Thyroid Dysfunction in Chronic Renal Failure

A STUDY OF THE PITUITARY-THYROID AXIS AND PERIPHERAL TURNOVER KINETICS OF THYROXINE AND TRIIODOTHYRONINE

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ABSTRACT Thyroid function was evaluated in 46 patients with end-stage kidney disease and 42 normal subjects. Patients were studied before and after the institution of maintenance hemodialysis (HD) and after renal transplantation (RT). Serum total triiodothyronine concentrations (TT₃, ng/100 ml, mean±SD) were 63±17 and 83±22 in the non-HD and HD groups, respectively. Values from normal subjects were 128±25 and from RT patients 134±20. The TT₃ was in the hypothyroid range (<78 ng/100 ml; 2 SD below normal mean) in 80% of non-HD and 43% of HD patients. Mean serum total thyroxine concentration (TT₄), although within the normal range, was lower than the control value. T₄-binding globulin capacity was also slightly lower but the difference was not statistically significant. Among patients whose TT₄ was 1 SD below the normal mean, the free T₄ index was equally depressed, suggesting that factors other than decreased binding capacity might be responsible for the low TT₄. In addition, there was a 37% incidence of goiter. Mean serum thyroid-stimulating hormone (TSH) was not elevated and the TSH response to thyrotropin-releasing hormone (TRH) was distinctly blunted, suggesting the possibility of pituitary dysfunction as well. In vivo ¹²₃I-L-T₄ and ¹³¹I-L-T₃ kinetics during 0.2 mg/day of L-T₄ replacement showed marked reduction in T₃ turnover rate in the uremic patients, both before and during HD; the values (μg T₃/day, mean±SD) for the different groups were as follows: normal, 33.8±6.1; non-HD, 13.5±2.6; HD, 12.9±3.1; and RT, 30.3±7.1. The low T₃ turnover rate was due to impaired extrathyroidal conversion of T₄ to T₃. The mean percent±SD of metabolized T₄ converted to T₃ was 37.2±5.8 in normal subjects, 15.7±3.1 in non-HD, 12.8±1.7 in HD, and 34.0±14.7 in RT patients. In contrast, thyroidal T₃ secretion rate was not different between the control and the three patient groups. Thus, it appears that uremia affects thyroid function at several levels: (a) subnormal pituitary TSH response to TRH; (b) possible intrathyroidal abnormalities as suggested by slightly decreased TT₄ and high incidence of goiter; and (c) abnormal peripheral generation of T₃ from T₄. Restoration of renal function with RT resulted in normalization of all parameters of thyroid function with the exception of blunted or absent TSH response to TRH. The latter may be a direct consequence of glucocorticoid administration.

INTRODUCTION

Patients with chronic renal failure often exhibit clinical features and laboratory findings suggestive of hypothyroidism. Despite extensive studies, thyroid status in uremia is still inconclusive due to the complexity of the system studied. For example, thyroidal radioiodide uptake is decreased because of reduced renal iodide clearance (1, 2). The serum hormonal concentration may be altered by changes in the binding capacity of serum proteins (3, 4), and abnormal serum constituents in uremia were thought to displace thyroid hormone from its protein-binding sites (5). Heparin, a standard hemodialysis (HD)¹ medication, has been shown to increase serum free thyroxine concentration (6, 7). Peritoneal dialysis is an effective way of

¹Abbreviations used in this paper: HD, hemodialysis; RT, renal transplantation; TT₃, serum total triiodothyronine; TT₄, serum total thyroxine; T₃, triiodothyronine; T₄, thyroxine; FTI, free thyroxine index; TBG, thyroxine-binding globulin; TSH, thyroid-stimulating hormone; TRH, thyrotropin-releasing hormone; MCR, metabolic clearance rate; D, turnover rate.

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removing thyroid hormones from the circulation (8, 9). Goiter may be induced by the high serum level of inorganic iodide (1, 2, 10) or retention of goitrogenic substances normally excreted by the kidney (11). Finally, malnutrition may be a contributing factor in altering thyroid function tests (12).

This communication presents the results of our attempts to study systematically the pituitary-thyroid axis as well as the peripheral metabolism of thyroid hormones in 46 patients with irreversible renal failure, before and during HD, and after renal transplantation (RT). Since recent studies from our laboratory (13) and by others (14–16) have demonstrated a reduction in the serum concentration of total triiodothyronine (TT₃) and since more than half of the circulating triiodothyronine (T₃) is derived from conversion of thyroxine (T₄) to T₃ in the periphery (17–19), emphasis was placed on the study of this particular pathway of T₄ metabolism. Additional studies include measurements of serum concentrations of total thyroxine (TT₄), TT₃, free thyroxine index (FTI), thyroxine-binding globulin (TBG) capacity, thyroid autoantibodies, and thyroid-stimulating hormone (TSH) as well as TSH response to thyrotropin-releasing hormone (TRH). The results indicate that uremia affects thyroid function in several ways, the most important of which is a profound impairment of extrathyroidal conversion of T₄ to T₃ resulting in a selective reduction in serum TT₃.

CLINICAL MATERIAL

46 patients (26 men and 20 women) with end-stage kidney disease were studied. Their ages ranged from 20 to 58 yr. The etiology of their renal failure included hypertensive nephrosclerosis, chronic glomerulonephritis, polycystic kidney disease, interstitial nephritis, and epithelial cell disease with glomerulosclerosis. 20 patients were studied before institution of HD, 37 during HD, and 11 after successful RT. 17 patients were studied more than once as they went through the various stages of treatment. At the time of this study, the mean duration of dialysis was 15 mo (range, 2–30 mo). Hemodialysis was performed three times weekly with EX-23 or EX-25 coil dialyzers. Of those patients studied before entrance into the dialysis program, blood samples were obtained during their clinic visits. In patients studied during maintenance HD, blood was routinely sampled immediately before HD and before heparin administration. After RT, blood samples were obtained periodically throughout the recovery phase, until thyroid function had either returned to normal or stabilized, representing a period of at least 6 mo. As both TT₄ and TT₃ increased progressively, only values that have reached a stable plateau were used in the computation of mean and standard deviation for this particular group. All renal failure patients received aluminum hydroxide, multivitamins, folic acid, and ferrous sulfate. In addition, many patients required antihypertensive medications including methyldopa, hydralazine, and propranolol. Patients who received Dilantin and androgens were excluded from this study. Diuretics were not prescribed. The RT subjects received prednisone and azathioprine in various doses. The control group consisted of 42 healthy subjects including laboratory personnel and blood donors. Their age and sex distribution were comparable to those of the patients.

