Thyroid Function in a Uremic Rat Model

EVIDENCE SUGGESTING TISSUE HYPOTHYROIDISM

VICTORIA S. LIM, CARLOS HENRIQUEZ, HISAO SEO, SAMUEL REFETOFF, and
ENIO MARTINO, Renal Division, Department of Medicine, Michael Reese Hospital
and Medical Center, and Thyroid Study Unit, Department of Medicine, University
of Chicago, Chicago, Illinois 60616

ABSTRACT The main objective of this study was to determine whether the principal abnormality of thyroid function observed in patients with chronic renal failure, low serum triiodothyronine (T3) concentration, causes hypothyroidism at the tissue level. A partially nephrectomized (Nx) uremic rat model was developed and the following parameters of thyroid function were assessed: serum total thyroxine (TT4), total T3 (TT3), and thyrotropin and liver T3 content, and activity of two thyroid hormone-dependent enzymes, mitochondrial a-glycerophosphate dehydrogenase (aGPD) and cytosol malate dehydrogenase (MDH). The results were compared to those of intact control (C), thyroidectomized (Tx), and nephrectomized-thyroidectomized (NxTx) littermates.

Results expressed as mean±SEM showed that Nx rats had a fivefold increase in blood urea nitrogen, (112±20 mg/dl in Nx, and 22±1 ng/dl in C) and manifested all the changes of thyroid function observed in uremic men, including a low serum TT3 level (30±7 ng/dl in Nx and 50±6 ng/dl in C). In the liver, T3 was significantly reduced (18±2 ng/total liver in Nx and 35±3 ng/total liver in C) as well as the activities of aGPD (8.8±1.0 and 16.1±1.5 AOD/min per total liver in Nx and C, respectively) and MDH (6.3±1.6 and 12.6±2.2 U/total liver in Nx and C, respectively). The reduction in liver enzyme activities correlated significantly with the decrease in T3 content.

The changes in Tx rats were as expected, showing a profound reduction in serum hormone levels, liver T3 content, and liver enzyme activities. Serum thyrotropin was markedly elevated to 2,390±212 ng/ml as compared to 703±61 in C and 441±87 ng/ml in Nx rats.

The NxTx rats showed the combined effects of nephrectomy and thyroidectomy; blood urea nitrogen was elevated to 203, and serum levels of TT4, TT3, and thyrotropin were 0.4, <10, and 2,525, respectively. Total liver T3 and aGPD and MDH were strikingly low; the corresponding values were 3.5, 2.4, and 2.5.

L-triiodothyronine replacement (0.4 μg/100 g body wt/d) for 4 wk in the Nx rats resulted in significant increases in liver enzyme activities, aGPD and MDH rose by 70 and 60% over their respective basal values without alteration in the severity of azotemia.

From these data, we conclude that the reduction of liver T3 content in the uremic rats, accompanied by a decrease in aGPD and MDH activity, indicates the presence of hypothyroidism at the tissue level. Restoration of enzyme activities toward normal levels after T3 administration provided further supporting evidence that the diminution in liver enzyme activity was causally related to tissue T3 deficiency.

INTRODUCTION

Patients with chronic renal failure have low serum levels of total triiodothyronine (TT3) due, in part, to a defect in the peripheral conversion of thyroxine (T4) to triiodothyronine (T3) (1–11). However, the physiologic significance of the low serum T3 concentration in a variety of conditions (4–6), including uremia, has not been clearly defined. Thus, the question of whether uremic patients are hypothyroid remains unanswered. Investigators who propose that uremic patients are euthyroid base their conclusion on the observation that serum thyrotropin (TSH) is not elevated and thyroid

1 Abbreviations used in this paper: BUN, blood urea nitrogen; C, control; FTI4, free thyroxine index; GPD, glycerophosphate dehydrogenase; MDH, malate dehydrogenase; Nx, nephrectomized; NxTx, nephrectomized-thyroidectomized; T3, triiodothyronine; T4, thyroxine; TSH, thyrotropin; TT4, total T4; TT3, total T3; Tx, thyroidectomized.
function tests, independent of hormone measurements such as the basal metabolic rate, the systolic time interval, and the deep tendon reflex relaxation time, are normal (7). The argument for hypothyroidism is purely theoretical and is based on the concept that T₃ is the hormone through which the major metabolic effects of T₄ are exerted (12, 13). The absence of TSH elevation should not be regarded as unequivocal evidence of euthyroidism because disturbances in the hypothalamic-pituitary axis are known to occur in uremia (14–17). The controversy regarding thyroid status in uremia is partly because most studies have concentrated on the measurement of thyroid hormone in serum. There is no information available on the content of T₃ in tissues. This is important because the bulk of extrathyroidal T₃ is generated in the peripheral tissues, at the sites of action (12, 13).

