

Inhibition of Sodium Transport by Prostaglandin E₂ across the Isolated, Perfused Rabbit Collecting Tubule

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ABSTRACT This study was designed to examine whether prostaglandin E₂ can directly affect sodium transport across isolated perfused rabbit renal collecting tubules. Changes in transepithelial potential and isotopic sodium fluxes in response to peritubular prostaglandin E₂ were measured. In addition, changes in transepithelial potential of the outer medullary collecting tubule in response to prostaglandin E₂ were also measured. With few exceptions, all rabbits received 5 mg/day desoxycorticosterone acetate for 4–11 days before experimentation. The results of the experiments show that: (a) prostaglandin E₂ inhibits the negative transepithelial potential in the cortical collecting tubule as well as the outer medullary collecting tubule; (b) prostaglandin E₂ inhibits net sodium transport out of the lumen by inhibiting efflux while backflux is unaffected; (c) prostaglandin E₂ produces this inhibition within 15 min, and the effects are dose dependent and reversible. These results suggest that prostaglandin E₂ may modulate sodium transport in vivo and may contribute to the final regulation of sodium excretion.

INTRODUCTION

The collecting duct participates in the final regulation of sodium excretion (1). Recent studies have demonstrated that the cortical collecting tubule (CCT)¹

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¹Abbreviations used in this paper: CCT, cortical collecting tubule; DOCA, desoxycorticosterone acetate; J_{Na,b}, bath-to-lumen sodium flux; J_{Na,l}, lumen-to-bath sodium flux; J_v, net water flux; MCT, outer medullary collecting tubule; PD, transepithelial potential difference; PGE₂, prostaglandin E₂.

is the segment sensitive to aldosterone (2, 3) and in this way is capable of regulating sodium reabsorption. Rapid natriuresis in response to a variety of stimuli such as acute volume expansion cannot be explained fully by inhibition of aldosterone because of its long duration of action. These studies were conducted to examine the possibility that a naturally occurring intrarenal substance could modulate sodium transport by the collecting tubule.

The renal medulla is a rich source of prostaglandins (4) and they have been implicated in sodium regulation by many investigators. No evidence currently exists to implicate prostaglandins in renal epithelial transport of salt though such evidence does exist for the jejunum (5), frog corneal epithelium (6), and toad bladder (7). It is difficult to interpret the effects of prostaglandins on renal epithelial transport with clearance techniques because of their well-demonstrated effects on renal blood flow (8, 9) and on vasopressin-induced osmotic water flow (10). The present studies were designed, therefore, to examine directly the effect of prostaglandin E₂ (PGE₂), the renal prostaglandin occurring in greatest quantity, on sodium transport across the in vitro-perfused rabbit collecting tubule.

METHODS

Segments of collecting tubule were dissected and perfused in vitro with the same techniques described previously (11). Briefly, female New Zealand white rabbits weighing 1.5–2.5 kg were used in all experiments. All rabbits were fed standard laboratory diet containing approximately 140 meq/kg sodium and 390 meq/kg potassium.

With the exception of a few rabbits so noted, all rabbits received 5 mg/day desoxycorticosterone acetate (DOCA) intramuscularly for 5–8 days before experimentation. This maneuver was done to insure a transepithelial potential difference (PD) lumen negative (2). All animals had free access to food and water before guillotine decapitation.

The kidney was quickly removed, a 1-mm slice was placed in chilled solution identical to the bath, and the appropriate segment of collecting tubule was dissected free.

The solution was maintained at pH 7.4 by continuous bubbling with 95% O₂ and 5% CO₂. Cortical collecting tubules (CCT) were 1.4–3.0 mm long and outer medullary collecting tubules (MCT) were 0.6–1.2 mm long. All studies were conducted at 37°C. Tubules were perfused with an artificial solution containing NaCl 105 mM, KCl 5 mM, NaHCO₃ 25 mM, Na₂HPO₄ 2.3 mM, Na acetate 10 mM, MgSO₄ 1 mM, CaCl₂ 1.8 mM, glucose 8.3 mM, and alanine 5 mM. The bath and dissection solutions were identical to the perfusion solution except for the addition of 5% fetal calf serum. PGE₂ was stored at –20°C in ethanol 10 mg/ml, and the bath solution containing PGE₂ was prepared immediately before use. PGE₂ prepared in this way is stable (12).