The experimental protocol was approved by the Committee on Human Experimentation and the Committee on the Use of Radioisotopes of the University of Chicago Hospitals. Informed written consent was obtained from the patients as well as from the normal volunteers.

METHODS

Thyroid function tests. Serum TT₄ was measured by the competitive binding procedure and the FTI was calculated (20). The T₄-binding capacity of TBG was measured by saturation (4) and the concentration of serum TT₃ by double-antibody precipitation radioimmunoassay (21). The T₃ antiserum was supplied by Dr. Gerald Burke, Cook County Hospital, Chicago, Ill. The characteristics of this antiserum have been previously described (21). The antibody was produced in the rabbit by immunization with T₃ coupled to human serum albumin; T₃ was displaced from TBG with 8-ami-nol-napthalene sulfonic acid. The method was sensitive to detect 10 ng T₃/100 ml of serum (21). The intra- and inter-assay coefficients of variation were <5 and 15%, respectively. Both T₃ and 3',3,5'-triiodo-L-thyr- 
onine (reverse T₃) in amounts equivalent to 50 μg/100 ml and 1,000 ng/100 ml of serum did not cross-react with T₃ in the radioimmunoassay. Further, 95–102% of known amounts of T₃ added to the serum of normal and uremic subjects were recovered. Serum concentration of TSH was also measured by the double-antibody precipitation radioimmunoassay (22) with one modification, in which the antiserum was incubated with 1,000 U of human chorionic gonadotropin at room temperature for 24 h. The anti-human TSH serum and purified human TSH for iodination were prepared by the National Pituitary Agency and supplied by the National Institutes of Health. The human TSH standard 68/38 was supplied by the Medical Research Council, London, England. Luteinizing hormone in amounts equivalent to 500 ng/ml (LER-907) did not interfere with the TSH assay. Thyro 
globulin and thyroid microsomal antibodies were determined by the sheep erythrocyte hemagglutination assay (23). For each test, the average value of several determinations obtained from the same patient was used in calculating the means and standard deviations of each group.

Estimation of thyroid gland size and iodine content. The size and consistency of the thyroid gland was assessed independently by two of us (V. S. Lim and S. Refetoff). The iodine content of the gland was estimated by fluorescent scanning (24).

TRH stimulation test. The test was performed during the morning in the fasting state in seven normal subjects and seven HD and three RT patients. For the HD patients, the test was carried out on the days without dialysis; for the RT patients, it was done 8–18 mo after restoration of renal function. Two basal blood samples were obtained 30 min apart before the intravenous injection of 400 μg of synthetic TRH (Abbott Laboratories, North Chicago, Ill.). Additional blood samples were obtained 15, 30, 45, 60, 90, 120, and 180 min after TRH administration; TSH was measured in all samples but TT₃ was measured only in samples obtained immediately before and 180 min after TRH.

Peripheral turnover kinetics of T₄ and T₃. The peripheral metabolism of T₄ and T₃ was studied simultaneously in 22 subjects: 5 healthy euthyroid volunteers, 12 uremic patients (4 before and 8 after institution of maintenance HD), and 5 patients after successful RT. ¹³¹I-L-Thyroxine (¹³¹I-T₄) and ¹³¹I-L-triiodothyronine (¹³¹I-T₃) were purchased from Industrial Nuclear Co., Inc., St. Louis.
Mo. Their specific activities were 100 and 85 µCi/µg, respectively. The radiochemical purity of both T4 and T3 in the isotope preparations was tested by descending paper chromatography in tertiary amyl alcohol:hexane:ammonia (25) and found to be >95%. The labeled hormones were mixed in sterile saline containing 5% salt-poor normal human serum albumin (Cutter Laboratories, Berkeley, Calif.). The final mixture was passed through a Millipore filter (Millipore Corp., Bedford, Mass.). Each patient received 2 ml of the saline-albumin mixture containing 10 µCi of 131I-T4 and 20 µCi of 131I-T3. This solution was injected intravenously and blood samples were obtained from the contralateral arm before, at 5 min, and at 2, 4, 6, 8, and 24 h after injection. Thereafter, samples were obtained every 24-48 h for another 8-10 days. The sera were stored at -20°C and assayed in batches with appropriate standards. Radioactivity was determined in all serum samples. Stable T4 and TT3 were measured in samples taken immediately before isotopic injection and on days 1-3, 5, 7, and 9; the mean value for each individual subject was used for the calculation of various kinetic parameters. In the HD patients, the isotope was injected 1 to 2 h after completion of HD treatment. The 24-h sample, as well as the subsequent blood samples, were obtained immediately before HD. Because of these patients' tendency to retain fluid and hemodilution, which may reduce thyroid hormone concentrations, total serum protein content was determined in every sample by refractometry and corrections were made for both T4 and TT3 concentrations as previously described (26). We used the protein concentration in the sample taken immediately before isotopic injection as base line because that was the time most HD patients were closest to their dry weight. To suppress endogenous thyroid hormone secretion and prevent recirculation of the labeled iodine, all subjects received 0.2 mg of L-thyroxine (Synthroid, Flint Laboratories, Deerfield, Ill.) daily, beginning 7 days before isotopic injection and during the entire study. Five drops of a saturated solution of potassium iodide was given ½ h before the administration of labeled hormones and was continued twice daily throughout the study period. After completion of each study, 2 ml of each serum sample was subjected to trichloroacetic acid (TCA) precipitation and the radioactivity in the precipitates was counted for 10 min in a dual channel gamma spectrometer (Packard Instrument Co. Inc., Downers Grove, Ill.). Contribution of 131I in the 131I channel as well as isotope decay were corrected by inclusion of appropriate standards. Only samples with isotope contents of at least fourfold the background were used in the calculations. Dose standards were prepared by addition of 0.1% of the dose to the preinjection serum sample of each patient and were subsequently handled in a manner identical to the postinjection samples.

