Regional Distribution and Alterations of Lectin Binding to Colorectal Mucin in Mucosal Biopsies from Controls and Subjects with Inflammatory Bowel Disease

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Abstract

Glycoconjugate composition of colorectal goblet cell mucin was characterized according to the anatomical distribution of lectin-binding sites in mucosal biopsies from 35 control subjects and 55 patients with inflammatory bowel disease. 24 of the controls had mucosal inflammation on biopsy, without clinical evidence of inflammatory bowel disease. These inflamed controls showed a similar rate of presence of lectin-binding sites as the normal noninflamed group. In the controls, the frequency of binding of Ricinus communis agglutinin I to galactosyl residues was consistently higher than that found with either Ulex europaeus agglutinin I to fucosyl or Dolichus biflorus agglutinin to N-acetyl galactosyl groups. A significant proximal to distal gradient of Ulex europaeus agglutinin I binding sites was identified in the control group. These binding sites were present four times more often in the proximal colon than in the distal colon ($P < 0.025$). In the ulcerative and Crohn's colitis groups, this gradient effect was lost, predominantly as a result of decreased availability of fucosyl residues in the proximal colon. In the descending colon of Crohn's colitis tissues, there was a complete absence of Dolichus biflorus agglutinin binding sites compared with the 62.5% incidence in the control group ($P < 0.05$). These results demonstrate that the expression of lectin-binding sites in human large intestinal goblet mucin is specifically altered in inflammatory bowel disease, indicating that there are changes in glycosylation of colorectal mucin consistent with alterations in goblet cell differentiation.

Introduction

Characterization of the biochemical abnormalities in the colonic mucosa of patients with inflammatory bowel disease has been attempted using a number of different techniques. Histochemical studies of colorectal mucin have demonstrated a decrease in goblet cell mucin in ulcerative colitis, but no specific qualitative difference between ulcerative colitis, Crohn's colitis, and normal patients (1, 2). Although increased sialomucin staining occurs when dysplastic changes are present (3), this does not seem to be clinically significant, since nonspecific increases in sialomucins, sulphomucins, and neutral muco-

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1. Abbreviations used in this paper: DBA, Dolichus biflorus agglutinin; FITC, fluorescein isothiocyanate; Fus, $\alpha$-l-fucose; Gal, $\beta$-galactose; GalNAc, $N$-acetyl galactosamine; PAS, periodic acid Schiff; RCA1, Ricinus communis agglutinin I; UEA1, Ulex europaeus agglutinin I.
lectin-binding sites, biopsies from all regions of the large intestine were examined. Tissue samples from a control group of subjects, comprising inflamed mucosal biopsies due to conditions other than inflammatory bowel and normal non-inflamed samples, were compared with the results found in mucosal biopsies from patients diagnosed as having either ulcerative or Crohn's colitis.

Methods

Study population. The clinical and histological material in this study was selected retrospectively. The majority of patients had been seen by the gastroenterology service at the University of California, Davis Medical Center, during the preceding 10-yr period. Additional clinical material was provided by other members of the faculty who had personally managed a number of patients with inflammatory bowel disease over several years. The clinical data are summarized in Table I. Subjects composing the control group had been investigated for large bowel disease, and had been found not to have inflammatory bowel disease. Patients with colonic carcinoma and/or polyposis were excluded, since previous studies indicate that intestinal mucin glycoconjugates may be altered in these conditions (19, 20). During these investigations, large bowel mucosal biopsies had been performed. According to prior histopathological evaluation, 11 of the 35 control biopsies were morphologically normal, while in the remaining 24 patients there was histological evidence of inflammation. 20 patients in the control group had evidence of mucosal inflammation by proctosigmoidoscopy. However, it was determined by clinical followup, as well as by endoscopic, radiological, and histological criteria, that none of these patients had ulcerative or Crohn's colitis. Among the clinical disorders constituting the control group were four patients with infective and noninfective forms of diarrhea, three with radiation damage, three with ischemia, seven with functional bowel disease, two with colonic pseudo-obstruction, three with colonic diverticular disease, four with infectious proctocolitis, and one with antibiotic-associated colitis. Eight more patients were included with self-limited colitis and/or proctitis, of brief duration and unknown etiology, which did not recur.

