Glucagon Receptor of Human Liver

Studies of its Molecular Weight and Binding Properties, and its Ability to Activate Hepatic Adenylyl Cyclase of Non-obese and Obese Subjects

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Abstract

The glucagon receptor and the adenylyl cyclase system of human liver membranes were studied in six non-obese and six obese subjects who had elevated insulin and plasma glucagon levels. Analysis of specific glucagon binding by the method of Scatchard demonstrated a linear (monocomponent) plot with a dissociation constant of 2–3 nM, and the binding at low hormone concentrations was sensitive to guanosine triphosphate (GTP). The molecular weight of the glucagon receptor was 63,000 D as determined by an affinity labeling procedure and sodium dodecyl sulfate gel electrophoresis. Affinity labeling of this structure was specific for glucagon and inhibited by GTP.

Glucagon stimulated the production of cyclic adenosine monophosphate (cAMP) by human membranes with half-maximal activation elicited by 6 nM hormone. The human cyclase system required GTP to facilitate an optimal glucagon response. NaF (10 mM) also activated the cyclase system and produced the same magnitude of response as maximum glucagon activation.

A comparison of the liver adenylyl cyclase system of non-obese and obese subjects was made using glucagon (5 nM and 1 μM) and NaF (10 mM). No significant differences in cAMP production were noted between the two groups, regardless of the agent used to activate the enzyme. These findings agree with the glucagon binding studies that showed similar amounts of binding activity in the membranes from the two groups. Also, there was no influence of either age or sex of the subjects on the adenylyl cyclase response.

In conclusion, human liver membranes contain a glucagon receptor and an adenylyl cyclase system that correspond closely to the well-studied system in animal liver. This system in human obesity is not altered by the twofold elevation in plasma glucagon that occurs in this metabolic disorder.

Introduction

Glucagon is an important contributor to the regulation of hepatic fuel production (1). Although the liver is a primary site of glucagon action (2), and several studies in man have demonstrated significant hepatic extraction of the hormone (3–5), very little direct information is available regarding the interaction of glucagon with the human glucagon receptor, the structure of this receptor, or the characteristics for the hormone activation of the adenylyl cyclase system. Two reports have shown the ability of glucagon to stimulate cyclic adenosine triphosphate (cAMP)$^1$ production by human liver membranes (6, 7), but studies of the glucagon receptor were not described.

Understanding these basic features of glucagon action is important, particularly since in certain diseases like diabetes, glucagon has been implicated in contributing to the overall pathophysiology (8). Obesity is another abnormality in which recent findings suggest an oversecretion of glucagon (9). It is possible that the oversecretion produces changes in glucagon receptor number and/or its ability to activate the adenylyl cyclase system. For example, in animal studies and in vitro experiments, elevations in glucagon reduce the number of glucagon receptors and decrease glucagon action (10, 12). Unfortunately, studies of glucagon action in human obesity have given ambiguous results, as have the studies of animal models for obesity (13).

In the present study, we examined the interaction of glucagon with the glucagon receptor of human liver membranes, the molecular weight of the glucagon receptor, and the ability of the hormone to stimulate adenylyl cyclase. In addition, studies of the effect of this hormone on adenylyl cyclase activity were conducted in liver membranes from obese subjects, to directly evaluate in this disorder an early step in glucagon action.

Methods

Materials. Carrier-free $^{125}$I-Na, [H]cAMP, and [α-$^{32}$P]ATP were purchased from Amersham Corp., Arlington Heights, IL. Guanosine triphosphate (GTP), ATP, glucagon, creatine phosphate, creatine phosphokinase (type I), myokinase (grade III), Dowex-50 (hydrogen form), and alumina (chromatographic, neutral) were supplied by Sigma Chemical Co., St. Louis, MO.