During all experiments, except those measuring bidirectional flux, the bath solution was continuously exchanged with a Delta micro-metering pump at 0.5 ml/min (Lab-line Instruments, Inc., Melrose Park, Ill.) while the bath fluid level was maintained by continuous suction at the opposite end. In this system >95% of the bath is exchanged in 5 min, and >99% is exchanged in 10 min. The bath solution can thus be changed continuously without inducing changes in turbulence or temperature. In addition, alterations in bath composition can be effected simply by changing the reservoir. Previous experiments have shown that solute concentration changes less than 2% in 2 h.

The PD was measured by techniques previously reported (13). Lumen-to-bath sodium flux ($J_{Na,lb}$) was calculated from measurements of the disappearance of ²²Na (10–20 μ Ci/ml) from the perfusate. Bath-to-lumen sodium flux ($J_{Na,bl}$) was determined by adding ²⁴Na (75–100 μ Ci/ml) to the bath (without bath perfusion) and measuring its appearance in the collected fluid. ²²Na and ²⁴Na (New England Nuclear, Boston, Mass.) were discriminated by using appropriate channels on a Packard model 3365 gamma spectrometer (Packard Instrument Co., Inc., Downers Grove, Ill.) and the sample was recounted 14 days later when ²⁴Na activity could not be detected. ²⁴Na activity was corrected to the time at which the counting began. ¹²⁵I-iothalamate (Glofil 125, Abbott Laboratories, N. Chicago, Ill.) was added to the perfusate (15 μ Ci/ml) and collections where the net water flux (J_v) was more negative than –0.1 nl/mm per min were discarded (5–10% of measurements) since this deviation reflects a leak in the seal at the collecting pipette. J_v was calculated according to the expression:

$$J_v = \frac{V_o}{L} \left[\frac{[^{125}I]_o}{[^{125}I]_i} - 1 \right],$$

where V_o is the collection rate, L is the length of the tubule, and $[^{125}I]_i$ and $[^{125}I]_o$ are the concentrations of ¹²⁵I in the perfused and collected fluid, respectively. $J_{Na,lb}$ was calculated according to the expression:

$$J_{Na,lb} = [Na]_i \frac{V_i}{L} \ln \frac{[^{22}Na]_i}{[^{22}Na]_o},$$

where $[Na]_i$ and $[^{22}Na]_i$ are the chemical and isotopic concentrations of Na in the perfusate, $[^{22}Na]_o$ is the isotopic concentration of sodium in the collected fluid, and V_i is the perfusion rate. $J_{Na,bl}$ was calculated according to the following expression which accounts for a significant efflux of sodium (modified from [14]):

$$J_{Na,bl} = \frac{[Na]_b [^{24}Na]_o}{[Na]_i [^{24}Na]_b} \left[\frac{J_{Na,lb}}{1 - \exp - (L \cdot J_{Na,lb} / V_i [Na]_i)} \right],$$

where $[Na]_b$ and $[^{24}Na]_b$ are the respective chemical and

isotopic concentrations in the bath and $[^{24}Na]_o$ is the isotopic concentration in the collected fluid.

Tubular perfusion rate was maintained between 3 and 6 nl/min by hydrostatic pressure. An equilibration period of 90–120 min was necessary to allow the control PD to stabilize. Three periods were recorded: control, PGE₂ addition to the bath, and recovery where PGE₂ was removed from the bath. Once the control PD was stable for 15 min, the bath was changed to the solution containing PGE₂.² The effect of PGE₂ became apparent within 2 min, and the maximum effect usually began to stabilize within 15 min. When the PD was stable for 10 min the bath was changed to the control bath. The return of a stable potential took 40–50 min.

To insure an unbiased interpretation of the results changes in PD were measured in nine “blind” experiments with solutions prepared by a person not connected with these investigations. The solution contained either 1.0 μ M PGE₂ or an equivalent amount of carrier (ethanol 0.004% vol/vol).

Sodium flux measurements were made for two to four collection periods in each of the three periods. Collections were initiated when the PD became stable, and the collected isotopes were counted. Bath perfusion during unidirectional flux experiments was constant, and distilled water was added to the bath during bidirectional flux experiments to replace evaporative losses. Before the beginning of the second period, an appropriate amount of PGE₂ was added to the bath to make the concentration 1.0 μ M. During the 40–60 min necessary for recovery of the PD, the bath exchange was continuous. It was again discontinued before the measurement of recovery bidirectional fluxes.

