Human Platelet-derived Growth Factor Stimulates Prostaglandin Synthesis by Activation and by Rapid *De Novo* Synthesis of Cyclooxygenase

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Abstract

Human platelet-derived growth factor (PDGF) stimulated prostaglandin (PG) E₂ synthesis in the cell cycle of Swiss 3T3 cells at two distinct time intervals, with a first plateau within 10 min and a second plateau within 2-4 h after addition of PDGF. At 4 h, the concentration of PGE₂ in PDGF-stimulated cultures exceeded the quiescent control cells by a factor of 10-15. Quiescent cells incubated with up to 16 μM exogenous arachidonic acid (AA) synthesized only small amounts of PGE₂. In contrast, 4 h after addition of PDGF, the concentration of PGE₂ synthesized from exogenous AA exceeded that in quiescent cultures by a factor of 28. The effect of PDGF stimulation on PG synthesis from exogenous AA could not be explained by growth factor-mediated increase in the cellular free AA pool as shown in experiments using [¹⁴C]AA. PDGF also stimulated synthesis of PG₁ (prostacyclin), thromboxane, and PGF₂α from exogenous AA. While inhibition of protein synthesis by 10 μg/ml cycloheximide had no effect on the early increase in PGE₂ synthesis, the second increase was completely prevented. Additionally, cycloheximide treatment at 6 h after PDGF stimulation resulted in rapid decline of PGE₂ synthesis from exogenous AA. Quiescent cultures pre-treated with 100 μM aspirin and stimulated by PDGF thereafter recovered from cyclooxygenase inhibition within 180 min. Our results suggest that phospholipase activation and resultant AA release is not sufficient to induce the burst of PG synthesis observed in PDGF-stimulated cells. Instead, PDGF stimulates PG synthesis by direct effects on the PG-synthesizing enzyme system, one involving a protein synthesis-independent mechanim and another that requires rapid translation of cyclooxygenase.

Introduction

The relation between phospholipase activation resulting in liberation of arachidonic acid (AA)¹ from complex glycerolipids and the cyclooxygenase pathway generating prostaglandins (PGs) and thromboxanes (TXs) is not entirely clear. Upon binding of hormones or other agonists to their respective cell surface receptors, AA is released from phospholipids through activation of phospholipases (1-9). In most if not all such instances, activation of phospholipases is associated with production of PGs and TXs (1, 2, 5, 9, 10). Therefore, it is generally believed that phospholipases control formation of PGs through release of AA. Since the intracellular pool of AA is low in mammalian cells (3, 6), there is little doubt that eicosanoid biosynthesis is dependent on phospholipase activity. However, it is less clear whether liberation of AA is sufficient to induce the burst of PG, TX, and leukotriene formation observed in many agonist response systems.

We have used quiescent Swiss 3T3 cells stimulated to divide by platelet-derived growth factor (PDGF), the principal mitogen for connective tissue-derived cells (11–13), as model systems to study the relation between growth factor-induced changes in phospholipid metabolism and AA release (14–16). Cells maintained in plasma-derived serum (PDS) become arrested in the G₀/G₁ phase of the cell cycle and can be stimulated to undergo cell cycle traverse after stimulation by PDGF in a partially synchronized fashion (17). We proposed that PDGF and epidermal growth factor stimulate AA release from phosphatidylinositol, a minor acidic phospholipid in mammalian cells, by activation of the phospholipase C/diglyceride lipase pathway (14, 15). This reaction sequence was first described by Bell et al. (18) and Majerus et al. (19) in thrombin-stimulated platelets.

The question we have addressed in the present report has been whether PDGF has direct effects on the PG synthesis pathway. It was of special interest to us whether PDGF has direct stimulatory effects on the key enzyme, the cyclooxygenase. Our results show that AA release from glycerolipids is not sufficient to induce the burst of PG synthesis in PDGF-stimulated cells (15, 20, 21). Instead, the growth factor greatly activates the PG synthetic enzyme system itself. This effect of PDGF appears to be mediated by both activation and rapid *de novo* synthesis of the cyclooxygenase.