Human subjects. Six non-obese and six obese subjects were studied (see Table I for clinical data). The non-obese subjects were admitted to the Surgical Department (Karolinska Institute, Stockholm, Sweden) for elective cholecystectomy; otherwise, they were healthy. The obese subjects were admitted for gastric operations for obesity. The body weight of all the subjects was constant during the 3-mo period before admittance, and none was receiving any drugs that affect liver or carbohydrate metabolism. The subjects were maintained as inpatients on a weight-maintaining diet (45% carbohydrate, 20% protein, and...

1. Abbreviations used in this paper: cAMP, cyclic adenosine monophosphate; GTP, guanosine triphosphate; M, relative molecular weight.
### Table 1. Clinical Characteristics of the Subjects

<table>
<thead>
<tr>
<th>Patient number</th>
<th>Sex</th>
<th>Age</th>
<th>Height (cm)</th>
<th>Weight (kg)</th>
<th>Glucagon (pg/ml)</th>
<th>Insulin (µU/ml)</th>
<th>Glucose (mM)</th>
<th>Liver biopsy</th>
</tr>
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<tbody>
<tr>
<td>1</td>
<td>M</td>
<td>47</td>
<td>181</td>
<td>78</td>
<td>56.1</td>
<td>6.8</td>
<td>4.5</td>
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</tr>
<tr>
<td>2</td>
<td>F</td>
<td>29</td>
<td>164</td>
<td>59</td>
<td>63.9</td>
<td>9.6</td>
<td>5.0</td>
<td>Normal</td>
</tr>
<tr>
<td>3</td>
<td>F</td>
<td>49</td>
<td>168</td>
<td>60</td>
<td>62.2</td>
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<td>4.0</td>
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</tr>
<tr>
<td>4</td>
<td>M</td>
<td>70</td>
<td>179</td>
<td>79</td>
<td>53.9</td>
<td>10.8</td>
<td>5.2</td>
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</tr>
<tr>
<td>5</td>
<td>F</td>
<td>30</td>
<td>162</td>
<td>61</td>
<td>48.5</td>
<td>10.9</td>
<td>4.8</td>
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</tr>
<tr>
<td>6</td>
<td>F</td>
<td>45</td>
<td>163</td>
<td>59</td>
<td>56.9</td>
<td>8.6</td>
<td>4.7</td>
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<td>45</td>
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<td>66</td>
<td>56.9</td>
<td>8.7</td>
<td>4.7</td>
</tr>
</tbody>
</table>

| SE             | 6   | 3   | 4          | 2.3          | 0.9              | 0.2           |
| 7              | F   | 30  | 161        | 145         | 77.8             | 20.8          | 4.4         | Steatose    |
| 8              | M   | 21  | 190        | 165         | 114.5            | 21.3          | 6.2         | Steatose    |
| 9              | F   | 45  | 162        | 143         | 120.4            | 14.4          | 3.9         | Steatose    |
| 10             | F   | 49  | 161        | 117         | 55.5             | 29.7          | 4.4         | Normal      |
| 11             | F   | 23  | 164        | 117         | 107.5            | 11.3          | 4.2         | Normal      |
| 12             | F   | 33  | 163        | 108         | 76.7             | 18.4          | 5.0         | Normal      |
| Mean           |     |     | 34          | 167         | 133              | 92.1          | 19.3        | 4.7         |
| SE             | 5   | 5   | 9          | 10.5         | 2.6              | 0.3           |

*p* Comparison between obese and non-obese subjects.

35% fat) for 3 d before surgery. After an overnight fast, general anesthesia was induced with a short-acting barbiturate and maintained by phenyural and a nitrous oxide-oxygen mixture. During surgery, a liver biopsy (1 g) was taken from the left lobe of the liver. The biopsy was immediately placed in ice-cold buffer and taken to the laboratory for the isolation of liver membranes.

This study was approved by the Ethical Committee of the Karolinska Institute. Informed consents were obtained from the subjects after they were given a detailed description of the study.