We studied thyroid function in a uremic rat model. T₃ concentrations in serum as well as in liver were measured. In addition, the activities of two liver enzymes regulated by thyroid hormone (18–21), mitochondrial α-glycerophosphate dehydrogenase (αGPD) and cytosol malate dehydrogenase (MDH), were determined. The results showed an early reduction in liver T₃ content accompanied by a significant diminution in both enzyme activities. These data, coupled with the correction of the observed enzyme deficiencies after T₃ treatment, suggest that hypothyroidism at the tissue level accompanies the uremic state.

**METHODS**

**Experimental design**

Four groups of rats, control (C), thyroidectomized (Tx), nephrectomized (Nx), and nephrectomized-thyroidectomized (NxTx), were examined. The Nx rats were studied 4, 5, and 6 wk after nephrectomy. Male Sprague Dawley rats, weighing 120–150 g at the start of each experiment, were obtained from Animal Resource Laboratory, Madison, Wis. Uremic rats were prepared by a two-stage 5/6 nephrectomy. The upper and lower poles of the left kidney were removed by ligation and the raw surfaces were cauterized to prevent bleeding and minimize compensatory hypertrophy. The entire right kidney was removed 5–7 d later. Only 50–60% of the Nx rats were used; 30–40% died within 72 h of the second stage of nephrectomy and another 10% were discarded because blood urea nitrogen (BUN) was insufficiently elevated (<50 mg/dl). Thyroidectomy was performed by a standard surgical technique. Mortality in the Tx group was negligible and hypothyroidism always ensued. In the NxTx rats, thyroidectomy was done 5 wk after nephrectomy. As these rats appeared very ill, studies were performed 3–5 d after thyroidectomy. The mortality in this group was ~75%. About 70% survived the second stage of nephrectomy but only 25% survived the subsequent thyroidectomy. Unless otherwise specified, rats were fed regular chow and given access to tap water ad lib. Tx and NxTx rats were given 0.9% calcium chloride as the drinking water. In addition, NxTx rats received 10 mg of calcium gluconate, by intraperitoneal injection, every other day after thyroidectomy.

In another experiment, only C and 6-wk Nx rats were used. Each group was further divided in half; one-half received no treatment whereas the other received exogenous L-triiodothyronine (Sigma Chemical Co., St. Louis, Mo.) supplement at 0.4 μg/100 g of body wt/d by intraperitoneal injections for 4 consecutive wk starting 2 wk after the second stage of nephrectomy. The dose was considered to be optimal for T₃ replacement based on the studies by Garcia et al. (22) on the effect of T₃ on TSH response to TSH-releasing hormone in Tx rats.

Rats were killed by cardiac exsanguination under light ether anesthesia. The liver was perfused in situ with normal saline solution until light tan. The entire liver was removed and weighed. 1 g was frozen on dry ice immediately and kept at −30°C until extraction was done on another day for measurement of T₃ content. Additionally, 1 gram of liver was processed freshly for quantitation of mitochondrial αGPD activity and another 2 g for determination of MDH activity. The two kidneys of C and Tx rats and the kidney remnants of Nx and NxTx rats were removed and weighed. Serum was saved for measurements of BUN, total thyroxine (TT₄), free thyroxine index (FT₄I), T₃, binding capacity, TT₃, and thyroid stimulating hormone (TSH). In the rats that received exogenous T₃ treatment, additional serum samples were obtained 2 h after the last T₃ injection for determination of TT₃ levels.

As serum thyroid hormone concentrations (23) and αGPD and MDH activities (24) vary with age, littersmates were used for each experiment. Execution of surgical procedures was staggered so that rats in each experimental group were of the same age at the time of study.