The serum disappearance rate of TCA-precipitable 131I activity from days 1 to 10 was used to calculate the T4 kinetics. For calculation of T3 turnover kinetics, the disappearance rate of TCA-precipitable 131I activity from days 1 to 3 was used. The fractional turnover, distribution space, and metabolic clearance rate (MCR) of T4 were calculated by the method of single compartment kinetics (27). The MCR of T3 was calculated by the noncompartmental method (19, 28). The turnover rate (D) of T4 and T3 was derived from the product of MCR and mean serum concentration of the respective iodothyronine. Assuming that during the study period administration of L-thyroxine completely suppressed endogenous thyroid hormone secretion, the production rate of T3 from monodeiodination of T4 in the peripheral tissue (percent conversion of T4 to T3) could be calculated (19). The percent of administered T4 absorbed was also calculated as described by Surks et al. (19).

Assuming that MCR of T4 and T3, as well as the percent of T4 to T3 conversion in each individual, were identical before and during T4 replacement, the T3 produced by monodeiodination and that derived from thyroidal secretion during the basal state (before T4 replacement) could be calculated by using serum TT4 and TT3 concentrations before T4 administration:

\[
T_3 \text{ derived from } T_4 \quad (\mu mol/day) = T_4 \text{ turnover (}\mu\text{mol/day}) \times \frac{\% \text{ } T_4 \rightarrow T_3 \text{ conversion}}{100}
\]

and

\[
T_3 \text{ derived from thyroidal secretion (}\mu\text{g/day}) = \text{Total } T_3 \text{ turnover (}\mu\text{g/day}) - T_3 \text{ derived from } T_4 \text{ (}\mu\text{g/day}).
\]

The thyroidal secretion as percent of the total T4 turnover rate was also estimated.

Serum volume was estimated from the percent of the 131I-T4 dose present in serum 5 min after injection of isotopes. Previous studies have shown that the dilutions of albumin and T4 are almost identical at this time of sampling and therefore the latter is suitable for measurement of serum volume (29, 30).

Statistical analysis. All grouped data were expressed as mean±SD and were analyzed by the analysis of variance; mean square within value and paired t test were used to assess the statistical difference between any two means as indicated in the text (31).

RESULTS

Thyroid function tests (Table I). The most striking abnormality in the uremic patients was a reduction in serum TT3 concentration. The mean level of the combined non-HD and HD groups, 70 ng/100 mg, was 2 SD lower than the normal mean of 128 ng/100 ml. In fact, in 16 of 20 non-HD patients and in 16 of 37 HD patients, TT3 was 2 SD below the normal mean, i.e., <78 ng/100 ml. The mean values of TT4, FTI, and TBG capacity for the two groups of uremic patients were also reduced, but none of the differences reached statistical significance, with the exception of TT4 in the HD group. Both TT4 and TBG capacity were highly correlated in all four groups of subjects, the correlation coefficients (not shown in the tables) being as follows: normal, 0.820 (P < 0.001); before HD, 0.686 (P < 0.05); during HD, 0.665 (P < 0.01); and after RT, 0.827 (P < 0.02). In the two uremic groups there were 7 non-HD and 11 HD patients whose TT4 was <6.1 µg/100 ml (1 SD below normal mean); their corresponding FTI was also depressed, being 4.9±1.5 and 5.5±1.1, respectively. The TT4/TT3 ratio was markedly elevated in the two groups of renal failure patients, suggesting a selective decrease in TT3. Despite markedly depressed TT3 and slightly low TT4 serum TSH was not elevated in either group of uremic patients, nor was there any negative correlation between TT3 and TSH when individual data were analyzed. Restoration of renal function by RT resulted in normalization of all thyroid function tests. The
were studied for function abnormalities in disease, and our reaction was positive. Absence of renal tissue was analyzed separately. The concentration of TT3 did not become normal until after HD treatment, possibly because of insufficient time. These values are not different from those of the normal subjects, and this is illustrated in Fig. 1. While serum creatinine fell precipitously after RT, TT3 did not become normal until about 3–4 mo later.

Data from eight anephric patients undergoing HD were analyzed separately to determine whether total absence of renal tissue may accentuate the observed abnormalities. The means±SD of the various thyroid function tests in these eight patients were as follows: TT4, 4.9±0.9 μg/100 ml; TT3, 78±18 ng/100 ml; and TSH, 4.4±4.5 μU/ml. These values were not different from those of the other HD patients.

Thyroglobulin and thyroid microsomal antibodies were absent in the 14 uremic subjects tested. A positive reaction with at least one of the two antigens is usually found in 10% of an unselected population without obvious clinical evidence of thyroid dysfunction and in 85–95% of patients with autoimmune thyroid disease (23). The failure to detect antibodies in our patients suggests that autoimmune thyroid disease is not likely to be the cause of the thyroid function abnormalities observed in uremia.

**Effect of HD on thyroid function tests.** 11 patients were studied both before and after HD. The reciprocal changes between serum creatinine and TT3 concentration in two patients are illustrated in Fig. 1. Reciprocal changes in serum creatinine and TT3 before and after RT in two patients, EM (○) and RM (▲). Mean±SD for normal subjects is represented by the bars on the left. 0 time on the abscissa corresponds to RT.