Methods

Materials. [¹-¹⁴C]AA (sp act, 56.9 mCi/mmol), [¹⁴C]6-keto-PGF₁α (sp act, 120 Ci/mmol), [¹³C]PGF₂α (sp act, 150 Ci/mmol), [¹⁴C]PGF₁α (sp act, 160 Ci/mmol), [¹³C]PGE₂ (sp act, 139 Ci/mmol) was obtained from New England Nuclear, Boston, MA; Dulbecco's modification of Eagle's medium from Gibco Laboratories, Grand Island, NY; methanol, ethanol, ethyl acetate, benzene, acetone, dioxane, acetic acid, 200-μm silica gel-coated aluminum thin-layer chromatograms from Merck Inc., Darmstadt, Federal Republic of Germany; standard PGs were purchased from Seragen, Boston, MA; antibodies against PGE₂, 6-keto-PGF₁α, TXB₂, and PGF₂α from the Institute Pasteur, Paris; cycloheximide from Sigma Chemie, Munich, Federal Republic of Germany; AA from P. L. Biochemicals, St. Goar, Federal Republic of Germany.

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1. Abbreviations used in this paper: AA, arachidonic acid; PDS, plasma-derivative serum; PG, prostaglandin; PDGF, platelet-derived growth factor; TX, thromboxane.

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Plasma proteins and PDGF. Human or calf PDS lacking platelet-derived growth promoting activity was prepared as described (17). PDGF was prepared as described by Raines and Ross (22). PDGF at 12 ng/ml maximally stimulated DNA synthesis in Swiss 3T3 cells.

Radioimmunoassay (RIA). The concentration of PGs was determined by RIA according to Granstrom and Kindahl (23) after reversed-phase extraction of the culture medium by octadecyl C18 silica gel column chromatography according to Powell (24) and thin-layer chromatography. Briefly, 1 ml culture medium was removed from the dish and acidified to pH 3.6. 900 µl of the culture medium was applied to an octadecyl C18 silica gel column equilibrated with methanol. The column was washed twice with 3 ml double distilled H2O, and the PGs were eluted with a total volume of 1.5 ml ethylacetate. The ethylacetate was dried down under a stream of N2, taken up in 1 ml methanol, and stored until further analysis at -26°C. Before thin-layer chromatography, the methanol was dried down and the extract taken up in 150 µl ethanol, applied to a 200 µm silica gel-coated aluminum thin-layer chromatogram, and developed in solvent system of benzene/acetone/methanol/dioxane/H2O, 120:80:6:3:2; vol/vol. The spot containing the respective PG was scraped from the plate and extracted with 2 ml methanol, dried down under a stream of N2, and taken up in phosphate-buffered saline. Procedural losses were monitored by adding tracer [3H]PGs to each sample. Recoveries of PGE2 ranged from 34-61%, for 6-keto-PGF1α, from 54-68%, and for TXB2, from 48-72%. Cross-reactivity at 50% binding of PGE2 was 10.1% with PGE1, <0.2% with PGA1, PGA2, PGB1, PGB2, PGD2, PGF2α, PGF2β, 6-keto-PGF1α, and TXB2; and <0.03% with AA. Cross-reactivity at 50% binding of 6-keto-PGF1α was 6.8% with 6-keto-PGE1, 2.2% with 6-keto-PGF2α, 0.7% with PGE1, 0.5% with PGE2, <0.1% with all other PGs tested, and <0.3% with AA. Water samples subjected to exactly the same treatment as the biological samples ranged between 0 and 7 pg/ml for PGE2 and <5-15 pg/ml for 6-keto-PGF1α. Intraassay variation for PGE2 was 2-9%; and for 6-keto-PGF1α, was 7-11%. Interassay variation for PGE2 was 5-12% and for 6-keto-PGF1α was 8-22%.