**Analytical procedures**

**Determination of T₃ content.** Liver T₃ was extracted using the method of Nejad et al. (25). Briefly, 1 g of frozen liver was homogenized in 2 ml of chilled water containing 50 μM propylthiouracil. 10 μl of the homogenate was saved for protein determination and the remainder was extracted twice with 6 ml of 95% ethanol. An equal volume of chloroform (12 ml) was added to the pooled ethanol extract placed in a separatory funnel. T₃ was extracted from the ethanol-chloroform mixture using 1/3 vol of 2 N NH₄OH three times. The pooled aqueous phase was concentrated in a flash evaporator and subsequently lyophilized. The residue was reconstituted in 0.5 M phosphate buffer containing 1% bovine serum albumin, pH 7.5, and was stored at −20°C until analyzed for T₃ content.

Recovery was monitored in every batch of T₃ extraction by addition of sufficient [¹³¹I]T₃ to a second 1 g piece of liver before extraction and corrections for recovery were carried out for each sample.

**Assessment of enzyme activity.** Mitochondrial αGPD (EC 1.1.1.5) activity was measured in the mitochondrial fraction prepared from 1 g of freshly perfused liver. Homogenization was done in 9 vol of 0.25 M sucrose, 50 mM Tris-HCl, pH 7.5. After centrifugation at 1,000 g, 4°C for 10 min, the supernate was again centrifuged at 12,000 g, 4°C for 10 min, and the resultant pellet containing mitochondria was resuspended in 1 ml of 0.125 M phosphate buffer, pH 7.5. αGPD activity was analyzed in triplicate (three sets of blanks and samples) according to the method of Lee and Lardy (19). Reactions were carried out for 15 min. The difference in the mean optical density in the assay tubes and in the blanks read at 500 nm (ΔOD) and the protein concentration, measured in an aliquot of the mitochondrial preparation, was used for the calculation of the enzyme activity expressed as ΔOD per minute per total liver and per mg of protein.

Cytosol MDH (EC 1.1.1.40) activity was measured in 2 g of fresh liver. Homogenization was done in 3 vol of chilled 0.25 M sucrose. The homogenate was centrifuged at 9,000 g 4°C for 10 min, and the supernate was recentrifuged at 100,000 g, transplanted to the rat liver. The liver was perfused in situ with normal saline solution until light tan. The entire liver was removed and weighed. 1 g was frozen on dry ice immediately and kept at −30°C until extraction was done on another day for measurement of T₃ content. Additionally, 1 gram of liver was processed freshly for quantitation of mitochondrial αGPD activity and another 2 g for determination of MDH activity. The two kidneys of C and Tx rats and the kidney remnants of Nx and NxTx rats were removed and weighed. Serum was saved for measurements of BUN, total thyroxine (TT₄), free thyroxine index (FT₄I), T₃, binding capacity, TT₃, and thyroid stimulating hormone (TSH). In the rats that received exogenous T₃ treatment, additional serum samples were obtained 2 h after the last T₃ injection for determination of TT₃ levels.

As serum thyroid hormone concentrations (23) and αGPD and MDH activities (24) vary with age, littersmates were used for each experiment. Execution of surgical procedures was staggered so that rats in each experimental group were of the same age at the time of study.

**Analytical procedures**

**Determination of T₃ content.** Liver T₃ was extracted using the method of Nejad et al. (25). Briefly, 1 g of frozen liver was homogenized in 2 ml of chilled water containing 50 μM propylthiouracil. 10 μl of the homogenate was saved for protein determination and the remainder was extracted twice with 6 ml of 95% ethanol. An equal volume of chloroform (12 ml) was added to the pooled ethanol extract placed in a separatory funnel. T₃ was extracted from the ethanol-chloroform mixture using 1/3 vol of 2 N NH₄OH three times. The pooled aqueous phase was concentrated in a flash evaporator and subsequently lyophilized. The residue was reconstituted in 0.5 M phosphate buffer containing 1% bovine serum albumin, pH 7.5, and was stored at −20°C until analyzed for T₃ content.