<table>
<thead>
<tr>
<th>TABLE I</th>
<th>Thyroid Function Tests in Normal Subjects Compared to Uremic and Renal Transplant Patients*</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>TT4 FTI TBG capacity TT4/TT3 TSH</td>
</tr>
<tr>
<td></td>
<td>μg/100 ml μg T4/100 ml ng/100 ml μg/100 ml μU/ml</td>
</tr>
<tr>
<td>Normal</td>
<td>6.6±1.4 7.1±1.0 18.3±5.4 128±25 55±17 2.8±1.2</td>
</tr>
<tr>
<td>Range</td>
<td>4.2–8.7 4.5–8.5 14–26 80–160 33–73 0.1–8</td>
</tr>
<tr>
<td>Before HD</td>
<td>6.2±2.5 7.0±2.4 16.4±3.0 63±17 102±43 3.2±1.5</td>
</tr>
<tr>
<td>P1</td>
<td>NS NS NS &lt;0.01 &lt;0.01 NS</td>
</tr>
<tr>
<td>During HD</td>
<td>5.6±1.2 6.6±1.5 16.5±4.0 83±22 72±28 2.7±2.5</td>
</tr>
<tr>
<td>P1</td>
<td>&lt;0.05 NS NS &lt;0.01 &lt;0.05 NS</td>
</tr>
<tr>
<td>After RT</td>
<td>7.9±1.6 7.8±1.1 22.0±6.4 134±20 58±13 3.3±2.1</td>
</tr>
<tr>
<td>P1</td>
<td>NS NS NS &lt;0.01 &lt;0.001 NS</td>
</tr>
<tr>
<td>F ratio</td>
<td>4.958 1.633 2.451 57.735 12.722 1.216</td>
</tr>
<tr>
<td>P</td>
<td>&lt;0.01 NS NS &lt;0.001 &lt;0.001 NS</td>
</tr>
</tbody>
</table>

* Values are given as mean±SD; in normal subjects, the absolute range is also included. The numbers in parentheses indicate patients studied in each category. Individual values from all four groups were included and tested by the analysis of variance. The F ratio, degree of freedom, and P values are indicated.

1 The significance of the difference between the means of each patient group and the control was tested by using mean square within value.
initiation of maintenance HD. Their thyroid function tests are summarized in Table II. Hemodialysis resulted in only a slight increase in TT3 without significant change in other thyroid functions.

**Thyroid gland size and iodine content.** Abnormal thyroid glands were found in 9 of 24 HD patients examined (37%). Two had nodular goiters and the other seven had diffusely enlarged thyroid glands. 14 HD patients were randomly selected for fluorescent thyroid scanning. The thyroidal iodine content estimated by this method was slightly decreased in two, normal in six, and markedly increased in the remaining six patients.

**TRH stimulation test.** After TRH injection, serum TSH concentration rose rapidly in seven normal subjects, reached a peak by 30 min, and returned to basal levels in 3 h. Of the seven HD patients tested, one showed no increase in serum TSH concentration, four had a blunted response, and the other two showed a low normal response. In the latter, the peak was delayed until 45–60 min, and TSH levels remained elevated for more than 3 h (Fig. 2). The basal TSH value was normal in both the control and the uremic subjects, but the latter appeared to be lower. After TRH injection, serum TT3 concentration increased in five HD patients and was unchanged in the other two. The mean basal value was 72.1 and rose to 93.3 ng/100 ml, representing an increment of 28%. The difference between these two means as computed by paired t test was significant at <0.05 level. Of three RT subjects, 1 showed a blunted response and the other two failed to show any serum TSH increment (Fig. 3). Despite minimal or no serum TSH elevation, serum TT3 increased from a mean baseline of 155 to 187 ng/100 ml, a 21% increment; the difference was not significant.

**Peripheral turnover kinetics of T4 and T3.** Table III presents data on the T4 and T3 metabolism in each subject studied, as well as mean values and statistical analyses. Administration of L-T4 elevated to a variable extent the serum TT4 concentration in all subjects. In some, TT4 increased to above 9.4 μg/100 ml, the upper limit of normal. There was no correlation between the TT4 concentration before and during L-T4 replacement. In contrast, the incremental rise in TT3 concentration during L-T4 replacement was proportional to the pretreatment TT3 value in all subjects from the four groups, r = 0.934, P < 0.001 (Fig. 4). Thus, although serum TT3 increased after L-T4

<table>
<thead>
<tr>
<th>TABLE II</th>
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<tbody>
<tr>
<td><strong>Thyroid Function Tests in Uremic Patients Studied before and 2 mo after Initiation of Maintenance Hemodialysis</strong></td>
</tr>
<tr>
<td><strong>TT3</strong></td>
</tr>
<tr>
<td>μg/100 ml</td>
</tr>
<tr>
<td>Before HD</td>
</tr>
<tr>
<td>(11)</td>
</tr>
<tr>
<td>After HD</td>
</tr>
<tr>
<td>(11)</td>
</tr>
<tr>
<td>P</td>
</tr>
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</table>

* Values are given as mean±SD. The number of patients is in parentheses. Statistical significance of difference was calculated by the paired t test.

![Figure 2](image1.png)

**FIGURE 2** Serum TSH response to 400 μg of TRH given intravenously in seven HD patients as compared to that of seven normal subjects whose ranges are represented by the shaded area.

![Figure 3](image2.png)

**FIGURE 3** Serum TSH response to 400 μg of TRH given intravenously in three RT patients. The shaded area represents the range obtained from seven normal subjects.
treatment in all uremic patients, in most it remained below the normal mean of 128 ng/100 ml. In normal subjects and RT patients, serum TT₄ increased often to above 160 ng/100 ml, the upper limit of normal. Individual fluctuations in serum TT₄ and TT₃ throughout the turnover study showed no distinct trend, indicating that a steady state has been achieved.

Also shown in Table III is the absorption rate of exogenous T₄ in uremic subjects, which ranged from 31 to 84%. These values are compatible with data obtained by Surks et al. (19) and by Hays (32) in normal subjects. The RT patients also absorbed L-T₄ normally.

The parameters of T₄ metabolism were similar in all groups studied with the exception of a decrease in K and a slightly higher DS in the non-HD uremic patients. This is probably due to the distribution of the labeled hormone into a large extracellular volume. As a result of these reciprocal changes, the T₄ turnover rate remained within normal limits. The MCR and D of the HD uremic patients were normal, but there was a trend for the fractional turnover to be lower and for the DS to be higher. The two patients who had the largest distribution space also had the largest serum volume. The T₃ kinetics of RT subjects were similar to that of controls.

