Basis for Defective Responses of Rheumatoid Arthritis
Synovial Fluid Lymphocytes to Anti–CD3 (T3) Antibodies

Martin Lotz, Constantine D. Tsoukas, Charles A. Robinson, Charles A. Dinarello,* Dennis A. Carson, and John H. Vaughan
Department of Basic and Clinical Research, Scripps Clinic and Research Foundation, La Jolla, California 92037; and *Tufts University
School of Medicine, Boston, Massachusetts 02112

Abstract

Synovial fluid mononuclear cells (SFMC) from patients with active rheumatoid arthritis characteristically respond poorly to mitogens. In this study, mitogenic antibodies reactive with the CD3(T3) antigen on human T lymphocytes were used to analyze the basis for the deficiency. OKT3-induced proliferation and release of interleukin 1 (IL-1) and interleukin 2 (IL-2) from SFMC were depressed in all patients. Purified IL-1 or recombinant IL-2 restored proliferative responses in SFMC and increased IL-2 receptor density. Exogenous IL-1 also enhanced IL-2 release.

Fractionation of SFMC supernatants on phosphocellulose columns revealed the presence of IL-1 and a potent IL-1 inhibitor. The monocyte-derived IL-1 inhibitor blocked IL-1-dependent responses of normal peripheral blood lymphocytes to OKT3, but had no effect on IL-2–dependent events. These results suggest that IL-1 inhibitor(s) in SFMC impair(s) OKT3-induced mitogenesis by interfering with the effects of IL-1 on T lymphocytes. The net result is deficient IL-2 secretion, IL-2 receptor expression, and impaired cellular proliferation. This novel inhibitory circuit provides a rational explanation for the diminished function of synovial fluid T lymphocytes in rheumatoid arthritis patients.

Introduction

The proliferative response of rheumatoid arthritis (RA)1 T lymphocytes to stimulation with plant mitogens is deficient (1–3). Peripheral blood lymphocytes from patients with active and severe disease are moderately hypo-responsive (1), whereas the effect of synovial fluid lymphocytes is much more profound (2, 3). The basis for this abnormality is unknown, and is difficult to study with nonspecific plant mitogens, because they react with sugar residues not selectively expressed on T lymphocytes (4, 5). Furthermore, phytohemagglutinin (PHA) reacts with at least two distinct surface molecules on T cells, the antigen receptor-T3 complex and the sheep red blood cell receptor (5, 6). Both these receptors are triggers for T cell activation.

Monoclonal antibodies against the CD3(T3) molecular complex, which is exclusively present on T lymphocytes (7, 8), are potent mitogens (9). The cellular interactions leading to anti-CD3–induced T cell proliferation are well defined. The anti-CD3 antibodies bind specifically to their corresponding antigen on the T cell surface, whereas the Fc portion binds to Fc receptors on monocytes. Both events are required for anti-CD3–triggered T cell activation (10–12). The T cell–monocyte interaction, presumably by a cross-linking event, induces interleukin 1 (IL-1) responsiveness and the presence of IL-1 is important for expression of receptors for interleukin 2 (IL-2), the release of IL-2, and ultimately IL-2–dependent cellular proliferation (13, 14).

In the present investigation we have used anti-CD3 antibodies in order to examine these activation events in lymphocytes from RA blood and synovial fluid. The results indicate that the depressed mitogenic responses of RA lymphocytes are mediated predominantly by a specific inhibitor of IL-1.

Methods

Patient selection and characteristics. Heparinized peripheral blood was obtained from 22 healthy volunteers and patients with classical RA according to American Rheumatism Association criteria (15). Paired synovial fluid and peripheral blood samples were obtained from 19 patients with RA. Their biological parameters and medications are shown in Table I. An additional nine RA synovial fluid specimens came from subjects whose blood was not available.