Recovery was monitored in every batch of T₃ extraction by addition of sufficient [¹³¹I]T₃ to a second 1 g piece of liver before extraction and corrections for recovery were carried out for each sample.

**Assessment of enzyme activity.** Mitochondrial αGPD (EC 1.1.1.5) activity was measured in the mitochondrial fraction prepared from 1 g of freshly perfused liver. Homogenization was done in 9 vol of 0.25 M sucrose, 50 mM Tris-HCl, pH 7.5. After centrifugation at 1,000 g, 4°C for 10 min, the supernate was again centrifuged at 12,000 g, 4°C for 10 min, and the resultant pellet containing mitochondria was resuspended in 1 ml of 0.125 M phosphate buffer, pH 7.5. αGPD activity was analyzed in triplicate (three sets of blanks and samples) according to the method of Lee and Lardy (19). Reactions were carried out for 15 min. The difference in the mean optical density in the assay tubes and in the blanks read at 500 nm (ΔOD) and the protein concentration, measured in an aliquot of the mitochondrial preparation, was used for the calculation of the enzyme activity expressed as ΔOD per minute per total liver and per mg of protein.

Cytosol MDH (EC 1.1.1.40) activity was measured in 2 g of fresh liver. Homogenization was done in 3 vol of chilled 0.25 M sucrose. The homogenate was centrifuged at 9,000 g 4°C for 10 min, and the supernate was recentrifuged at 100,000 g,
4°C for 60 min. This supernate, containing the cytosol fraction, was passed through a 0.22 μMillipore filter (Millipore Corp., Bedford, Mass.) and diluted 5- or 10-fold with a solution containing 10 mM EDTA, 50 mM Tris HCl, 200 mM magnesium acetate, and 2 mM 2-mercaptoethanol, pH 7.4. MDH activity was then measured using the method of Ochoa (26) as modified by Hsu and Lardy (27), by monitoring the reduction of NADP to NADPH, producing changes in the optical density at 340 nm. Appropriate blanks, without added substrate, and a standard preparation of purified chicken liver enzyme (Sigma Chemical Co., St. Louis, Mo.) were used in each assay. Enzyme activity was expressed as units of MDH per milligram of cytosol protein and per total liver.

Other measurements. Serum TT₄ was measured by a competitive binding assay and the results of a resin T₄ uptake test were used to calculate FT₄I as described (28).

Serum and tissue T₃ were measured by a radioimmunoassay (29) using a specific antiserum, a gift from Dr. G. Burke (Cook County Hospital, Chicago, Ill.). Determinations in serum were carried out in the presence of 8-anilino-1-naphthalene sulfonic acid and those in tissues were done in the absence of this substance. Corresponding standard curves contained hypothyroid rat serum or buffer only. The sensitivity of the assay was 10 ng/dl and the intra- and interassay coefficients of variation were 7 and 15%, respectively.

Serum T₄ binding capacity was measured by saturation (30) and expressed as percent of control.

Serum TSH was measured by a radioimmunoassay (31) using materials from two kits (Nos. 3 and 4) obtained from National Institute of Arthritis Metabolism and Digestive Diseases. The assay sensitivity was 100 ng/ml. The intra- and interassay coefficients of variation were 3.1 and 16.4%, respectively.

The BUN was determined by the standard microcolorimetric technique with diacetyl monoxime and ferric nitrate reagents.

Protein concentration was measured using the method of Lowry (32).

Statistical analyses, including Student's t test, analysis of variance, and linear regression analysis, were done by using the Biomedical Computer Program package (Health Science Computer Facility, University of California, Los Angeles, Calif.). Group data are presented as mean±SEM.