In contrast to the T₄ kinetics, T₃ kinetics were distinctly abnormal in both groups of uremic patients. The mean serum TT₃ concentration, T₃ turnover rate, and percent T₄ to T₃ conversion were drastically

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**Table III**

*Metabolism of T₄ and T₃ during L-T₄ Replacement*

<table>
<thead>
<tr>
<th>Group and patient</th>
<th>Creatinine</th>
<th>Weight</th>
<th>Serum volume</th>
<th>T₄ metabolism</th>
<th>T₃ absorption</th>
<th>T₃ metabolism</th>
<th>T₄ to T₃ conversion</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>mg/100 ml</td>
<td>kg</td>
<td>liters</td>
<td>µg/ml</td>
<td>%/day</td>
<td>liters/day</td>
<td>µg/day</td>
</tr>
<tr>
<td>Normal</td>
<td></td>
<td></td>
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<tr>
<td>D. B.</td>
<td>1.0</td>
<td>70</td>
<td>3.79</td>
<td>8.3</td>
<td>11.5</td>
<td>12.8</td>
<td>1.47 ± 0.71</td>
</tr>
<tr>
<td>T. C.</td>
<td>0.8</td>
<td>59</td>
<td>3.30</td>
<td>9.2</td>
<td>9.8</td>
<td>10.1</td>
<td>0.99 ± 0.91</td>
</tr>
<tr>
<td>F. K.</td>
<td>1.1</td>
<td>77</td>
<td>3.86</td>
<td>8.6</td>
<td>9.8</td>
<td>12.4</td>
<td>1.21 ± 0.43</td>
</tr>
<tr>
<td>W. S. T.</td>
<td>1.2</td>
<td>74</td>
<td>4.64</td>
<td>8.4</td>
<td>10.9</td>
<td>12.3</td>
<td>1.34 ± 0.11</td>
</tr>
<tr>
<td>E. S.</td>
<td>0.9</td>
<td>46</td>
<td>3.39</td>
<td>11.2</td>
<td>9.2</td>
<td>11.1</td>
<td>1.02 ± 0.18</td>
</tr>
<tr>
<td>Mean ± SD</td>
<td>1.0 ± 0.2</td>
<td>65 ± 13</td>
<td>3.80 ± 0.53</td>
<td>9.1 ± 1.2</td>
<td>10.2 ± 0.9</td>
<td>11.7 ± 1.1</td>
<td>1.21 ± 0.21</td>
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<tr>
<td>Before HD</td>
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<td></td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>W. S.</td>
<td>11.5</td>
<td>76</td>
<td>5.02</td>
<td>9.2</td>
<td>5.9</td>
<td>18.5</td>
<td>1.10 ± 0.10</td>
</tr>
<tr>
<td>W. A.</td>
<td>6.5</td>
<td>44</td>
<td>3.03</td>
<td>10.4</td>
<td>7.9</td>
<td>12.8</td>
<td>1.01 ± 0.10</td>
</tr>
<tr>
<td>D. M.</td>
<td>11.0</td>
<td>80</td>
<td>4.23</td>
<td>9.9</td>
<td>6.2</td>
<td>16.7</td>
<td>1.03 ± 0.10</td>
</tr>
<tr>
<td>M. A. S.</td>
<td>10.8</td>
<td>61</td>
<td>3.66</td>
<td>8.5</td>
<td>8.0</td>
<td>15.2</td>
<td>1.21 ± 0.10</td>
</tr>
<tr>
<td>Mean ± SD</td>
<td>10.0 ± 2.3</td>
<td>65 ± 16</td>
<td>3.99 ± 0.85</td>
<td>9.5 ± 0.8</td>
<td>7.0 ± 1.1</td>
<td>15.8 ± 2.4</td>
<td>1.09 ± 0.09</td>
</tr>
<tr>
<td></td>
<td></td>
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<td></td>
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<td></td>
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<tr>
<td>During HD</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C. L.</td>
<td>10.6</td>
<td>75</td>
<td>2.84</td>
<td>6.1</td>
<td>7.8</td>
<td>12.8</td>
<td>1.00 ± 0.61</td>
</tr>
<tr>
<td>S. R.</td>
<td>9.6</td>
<td>60</td>
<td>2.31</td>
<td>6.2</td>
<td>11.2</td>
<td>15.2</td>
<td>1.69 ± 1.05</td>
</tr>
<tr>
<td>W. C.</td>
<td>6.5</td>
<td>75</td>
<td>3.79</td>
<td>12.7</td>
<td>11.0</td>
<td>9.8</td>
<td>1.08 ± 137</td>
</tr>
<tr>
<td>J. S.</td>
<td>15.6</td>
<td>79</td>
<td>6.44</td>
<td>7.7</td>
<td>7.3</td>
<td>25.0</td>
<td>1.82 ± 140</td>
</tr>
<tr>
<td>E. J.</td>
<td>13.2</td>
<td>79</td>
<td>3.01</td>
<td>12.2</td>
<td>8.3</td>
<td>12.2</td>
<td>1.01 ± 123</td>
</tr>
<tr>
<td>W. B.</td>
<td>11.0</td>
<td>82</td>
<td>3.43</td>
<td>7.2</td>
<td>12.6</td>
<td>13.2</td>
<td>1.66 ± 119</td>
</tr>
<tr>
<td>R. J.</td>
<td>17.4</td>
<td>71</td>
<td>6.40</td>
<td>8.2</td>
<td>9.8</td>
<td>20.8</td>
<td>2.05 ± 168</td>
</tr>
<tr>
<td>M. T.</td>
<td>13.8</td>
<td>76</td>
<td>4.25</td>
<td>12.7</td>
<td>8.6</td>
<td>11.2</td>
<td>0.96 ± 122</td>
</tr>
<tr>
<td>Mean ± SD</td>
<td>12.2 ± 3.5</td>
<td>75 ± 7</td>
<td>4.06 ± 1.57</td>
<td>9.1 ± 2.9</td>
<td>9.6 ± 1.9</td>
<td>15.0 ± 5.2</td>
<td>1.41 ± 0.44</td>
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<td></td>
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<td></td>
</tr>
</tbody>
</table>

* Individual values from all four groups were analyzed together by the method of analysis of variance and reported as F ratio, degree of freedom, and P values.  
† The difference between the means of each patient group and the controls (P) were analyzed by using the mean square within value.
After normal, RT, the MCR, D, and percent of metabolized T₄ converted to T₃ became normal. The MCR also tended to be lower, but the difference was significant only in the HD group. All subjects in the two uremic groups had D and percent T₄ to T₃ conversion at least 2 SD below the normal means, and the latter was reduced to about a third of normal. After RT, the MCR, D, and percent of metabolized T₄ converted to T₃ became normal.

