by carbonic anhydrase infusion. Within approximately 50 min, the U-B PCO₂ in the hyperventilated and carbonic anhydrase infused rats approached zero. Consequently, equilibrium between collecting duct urine and arterial blood PCO₂ was then presumed to exist. This provided the basis for the subsequent studies on a series of rats. The U-B PCO₂ decreased from a control of 22±1 mm Hg (mean±SEM) to 11±2 mm Hg (mean±SEM) with hypocapnia, and rose again to its control value when the blood PCO₂ returned to prehyperventilation values. This decline in U-B PCO₂ with acute hyperventilation could not be attributed to changes in urine flow, phosphate, or bicarbonate excretion, suggesting, therefore, a decrease in distal nephron (probably collecting duct) hydrogen ion secretion with acute hyperventilation. Possible pitfalls in the interpretation of the U-B PCO₂ are illustrated.

Introduction

The effect of hyperventilation on the rate of hydrogen ion secretion in the terminal segments of the nephron has not been elucidated. Stanbury and Thomson (1) suggested that the electrolyte excretion patterns observed during hypocapnia might be due to inhibition of hydrogen ion secretion in the kidney. Micropuncture studies have shown that hyperventilation decreases hydrogen ion secretion in the proximal and distal convoluted tubules (2), but information about the response of the collecting duct to this maneuver is not available.

The collecting duct is probably the most important segment of the nephron for achieving the minimum urine pH and thereby for augmenting acid excretion. In addition, the collecting duct has a major regulatory role in homeostasis under a variety of conditions (3).

In the present study, we have attempted to characterize the effect of hyperventilation on the distal nephron. To accomplish this, we have estimated distal nephron hydrogen ion secretion by utilizing the urine minus blood (U-B)¹ PCO₂ gradient (4). The U-B PCO₂ decreased during hyperventilation. This result cannot be explained by changes in urinary volume, bicarbonate, or buffer excretion. We propose that hyperventilation results in a reduction in distal nephron hydrogen ion secretion, presumably in the collecting duct.

Methods

Experimental protocol. Male Wistar rats (250–300 g), obtained from Woodlyn Farms, Guelph Ontario, were allowed free access to food (Purina Lab Chow) and water before the experiment. Sodium bicarbonate, (20 g/l) and glucose (50 g/l) were added to the drinking water 2 days before the experiment to facilitate alkalization of the urine. Rats were anesthetized with inactin, 100 mg/kg intraperitoneally. Intravenous infusions were administered into the jugular vein. Blood pressure was monitored by a femoral artery catheter connected to a mercury manometer. Blood samples were collected from the femoral artery into heparinized capillary tubes. The urine for pH and PCO₂ was collected anaerobically into capillary tubes via a suprapubic catheter in the bladder. pH and PCO₂ determinations were performed immediately after the collection as previously described (5). Urine for flow rate and all other parameters was collected into prelabeled vessels.

After induction of anaesthesia, 0.3 M sodium bicarbonate was infused at a rate of 100 µl/min throughout the experiment. When the urine pH and PCO₂ remained constant for three consecutive periods, a steady state was assumed to exist and blood was collected for pH and PCO₂. After the

¹Abbreviations used in this paper: GFR, glomerular filtration rate; U-B, urine minus blood.
control period, hyperventilation was induced with a mechanical respirator (Phipps and Bird small animal respirator, model no 7088-600, Phipps and Bird Inc., Richmond, Va.). In the experiments reported in Figs. 1, 2–4, samples were obtained after the blood Pco₂ had declined to a steady-state value (blood Pco₂ in two consecutive samples stable within 3–5 mm Hg). In the experiments reported in Tables I and II, by contrast, samples were collected after the urine Pco₂ had remained constant for three consecutive periods. The animals were hyperventilated for approximately 90 min before the experimental specimens were taken. The respirator then disconnected and the measurements were repeated after 15–30 min of spontaneous ventilation. If the blood Pco₂ remained low, the animal was placed in a covered box where the inspired gas contained humidified 5% CO₂; 95% O₂. Two rats hyperventilated spontaneously during the first period. In these animals steady-state values were obtained during hyperventilation and then normocapnia was induced by exposing them to the 5% CO₂; 95% O₂ gas mixture. Subsequently, they were removed from the box and allowed to resume spontaneous hyperventilation. Since the results were similar with both methods of hyperventilation, the data from both groups were combined.