Patients were diagnosed as having inflammatory bowel disease according to a typical clinical history, abnormal barium studies, and colonoscopy or proctosigmoidoscopy with biopsies, showing an appearance consistent with colitis. More specifically, in those patients with ulcerative colitis, there was absence of fistulization and stricturing. This contrasted with those patients with Crohn's disease of the colon, in whom the presence of strictures, fistulae, segmental involvement, and the presence of granulomas were all documented (Table I).

Histological staining of large bowel mucosal biopsies with fluorescein isothiocyanate (FITC)-conjugated lectins. Large intestinal biopsies were categorized according to their anatomical sites of origin; namely, as either ascending, transverse, descending, or sigmoid colon, and rectum. Biopsies had been previously fixed in formaldehyde and imbedded in paraffin. The retrospectively selected paraffin blocks were then used to prepare sections 4-5 μm thick. Deparaffinization and lectin staining were carried out as previously described (15). In brief, sections were first deparaffinized in xylene before sequential passage through 95, 80, and 70% ethanol, after which slides were rinsed in 0.01 M phosphate-buffered saline, pH 7.4. Then, under a dim light, the slides were carefully blotted before treatment with 25 μl (1 mg/ml) of the FITC-conjugated lectin. A solution of the same lectin concentration containing 0.15 M sugar hapten to inhibit lectin-binding was applied directly to the surface of adjacent serial tissue sections. After 30 min in a dark, humid atmosphere, the slides were rinsed with 0.01 M phosphate-buffered saline, pH 7.4, carefully blotted, and then covered with phosphate-buffered glycerol before applying a cover slip. The pattern of fluorescence was then examined with a fluorescence photomicroscope (II; Carl Zeiss, Inc., Thornwood, NY) and photographed for further documentation. Slides were coded so that the observer (P.W.H.) did not know their identity. A number of randomly selected biopsies were reexamined with FITC-lectins at the end of the study to control for intraobserver variation and to confirm the reproducibility of earlier observations. Morphology was assessed on hematoxylin and eosin stained sections, and the presence of goblet mucin was confirmed with periodic acid Schiff (PAS) as shown in Fig. 1 A. Only those areas containing goblet cells with mucin present were examined. Results were recorded according to the intensity of fluorescence in individual goblet cells. Biopsies were considered to show positive lectin-binding if more than half the total crypts per field demonstrated lectin staining, with a minimum of 10 labeled goblet cells per crypt. Thus, if fluorescence was only present in a patchy distribution, this was classified as 0. The intensity of individual goblet cell fluorescence was graded using a semiquantitative scale with 0 representing no detectable labeling; 1+, trace present; 2+, weak intermediate; 3+, strong intermediate; 4+, intense (16). Examples of fluorescein-conjugated lectin-binding to intestinal biopsies are shown in Fig. 1.

Many patients had a series of biopsies taken from multiple sites throughout the large intestine. Some subjects had several biopsies available from the same anatomical region, either taken at the same time or alternatively over a period of several years. Although all available biopsies were subjected to lectin-binding studies, only one biopsy per anatomic region per patient was included in the final subject analysis. The selection of a representative sample was easily determined since the absence or presence of lectin-binding sites was remarkably consistent between multiple biopsies obtained from the same region in the same patient. This was in contrast to the grade of fluorescence intensity, which showed some variation over time, possibly related to disease activity. However, the available clinical data and the limited number of suitable samples did not permit a valid correlation analysis to be undertaken between disease activity and intensity of FITC-lectin fluorescence. The majority of biopsies were obtained in the preceding four years. There was no evidence of a loss of lectin-binding activity as a function of age of the biopsy. The oldest biopsy was 9 yr old and revealed a pattern of lectin-binding similar to that seen with the other biopsies from the same clinical group, some of which were less than 1 yr old. Areas of ulceration and those with loss of mucosal architecture were not included in the final data analysis, since such changes result in a loss of goblet cell number and disruption of normal crypt structure.

Lectins. Plant lectins, purified by affinity chromatography and conjugated with FITC, were obtained from E-Y Laboratories, San