To determine the thyroidal contribution to T₃ metabolism by direct secretion of T₃, data from the MCR of T₄ and T₃ during L-T₄ replacement and the serum concentration of TT₄ and TT₃ before L-T₄ replacement were used as described in the section on Methods and the results are summarized in Table IV. The mean thyroidal secretion of T₃ ranged from 1.7 to 3.7 μg/day and was not significantly different in the various groups. However, due to decreased peripheral conversion of T₄ to T₃ in uremia, the relative contribution of direct thyroidal T₃ secretion to total T₃ turnover was greater, being 33.5±10.5% in the non-HD and 16.6±10.5% in the HD patients. The former was significantly higher than that of normal controls (6.9 ±4.4%).

Table IV also lists D for T₄ and T₃ in the basal state (without L-T₄ replacement). T₄ metabolism was not different from the controls whereas T₃ D was markedly decreased in both non-HD and HD patients.

**DISCUSSION**

These studies demonstrate that disturbances of thyroid function, the most important of which is a marked reduction in serum TT₃ concentration, are common in chronic renal failure. Decreased serum TT₃ was previously reported by us (13) and by Silverberg et al. (14) and was subsequently confirmed by several other investigators (15, 16, 33, 34). Circulating TT₃ was reduced into the hypothyroid range (<78 ng/100 ml) in 80% of the non-HD and 43% of the HD patients. While L-T₄ administration effectively raised serum TT₃ to levels often above the normal range, TT₃ remained subnormal. This observation, coupled with the high TT₄/TT₃ ratio in the serum, suggested that low TT₃ concentration in uremia may represent a selective deficiency. In our patient population, low TT₃ could
### TABLE IV

**Turnover Rate of T₄ and T₃ and Thyroidal Secretion of T₃ before L-T₄ Replacement**

<table>
<thead>
<tr>
<th>Group and patient</th>
<th>T₄ metabolism</th>
<th>T₃ metabolism</th>
<th>T₃ secreted by thyroid</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>TT, D</td>
<td>TT, D</td>
<td>% of D (T₃)</td>
</tr>
<tr>
<td></td>
<td>µg/100 ml</td>
<td>µg/day</td>
<td>ng/100 ml</td>
</tr>
<tr>
<td>Normal</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>D. B.</td>
<td>6.0</td>
<td>88</td>
<td>136</td>
</tr>
<tr>
<td>T. C.</td>
<td>6.7</td>
<td>66</td>
<td>146</td>
</tr>
<tr>
<td>F. K.</td>
<td>5.6</td>
<td>77</td>
<td>142</td>
</tr>
<tr>
<td>W. S. T.</td>
<td>6.5</td>
<td>87</td>
<td>130</td>
</tr>
<tr>
<td>E. S.</td>
<td>8.0</td>
<td>82</td>
<td>145</td>
</tr>
<tr>
<td>Mean±SD</td>
<td>6.6±0.9</td>
<td>80±9</td>
<td>140±7</td>
</tr>
<tr>
<td>Before HD</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>W. S.</td>
<td>5.4</td>
<td>59</td>
<td>72</td>
</tr>
<tr>
<td>W. A.</td>
<td>4.3</td>
<td>43</td>
<td>55</td>
</tr>
<tr>
<td>D. M.</td>
<td>5.2</td>
<td>53</td>
<td>88</td>
</tr>
<tr>
<td>M. A. S.</td>
<td>3.4</td>
<td>41</td>
<td>58</td>
</tr>
<tr>
<td>Mean±SD</td>
<td>4.6±0.9</td>
<td>49±9</td>
<td>68±15</td>
</tr>
<tr>
<td>P1</td>
<td>NS</td>
<td>NS</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>During HD</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>C. L.</td>
<td>4.6</td>
<td>46</td>
<td>70</td>
</tr>
<tr>
<td>S. R.</td>
<td>3.5</td>
<td>49</td>
<td>64</td>
</tr>
<tr>
<td>W. C.</td>
<td>7.6</td>
<td>82</td>
<td>102</td>
</tr>
<tr>
<td>J. S.</td>
<td>6.0</td>
<td>109</td>
<td>61</td>
</tr>
<tr>
<td>E. J.</td>
<td>7.5</td>
<td>76</td>
<td>98</td>
</tr>
<tr>
<td>W. B.</td>
<td>5.4</td>
<td>90</td>
<td>95</td>
</tr>
<tr>
<td>R. J.</td>
<td>6.0</td>
<td>123</td>
<td>74</td>
</tr>
<tr>
<td>M. T.</td>
<td>6.4</td>
<td>61</td>
<td>100</td>
</tr>
<tr>
<td>Mean±SD</td>
<td>5.9±1.4</td>
<td>81±26</td>
<td>83±17</td>
</tr>
<tr>
<td>P1</td>
<td>NS</td>
<td>NS</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>After RT</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>J. R.</td>
<td>4.1</td>
<td>67</td>
<td>120</td>
</tr>
<tr>
<td>R. T.</td>
<td>8.1</td>
<td>89</td>
<td>112</td>
</tr>
<tr>
<td>M. S.</td>
<td>7.0</td>
<td>88</td>
<td>145</td>
</tr>
<tr>
<td>S. W.</td>
<td>7.2</td>
<td>68</td>
<td>148</td>
</tr>
<tr>
<td>N. G.</td>
<td>8.0</td>
<td>64</td>
<td>125</td>
</tr>
<tr>
<td>Mean±SD</td>
<td>6.9±1.6</td>
<td>75±12</td>
<td>130±16</td>
</tr>
<tr>
<td>P1</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
</tr>
</tbody>
</table>

* The turnover rates (D) of T₄ and T₃ were calculated from the respective MCR determined during L-T₄ replacement and the TT₄ and TT₃ concentrations measured before L-T₄ treatment. The amount of T₃ secreted by the thyroid gland was derived from turnover rates of T₄ and T₃ and the percent of T₄ converted to T₃. Individual values from all four groups were analyzed by the analysis of variance and summarized as F ratio, degree of freedom, and P values.