Lymphocyte isolation and culture. Mononuclear cells were isolated from synovial fluid and peripheral blood by Ficoll-Hypaque density-gradient sedimentation (16). Lymphocytes were cultured in RPMI-1640 medium supplemented with 5% heat-inactivated (56°C, 30 min) fetal bovine serum, 100 U/ml penicillin, 100 μg/ml streptomycin, 2 mM L-glutamine (all from M.A. Bioproducts, Los Angeles, CA) and 12.5 μg/ml polymyxin B. Aliquots of cells (200 μl each) at a density of 106/ml were distributed among the wells of microtiter trays (Costar No. 3596, Data Packaging Corp., Cambridge, MA), and were incubated for 72 h in an atmosphere of 5% CO2-air in the presence or absence of the anti-CD3 antibodies, OKT3 (0.001–10 ng/ml) or Leu 4 (0.01–200 ng/ml). [3H]Thymidine (1 μCi/well) was added during the last 4 h of incubation. The cells were collected on glass fiber filters, and radioactivity was assessed by liquid scintillation spectrometry.

Sources of IL-1, IL-2, and antibodies. The IL-1 used in these studies was purified from cultures of human adherent blood monocytes as previously described using a combination of immunoadsorption and gel filtration (17). When separated by charge (chromatofocusing or isoelectric focusing), this IL-1 contained predominantly the p7 form. The purified material was assayed for the production of fever in rabbits and for lymphocyte-activating factor (LAF). The IL-1 used in these studies contained ~100 U/ml where a unit is defined as the amount of IL-1/ml which doubled the proliferative responses of murine thymocytes to suboptimal concentrations of PHA. The antibody to IL-1 has been described previously (18). It was absorbed against immobilized human serum and human mononuclear cells at 4°C. It did not react with human IL-2, B cell growth factor, human CS, or interferons-α or -γ. A 1:100 dilution of the absorbed anti–IL-1 neutralized 100 U/ml of IL-1 in the standard

This is Publication No. 3926BCR from the Research Institute of Scripps Clinic.

Address reprint requests to Dr. Lotz, Department of Basic and Clinical Research-BCR4, Scripps Clinic and Research Foundation, 10666 North Torrey Pines Road, La Jolla, CA 92037.

Received for publication 8 July 1985 and in revised form 22 April 1986.

Abbreviations used in this paper: IL-1 and IL-2, interleukins 1 and 2; PBMC, peripheral blood mononuclear cell(s); PHA, phytohemagglutinin; RA, rheumatoid arthritis; SFMC, synovial fluid mononuclear cell(s).