Table I. Characteristics of Patient Population

<table>
<thead>
<tr>
<th></th>
<th>Controls</th>
<th>Ulcerative colitis</th>
<th>Crohn's colitis</th>
</tr>
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<tbody>
<tr>
<td>No. of subjects</td>
<td>35</td>
<td>28</td>
<td>27</td>
</tr>
<tr>
<td>Male/female</td>
<td>22:13</td>
<td>14:14</td>
<td>12:15</td>
</tr>
<tr>
<td>Age (yr)</td>
<td></td>
<td></td>
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<tr>
<td>Mean</td>
<td>39.1</td>
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<tr>
<td>Range</td>
<td>17–94</td>
<td>17–71</td>
<td>21–68</td>
</tr>
<tr>
<td>Disease duration (yr)</td>
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<td></td>
<td></td>
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<tr>
<td>Mean</td>
<td></td>
<td>10.1</td>
<td>8.0</td>
</tr>
<tr>
<td>Range</td>
<td></td>
<td>0.25–30</td>
<td>0.08–21</td>
</tr>
<tr>
<td>Site of intestinal inflammation</td>
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<tr>
<td>Ileum/colon</td>
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<td>0</td>
<td>10</td>
</tr>
<tr>
<td>Total colon</td>
<td>3</td>
<td>12</td>
<td>2</td>
</tr>
<tr>
<td>Distal colon/rectum</td>
<td>17</td>
<td>16</td>
<td>15</td>
</tr>
<tr>
<td>Granulomas</td>
<td>0</td>
<td>0</td>
<td>11</td>
</tr>
<tr>
<td>Strictures/fistulæ</td>
<td>0</td>
<td>0</td>
<td>5/5</td>
</tr>
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Figure 1. Human colonic mucosal biopsies. (A) Biopsy from a patient with ulcerative colitis, stained with PAS to show the presence of goblet cell mucin (see arrow); × 240. (B) Mucosal biopsy from the colon of a subject with ischemic colitis, stained with FITC-conjugated DBA to show goblet cell mucin (see arrow); × 240. FITC-lectins were incubated with histological sections and examined by fluorescence microscopy as described in Methods. (C) Biopsy from a patient with ulcerative colitis, stained with FITC-DBA to show the presence of binding sites in goblet cell mucin (see arrow); × 240. (D) Mucosal biopsy from the colon of a subject with Crohn's colitis, stained with FITC-DBA, which did not bind to goblet cell mucin (see arrow). The presence of mucin was confirmed by PAS staining and binding of RCA; × 240.

Mateo, CA. The lectins and their sugar specificities were as follows: UEA₁, specifically binds with α-L-fucose (Fuc); RCA₁, with D-galactose (Gal); and DBA, with N-acetyl galactosamine (GalNAc). The inhibitor carbohydrates Fuc, Gal, and GalNAc (Sigma Chemical Co., St. Louis, MO) were all used at a 0.15-M concentration.

Statistical analyses. The results were analyzed using χ² and Fisher's Exact Tests (21).

Results

Although the intensity of fluorescence observed was graded for each slide, such a subjective semiquantitative assessment might not be reproducible by other observers. Therefore, the data were analyzed only to determine the rate of presence of lectin-binding sites. This was achieved by classifying biopsies either as being completely devoid of lectin-binding sites (grade 0), or as exhibiting the presence of lectin-binding sites (grades 1+ or greater). The control group of biopsies was divided into inflamed and noninflamed tissues, according to previous pathological interpretation. The results were expressed as either the number of patients or the number of mucosal biopsies with positive lectin-binding. No statistical differences in the rate of presence of lectin-binding sites between inflamed control patients and those control patients with noninflamed tissues could be demonstrated (results not shown). Because of this, these two subsets were pooled together to form a single control group for further comparison with the tissues from patients with inflammatory bowel disease. Fig. 2 A shows the percent of control subjects with lectin-positive biopsies, according to the site of tissue sampling, for the three different lectins used.
In the three most proximal regions of the colon, the frequency of RCA1 binding was 100%, dropping to 81.3% in the sigmoid colon, and only 12.5% in the rectum. In contrast to the regional gradient of lectin-binding sites demonstrated for RCA1 and UEA1 (maximal binding in the proximal colon and minimum binding in the rectosigmoid), the pattern for DBA binding showed no evidence of such a gradient effect. In the proximal colon, the frequency of subjects with positive DBA staining was <20%, while maximal binding was observed in the descending colon with 62.5% of subjects showing positive lectin staining, and less than a 25% positive rate in the rectosigmoid. The frequency of DBA staining was significantly lower than that found with RCA1 in all regions except the descending colon (P < 0.025 or better), while the frequency of UEA1 staining was lower than that with RCA1 only in the descending colon, sigmoid, and rectum (P < 0.005).