† The significance of the difference between the means of each patient group and the controls (P) was derived by using the mean square within value.
theoretically result from removal of circulating hormone through HD, decreased T\textsubscript{3} binding capacity, increased hormone catabolism, reduced thyroidal T\textsubscript{3} secretion, and lastly, diminished extrathyroidal T\textsubscript{3} to T\textsubscript{3} conversion. The results of our studies provide evidence for the latter mechanism.

Peritoneal dialysis has been shown by Hermann et al. to be effective in removing T\textsubscript{4} in patients with thyrrotoxic storm (8) and in rats made thyrrotoxic by hormone administration (9). There are no data showing a decrease in serum TT\textsubscript{4} after conventional HD, using either cuprophane or cellulose membrane. Both Carter's report (15) and our study of the uremic patients before and 2 mo after institution of HD failed to show significant changes in the serum TT\textsubscript{4} and FTI (Table II). On the contrary, serum-free T\textsubscript{4} was reported to be higher after HD, an effect attributed to heparin (7, 35), which is known to alter hormone binding and increase the concentration of free T\textsubscript{4} (6).

Although we have not measured circulating free T\textsubscript{3}, Carter et al. (15), in a report of T\textsubscript{3} deficiency syndrome associated with a variety of nonthyroidal diseases, including renal failure, found that free T\textsubscript{3} and TT\textsubscript{3} were similarly decreased. Chopra et al. (16), in a study of the low T\textsubscript{3} syndrome with liver and kidney diseases, also reported that free T\textsubscript{3} was significantly reduced. Bermudez et al. (36) described 24 patients with decreased serum T\textsubscript{3} concentrations without thyroidal disease; 8 were due to low T\textsubscript{3}-binding capacity, 5 were attributable to their advanced age, and the remaining 11 were unexplained. Spector et al. reported that 54% of renal failure patients had low serum-free T\textsubscript{3} concentrations (33). If our finding of low TT\textsubscript{3} concentration was due entirely to decreased binding capacity, it should have affected TT\textsubscript{4} to a greater extent. Furthermore, changes in TBG capacity were small and statistically not significant. The fact that TT\textsubscript{4}/TT\textsubscript{3} ratio was markedly elevated in our patients suggests that factors other than low binding capacity must be responsible for the low T\textsubscript{3} deficiency.

Our kinetic studies showed that T\textsubscript{3} degradation was not accelerated. In fact, the mean MCR was slightly decreased in both groups of uremic patients. As for the mean T\textsubscript{3} secretion by the thyroid gland, it ranged only from 1.7 to 3.7 μg/day in the four groups of subjects—representing only 7–14% of the total daily turnover in normal subjects. Hence, the observed depression in serum TT\textsubscript{3} concentration cannot be explained on the basis of reduction, or even cessation, of thyroidal T\textsubscript{3} secretion. Actually, the mean percentage of thyroidal T\textsubscript{3} contribution to the overall amount of T\textsubscript{3} metabolized was higher in patients with renal failure because of the marked reduction in their total T\textsubscript{3} turnover rate. The T\textsubscript{3} degradation was drastically reduced to about 40% of control value. The mean percent of metabolized T\textsubscript{4} converted to T\textsubscript{3} was decreased to approximately one-third of normal. Since under our experimental conditions it could be assumed for practical purposes that the only source of T\textsubscript{3} was from extrathyroidal monodeiodination of T\textsubscript{4}, these results indicate that uremic patients have a profound impairment in the ability to generate T\textsubscript{3} from T\textsubscript{4}. This conclusion is in agreement with the observation that their serum TT\textsubscript{3} remained subnormal when serum TT\textsubscript{4} was raised after L-T\textsubscript{4} replacement.

Surks and co-workers have shown that a portion of radioiodine-labeled T\textsubscript{3} administered to animals or humans is present in serum and tissue in a TCA-precipitable but ethanol nonextractable form. It has turnover kinetics different from T\textsubscript{3} and a more prolonged half-life of 14 days (37, 38). If not accounted for, especially in samples obtained 2 or more days after administration of the tracer, it may lead to falsely high 131\textsubscript{I}-T\textsubscript{3} concentrations and consequently underestimation of MCR for T\textsubscript{3}. A comment is therefore in order to validate our T\textsubscript{3} kinetic study, which did not take into account the nonextractable form of iodine. We believe that our observations are valid for several reasons: (a) In the plotting and calculation of MCR, we used only samples obtained up to 48 h when nonextractable iodine represents a relatively small fraction of serum TCA-precipitable radioactivity. (b) The MCR of T\textsubscript{3} (18.6 liters/day) in our control subjects was comparable to that reported by Nomura et al. (39), Surks et al. (19), and Inada et al. (40). (c) The magnitude of suppression of T\textsubscript{3} turnover rate was so striking that even with a 10% correction for possible underestimation of MCR, the daily degradation would still be subnormal. Analogous to our results, in cirrhotic patients studied by Nomura et al. (39), MCR was only minimally decreased, but the T\textsubscript{3} disposal rate was markedly reduced due to the low serum TT\textsubscript{3} concentration.