Results

Kinetics of PGE2 synthesis in the cell cycle of Swiss 3T3 cells stimulated by PDGF. The synthesis kinetics of PGE2 in quiescent and PDGF-stimulated cells is shown in Fig. 1, A and B. We consistently observed in five experiments an early burst of PGE2 formation that was complete within 10 min. At 2 min, PDGF caused a statistically significant increase in PGE2 formation (Fig. 1 B). No further increase was observed until 2-4 h (Figs. 1 and 3). At that time, another sharp rise in PGE2 levels was observed. No additional increase was detected for the next 18 h. Thus, PDGF stimulated PGE2 synthesis in the cell cycle during two distinct time intervals: a first early increase that was complete within 10 min, and a second increase between 2 and 4 h.

PDGF stimulates PGE2 synthesis from exogenous AA. To test the ability of the cells to metabolize exogenous AA to PGE2, we incubated quiescent and PDGF-stimulated cells with increasing concentrations of AA (Fig. 2). Though the quiescent cultures were receiving concentrations of AA as high as 16 µM, they synthesized only small amounts of PGE2. In contrast, cells stimulated by PDGF synthesized up to 100 ng PGE2/10⁶ cells in the first 60 min of the cell cycle in all of five experiments. Concentrations between 4 and 16 µM AA did not further increase PGE2 synthesis both in quiescent and PDGF-stimulated cells. Therefore, a concentration of 10 µM AA was chosen for all future experiments. The ability of PDGF to stimulate PGE2 biosynthesis from exogenous AA might have been due to a PDGF-mediated increase in the cellular free AA pool, thus making more AA available to the cyclooxygenase. Therefore, we determined the effects of PDGF on this pool after various times of stimulation with the growth factor (Table 1). PDGF had only minor effects on free [14C]AA. Analysis of individual phospholipids by two-dimensional thin-layer chromatography indicated that PDGF stimulated [14C]AA incorporation into phosphatidic acid, phosphatidylglycerol, and triacylglycerol (16).

We next determined the cell cycle kinetics of PGE2 synthesis from exogenous AA. As shown in Fig. 3, the second rise in endogenous PGE2 synthesis which occurred between 2 and 4 h was associated with a second sharp increase in the formation of PGE2 from exogenous AA. The concentration of PGE2 in PDGF-stimulated cultures incubated with 10 µM AA for 60 min rose from 60 to 100 ng/10⁶ cells in the first 2 h up to 280 ng/10⁶ cells at 4 h in all of four experiments. Cells that had been stimulated by PDGF at time 0 and were incubated with 10 µM AA from 5 to 6 h synthesized up to 90 times more

Figure 1. Kinetics of PGE2 synthesis in the cell cycle of Swiss 3T3 cells stimulated by PDGF. Swiss 3T3 cells were plated at a density of 1.25 × 10⁴ cells in 35-mm Costar plates (Costar, Cambridge, MA) containing 1 ml of Dulbecco's modification of Eagle's medium (Gibco Laboratories) supplemented with 100 U/ml penicillin, 100 µg/ml streptomycin, and 1.25 mg/ml FBS protein. On day 3, the cultures were fed with the same medium and divided into two groups, and received either 50 µl 10 mM acetic acid (o) or 12 mg/ml PDGF in a concentrated solution of 50 µl 10 mM acetic acid (e). 12 ng/ml PDGF stimulated 1.25 × 10⁴ cells to double within 36 h. The same amount of PDGF stimulated incorporation of [methyl-3H]thymidine into trichloroacetic acid-precipitable material by a factor of 12-35 when assayed between 23 and 24 h. At the time points indicated in the figure (A, hours; B, minutes), 1 ml culture medium was removed from the dish and acidified to pH 3.6. The concentration of PGE2 was determined by RIA as described under Methods. Treatment of the cultures using 100 µM aspirin completely prevented formation of PGE2. The points indicated represent the means of three parallel cultures ± SD.
acetic acid and increasing concentrations of AA (c). Cultures that were not incubated with AA received 10 μl ethanol instead. AA was obtained from P. L. Biochemicals (St. Goar, Federal Republic of Germany), and multiple aliquots of a 10-mM stock solution were stored in ethanol under argon at −85°C. After 60 min, upon addition of PDGF and AA/ethanol, PGE_2_ levels were determined as described under Methods. The data points represent the means of three parallel cultures±SD.