All analyses were done with the Student's t test for paired values, and mean values are reported \pm standard error.

RESULTS

Effect of PGE₂ on the transepithelial potential of the cortical collecting tubule and outer medullary collecting tubule. The addition of PGE₂ to the bath caused the magnitude of the negative PD of the CCT and MCT to decrease. No effect on PD was seen when comparable doses of PGE₂ were added to the perfusate ($n = 5$). The peritubular effect was rapid, requiring only minutes to become apparent with a maximal response beginning to stabilize within 15 min. Though on rare occasion the PD became transiently more negative, the majority of responses simply demonstrated an inhibition. The onset of recovery required at least 15 min after complete elimination of PGE₂ from the bath and stable recovery generally required 40–60 min. Fig. 1 displays the change in PD of CCT as a function of PGE₂ concentration of the bathing solution.

Doses of 10 nM resulted in a barely detectable response while doses of 0.1–10 μ M effected an increasing response such that at 10 μ M the mean inhibition was greater than 80%. The PD of CCT from normal rabbits is difficult to predict and is dependent, at least in part, on the mineralocorticoid activity

² Kindly provided by Dr. John Pike, The Upjohn Co., Kalamazoo, Mich.

(2). The administration of DOCA insures a negative PD which is invariably responsive to PGE_2 in doses of $0.1 \mu\text{M}$ or greater. The pretreatment of the rabbit with DOCA appeared to have no effect on the PD response when compared to tubules of equivalent base-line PD not so treated.

Nine experiments were conducted in a blinded fashion where the experimenter was unaware whether the solution contained PGE_2 ($1.0 \mu\text{M}$, $n = 6$) or the carrier ($n = 3$). PGE_2 in these experiments caused a reduction in the magnitude of the PD by $>26\%$ while the carrier had no effect ($P < 0.001$).

Since PGE_2 is found in high concentrations in the medulla (4), the response of the PD in the MCT was also examined. This segment is morphologically indistinguishable from the CCT at $\times 400$ during the experiment. Fig. 2 demonstrates the response of the negative PD to various doses of PGE_2 in the bath. The response to 1 and $10 \mu\text{M}$ PGE_2 was similar to that found in the CCT. However, the tubule failed to recover from 0.1 mM in 1 h, the time at which

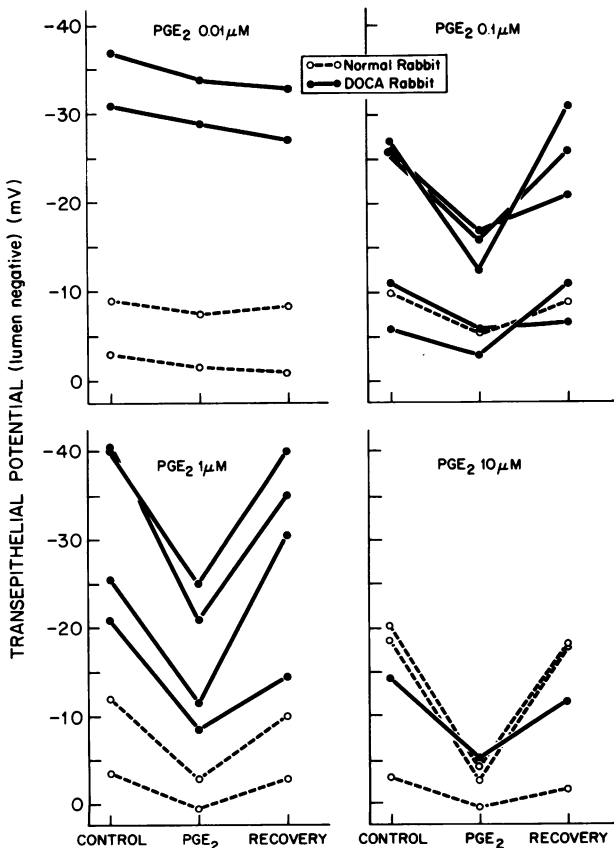


FIGURE 1 Effect of PGE_2 on rabbit CCT PD. PGE_2 was added to the bathing solution in the concentrations shown. In all experiments a stable PD was obtained after 90–120 min. The stable PD resulting from PGE_2 addition is recorded. Recovery was complete within 60 min.