Carbonic anhydrase was injected intravenously as a 2-mg bolus followed by a continuous infusion at a rate of 20 μg/min. Glomerular filtration rates (GFR) were measured with tritium labeled inulin using a 5-μCi priming dose followed by a sustaining dose of 6 μCi/h.

Analytical Methods. The blood and urine samples for pH and Pco₂ were measured on a Radiometer model PHM 72 digital acid base analyzer. Urine bicarbonate concentrations were calculated from the pH and Pco₂ using a pK as determined from the formula: pK = 6.33 – 0.5 V/n and a solubility coefficient of 0.0309. Sodium and potassium were measured by flame photometry as previously described (4). Phosphorus was measured by a modified method for the photoelectric colorimeter (7). Organic anions were measured by the titrimetric method of Chan (8). Tritium was measured on a Beckman LS 230 liquid scintillation counter (Beckman Instruments, Inc., Cedar Grove, N. J.) as previously described (5). Statistical analyses were performed by using a t test on paired observations.

Materials and supplies. All chemicals were of analytical grade. Tritium labeled inulin was obtained from New England Nuclear, Boston, Mass.; inactin from Henley and Co., Inc., New York; gas mixtures from Gas Dynamics, Toronto, Ontario; carbonic anhydrase from Sigma Chemical Co., St. Louis, Mo.; and reference buffers from Radiometer Co., Copenhagen, Denmark.

RESULTS

With acute hyperventilation, the blood Pco₂ decreased rapidly and stabilized while the urine Pco₂ declined more slowly. This resulted in an immediate rise in the U-B Pco₂ of 13.7±4.6 mm Hg; P < 0.025 (control U-B Pco₂ was 30±6.9 mm Hg, acute hyperventilation U-B Pco₂ was 43.3±8.1 mm Hg, mean±SEM, n = 7) (Fig. 1). To evaluate the possibility that the initial increase in the U-B Pco₂ was due to the failure of the arterial blood to reflect the medullary Pco₂, or to mixing of urines with a higher and lower Pco₂, carbonic anhydrase was infused into control and acutely hyperventilated rats. These results are shown in Fig. 2 and demonstrate that the U-B Pco₂ gradient was immediately abolished in the control rats (1±0.2 mm Hg) (n = 4) but not in the acutely hyperventilated rats. Therefore, the initial increase in the U-B Pco₂ could not be attributed to a distal nephron mechanism involving delayed dehydration.

of carbonic acid, that is to an increase in distal nephron hydrogen ion secretion. In a separate group of rats, carbonic anhydrase was infused for 50 min after acute induction of hyperventilation. The U-B Pco2 in hyperventilating rats was 32±4 mm Hg (mean±SEM, n = 6), 10 min after carbonic anhydrase infusion but decreased virtually to zero after approximately 50 min (2.5±1.0 mm Hg, n = 6) when equilibrium between arterial blood and renal medulla was assumed to exist. Therefore, the control U-B Pco2 values were compared to values obtained after at least 50 min of hyperventilation in subsequent studies. As shown in Fig. 3, the U-B Pco2 gradient decreased significantly (P < 0.01) as a result of steady-state hyperventilation. Furthermore, the results reported in Table I demonstrate that U-B Pco2 gradient returned to control values when the blood Pco2 returned to prehypocapnia values.

Other factors known to affect the U-B Pco2 are the rate of buffer excretion, urine flow, and bicarbonate excretion. There was no significant fall in the urine phosphate excretion as a result of hyperventilation when the control and experimental periods were compared by the paired t-test (Table II). The analysis of paired data, however, reveal that there were significant decreases in both urinary flow rate and bicarbonate excretion rate during hyperventilation (Table II). GFR’s were measured in three experiments and the results were similar in both states (2.20±0.5 vs. 2.01±0.6 ml/min in normal and hyperventilated rats, respectively). Calculated bicarbonate reabsorption rates in these experiments were 33.2 meq/l GFR in the control period and 27.5 meq/l GFR after 60 min of hyperventilation.