**Preparation of liver membranes.** The liver biopsy was homogenized in 50 ml of a 1-mM NaHCO₃ buffer, pH 7.5, that contained 0.5 mM CaCl₂ (14). The homogenate was subjected to centrifugation at 10,000 g for 20 min, and the membranes were washed in 0.25 M sucrose, 10 mM Tris buffer, pH 7.4. The membranes were resuspended in this buffer, frozen at −80°C, and shipped to Rochester, NY for further analysis. Protein content in the membrane preparation was determined as previously described (15).

Rat liver plasma membranes were prepared by the two-phase polymer method of Lesko et al. (14).

**Glucagon binding studies.** Glucagon was radiolabeled with ¹²⁵I-Na to a specific activity of 0.8-1.0 Ci/μmol by the chloramine-T method (16). Liver membranes (10-20 μg protein) were incubated for 30 min at 21°C in 200 μl of a 20-mM Tris buffer, pH 7.4, that contained 1 mM EDTA, 0.1% g/w/vol bovine serum albumin, and 1 mg/ml bacitracin. ¹²⁵I-glucagon (0.33-33 nM) was added in the presence or absence of 1 μM native glucagon used to estimate nonspecific glucagon binding. The incubations were terminated and the membranes isolated by the centrifugation method of Rodbell et al. (17). Degradation of ¹²⁵I-glucagon was estimated by a trichloroacetic acid precipitation method (18).

**Adenyl cyclase assay.** The conditions of the adenyl cyclase assay were essentially those used by Rojas et al. (19). In brief, liver membranes (10-25 μg protein) were incubated for 10 min at 32°C in 50 μl of the following buffer system: 0.1 mM [α-³²P]ATP (2,000 cpm/pmol), 5 mM MgCl₂, 1 μM GTP, 1 mM EDTA, 0.1% bovine serum albumin, 1 mM [³H]cAMP (10,000 cpm), 20 mM creatine phosphate, 0.2 mg/ml creatine phosphokinase, 0.02 mg/ml myokinase, and 25 mM Tris–HCl, pH 7.5. Termination of the reaction and quantitation of the cAMP produced from ATP were carried out as described in method C of Salomon et al. (20).

**Photoaffinity labeling and sodium dodecyl sulfate (SDS) polyacrylamide gel electrophoresis of the glucagon receptor.** Liver membranes (1.0 mg protein) were incubated with ¹²⁵I-glucagon and other agents, as indicated in Results, in 1 ml of the buffer used in studies of glucagon binding. After a 20-min incubation at 21°C, the membranes were rapidly chilled to 4°C and pelleted by a 3-min centrifugation step in a microfuge (Beckman Instruments, Inc., Fullerton, CA). The membrane pellet was washed twice by resuspension in 1 ml of an ice-cold 20-mM sodium phosphate buffer, pH 7.4, followed by microfuge centrifugation. After the second wash, the pellet was resuspended in the dark in 1 ml of the phosphate buffer that contained 20 μl of a solution of 6.5 mg of N-hydroxysuccinimyl-4-azidobenzoate in 1 ml of dimethyl sulfoxide (21). The membranes were incubated in the dark at 4°C for 10 min. The reaction was terminated by the addition of 20 μl of a 2-M Tris-HCl solution, pH 7.4. The membrane suspension was placed in a small petri dish (35-mm diam) on ice. The dish was positioned 15 cm under a sunlamp (275 W and 110-125 V; General Electric Co.) and exposed to light for 10 min with constant shaking. The membranes were pelleted by microfuge centrifugation and dissolved in 2% SDS, 40 mM Tris–HCl buffer, pH 6.8, that contained 5% 2-mercaptoethanol. After being heated at 100°C for 5 min, the samples were subjected to polyacrylamide gel electrophoresis using a 5-12% gradient resolving gel as described by Laemmli (22). After electrophoresis, the gels were fixed in 10% trichloroacetic acid, stained with Coomassie Blue R-250, destained, and dried. The gels were exposed to preflected film (X-Omatic; Eastman Kodak Co., Rochester, NY) using intensifying screens (X-Omatic; Eastman Kodak Co.). High molecular weight standards (relative molecular weight [Mr] range, 200,000–45,000; Bio-Rad Laboratories, Richmond, CA) were used to estimate the molecular weight of the glucagon receptor.