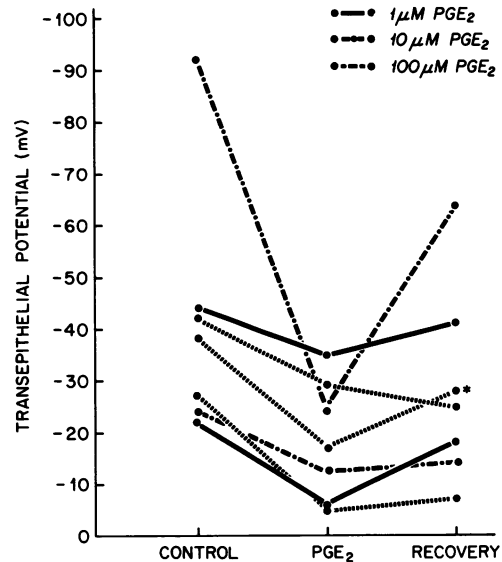


FIGURE 2 Effect of PGE_2 on outer medullary collecting tubule PD. Technique was identical to that used in Fig. 1. All rabbits received DOCA. *Indicates one experiment where recovery from 0.1 mM PGE_2 was allowed to continue for 180 min rather than the usual 60 min. At 0.1 mM no tubule recovered within 60 min.

recovery was virtually complete when smaller doses were used. The one tubule demonstrating recovery after 0.1 mM did so only after 3 h. The finding that PGE_2 inhibits the negative PD in the MCT in a fashion similar to CCT provides stronger evidence that PGE_2 may act similarly *in vivo*.

Effect of PGE_2 ($1.0 \mu\text{M}$) on sodium flux. Table I depicts the alterations in simultaneous bidirectional sodium fluxes across the CCT induced by the addition of $1.0 \mu\text{M}$ PGE_2 to the bath. There was no significant change in backflux ($J_{\text{Na},\text{bl}}$) while efflux was significantly inhibited. In an additional six experiments unidirectional lumen-to-bath fluxes were measured under conditions identical to the PD experiments (i.e., with the bath pump on). PGE_2 inhibited efflux in a similar fashion: 25.5 ± 4.0 (control), 15.1 ± 3.5 (PGE_2), and 21.3 ± 4.1 (recovery). The differences are highly significant ($P < 0.001$).

The resulting alteration in sodium transport can be expressed as a change in flux ratio (Table I) or in a net sense. If one assumes a perfusion rate of 2 nl/min of isotonic fluid and a net efflux of $21.2 \text{ pEq. cm}^{-1} \text{ per s}$ ($J_{\text{Na},\text{lb}} - J_{\text{Na},\text{bl}}$), in 2 mm of tubular length, 45% of the sodium load would be reabsorbed. PGE_2 ($1.0 \mu\text{M}$) would decrease this reabsorption by an average of 40–50%.

There is a greater fall in $J_{\text{Na},\text{lb}}$ in these bidirectional flux experiments than in the unidirectional flux experiments, and the recovery in the bidirectional experiments is less complete. These findings are probably

TABLE I
Effect of 1.0 μ M PGE₂ Applied to Peritubular Surface of CCT on Transepithelial Potential and Bidirectional Sodium Flux

Experiment	Potential			J _{Na,b}			J _{Na,a}			J _{Na,b} /J _{Na,a}		
	Control	Exp.	Recovery	Control	Exp.	Recovery	Control	Exp.	Recovery	Control	Exp.	Recovery
	mV			peq·cm ⁻¹ ·s ⁻¹			peq·cm ⁻¹ ·s ⁻¹			peq·cm ⁻¹ ·s ⁻¹		
1	-38	-19	-38	21.6	8.7	13.3	4.8	3.6	4.4	4.5	2.4	3.0
2	-49	-16	-46	30.7	7.0	14.9	4.7	3.9	3.6	6.5	1.8	4.1
3	-35	-14	-66	33.2	8.2	16.6	4.0	4.2	4.1	8.3	1.9	4.0
4	-38	-12	-39	18.9	6.7	7.0	6.2	6.5	4.6	3.0	1.0	1.5
Mean	-40	-15	-47	26.1	7.6	12.9	4.9	4.5	4.2	5.6	1.8	3.2
SEM	±3.1	±1.5	±6.5	±3.4	±0.5	±2.1	±0.5	±0.7	±0.2	±1.2	±0.3	±0.6
P	<0.0025			<0.01			NS			<0.025		

Each flux value is the mean of two to four collections. Exp., experimental.

due to the fact that bidirectional experiments take longer to complete than do unidirectional ones. A longer exposure to PGE₂ and tubule fatigue most likely account for the differences. That this fatigue does not affect membrane permeability is evidenced by the sodium backflux which tended to fall insignificantly with time.