The U-B Pco2 was decreased during hypocapnia when the results were compared at similar rates of urine flow, and bicarbonate, and phosphate excretion (Table III). For these data, a group of bicarbonate loaded rats with bicarbonate excretion rates between 1.5 and 8.0 meq/min and phosphate excretion rates between 0.5 and 1.2 μmol/min was chosen for the normocapnic group for comparison with the hyperventilation data. (Methods for

### Table I

Effect of Hyperventilation on U-B Pco2

<table>
<thead>
<tr>
<th>Rat</th>
<th>Normocapnia B Pco2</th>
<th>Hypocapnia B Pco2</th>
<th>Normocapnia U-B Pco2</th>
<th>Hypocapnia U-B Pco2</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>32</td>
<td>19</td>
<td>25</td>
<td>6</td>
</tr>
<tr>
<td>2</td>
<td>32</td>
<td>20</td>
<td>19</td>
<td>9</td>
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<tr>
<td>5</td>
<td>29*</td>
<td>16</td>
<td>40</td>
<td>25</td>
</tr>
<tr>
<td>6</td>
<td>28*</td>
<td>17</td>
<td>36</td>
<td>24</td>
</tr>
</tbody>
</table>

B, blood.
* These animals were hyperventilating spontaneously and therefore the initial period was treated as a hypocapnia one. Each value represents the mean of the values in three consecutive periods.

### Table II

Effect of Hyperventilation on the U-B Pco2 and on Parameters Influencing U-B Pco2

|-------|----|------|-------|-----|----|------|---------|-------|-----------|         |      |      |
|       |    |      |       |     |    |      |         |       |           | anions  |      |      |
|       |    |      |       |     |    |      |         |       |           |         |      |      |
| Control | 7.58 | 36 | 33 | 7.81 | 58 | 104.8 | 11.2 | 0.96 | 14.1 | 120 | 22 |
|         | ±0.04 | ±1 | ±2.2 | ±0.06 | ±2 | ±19.6 | ±2.2 | ±0.23 | ±2.5 | ±31 | ±1 |
| Hyperventilation | 7.68* | 26 | 30 | 7.84 | 37 | 71.9 | 4.0* | 0.72 | 8.0* | 72 | 111 |
|         | ±0.02 | ±1 | ±1.3 | ±0.07 | ±3 | ±9.0 | ±0.5 | ±0.14 | ±1.5 | ±16 | ±2 |

These values represent the means and standard error of the means for control and hyperventilation data on animals from Table 1.

* P < 0.05 for paired observations.

† P < 0.01.
analyses were performed on the U-B data. For this reason, the results are shown as a comparison of rates (Table 1) obtained in normocapnic animals, confirming the results of others (9, 11, 12). The U-B Pco2 levels which were observed in acutely hyperventilated rats were comparable with control levels (Fig. 2). In contrast, the same dose of carbonic anhydrase caused the urine Pco2 to virtually equal the blood Pco2 in the normocapnic animals, allowing for steady state conditions (Table 1).

Assuming steady-state conditions, the degree of elevation of the U-B Pco2 is influenced by the interplay of several factors in addition to distal hydrogen ion secretion. These include rates of buffer excretion, urine

**DISCUSSION**

The U-B Pco2 gradient is a qualitative index of distal nephron hydrogen ion secretion. The physiologic basis for this assumption has been discussed previously (4). Briefly, carbonic acid is formed in the lumen of the distal nephron from the interaction of secreted hydrogen ions with the luminal bicarbonate. Since carbonic anhydrase activity is virtually absent in this area of the nephron (9), the dehydration of carbonic acid is delayed until it reaches the lower urinary tract, where, because of unfavorable volume relationships, little CO2 is reabsorbed. It has been shown (10) that blood Pco2 is a reasonable estimate of renal vasa recta Pco2 in rats infused with sodium bicarbonate. Therefore, the U-B Pco2 is an index of distal nephron net hydrogen ion secretion.