A higher frequency of RCA1 binding to colorectal mucin was also observed when ulcerative (Fig. 2 B) and Crohn’s colitis (Fig. 2 C) samples were studied. In ulcerative colitis (Fig. 2 B), the frequency of RCA1 binding was significantly higher than that of UEA1 or DBA in all five regions of the intestine examined (P < 0.05 or better). In Crohn’s colitis (Fig. 2 C), the frequency of RCA1 binding to mucin was significantly higher than that of UEA1 in all regions except the descending colon (P < 0.025 or better), and significantly greater than with DBA1 in all regions except the transverse colon (P < 0.05 or better). In both forms of colitis (Figs. 2 B and C), the existence of a proximal-to-distal gradient in lectin-binding was much less evident than that found for the control group (Fig. 2 A).

In all three sets of patients studied (Fig. 2), lectin-binding frequencies in the ascending and transverse regions of the proximal colon were not significantly different. Similarly, the two most distal segments (sigmoid and rectum) revealed comparable patterns and levels of lectin-binding. For these reasons, biopsies from the two most proximal and two most distal segments were pooled and designated as constituting the proximal and distal colon, respectively. Using this anatomical definition, the results for UEA1-binding to control biopsies were further analyzed, as shown in Fig. 3 A. This confirmed the impression gained from Fig. 2 A that there was a significant gradient in the frequency of lectin-binding between the proximal and distal colon. In the proximal colon, 58.3% of the biopsies were positive, compared with 14.3% in the distal colon (P < 0.025). Although there was also evidence of a proximal-to-distal gradient in ulcerative and Crohn’s colitis (Fig. 3 A), this was much less marked than in the control patients, and was not statistically significant. A similar analysis for the results of RCA1 (Fig. 3 B) and DBA (Fig. 3 C) binding demonstrated that lectin-binding in all groups was greater in the proximal than in the distal colon. However, none of these differences was statistically significant.

Because biopsies from the area of the descending colon showed lectin-binding characteristics that were sometimes dissimilar from those observed in the proximal or distal colon (Fig. 2), these data were analyzed separately (Fig. 4). The results showed that with UEA1, 50% of the control subjects demonstrated positive lectin-binding, compared with 44.4% in the ulcerative colitis, and 20% in the Crohn’s colitis groups. With RCA1, 100% of the control and ulcerative colitis subjects showed positive lectin-binding, compared with 80% of the Crohn’s patients. With DBA, 62.5% of the control group and 40% of the ulcerative colitis group showed positive binding,
compared with the Crohn’s colitis group, in which there was a complete absence of any lectin-binding (Figs. 1 D and 4) \( P < 0.05 \). The specificity of this alteration in the Crohn’s colitis biopsies was further strengthened by the fact that 80% of these same tissues demonstrated binding sites for \( \text{RCA}_1 \), and 100% of the biopsies showed a normal-appearing amount of mucin by PAS staining.

**Discussion**

These results provide evidence that the expression of lectin-binding sites in human large intestinal goblet cell mucin is altered in patients with inflammatory bowel disease. These changes seem to be disease specific, since analysis of the rate of presence of lectin-binding sites in the control population showed no effect that could be attributed to inflammatory changes alone. Although all lectins tended to show a higher frequency rate of binding to proximal than to distal colorectal mucin, this was only statistically significant for binding to fucosyl residues (\( \text{UEA}_1 \)) in the control group. The loss of this gradient effect in inflammatory bowel disease seemed to result from a decreased availability of fucosyl residues in the proximal colon of the ulcerative and Crohn’s colitis groups (Fig. 3 A).

Inflammatory bowel disease samples showed additional evidence of a change in colonic mucin differentiation, as measured by the alteration in the relative frequency of lectin-binding sites expressed in the ascending and transverse colon where fucosyl residues (\( \text{UEA}_1 \) binding) were significantly reduced relative to galactosyl groups (\( \text{RCA}_1 \) binding). This contrasted with proximal colonic samples from the control groups, in which no significant differences between lectin-binding to galactosyl and fucosyl residues could be shown. Ulcerative colitis samples showed uniform changes in every anatomic region from the ascending colon through to the rectum, with higher rates of available galactosyl residues compared with either fucosyl or \( \text{N-acetyl galactosyl} \) groups. The complete absence of \( \text{DBA} \) binding to \( \text{N-acetyl D-galactosamine} \) residues in mucin from a small number of descending colon biopsies from Crohn’s colitis patients seems to deserve further investigation. Confirming this result in a prospective study, using a larger number of patients, will help determine the value of \( \text{DBA} \) binding as a sensitive marker for Crohn’s colitis. However, the present results indicate that the absence of \( \text{DBA} \) binding was not very specific, since 37.5% of controls and 60% of ulcerative colitis patients also demonstrated absence of \( \text{DBA} \) binding in the descending colon. Unpublished observations from this laboratory on several patients with Crohn’s Disease, limited to the small bowel, indicate that \( \text{GalNAc} \) residues continue to be expressed in the descending colon. Because of the retrospective design of this study, it was not possible to reliably correlate disease activity with lectin binding. However, based on these