DISCUSSION

The present studies provide the first evidence that PGE₂ can affect renal tubular sodium transport directly. In the setting of stimulated sodium transport across the cortical collecting tubule and outer medullary collecting tubule PGE₂ in doses ranging from 0.1 μ M to 0.1 mM when applied to the peritubular surface causes an inhibition of the negative PD (Figs. 1 and 2) and a decrease in the outward sodium flux without a significant change in the influx of sodium (Table I). The effect is rapid, reversible, and dose dependent. The net efflux of sodium can be inhibited by as much as 85% by 1.0 μ M PGE₂.

Since the discovery of large quantities of prostaglandins in the renal medulla (4), many attempts have been made to elucidate its mechanisms of action. The general approaches that have been utilized to assess the effect of prostaglandins on renal function in the intact kidney include: (a) infusion of prostaglandins into the renal artery (8, 9, 15, 16), (b) infusion of sodium arachidonate into the renal artery to stimulate endogenous prostaglandins (17, 18), and (c) reduction of endogenous intrarenal prostaglandins by drugs or dietary deficiency (19, 20). Because of concomitant changes in renal blood flow and water excretion, the effects of prostaglandins on renal tubular transport are difficult, if not impossible, to interpret. Because of these difficulties and the controversies surrounding the role of prostaglandins in sodium excretion, we have chosen to study their ef-

fects in the *in vitro*, isolated, perfused tubule. The advantage of the technique is that one can control completely the epithelial environment and thus eliminate such conditions as intrarenal hydrostatic pressure, regional blood flow, and circulating factors which might cause uninterpretable changes.

The sodium fluxes reported in these CCT experiments are two to three times higher than those reported by other investigators (21–24) when they have examined the CCT from rabbits not receiving a mineralocorticoid hormone. These flux data support the electrophysiologic data implying that the collecting tubule is the site where aldosterone exerts its action (2, 3). The capacity of the CCT taken from these DOCA-stimulated rabbits to transport sodium is second only to the proximal convoluted tubule which can transport sodium at a rate of 37 peq·cm⁻¹ per s (25). No other segment when evaluated under similar conditions has been found to transport more than 13 peq·cm⁻¹ per s (26). The range of fluxes exhibited by the collecting tubule (ranges presumably due to the effect of mineralocorticoid hormones) and the magnitude of its capacity are entirely reasonable considering the teleological importance of maintaining extracellular volume.

The inhibition of sodium transport in the rabbit collecting tubule by PGE₂ may appear to be at variance with the stimulation of sodium transport in the toad bladder by PGE₁, since the toad bladder is sometimes compared with the mammalian collecting tubule. However Besley et al. (27) may have resolved this issue since they have demonstrated an inhibition of sodium transport by PGE₂ while confirming the stimulatory effect of PGE₁ in the theophylline-treated toad bladder.

The evidence that this action of PGE₂ is physiologically significant is circumstantial but supported by four studies. (a) Recent determinations of renal PGE₂ by gas chromatography-mass spectrometry (28) demon-

strate levels of $0.19 \pm 0.04 \mu\text{g} \cdot \text{g}^{-1}$ in the cortex and $4.36 \pm 1.04 \mu\text{g} \cdot \text{g}^{-1}$ in the medulla, concentrations which fall well within the effective dose range utilized in our experiments. (b) Histochemical studies have demonstrated prostaglandin dehydrogenase (the enzyme responsible for the degradation of PGE_2) activity in the thick ascending limb of Henle, the distal convoluted tubule, and the inner medullary collecting tubule, a distribution which skips the segments of the collecting tubule examined in this study (29). (c) Inhibition of prostaglandin synthesis is associated with a striking increase in medullary NaCl concentrations (30), a finding which is compatible with the thesis that prostaglandins normally inhibit sodium reabsorption in the collecting tubule. (d) Bartter's syndrome, a pathological state identified by hypokalemic alkalosis; high circulating renin and aldosterone; a tendency to renal salt wasting; and a normal blood pressure which is resistant to angiotensin II infusions is also accompanied by high urinary prostaglandins (31–33). Several patients have been reported whose metabolic and pathologic syndrome has been reversed with indomethacin, an inhibitor of prostaglandin synthesis (31, 32). Taken in the aggregate, these studies strongly suggest that PGE_2 may inhibit the reabsorption of sodium by the collecting tubule in vivo.