PGE_2_ when compared with quiescent cells and up to 28 times when compared with quiescent cells incubated with 10 μM AA.

**PDGF stimulates prostacyclin and TXB_2_ synthesis from exogenous AA.** Since PDGF greatly stimulated PGE_2_ synthesis from exogenous AA without comparable stimulation of the intracellular [1^4]AA pool, it seemed that the growth factor had direct stimulatory effects on one or more enzymes involved in PG synthesis. To test this hypothesis further, we determined the levels of 6-keto-PGF_1α_ (the stable hydrolysis product of prostacyclin), another major cyclooxygenase-derived product of AA in Swiss 3T3 cells (25), over a period of 12 h (Fig. 4). 6-keto-PGF_1α_ levels increased up to 2 h upon stimulation by PDGF and then remained relatively constant or increased slightly for the next 10 h. In addition, PDGF-stimulated 6-keto-PGF_1α_ formation from exogenous AA (Fig. 4). When PDGF-stimulated cells incubated with AA were compared with quiescent cells incubated with AA, it became apparent that the growth factor stimulated 6-keto-PGF_1α_ synthesis by a factor of 27–35 in two experiments. Thus, the degree of stimulation of 6-keto-PGF_1α_ formation from exogenous AA was similar to that of PGE_2_ synthesis (Fig. 3). Other experiments demonstrated comparable effects of PDGF on synthesis of TXB_2_ and PGF_2α_ from exogenous AA (results not shown).

**PDGF stimulates PG synthesis by two distinct mechanisms.** To obtain further information on the mechanism of PG synthesis stimulation by the growth factor, we examined the effect of the protein synthesis inhibitor cycloheximide on synthesis of PGE_2_ (Table II). The concentration of cycloheximide used inhibited protein synthesis >90% as determined by [1^4]C]leucine incorporation into trichloroacetic acid-precipitable material. While cycloheximide did not inhibit the early increase of endogenous PGE_2_ synthesis, the second increase was completely prevented. Since this effect of cycloheximide might have been due to protein synthesis-sensitive phospholipase activation and AA release, we determined PGE_2_ synthesis after cycloheximide treatment from exogenous AA (Table II). Cycloheximide did not alter the early increase of PGE_2_ synthesis from exogenous AA while it completely prevented the second increase. In other experiments, PDGF-stimulated cells were

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**Table I. Effect of PDGF on Radioactivity of the Cellular Free Fatty Acid Pool Using [1^4]C]AA**

<table>
<thead>
<tr>
<th>Additions</th>
<th>1 h</th>
<th>2 h</th>
<th>16 h</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>dpm × 10^4/dish</td>
<td>dpm × 10^4/dish</td>
<td>dpm × 10^4/dish</td>
</tr>
<tr>
<td>Control</td>
<td>4.8±0.6</td>
<td>6.6±2.2</td>
<td>10.2±1.2</td>
</tr>
<tr>
<td>+PDGF</td>
<td>5.8±0.6</td>
<td>10.4±0.8</td>
<td>11.6±0.6</td>
</tr>
</tbody>
</table>