RESULTS

Kidney and thyroid function of all groups of rats are compared in Table I. After nephrectomy, mean BUN in the Nx rats increased fivefold and the mean weight

<table>
<thead>
<tr>
<th>Table I</th>
<th>Thyroid and Renal Function in C, Tx, Ns, and NxTx Rats</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body weight</td>
<td>Kidney weight</td>
</tr>
<tr>
<td>g</td>
<td>mg/dl</td>
</tr>
<tr>
<td>C (11)</td>
<td>373</td>
</tr>
<tr>
<td>±12</td>
<td>±0.1</td>
</tr>
<tr>
<td>Tx (8)</td>
<td>349</td>
</tr>
<tr>
<td>±8</td>
<td>±0.1</td>
</tr>
<tr>
<td>P</td>
<td>NS</td>
</tr>
<tr>
<td>Nx4 (14)</td>
<td>283</td>
</tr>
<tr>
<td>±13</td>
<td>±0.1</td>
</tr>
<tr>
<td>P</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Nx5 (12)</td>
<td>276</td>
</tr>
<tr>
<td>±13</td>
<td>±0.1</td>
</tr>
<tr>
<td>P</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Nx6 (11)</td>
<td>256</td>
</tr>
<tr>
<td>±25</td>
<td>±0.1</td>
</tr>
<tr>
<td>P</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>NxTx (6)</td>
<td>173</td>
</tr>
<tr>
<td>±8</td>
<td>±0.1</td>
</tr>
<tr>
<td>P</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>P</td>
<td>&lt;0.0001</td>
</tr>
</tbody>
</table>

All values are given as mean±SEM. (Nx), 4, 5, and 6 indicate weeks postnephrectomy. The number of rats per group is in parenthesis. The significance of the difference between the means of each experimental group and the control was assessed by the Student's t test.

* Individual values from all six groups were tested by the analysis of variance; DF, the degree of freedom, F ratio, and P values are listed.
of the kidney remnant was <40% of the combined kidney weights of the control littermates. Mean body weight was also decreased by ~30%. The postnephrectomy hypertrophy of the kidney appeared to have been completed during the first 4 wk after the second stage operation as there was no difference in either the mean kidney weights or the mean BUN levels among the 4-, 5-, and 6-wk Nx rats. Thyroid function also differed significantly in the Nx rats. Mean serum TT₄ and FT₄I were decreased by ~30% 4 wk postnephrectomy and remained stable during the subsequent 2 wk of observation. Serum T₃ binding capacity was not affected by nephrectomy, being 98±6% in the 6-wk Nx rats as compared to 100±4% in the C group. The mean serum TT₃ showed a declining trend during the 4th and 5th wk, but the level became significantly reduced to 30 ng/dl at 6 wk postnephrectomy. Mean serum TSH concentrations were significantly lower in all three Nx rat groups. The Tx rats showed changes typical of severe hypothyroidism: mean serum TT₄ and FT₄I were reduced to <10%, mean serum TT₃ to about 35%, and mean serum TSH increased by 300% of control littermates. The slight but significantly elevated BUN in the Tx rats is consistent with the known effect of thyroid hormone in augmenting glomerular filtration rate (33). The NxTx rats showed the combined effects of nephrectomy and thyroidectomy. Mean serum TT₄ and FT₄I were reduced to the same extent as the Tx littermates, but the reduction in serum TT₃ to <10 ng/dl was more severe than could be accounted for by thyroidectomy alone. Mean serum TSH was elevated to a comparable degree as in the Tx rats. When compared to Nx rats, NxTx rats had a mean BUN level twofold higher, and the kidney remnant weighed an average of 20% less. These rats generally appeared very ill and their mean body weight was 40% less than the nonthyroidectomized uremic rats.

T₃ content and the activity of the two thyroid-hormone dependent enzymes in the liver are summarized in Table II. Nephrectomy resulted in a significant reduction in liver T₃ content as well as the activities of the thyroid hormone-dependent enzymes. Mean total

### Table II

Liver T₃ Content, Mitochondrial αGPD, and Cytosol MDH Activities in C, Tx, Nx, and NxTx Rats