**In vivo determinations.** Venous blood for measurements of plasma glucose, insulin, and pancreatic glucagon was obtained after an overnight
fast. Plasma insulin levels were measured using a commercial radioim-
munoassay (Pharmacia Fine Chemicals, Div. of Pharmacia, Inc., Piscataway, NJ). Pancreatic glucagon was determined in plasma prepared
with Trasylol (400 IU/ml) by the method of Heiding (23) using
antiserum (E7) supplied by Von Schenck (24). The analyses of glucagon
levels were performed on the same occasion for all the subjects.

Statistical methods. The values given are mean±SE. Student's
unpaired t test was used for statistical comparison.

Results

Human subjects. Table I lists the characteristics of the two
groups of human subjects studied. The weight of the subjects
in the obese group was ~100% over that of the non-obese
group. The fasting insulin and glucagon levels in the obese
subjects were approximately double those found in the normal
group. One obese subject had a slightly enhanced fasting blood
glucose level at the time of the study. However, on two other
occasions immediately before the study, his blood glucose
concentration was normal. All of the remaining subjects had
normal fasting blood glucose levels at the time of the study.

Glucagon binding and the activation of adenylyl cyclase
activity. Initial studies indicated that steady-state binding of

glucagon (0.33 nM) by human liver membranes was reached
by 30 min at 21°C (data not shown). Glucagon degradation
under these conditions was ~2%. In these studies, specific
glucagon binding was 50% of the total amount of binding. In
contrast, specific binding by rat liver membranes was 90% of
the total amount bound.

Fig. 1 shows a Scatchard plot of glucagon binding to liver
membranes from a non-obese subject. As illustrated, the plot
is linear for human membranes and is quite similar to the
linear plot found with glucagon binding to rat liver membranes
(Fig. 1, insert). The membranes from both species have disas-
soociation constant values for glucagon binding of 2–3 nM.
The major difference lies in the amount of glucagon bound
per milligram of protein, with the human preparation having
much less total glucagon binding activity than the rat mem-
branes. This difference may be due in part to the use of a
highly purified preparation of rat liver plasma membranes,
whereas the human preparation is not as enriched in this
fraction. It is also likely that the difference in the ratio of
specific vs. total glucagon binding in the two membrane
preparations arises from the difference in purity.

It was not possible to carry out complete glucagon binding
studies on all of the human membrane preparations because
of the limited amount of material. Instead, the ability of the
membrane fractions from six non-obese and five obese subjects
to specifically bind 3 nM 125I-glucagon was compared. The
mean for glucagon binding by the non-obese preparations was
1.9±0.3 fmol/20 μg of protein versus a value of 1.5±0.3 fmol
for the membranes from the obese subjects. The difference
in these values was not statistically significant.

Initial studies of cAMP production in the absence or
presence of glucagon demonstrated a linear relationship with
respect to the time of incubation (0–20 min) and to the
concentration of membranes in the assay (3–24 μg protein)
(data not shown). In these studies we also examined the effect
of theophylline (3 mM) and found that it did not alter the
rate of cAMP production (data not shown). Thus, phospho-
diesterase did not destroy a fraction of the cAMP formed
during these assays.

The effect of GTP formation on cAMP was studied (Table
II), and as indicated, this nucleotide is required to obtain the
full stimulation of the cyclase system in the human membranes.
In the absence of GTP, glucagon elicited only a 50% stimulation
compared with the stimulation found in the presence of the
nucleotide. Basal cAMP formation was also affected, being
reduced to the lower limit of our ability to accurately measure
cAMP formation. Furthermore, GTP at a concentration of 1
μM was sufficient to allow glucagon to maximally activate the
cyclase system. Higher nucleotide concentrations did not sig-
nificantly change basal or glucagon-stimulated cAMP produc-
tion from the rates found with 1 μM GTP.