Swiss 3T3 cells were cultured as described in Fig. 1. On day 3, the cultures were divided into two groups and either received 50 μl 10 mM acetic acid (control) or 4 μg/ml partially purified PDGF dissolved in a concentrated solution of 50 μl 10 mM acetic acid. The amount of PDGF present in this preparation is equivalent to the amounts of highly purified PDGF used in the experiments in Figs. 1–3 and 5–7. At time 0, the cultures were incubated with 11.7 μM [1^4]C]AA dissolved in 10 μl ethanol. At the time points indicated, the culture medium was removed and the cell monolayer washed three times with ice-cold phosphate-buffered saline. Free fatty acids were extracted and separated by thin-layer chromatography as described (14). The silica gel was transferred to a scintillation vial and radioactivity was determined in a PW 4700 Philips Scintillation Spectrometer (Philips, Eindhoven, The Netherlands). The numbers represent the means of three parallel cultures±SD.
of amounts.

Exogenous cyclooxygenase.

To determine whether PDGF had direct effects on cyclooxygenase, we preincubated quiescent cells with aspirin (26). We then determined the kinetics of recovery by measurement of PGE2 synthesis from exogenous AA upon removal of aspirin and stimulation with PDGF (Fig. 6). The stimulated cells largely recovered between 2 and 4 h. Furthermore, aspirin treatment resulted in up to 45% enhanced PGE2 synthesis from exogenous AA when compared with PDGF-stimulated

Table II. Effects of Cycloheximide on PGE2 Synthesis from Endogenous and Exogenous AA in Quiescent and PDGF-stimulated Swiss 3T3 Cells

<table>
<thead>
<tr>
<th>Hours after addition of PDGF</th>
<th>PDS</th>
<th>PDS plus PDGF</th>
<th>PDS plus cycloheximide</th>
<th>PDS plus PDGF plus cycloheximide</th>
<th>PDS plus AA</th>
<th>PDS plus PDGF plus AA</th>
<th>PDS plus AA plus cycloheximide</th>
<th>PDS plus AA plus PDGF plus cycloheximide</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>2.1±0.3</td>
<td>24.0±3.5</td>
<td>2.8±0.1</td>
<td>22.0±1.1</td>
<td>7.9±1.5</td>
<td>109.0±5.9</td>
<td>6.4±0.8</td>
<td>104.2±6.9</td>
</tr>
<tr>
<td>3</td>
<td>3.8±0.3</td>
<td>47.2±1.4</td>
<td>3.6±1.7</td>
<td>26.4±3.2</td>
<td>7.2±0.8</td>
<td>216.0±17.3</td>
<td>6.2±2.8</td>
<td>102.4±20.8</td>
</tr>
</tbody>
</table>

Swiss 3T3 cells were cultured as described in Fig. 1. On day 4, the cultures were fed with medium containing 1.25 mg/ml PDS protein, divided into two groups, and received either 50 μl 10 mM acetic acid or 12 ng/ml PDGF in a concentrated solution of 50 μl 10 mM acetic acid. 60 min before the time points indicated, parallel cultures were incubated with 10 μM AA dissolved in 10 μl ethanol. Control cultures received 10 μl ethanol instead. PGE2 levels were determined as described in Fig. 1. The data represent the means of three parallel cultures±SD.
Figure 6. Recovery of PGE₂ synthesis from exogenous AA after inhibition of cyclooxygenase by aspirin in quiescent Swiss 3T3 cells stimulated by PDGF. Swiss 3T3 cells were cultured as described in Fig. 1. On day 4, the cultures were fed with medium containing 1.25 mg/ml PDS protein. 30 min before the medium was changed, parallel cultures were incubated with 50 μl phosphate-buffered saline or 100 μM aspirin (ASA pr.) dissolved in 50 μl phosphate-buffered saline. Before replacement of the medium containing PDS protein, the monolayers were washed twice with 2 ml phosphate-buffered saline to remove residual aspirin. At the time of the medium change, the cultures were divided into two groups and received either 50 μl 10 mM acetic acid or 12 ng/ml PDGF in a concentrated solution of 50 μl 10 mM acetic acid. 60 min before each time point indicated in the figure, the cultures were incubated with 10 μM AA dissolved in a concentrated solution of 10 μl ethanol. PGE₂ levels were determined as described under Methods. The data represent the means of three parallel cultures±SD.

cells in three experiments. No significant recovery from aspirin treatment was observed within the first 2 h after PDGF stimulation.