Several criteria must be met before one can use the U-B Pco2 to assess distal nephron hydrogen ion secretion. The U-B Pco2 must be measured during steady-state conditions. As shown in Fig. 1, the U-B Pco2 gradient was increased after acute hyperventilation. This would indicate an increased distal nephron hydrogen ion secretion only if the blood Pco2 accurately reflected renal medullary Pco2. In acute hypocapnia this may not be the case if there is inadequate time for equilibration. The U-B Pco2 would overestimate distal nephron hydrogen ion secretion in this circumstance. The U-B Pco2 could also overestimate distal hydrogen ion secretion if the collection contained urine with a higher Pco2 formed before hyperventilation.

The elevation in the urine minus vasa recta Pco2 due to distal nephron hydrogen ion secretion is abolished by carbonic anhydrase infusion (11). Carbonic anhydrase infused into acutely hyperventilated rats resulted in a fall in the U-B Pco2 but not to levels which approximated the blood Pco2 (Fig. 2). In contrast, the same dose of carbonic anhydrase caused the urine Pco2 to virtually equal the blood Pco2 in the normocapnic animals, confirming the results of others (9, 11, 12). The U-B Pco2 approached zero after 50 min in acutely hyperventilated rats infused with carbonic anhydrase, indicating that a steady state had been achieved between arterial blood and collecting duct urine by this time. Detailed studies were then carried out to study the effects of hyperventilation on U-B Pco2 after allowing 50 min equilibration time in order to achieve this steady state. There was a significant reduction in the U-B Pco2 gradient during hypocapnia which returned to control values with cessation of hyperventilation (Table 1).

Assuming steady-state conditions, the degree of elevation of the U-B Pco2 is influenced by the interplay of several factors in addition to distal hydrogen ion secretion. These include rates of buffer excretion, urine

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**Table III**

**Effect of Hyperventilation on the U-B Pco2 at Matched Rates of Urine Flow Phosphate and Bicarbonate Excretion**

<table>
<thead>
<tr>
<th></th>
<th>Normocapnia</th>
<th>Hypocapnia</th>
</tr>
</thead>
<tbody>
<tr>
<td>Blood Pco2, mm Hg</td>
<td>40.7±1.7</td>
<td>26±1*</td>
</tr>
<tr>
<td>Bicarbonate excretion, μeq/min</td>
<td>4.7±0.6</td>
<td>4.0±0.5</td>
</tr>
<tr>
<td>Phosphate excretion, μmol/min</td>
<td>0.80±0.08</td>
<td>0.72±0.14</td>
</tr>
<tr>
<td>Urine flow rate, μl/min</td>
<td>52±10</td>
<td>72±16</td>
</tr>
<tr>
<td>(U-B) Pco2, mm Hg</td>
<td>26.2±1.5</td>
<td>11±2*</td>
</tr>
</tbody>
</table>

The animals selected in the normocapnia group (n = 16) were those in which the urine flow rate phosphate excretion rate, and bicarbonate excretion rate were comparable to those observed in the hypocapnia group (n = 6) reported in Table II. The methods for preparation of animals, infusion protocols, and sample analyses were identical for both groups. Results are reported as mean±SEM.