GTP also affected glucagon binding by human liver mem-
branes, as reported for membranes from rat liver (19). The
specific binding by human membranes of 125I-glucagon (0.33
nM) was reduced to 48±8% of the control value (100%) by
100 μM GTP. At a high hormone concentration of 33 nM,
glucagon binding was not altered by the nucleotide (data not
shown). Thus, GTP apparently reduces the binding affinity of

Table II. Effects of GTP on Adenylyl Cyclase Activity*

<table>
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<th>GTP μM</th>
<th>− Glucagon</th>
<th>+ Glucagon</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
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<td>0.60</td>
</tr>
<tr>
<td>1</td>
<td>0.21</td>
<td>1.33</td>
</tr>
<tr>
<td>10</td>
<td>0.33</td>
<td>1.36</td>
</tr>
<tr>
<td>100</td>
<td>0.26</td>
<td>1.35</td>
</tr>
</tbody>
</table>

* pmol/10 μg protein per 10 min. Glucagon concentration was 1 μM.

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the glucagon receptor, and enhances glucagon-stimulated activation of human adenyl cyclase activity.

A comparison of the glucagon dose-response relationship between human and rat liver membranes is shown in Fig. 2. Maximum activation of the adenyl cyclase system is produced by 0.33 μM glucagon in both membrane preparations. Half-maximal activation in human membranes was produced by a glucagon concentration of ~6 nM, compared with a concentration of 8 nM for rat liver membranes. Note that the amount of protein and the incubation period used in these experiments differed for the two membrane preparations. When standardized to the same membrane concentration and incubation period, basal cAMP formation by human membranes is approximately one-half that found in the rat preparation, and maximal glucagon-stimulated activity is one-third. Again, these quantitative differences between the two membrane preparations are likely due in part to different degrees of enrichment in plasma membranes. As subsequently demonstrated (see Fig. 3), the response of human membranes to 10 mM NaF is quite similar to that produced by maximal glucagon stimulation. This relationship is well established in highly enriched preparations of rat liver plasma membranes (25), which suggests that the lack of purity of the human preparations does not alter their ability to respond to the hormone.

Fig. 3 compares the ability of membranes from non-obese and obese subjects to respond to glucagon and NaF. Two glucagon concentrations were used; one that produces approximately half-maximal stimulation, and another that maximally activates the cyclase system. NaF was included in the study to provide an independent estimate of cyclase content and a separate parameter other than membrane protein to standardize glucagon stimulation. The results indicate similar basal adenyl cyclase activity between both groups, with no significant differences in cAMP formation with either concentration of glucagon or with NaF stimulation. Thus, the membranes isolated from the obese subjects have a normal response to glucagon, a finding that agrees with normal 125I-glucagon binding.

Studies of the structure of the human glucagon receptor.
Because of the large amount of membranes required by the structural studies, membranes from four normal subjects were pooled for these studies. Fig. 4 shows an autoradiogram of a polyacrylamide gel after the affinity labeling procedure described in Methods was carried out in human and rat liver membranes. Both membranes contain several structures that are labeled with 125I-glucagon by the photoaffinity labeling technique. However, only one structure in each membrane preparation is specifically labeled with 125I-glucagon, i.e., one whose labeling is markedly reduced by the presence of 1 μM native glucagon. The molecular weight of this structure in rat liver membranes is ~58,000, whereas in human membranes, the affinity labeled component is slightly larger, having a molecular weight of 63,000.