<table>
<thead>
<tr>
<th></th>
<th>T₃</th>
<th>αGPD</th>
<th>MDH</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>ng/total liver</td>
<td>ng/g</td>
<td>pg/mg protein</td>
</tr>
<tr>
<td>C (11)</td>
<td>35.4</td>
<td>2.47</td>
<td>19</td>
</tr>
<tr>
<td></td>
<td>±3.1</td>
<td>±0.21</td>
<td>±1</td>
</tr>
<tr>
<td>Tx (8)</td>
<td>5.3</td>
<td>0.45</td>
<td>4</td>
</tr>
<tr>
<td></td>
<td>±0.9</td>
<td>±0.05</td>
<td>±0.4</td>
</tr>
<tr>
<td>P</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Nx4 (14)</td>
<td>28.8</td>
<td>2.32</td>
<td>20</td>
</tr>
<tr>
<td></td>
<td>±3.1</td>
<td>±0.20</td>
<td>±0.2</td>
</tr>
<tr>
<td>P</td>
<td>NS</td>
<td>NS</td>
<td>NS</td>
</tr>
<tr>
<td>Nx5 (12)</td>
<td>20.5</td>
<td>1.62</td>
<td>13</td>
</tr>
<tr>
<td></td>
<td>±3.5</td>
<td>±0.19</td>
<td>±2</td>
</tr>
<tr>
<td>P</td>
<td>&lt;0.01</td>
<td>&lt;0.01</td>
<td>&lt;0.01</td>
</tr>
<tr>
<td>Nx6 (11)</td>
<td>18.5</td>
<td>1.45</td>
<td>12</td>
</tr>
<tr>
<td></td>
<td>±2.4</td>
<td>±0.14</td>
<td>±1</td>
</tr>
<tr>
<td>P</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>NxTx (6)</td>
<td>3.5</td>
<td>0.44</td>
<td>5</td>
</tr>
<tr>
<td></td>
<td>±0.3</td>
<td>±0.04</td>
<td>±0.4</td>
</tr>
<tr>
<td>P</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>P</td>
<td>&lt;0.0001</td>
<td>&lt;0.0001</td>
<td>&lt;0.0001</td>
</tr>
</tbody>
</table>

All values are represented as means±SEM. (Nx)4, 5, and 6 refer to weeks postnephrectomy. The number of rats per group is in parenthesis. The significance of the difference between the means of each experimental group and the control was assessed by the Student t test.

* Individual values from all six groups were tested by the analysis of variance; DF, the degree of freedom, F ratio, and P values are summarized.

Tissue Hypothyroidism in a Uremic Rat Model
liver T₃ content and total liver mitochondrial αGPD activity were reduced on the fourth week; the decrease became statistically significant on the 5th wk and further declined on the 6th wk postnephrectomy. Results are similar when expressed in terms of total tissue or mitochondrial protein concentration. Cytosol MDH activity was examined only in the 6-wk Nx rats; both total liver MDH and MDH/milligrams of cytosol protein were significantly reduced. The changes after Tx were as expected: liver T₃ and αGPD and MDH activities were all markedly reduced. In the NxTx rats, mean liver weight was ~50% that of control rats so that total liver T₃ content and total liver enzyme activities were further decreased to levels lower than either Tx or Nx alone. As the protein concentration in the liver and the subcellular fractions were also greatly reduced, T₃ content and enzyme activities in this group of rats when corrected for protein concentration were reduced to the same level as in the Tx littermates.

It appeared that the reduction in αGPD and MDH activities in the Nx rats correlated best with the reduction in liver T₃ content (Figs. 1 and 2). Significant positive correlations were found when the Nx rats were examined independently of the C and Tx rats. There was no significant difference when the two regression lines were compared. Control rats having higher T₃ content and greater enzyme activities were grouped in the upper section whereas the Tx rats with the least amount of T₃ and the lowest enzyme activities were tightly packed in the left lower corner. The Nx rats having intermediary values were scattered in between.

The effects of T₃ treatment in the Nx rats are depicted in Figs. 3 and 4. Both C and 6-wk Nx rats were either not treated or given 0.4 µg T₃/100 g of body wt by daily intraperitoneal injections for 4 consecutive wk starting 2 wk after nephrectomy. In both groups of rats, mean serum TT₃ concentration increased to 185 and 180 ng/dl, respectively, 2 h after Tx injection and serum TSH was suppressed to levels <100 ng/ml. Mean concentrations of BUN were not changed by Tx treatment. At the tissue level, Tx treatment did not significantly alter the mean T₃ content, αGPD, or MDH activities of the C rats. The only change was an increase in the specific activity of αGPD. In the Nx rats, changes were more apparent. Total liver αGPD and MDH rose by 70 and 60% over their respective base-line values. These increases became more significant when corrected for protein concentration. In fact, the reduced enzyme activities of Nx rats were normalized to values comparable to untreated controls. These changes in enzyme activities were in agreement with the corresponding normalization of the liver T₃ content, which rose from 12 to 17 pg/mg protein. Other important information not shown in the figures is that of body weights and serum TT₃. Despite improvement in thyroid function, Tx treatment had no beneficial effect on body weight, which was 340±8 g in C and 251±13 g in Nx rats (P < 0.001). Mean serum TT₃ was decreased in both groups of Tx treated rats, being 2.5±0.2 µg/dl in the C and 1.5±0.2 µg/dl in the Nx rats (P < 0.001).