* P < 0.01.
flow, and bicarbonate delivery. The major urine buffer is phosphate. There is a direct correlation between the U-B Pco2 and the urine phosphate concentration (13). Phosphate infusion elevates the urine Pco2, presumably by delaying the dehydration of carbonic acid (14). There was no decline in urine phosphate concentration or excretion rates, however, during acute hyperventilation (Table II). Therefore, alterations in urine phosphate excretion are excluded as an important factor in our studies. Urine flow rate is another factor requiring consideration. Reid and Hills demonstrated that the urine Pco2 varies inversely with urine flow (15). In these studies, both the U-B Pco2 and urine flow rate fell with hyperventilation (Table I). Therefore, these results cannot be attributed to the changes in flow rate. Bicarbonate excretion rate can also influence the urine Pco2 (16). Although the urine pH exceeded 7.55 in our studies, the urine bicarbonate excretion rates decreased with hyperventilation (Table II). We therefore examined the U-B Pco2 at comparable rates of bicarbonate excretion in both normo- and hypocapnia (Table III). The U-B Pco2 was 26.2±1.5 mm Hg during normocapnia when the bicarbonate excretion rate was 4.7±0.6 μeq/min whereas the U-B Pco2 was significantly reduced to 11±2 mm Hg when the blood Pco2 was 28±1 mm Hg and bicarbonate excretion 4.0±0.5 μeq/min. Urine flow rate and phosphate excretion rates were comparable between groups. Furthermore, examination of the data of Thompson and Barrett (17) demonstrates that when the bicarbonate excretion rate is acutely reduced by constriction of the aorta in bicarbonate loaded animals, the U-B Pco2 did not decrease. Similar results were obtained in our laboratory when the bicarbonate excretion was decreased by acute unilateral ureteral constriction or by sodium nitroprusside infusion (unpublished observations). Thus, it appears unlikely that the hypocapnia-induced fall in U-B Pco2 was caused by the reduced rate of bicarbonate excretion.

Another possible cause for the decreased U-B Pco2 during hyperventilation is an increased rate of reabsorption of organic acids that were titrated in the distal nephron (18). This is unlikely in these experiments because the rate of organic anion excretion did not increase in bicarbonate-infused rats with superimposed hyperventilation.

The exact nephron segment responsible for the decreased U-B Pco2 noted in our studies cannot be precisely determined. However, as the transit time from the distal tubule to renal pelvis is approximately 20 s, (19) and the H2CO3 dehydration time is quite rapid in urine of low buffer capability (20), we consider it unlikely that the U-B Pco2 originates from areas other than the terminal collecting duct (18). It is conceivable that a reduction in the U-B Pco2 during hyperventilation, rather than being due to decreased hydrogen ion secretion, might occur as the consequence of carbonic acid reabsorption in the collecting duct or to an increased rate of its dehydration within the lumen of this nephron segment. We feel that these latter possibilities are less likely and therefore interpret our data to indicate that the collecting duct responds to hypocapnia with a decrease in hydrogen ion secretion. If one assumes a semi-quantitative dimension for the U-B Pco2, the data presented would suggest that the largest proportional decrease in hydrogen ion secretion during hyperventilation occurs in the collecting duct. That is, the U-B Pco2 decreased from 22±1 to 11±2, a 50% decline while the total bicarbonate reabsorption decreased only 17% from 33.2 to 27.5 mM/1 GFR. This type of analysis, however, is potentially misleading in view of the lack of information regarding the quantitative aspect of the relationship between the collecting duct secretion and the U-B Pco2.

Previous investigators have provided indirect evidence that distal hydrogen ion secretion might be reduced in hypocapnia. Stanbury and Thomson (1) found an increased urine sodium and potassium excretion in respiratory alkalosis and suggested that this might reflect decreased distal nephron hydrogen ion secretion. Gennari et al. (21) demonstrated that net acid excretion was decreased in dogs with chronic hyperventilation. Malnic et al. (22) demonstrated in micropuncture studies that hypocapnia in rats reduced fractional and absolute bicarbonate reabsorption in both the proximal and distal tubules. They also note that hyperventilation reduced the dysequilibrium pH in the distal convoluted tubule of bicarbonate-infused rats. Their results strongly suggest a decrease in distal tubular hydrogen ion secretion.

Two major implications result from our studies. One is that the U-B Pco2 provides a powerful tool to assess collecting duct hydrogen ion secretion in vivo provided the critical criteria are satisfied. In addition to controlling the excretion rates of phosphate, bicarbonate, and water, one must also control for the blood Pco2 and ensure that measurements are made under steady state conditions. The other is that acute hyperventilation reduces collecting duct hydrogen ion secretion. This might be the basis for the reduced net acid excretion observed with more prolonged hypocapnia, leading to the greater fall in blood bicarbonate concentration associated with chronic hypocapnia (21, 23-25). In conclusion, we interpret our data to indicate that acute steady-state hyperventilation causes a decrease in distal nephron (presumably collecting duct) hydrogen ion secretion.

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