We next examined the kinetics of recovery from cyclooxygenase inhibition in PDGF-treated cells (Fig. 7). Complete recovery was observed within 180 min in PDGF-stimulated cells when cyclooxygenase inhibition was initiated between 5.5 and 7 h. The kinetics of recovery from cyclooxygenase inhibition (Fig. 7) was similar to the kinetics of inhibition of PGE₂ synthesis from exogenous AA after protein synthesis inhibition (Fig. 5). In addition, recovery from cyclooxygenase inhibition by aspirin was completely prevented by cycloheximide (results not shown).

Discussion

The experiments performed in this investigation focused on the possibility that human PDGF stimulated synthesis of PGs by direct activation of the cyclooxygenase pathway.

The generally accepted view of PG synthesis regulation has been that AA hydrolysis from complex glycerolipids represents the major control mechanism of PG biosynthesis (1-3, 6, 9, 18). However, this concept is based on circumstantial evidence; the intracellular pool of free AA, the substrate for the cyclooxygenase, is low in mammalian cells (2, 3, 5, 6, 18) and inhibitors of phospholipase activity have been shown to suppress PG synthesis from endogenous but not exogenous AA (27). Furthermore most if not all agonists eliciting increased PG synthesis concomitantly induce release of AA from phospholipids (3, 9, 14, 15, 20, 21, 28-30). While these studies are consistent with a critical role of phospholipase activity for PG synthesis, they do not provide information on several important questions: First, is AA release induced by phospholipases sufficient to induce the burst of PG synthesis observed in many agonist response systems? Second, what are the determining factors of regulation of the PG synthetic enzymes, and particularly, the cyclooxygenases? Work on mechanisms of regulation of the cyclooxygenase on the other hand has been difficult because this enzyme is subject to rapid inactivation by endoperoxide intermediates in cultured cells (31).

Several lines of published evidence suggest, however, that PG synthesis regulation is not simply the result of increased phospholipase activity. Thus, Bonser et al. (32) obtained evidence in cultured Swiss 3T3 cells that both bradykinin and thrombin stimulate PG synthesis from exogenous AA and Hyman et al. (25) showed that quiescent Swiss 3T3 cells form less PGs from exogenous AA when compared with their rapidly proliferating counterparts maintained in high serum concentrations. Furthermore, Ogekawa et al. (33) demonstrated that both the perfused hydrenephrotic kidney and microsomes prepared from the renal cortex of ureter-obstructed kidneys
show greatly enhanced PGE₂ synthesis from exogenous AA. Enhanced PGE₂ synthesis from exogenous AA appears to correlate best with the proliferation of fibroblast-like cells in this system. These results are consistent with the possibility that PG synthesis regulation involves activation of the PG synthesis enzyme system independent of phospholipase activity.

In the first series of experiments presented above we showed that PDGF stimulated PGE₂ formation in the cell cycle of Swiss 3T3 cells at two distinct time intervals: a first increase that was complete within 10 min and a second increase that occurred between 2 and 4 h. These results confirmed and extended the findings of Shier (28) and Coughlin et al. (20) who showed that PDGF stimulates PG synthesis in cultured cells.

Our results using exogenous AA (Figs. 2 and 3, Table II) suggested that PDGF stimulated PG synthesis by mechanisms that are independent of phospholipase; the effect of PDGF on PGE₂ synthesis from exogenous AA did not appear to be due to the PDGF-dependent increase in the AA pool (Table I). Furthermore, quiescent cells incubated with concentrations of AA as high as 16 μM (Fig. 2) were unable to synthesize significant amounts of PGE₂ although AA was taken up by the cells.