**Figure 1** Correlation between total liver T₃ and total liver mitochondrial αGPD. Two linear regression lines are presented. The solid line was derived from C (●) and Tx (▲) rats (n = 19, r = 0.896, and P < 0.01). The dotted line was constructed from 5- and 6-wk Nx (○) rats (n = 29, r = 0.666, and P < 0.01). There is no statistical difference between the two lines.

**Figure 2** Total liver cytosol MDH was plotted as a function of total liver T₃ showing excellent correlations between these two parameters. The solid line (n = 13, r = 0.688, and P < 0.01) was derived from C (●) and Tx (▲) rats, whereas the dotted line (n = 6, r = 0.946, and P < 0.01) was based on the data of 6-wk Nx (○) rats. The two regression lines are not different statistically.
associated with chronic renal failure, low serum T₃ concentration (1, 4–9), by searching for evidence of hypothyroidism at the tissue level. We attempted to answer the question by quantitation of T₃ content in the liver and by measurement of the activities of two thyroid hormone-dependent hepatic enzymes. A uremic rat model produced by a two-stage 5/6 nephrectomy was developed. After an initial 30% loss of body weight, a stable and reproducible form of chronic azotemia was achieved with a five- to sixfold elevation of BUN during the entire period of observation. More importantly, the uremic rat exhibited changes of thyroid function typical of man with chronic renal failure (1–11) and thus constituted a suitable experimental model. There was a slight decrease in serum TT₄, FT₄Iₐ, and a marked diminution in serum TT₃ concentration; serum TSH level was normal or slightly reduced (Table I). In the NxTx rats, serum TT₄ and FT₄Iₐ were decreased to the same extent as in the Tx rats, but serum TT₃ was reduced to a greater degree. This observation indicated that the low serum TT₃ in NxTx rats is not only due to lack of T₄ but also to an impairment in T₄ to T₃ conversion caused by uremia.

An important finding in this study was a reduction in liver T₃ content in the Nx rats. The decrease was most marked when expressed as T₃ content per total liver, an average of 50% of the control values 6 wk after nephrectomy (Table II). The reduction in liver T₃ was also significant when expressed as nanograms per milligram protein. It is unlikely that the decrease in liver T₃ is due to a lack of T₄, as serum TT₄ was only slightly...
reduced. Moreover, while the reduction in serum TT₄ occurred early and remained stable, a significant decrease in liver T₃ was not noted until 5 wk after nephrectomy with further diminution at 6 wk. Using isotopically labeled T₄ and T₃ we have previously shown that uremic patients have impaired ability in T₄ to T₃ conversion (9), a mechanism most likely responsible for the low liver T₃ content in the present experimental model. Reduction in tissue T₃ content is not unique to uremia; starvation and partial hepatectomy in the rat have been shown to produce a decrease in nuclear T₃ binding capacity and T₃ content (34, 35). In humans, Reichlin et al. (36) found decreased T₃ concentration in liver and kidney obtained at necropsy from patients who died from a variety of chronic illnesses.

T₃ concentration in the liver was several times higher than that in the serum and reduction in liver T₃ content preceded the appearance of significant diminution in serum T₃ concentration. Whereas the finding of low tissue T₃ content is not necessarily indicative of hypothyroidism, the concomitant reduction of mitochondrial αGPD and cytosol MDH in the Nx rats suggested the presence of metabolic consequences of T₃ deficiency. As shown in Fig. 1, changes in αGPD paralleled those of T₃ content. A similar correlation was found between the MDH activity and liver T₃ content in the 6-wk Nx rats (Fig. 2). The linear regressions correlating liver enzyme activities and T₃ content were not different in azotemic rats when compared to Tx and C rats with intact renal function.