In summary, we have shown that PGE_2 can inhibit sodium efflux from the lumen of cortical collecting tubules taken from rabbits stimulated by DOCA. The inhibition is rapid, reversible, and dose dependent. In addition, PGE_2 exerts a similar effect on the outer medullary collecting tubule, a fact which greatly enhances the probability that PGE_2 acts in vivo to inhibit directly sodium reabsorption by the collecting tubule.

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REFERENCES

- Stein, J. H., and H. J. Reineck. 1974. The role of the collecting duct in the regulation of excretion of sodium and other electrolytes. *Kidney Int.* **6**: 1–9.
- Gross, J. B., M. Imai, and J. P. Kokko. 1975. A functional comparison of the cortical collecting tubule and the distal convoluted tubule. *J. Clin. Invest.* **55**: 1284–1294.
- Gross, J. B., and J. P. Kokko. 1977. Effects of aldosterone and potassium-sparing diuretics on electrical potential differences across the distal nephron. *J. Clin. Invest.* **59**: 82–89.
- Lee, J. B., K. Crowshaw, B. H. Takman, K. A. Attrep, and J. Z. Gougoutas. 1967. The identification of prostaglandins E_2 , $\text{F}_2\alpha$ and A_2 from rabbit kidney medulla. *Biochem. J.* **105**: 1251–1260.
- Kimberg, D. V., M. Field, J. Johnson, A. Henderson, and E. Gershon. 1971. Stimulation of intestinal mucosal adenyl cyclase by cholera enterotoxin and prostaglandins. *J. Clin. Invest.* **50**: 1218–1230.
- Beitch, B. R., I. Beitch, and J. A. Zadunaisky. 1974. The stimulation of chloride transport by prostaglandins and their interaction with epinephrine, theophylline, and cyclic AMP in the corneal epithelium. *J. Membr. Biol.* **19**: 381–396.
- Lipsom, L. C., and G. W. G. Sharp. 1971. Effect of prostaglandin E_1 on sodium transport and osmotic water flow in the toad bladder. *Am. J. Physiol.* **220**: 1046–1052.
- Gross, J. B., and F. C. Bartter. 1973. Effects of prostaglandins E_1 , A_1 and $\text{F}_2\alpha$ on renal handling of salt and water. *Am. J. Physiol.* **225**: 218–224.
- Martinez-Maldonado, M., N. Tsaparas, G. Eknayan, and W. N. Suki. 1972. Renal actions of prostaglandins: Comparison with acetylcholine and volume expansion. *Am. J. Physiol.* **222**: 1147–1152.
- Grantham, J. J., and J. Orloff. 1968. Effect of prostaglandin E_1 on the permeability response of the isolated collecting tubule to vasopressin, adenosine 3', 5'-monophosphate and theophylline. *J. Clin. Invest.* **47**: 1154–1161.
- Kokko, J. P. 1970. Sodium chloride and water transport in the descending limb of Henle. *J. Clin. Invest.* **49**: 1838–1846.
- Brummer, H. C. 1971. Storage life of prostaglandin E_2 in ethanol and saline. *J. Pharm. Pharmacol.* **23**: 804–805.
- Kokko, J. P. 1973. Proximal tubule potential difference. Dependence on glucose, HCO_3^- and amino acids. *J. Clin. Invest.* **52**: 1362–1367.
- Imai, M., and J. P. Kokko. 1974. Sodium chloride, urea, and water transport in the thin ascending limb of Henle. Generation of osmotic gradients by passive diffusion of solutes. *J. Clin. Invest.* **53**: 393–402.
- Strandhoy, J. W., C. E. Ott, E. G. Schneider, L. R. Wibis, N. P. Beck, B. B. Davis, and F. G. Knox. 1974. Effects of prostaglandins E_1 and E_2 on renal sodium reabsorption and Starling forces. *Am. J. Physiol.* **226**: 1015–1021.
- Fülgraff, G., and A. Meiforth. 1971. Effects of prostaglandin E_2 on excretion and reabsorption of sodium and fluid in rat kidneys (micropuncture studies). *Pfluegers Arch. Eur. J. Physiol.* **330**: 243–256.
- Tannenbaum, J., J. A. Splawinski, J. A. Oates, and A. S. Nies. 1975. Enhanced renal prostaglandin production in the dog. I. Effects on renal function. *Circ. Res.* **36**: 197–203.
- Chang, L. C. T., J. A. Splawinski, J. A. Oates, and A. S. Nies. 1975. Enhanced renal prostaglandin production in the dog. II. Effects on intrarenal hemodynamics. *Circ. Res.* **36**: 204–207.
- Rosenthal, J., P. G. Simone, and A. Silbergleit. 1974. Effects of prostaglandin deficiency on natriuresis, diuresis, and blood pressure. *Prostaglandins*. **5**: 435–440.
- Kirschenbaum, M. A., and J. H. Stein. 1976. The effect of inhibition of prostaglandin synthesis on urinary sodium excretion in the conscious dog. *J. Clin. Invest.* **57**: 517–521.
- Frindt, G., and M. B. Burg. 1972. Effect of vasopressin on sodium transport in renal cortical collecting tubules. *Kidney Int.* **1**: 224–231.
- Stoner, L. C., M. B. Burg, and J. Orloff. 1974. Ion