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**Figure 3.** Rate of presence of (A) \( \text{UEA}_1 \) binding sites, (B) \( \text{RCA}_1 \) binding sites, and (C) \( \text{DBA} \) binding sites in proximal (solid bars) and distal (open bars) colorectal mucin from control subjects and patients with inflammatory bowel disease. Lectin binding and its analysis was performed as described in Fig. 2 and Methods. \( \text{**P} < 0.025 \), proximal compared with distal.

**Figure 4.** Rate of presence of FITC-lectin binding to descending colon mucin from controls and patients with inflammatory bowel disease. Binding of \( \text{UEA}_1 \), \( \text{RCA}_1 \), and \( \text{DBA} \) to histological sections was carried out as described in Fig. 2 and Methods. \( \text{**P} < 0.05 \), for the Crohn’s colitis compared with control group. \( \text{■} \), controls; \( \text{□} \), ulcerative colitis; \( \text{●} \), Crohn’s colitis.
The effects of mucosal inflammation on colorectal mucin lectin-binding sites has not to our knowledge previously been investigated. Using cytochemical techniques, Ehsanullah et al. (3) studied mucin chemistry in nonspecific proctitis, and as with the present study, found no change from normal patients, in contrast to the abnormal patterns seen in Crohn's Disease and ulcerative colitis. Yonezawa et al. (17, 19) examined the nonneoplastic mucosa of patients with colorectal tumors and found that with right-sided tumors, all patients have demonstrable UEA-I-binding sites in the right colon, whereas with left-sided tumors, the distal nonneoplastic mucosa contained binding sites in only 10.3% of cases, while in patients with rectal carcinoma, the nonneoplastic mucosa was completely devoid of UEA-I-binding sites. In comparison, our own results for UEA I binding show far less specificity. This is probably due to differences in clinical material, patients with any history or evidence of colorectal neoplasia having been excluded from our control group, whereas the control biopsies in the study of Yonezawa et al. were taken from the same subjects with colorectal cancer. Similarly, the present results cannot be validly compared with those of Boland et al. (16), who obtained their normal tissues predominantly from patients with colon cancer. These authors (16) did not analyze their results according to anatomic region, nor did they estimate the rate of presence of lectin-binding sites. Control groups of this sort (16, 17, 19) can no longer be regarded as reliably representing normal mucosa, since it has now been well established that even the histologically normal mucosa of human large bowel bearing carcinomas frequently shows abnormal mucin composition (20, 22, 23). It has further been suggested that such changes in mucin glycoprotein synthesis antedate malignant changes and may serve as markers for premalignant epithelium (20, 22, 23). The variations in mucin composition that have been reported in ulcerative colitis associated with carcinoma (24, 25) and dysplasia (3) are not present when the disease is in remission, and are not found in areas of reparative hyperplasia (24, 25). Note that none of the ulcerative colitis patients in the present study had any history or evidence of colorectal cancer.

It is generally accepted that there is a sequential scheme for goblet cell mucin glycosylation, in which carbohydrate residues such as Fuc, Gal, and GalNAc may be added terminally (26). However, expression of these terminal carbohydrate residues may be further modified by the enteric microflora, which have been shown to extensively degrade mucin carbohydrate moieties (27). Thus, the final availability of lectin-binding sites in human colorectal mucin seems to be determined not only by biosynthetic factors, but also by those that produce degradation. As a result of the more rapid rate of cell turnover in inflammatory bowel disease (6, 7), less time may be available to allow full differentiation of mucin, thus resulting in a relative reduction in certain terminal carbohydrate residues such as fucose. This might also explain the decreased rate of presence of UEA-I binding sites in the proximal colon of Crohn's and ulcerative colitis, resulting in a loss of the normal proximal-to-distal gradient of mucin fucosyl expression.

The results of the present study add further weight to the growing body of evidence that indicates that there are specific biochemical alterations in the colorectal mucin of patients with Crohn's and ulcerative colitis. Whether these changes are the basis or simply just another manifestation of the disease process still remains to be determined.

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