It is unlikely that starvation was responsible for the reduction in enzyme activities in the uremic rat as enzymes were restored to normal despite failure of these animals to achieve a higher body weight during the 4-wk period of T₃ replacement. The possibility that metabolic consequences directly related to uremia, rather than reduced T₃ supply, were responsible for the observed enzymatic changes, was considered. In fact, although thyroid hormone replacement in the Tx rats is known to correct the activity of these two liver enzymes (20), it has been shown that the apparently T₃-deficient severely starved and partially hepatectomized rats are resistant to the induction of liver MDH by thyroid hormone (35). Relative resistance of the starving rat to a single dose of T₃ by augmentation in resting oxygen consumption has also been demonstrated (37). Nevertheless, chronic administration of physiologic doses of T₃ prevented the diminution of αGPD and MDH activities seen in the uremic rat. Such an observation underscores the causal relationship between tissue T₃ and enzyme induction in this animal model, especially when the increase in enzyme activities was associated with a repletion of T₃ content (Fig. 4). The slightly higher liver T₃ content in the C rats given T₃ is either due to a more efficient endogenous generation of T₃ from the residual nonsuppressible T₄ in these rats or, alternatively, to abnormal catabolism of T₃ in the Nx rats. It is important to note that although T₃ treatment prevented the decline in liver enzyme activity in the uremic rat, it had no effect on the magnitude of weight loss or the degree of uremia associated with nephrectomy.

Although these data clearly indicate that the reduction of liver αGPD and MDH activities in uremia is causally related to tissue T₃ deficiency, they are in apparent contradiction with results reported in the starving and partially hepatectomized rats, models bearing similarities with the uremic rat. The resistance to T₃ in the starving and partially hepatectomized rats was manifested by the failure to observe an increase in MDH but not αGPD activity after the administration of a single pharmacologic dose of T₃ (35). Unlike uremic and hypothyroid rats (20), basal enzyme activity in these acute experimental models was not decreased (35). No resistance to T₃ was observed in fasting human subjects with low serum TT₃ when the hormone was given chronically and in physiologic doses (38).

The lack of TSH elevation in the presence of tissue hypothyroidism and low serum levels of TT₃ and TT₄ in the uremic rats is puzzling. It certainly could not be attributable to low TSH reserve in the pituitary as serum TSH in the NxTx rats reached levels as high as in the nonuremic Tx rats (Table I). It is possible that intrapituitary T₃ content is normal in the uremic rats as monodeiodination of T₃ tends to be more efficient in the pituitary (39, 40). In such an instance, hypothyroidism would only be confined to the peripheral tissue, including the liver. Alternatively, there may be concomitant disturbances in the hypothalamic-pituitary axis with regard to TSH secretion associated with uremia. Impaired TSH response to TSH-releasing hormone is well documented in patients with chronic renal failure (8, 9). A resetting of the feedback regulation of TSH secretion at a lower level, proposed as the mechanism responsible for the failure to observe serum TSH elevation in response to starvation-induced decrease in T₃ generation in man (38), may also be operational in the uremic rat model.

In summary, the partially nephrectomized rat was an excellent experimental model to examine thyroid function in renal failure because uremia was chronic and stable and the changes in thyroid hormone profile in the circulation were surprisingly similar to that found in patients with chronic renal failure. In this model, T₃ content was markedly reduced and the αGPD and MDH activities were also significantly decreased. These data were interpreted as evidence consistent with hypothyroidism at the tissue level. Restoration of enzyme activity after T₃ treatment lent further support to this hypothesis. The physiologic significance of these findings with regard to total body economy and energy metabolism remains to be elucidated.
ACKNOWLEDGMENTS

We appreciate the advice and encouragement of Dr. Leslie DeGroot and Dr. Fredric L. Cee. We thank Mr. Sven R. Hagen for his technical assistance and Ms. Amy Strauss for her help in statistical analysis.

The work was supported by U. S. Public Health Service grants HD 10918 and AM 15070 and by the Michael Reese Research Institute.

REFERENCES