Our studies do not indicate whether the defect in peripheral T\textsubscript{3} generation is due to the reduction of functionally normal tissue, to the metabolic derangement of uremia, or to the nonspecific effect of chronic illness. Experimental evidence of T\textsubscript{3} production from monodeiodination of T\textsubscript{4} has been obtained in various tissues (25, 41, 42), including the kidney (43). In our patients with renal failure, it does not appear that the conversion defect can be attributed to the lack of renal tissue, as serum TT\textsubscript{3} concentration was not reduced further in anephric patients. A preponderant decrement in serum TT\textsubscript{3} has been shown to occur in a variety of nonrelated chronic illnesses including liver cirrhosis, chronic obstructive pulmonary disease, disseminated malignancy, and malnutrition (15, 16, 36, 44). Diminished peripheral conversion of T\textsubscript{4} to T\textsubscript{3} is responsible for the decreased serum TT\textsubscript{3} concentration in patients with liver failure (39) and starvation (45) and may be the common denominator in the selective T\textsubscript{3} deficiency syndrome.
In addition to the low serum TT₃, uremia seems to be associated with other abnormalities of thyroid function, including slightly decreased TT₄, high incidence of goiter, and abnormal TSH response to TRH. We found that serum TT₄ concentrations, although within the normal range, tended to be lower than in controls, in agreement with the findings of other investigators (5, 14, 33, 46). Neuhaus et al. (46) attributed the low TT₄ to decreased serum albumin and prealbumin. Joasso et al. (5) found that uremic patients had low serum TT₄ and elevated T₃ resin uptake, suggesting a decrease in TBG. However, actual measurement of TBG was normal. They postulated that uremic toxin(s) might have displaced T₄ from TBG (5). In the 46 patients that we studied, only one had serum albumin lower than 3.5 g/100 ml, and TBG capacity was not significantly different from normal. Of those patients whose TBG capacity was decreased, their TT₄ was always low, but low TT₄ was not necessarily accompanied by a reduction in TBG capacity, suggesting that factors other than decreased binding might, in part, be responsible for the slightly decreased serum TT₄ concentration observed in some patients. Despite a marked reduction in the amount of T₄ converted to T₃ in patients with renal failure, the mean T₄/D was not decreased, indicating that other metabolic pathways of degradation were not inhibited. The possibility that monodeiodination occurred preferentially in the inner benzene ring, resulting in the formation of reverse T₃, should also be considered, as Chopra et al. (16) found that the serum reverse T₃ concentration was elevated in patients with renal failure.

A 37% incidence of goiter is considerably higher than expected for the Chicago area. Increased frequency of goiter in uremia was first described by Ramirez et al. (11) and was confirmed by Zerefos et al. (47) in an autopsy series. Thyroid enlargement could result from increased TSH stimulation related to excessive iodine accumulation, or the presence of goitrogenic substances. In a preliminary communication, we reported slightly elevated TSH levels in patients with renal failure (13), but we found subsequently that our anti-human TSH serum had considerable cross-reactivity with luteinizing hormone. Since we modified our assay procedures by prior absorption with human chorionic gonadotropins, the TSH values of our uremic patients had been consistently normal. Normal TSH values in renal failure were also reported by Silverberg et al. (14) and by Ramirez et al. (11).

Renal failure patients tend to retain iodide as shown by their high plasma inorganic iodide level (1, 2, 14). Our finding of increased iodine content in the thyroid gland suggests that there might be a causal relationship between the high iodide stores and the increased incidence of goiter in uremia, particularly in the Chicago area, where the average dietary iodine intake is 420 µg/day.² Ramirez et al. (11) proposed that retention of unknown goitrogens might be the explanation, and more recently Grantham et al. reported that aryl acid was markedly elevated in uremic patients and suggested that these aromatic compounds might be potentially goitrogenic (34, 48).

Although low T₃ syndrome is now well recognized in uremia, its significance remains unclear. Assuming that T₃ is responsible for about one-half to two-thirds of the metabolic effects of the thyroid hormones (49), it is surprising that T₃ deficiency of the magnitude observed in our patients does not produce clinical hypothyroidism. In fact, studies designed to evaluate the metabolic effects of thyroid hormones in uremia have not revealed any abnormal results (33). Nevertheless, one should be cautious in interpreting some of these findings, since uremia per se or the presence of an arteriovenous fistula may alter some of the commonly used clinical tests of thyroid function such as basal metabolic rate and QK₄₄ interval measurements. The absence of TSH elevation is generally regarded as evidence against hypothyroidism, yet hypothalamic-pituitary dysfunction may also be present, as suggested by the subnormal TSH response to TRH. The delayed peak response and sustained elevation might be related to the decreased renal clearance of either TSH (50) or TRH (51). Blunted TSH secretion after TRH administration was also reported by Ramirez et al. (52), Alvarez-Ude et al. (53), and Czernichow et al. (54). Furthermore, abnormal prolactin (55) and growth hormone (56) secretion, as well as inadequate gonadotropin compensation (57), have been demonstrated in patients with renal insufficiency.

We have not fully assessed the functional integrity of the thyroid gland. The T₃ increment of 20 ng/100 ml or 28% after TRH administration was lower than that reported by Chopra et al. (58) in normal individuals, who at the end of 2 h had a mean T₃ increment of 63 ng/100 ml serum, representing a 70% rise over baseline. The smaller rise in serum TT₃ may not represent an intrinsic thyroid abnormality as their TSH response to TRH was subnormal. Ramirez et al. (52), however, reported that serum TT₄ and TT₃ increments after exogenous TSH administration were less than in controls, suggesting intrinsic thyroidal dysfunction.

After RT, serum thyroid hormone concentrations became normal. The rise in serum TT₄ was in part due to higher TBG capacity while the increase in serum TT₃ was due mostly to improved peripheral T₄ to T₃ conversion. The two patients whose T₃ turnover and T₄ to T₃ conversion rates were the lowest also had the

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poorest renal function, their serum creatinine being
>2 mg/100 ml. With restoration of renal function,
serum creatinine fell precipitously, but serum TT₄ rose
very slowly over a period of 3–4 mo. Before this,
however, serum TT₄ transiently decreased to below the
pretransplant value, a possible consequence of the
high-dose steroid treatment. Glucocorticoids are known
to affect thyroid function by inhibition of extrathyroidal
T₄ to T₃ conversion (59), decreased basal TSH secretion
and TSH response to TRH stimulation (60, 61), and possibly suppression of TRH as well (62). When the
dose of prednisone was tapered, circulating thyroid hormones gradually increased, but TSH response to
TRH remained subnormal.

In summary, these studies indicate that uremia affects
thyroid function at multiple levels: (a) There is a
blunted TSH response to TRH, suggesting pituitary
dysfunction or hypersensitivity to hormonal feedback.
(b) There may be an intrathyroidal defect in hormono-
genesis, hormonal secretion, or both, as evidenced by
high incidence of goiter, increased thyroidal iodine
content, and, in some patients, low serum TT₄, not
accountable for by depressed TBG capacity. The sub-
normal TT₄ response to exogenous TSH administration
reported by Ramirez et al. (52) lends support to this
possibility. (c) Abnormal peripheral metabolism is
characterized by a profound impairment of T₄ to T₃
conversion in extrathyroidal tissues, which results in
a selective and marked reduction in serum TT₄ con-
centration. The metabolic significance of these ab-
normalities remains unclear and deserves further
study.

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