- transport in cortical collecting tubule; effect of amiloride. *Am. J. Physiol.* **227**: 453-459.
23. Grantham, J. J., M. B. Burg, and J. Orloff. 1970. The nature of transtubular Na and K transport in isolated rabbit renal collecting tubules. *J. Clin. Invest.* **49**: 1815-1826.
 24. Fine, L. G., J. J. Bourgoinie, K. W. Hwang, and N. S. Bricker. 1976. On the influence of the natriuretic factor from patients with chronic uremia on the bioelectric properties and sodium transport of the isolated mammalian collecting tubule. *J. Clin. Invest.* **58**: 590-597.
 25. Kokko, J. P., M. B. Burg, and J. Orloff. 1971. Characteristics of NaCl and water transport in the renal proximal tubule. *J. Clin. Invest.* **50**: 69-76.
 26. Burg, M. B., and N. Green. 1973. Function of the thick ascending limb of Henle's loop. *Am. J. Physiol.* **224**: 659-668.
 27. Besley, G. T. N., D. A. Frith, and R. S. Snart. 1974. An inhibitor of the toad bladder cyclic AMP system and theophylline-stimulated Na⁺ transport. *Steroids Lipids Res.* **5**: 365-379.
 28. Larsson, C., and E. Ånggård. 1976. Mass spectrometric determinations of prostaglandin E₂, F₂, and A₂ in the cortex and medulla of the rabbit kidney. *J. Pharm. Pharmacol.* **28**: 326-328.
 29. Nissen, H. M., and H. Andersen. 1968. On the localization of a prostaglandin-dehydrogenase activity in the kidney. *Histochemie.* **14**: 189-200.
 30. Ganguli, M., L. Tobian, S. Azar, and M. O'Donnell. 1976. Effect of indomethacin, a prostaglandin synthetase inhibitor, and dietary NaCl on sodium concentration and plasma flow in renal papilla. *Clin. Res.* **24**: 467A. (Abstr.)
 31. Verberckmoes, R., B. Van Damme, J. Clement, A. Amery, and P. Michielsen. 1976. Bartter's syndrome with hyperplasia of renomedullary cells: Successful treatment with indomethacin. *Kidney Int.* **9**: 302-307.
 32. Fichman, M. P., N. Telfer, P. Zia, P. Speckart, M. Golub, and R. Rude. 1976. Role of prostaglandins in the pathogenesis of Bartter's syndrome. *Am. J. Med.* **60**: 785-797.
 33. Gill, J. R., J. C. Frölich, R. E. Bowden, A. A. Taylor, H. R. Keiser, H. W. Seyberth, J. A. Oates, and F. C. Bartter. 1976. Bartter's syndrome: A disorder characterized by high urinary prostaglandins and a dependence of hyperreninemia on prostaglandin synthesis. *Am. J. Med.* **61**: 43-51.