To gain further insight into the mechanism of PDGF-dependent PGE₂ synthesis stimulation from both endogenous and exogenous AA, we next determined whether other cyclooxygenase products were stimulated by PDGF. The results presented in Fig. 4 demonstrate that PDGF stimulated 6-keto-PGF₁α synthesis from both endogenous and exogenous AA and other experiments revealed similar effects of the growth factor on TXB₂ and PGF₂α synthesis (results not shown). Since the effect of PDGF on stimulation of PG synthesis from exogenous AA included all of the major cyclooxygenase-derived products, i.e., PGE₂, 6-keto-PGF₁α, TXB₂, and PGF₂α, these results suggest that the effect of PDGF was mediated by activation of the key enzyme of PG and TX synthesis, the cyclooxygenase.

In experiments using protein synthesis inhibition, we showed that the first increase in PGE₂ synthesis from both endogenous and exogenous AA was protein synthesis independent while the second increase was completely suppressed in the presence of cycloheximide (Table II). The protein synthesis insensitivity of the first increase in PGE₂ formation might, therefore, reflect rapid activation of the cyclooxygenase pathway by posttranslational modification mechanisms. Although these results are consistent with activation of the cyclooxygenase pathway, the mechanism of activation remains to be elucidated. It is possible that phosphorylation reactions are involved since PDGF has been shown to induce, within minutes, phosphorylation of several proteins (34).

The second increase of PGE₂ synthesis observed between 2 and 4 h after stimulation of the cells was totally dependent on protein synthesis (Table II). To test whether protein synthesis was required for maintaining high PG synthesis rates from exogenous AA, we determined the kinetics of decline of PGE₂ synthesis from exogenous AA in PDGF-stimulated cells (Fig. 5). The results show that protein synthesis inhibition resulted in rapid inactivation of the cyclooxygenase pathway with approximate half-maximal inhibition between 90 and 120 min. This decline might reflect rapid turnover of enzymes involved in PG synthesis. The results are also consistent with rapid inactivation of the cyclooxygenase pathway as has been observed by Brotherton and Hoak (31) in cultured endothelial cells. In both cases, maintenance of high PG synthesis rates required translational activity.

The requirement for protein synthesis appeared to be closely associated with cyclooxygenase activity. Thus, quiescent cells pretreated with aspirin (Fig. 6) and stimulated by PDGF thereafter largely recovered during the protein synthesis-sensitive time period, i.e., between 2 and 4 h. No significant recovery was observed during the first 2 h of the cell cycle. Furthermore, PDGF-stimulated cells that were incubated with aspirin, upon removal of aspirin, recovered from cyclooxygenase inhibition within 180 min. In three independent experiments we observed that aspirin-treated cells, after recovery from cyclooxygenase inhibition, synthesized up to 45% more PGE₂ from endogenous (not shown) and exogenous AA as compared with the control PDGF-stimulated cultures (Figs. 6 and 7). Since recovery from aspirin treatment was totally dependent on protein synthesis (results not shown), the enhanced PG synthesis rate after short-term inhibition with aspirin seemed to be due to enhanced de novo synthesis of cyclooxygenase after aspirin treatment. The mechanism underlying this phenomenon is presently unclear. Taken together, the results presented in Figs. 5–7 show that the PDGF-dependent stimulation of PGE₂ occurring between 2 and 4 h is mediated by rapid de novo synthesis of cyclooxygenase.

In conclusion, our results disclose that PDGF has profound direct stimulatory effects on the PG synthesis system in addition to the earlier observed activation of the phospholipase C/diglyceride lipase pathway (14–16).

Recently, Bailey et al. (35) reported that epidermal growth factor shares some of the activities of PDGF. It will be of major general interest to determine whether activation of cyclooxygenase by agonists represents a general mechanism of PG biosynthesis regulation. If so, our results would have important implications for a variety of physiological as well as pathophysiological conditions. These include the response of tissues to hormones and the regulation of PG, leukotriene, and lipoxin biosynthesis in inflammatory diseases.

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