To further characterize the specificity of this binding component for glucagon and to compare it with the established features of the glucagon receptor, studies using native insulin, GTP, and different concentrations of native glucagon were conducted (Fig. 5). As shown, when the membranes were
incubated in the presence of increasing concentrations of native glucagon, there was a progressive decrease in the amount of the affinity labeled 63,000 band. Laser density spectrophotometric scanning of the autoradiogram demonstrated a 50% reduction in $^{125}$I-glucagon labeling of the 63,000 Mr band by 5 nM native glucagon. A high concentration of native insulin, however, did not reduce the glucagon labeling of this structure, in contrast to the incubation with GTP, which caused a marked decrease. These results are compatible with those expected of a glucagon receptor, and provide a strong argument that by polyacrylamide gel electrophoresis, the human glucagon receptor is a structure of 63,000 Mr.

**Discussion**

These findings demonstrate a glucagon receptor and a responsive adenylyl cyclase system in human liver membranes that are quite similar to the well-characterized system in rat liver membranes (17, 19, 25). Similarities are apparent in the glucagon binding characteristics between human and rat receptors, and both structures also have similar molecular weights. The Mr value of 63,000 for the human receptor is only slightly larger than the Mr 58,000 found in the rat liver membranes. These values are much larger than the 23,000 Mr originally reported by Bregman and Levy (26); however, they agree in general with the more recent reports of the size of the rat glucagon receptor, which range from 52,000 to 70,000 Mr (21, 27, 28).

The adenylyl cyclase system of human membranes also had a similar sensitivity to glucagon stimulation as rat liver membranes. The median effective dose value for stimulation by glucagon was 6 nM, and GTP was required for glucagon to fully activate the cyclase system. This GTP effect differs from the results described by Pecker et al. (7), who stated that the nucleotide did not affect the ability of glucagon to stimulate cAMP production. The reason for this difference is not clear,
since our data clearly show an effect of GTP both on the glucagon receptor binding activity and its ability to activate the adenylyl cyclase system.

The ability of human liver membranes from obese subjects to respond to glucagon was examined because recent findings indicate the presence of hyperglucagonemia in this disorder (9). Although other work has given conflicting results (reviewed in 13), the careful studies by Stark et al. (9) clearly demonstrate elevated pancreatic glucagon in obese subjects, including those who were glucose tolerant or intolerant. Our obese subjects also showed evidence of increased secretion. We measured plasma glucagon with a sensitive and specific radioimmuno-logical method (23, 24) and observed almost a 100% increase in the fasting glucagon level in obese subjects compared with non-obese subjects.

The presence of hyperglucagonemia in obesity raises the possibility that alterations exist in the number of glucagon receptors and glucagon action in much the same manner that hyperinsulinemia is associated with insulin resistance and loss of insulin receptors (29). Our findings show, however, that the membranes from obese subjects are as glucagon-responsive as those from normal weight subjects, regardless of the basis for expressing the glucagon effect, i.e., per milligram of protein, or the ability of NaF to stimulate cAMP production. These findings, along with the similar amount of glucagon binding activity in the two groups, indicate an adequate population of glucagon receptors that are appropriately coupled to the adenylyl cyclase system in obesity. Thus, unlike in vitro studies of hepatocytes, in which high concentrations of glucagon cause “down-regulation” of the glucagon receptor and a loss of response to the hormone (10), the moderate hyperglucagonemia found in obesity is not sufficient to elicit changes in receptor number or alterations in hormone action. It is also possible that the mild degree of hyperglucagonemia and the pulsatile delivery of the hormone that occurs in normal and obese subjects (30) helps prevent receptor loss and the development of glucagon resistance.

In summary, human liver membranes contain a glucagon receptor of Mr 63,000 and an adenylyl cyclase system that corresponds closely to the well-studied system in rat liver membranes. Furthermore, the hyperglucagonemia that develops in obesity does not alter the ability of this system to respond to glucagon. These findings do not eliminate the possibility of alterations at sites beyond the cyclase system, e.g., elevated phosphodiesterase activity (31), but they show that in obesity the initial steps in hepatic glucagon action are intact.

Acknowledgments

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References