Table I. Biological Parameters from RA Patients

<table>
<thead>
<tr>
<th>Patient</th>
<th>Sex</th>
<th>Age</th>
<th>ESR</th>
<th>Disease duration</th>
<th>Medications</th>
<th>PBMC</th>
<th>OKT3</th>
<th>SFMC</th>
<th>OKT3</th>
</tr>
</thead>
<tbody>
<tr>
<td>B.A.</td>
<td>Female</td>
<td>63</td>
<td>68</td>
<td>12</td>
<td>NSAID, prednisone</td>
<td>341</td>
<td>24,536</td>
<td>294</td>
<td>5,220</td>
</tr>
<tr>
<td>V.M.</td>
<td>Female</td>
<td>52</td>
<td>59</td>
<td>16</td>
<td>NSAID</td>
<td>673</td>
<td>59,310</td>
<td>559</td>
<td>7,818</td>
</tr>
<tr>
<td>D.M.</td>
<td>Female</td>
<td>63</td>
<td>69</td>
<td>6</td>
<td>NSAID, plaquenil</td>
<td>2,460</td>
<td>83,969</td>
<td>522</td>
<td>9,395</td>
</tr>
<tr>
<td>E.S.</td>
<td>Male</td>
<td>64</td>
<td>48</td>
<td>11</td>
<td>Prednisone, NSAID</td>
<td>912</td>
<td>122,525</td>
<td>3,319</td>
<td>11,861</td>
</tr>
<tr>
<td>H.W.</td>
<td>Male</td>
<td>68</td>
<td>32</td>
<td>7</td>
<td>NSAID</td>
<td>3,374</td>
<td>82,206</td>
<td>1,640</td>
<td>23,613</td>
</tr>
<tr>
<td>J.S.</td>
<td>Female</td>
<td>62</td>
<td>91</td>
<td>4</td>
<td>NSAID</td>
<td>2,488</td>
<td>92,937</td>
<td>634</td>
<td>24,342</td>
</tr>
<tr>
<td>M.F.</td>
<td>Female</td>
<td>48</td>
<td>52</td>
<td>10</td>
<td>NSAID, gold</td>
<td>1,096</td>
<td>56,382</td>
<td>1,936</td>
<td>14,364</td>
</tr>
<tr>
<td>E.D.</td>
<td>Female</td>
<td>72</td>
<td>117</td>
<td>8</td>
<td>d-Penicillamine</td>
<td>1,930</td>
<td>49,387</td>
<td>2,876</td>
<td>12,392</td>
</tr>
<tr>
<td>R.P.</td>
<td>Female</td>
<td>64</td>
<td>96</td>
<td>12</td>
<td>NSAID</td>
<td>876</td>
<td>38,492</td>
<td>1,392</td>
<td>9,845</td>
</tr>
<tr>
<td>L.J.</td>
<td>Male</td>
<td>52</td>
<td>83</td>
<td>3</td>
<td>Gold, prednisone, NSAID</td>
<td>218</td>
<td>42,836</td>
<td>1,231</td>
<td>19,348</td>
</tr>
<tr>
<td>L.C.</td>
<td>Female</td>
<td>54</td>
<td>46</td>
<td>8</td>
<td>Prednisone, NSAID</td>
<td>2,046</td>
<td>68,392</td>
<td>1,796</td>
<td>16,839</td>
</tr>
<tr>
<td>L.K.</td>
<td>Female</td>
<td>28</td>
<td>108</td>
<td>5</td>
<td>d-Penicillamine, plaquenil, prednisone, NSAID</td>
<td>3,151</td>
<td>39,569</td>
<td>2,046</td>
<td>14,834</td>
</tr>
<tr>
<td>L.K.</td>
<td>Female</td>
<td>65</td>
<td>48</td>
<td>14</td>
<td>Plaquenil, NSAID</td>
<td>734</td>
<td>58,304</td>
<td>1,124</td>
<td>8,837</td>
</tr>
<tr>
<td>J.D.</td>
<td>Male</td>
<td>43</td>
<td>104</td>
<td>1</td>
<td>NSAID, gold</td>
<td>782</td>
<td>148,580</td>
<td>582</td>
<td>77,988</td>
</tr>
<tr>
<td>P.C.</td>
<td>Female</td>
<td>63</td>
<td>62</td>
<td>8</td>
<td>NSAID</td>
<td>1,832</td>
<td>42,893</td>
<td>1,413</td>
<td>17,922</td>
</tr>
<tr>
<td>M.Z.</td>
<td>Male</td>
<td>65</td>
<td>34</td>
<td>11</td>
<td>Plaquenil, NSAID</td>
<td>1,123</td>
<td>39,845</td>
<td>1,780</td>
<td>9,438</td>
</tr>
<tr>
<td>B.G.</td>
<td>Female</td>
<td>60</td>
<td>73</td>
<td>15</td>
<td>Cytoxan</td>
<td>1,481</td>
<td>79,445</td>
<td>2,034</td>
<td>13,942</td>
</tr>
<tr>
<td>P.M.</td>
<td>Male</td>
<td>63</td>
<td>82</td>
<td>8</td>
<td>Imuran, NSAID</td>
<td>987</td>
<td>56,832</td>
<td>1,684</td>
<td>12,396</td>
</tr>
<tr>
<td>D.E.</td>
<td>Male</td>
<td>66</td>
<td>45</td>
<td>3</td>
<td>d-Penicillamine, NSAID</td>
<td>2,033</td>
<td>68,794</td>
<td>986</td>
<td>16,341</td>
</tr>
</tbody>
</table>

Abbreviations: ESR, erythrocyte sedimentation rate; NSAID, nonsteroidal anti-inflammatory drugs.

thymocyte assay. Recombinant IL-2 produced in Escherichia coli (19) was generously provided by Cetus Corp. (Emeryville, CA). IL-2 lot LP227 was shown by the supplier to be 93% pure by sodium dodecyl sulfate-polyacrylamide gel electrophoresis and contained <0.02 ng/mg endotoxin. One unit of IL-2 is the amount producing half maximal growth of an IL-2-dependent cell line (20). Monoclonal anti-IL-2 antibody (DMS-3) (21) was kindly provided by Dr. Kendall Smith (Dartmouth Medical School, Hanover, NH). The anti-CD3 monoclonal antibodies used were OKT3 (Ortho Diagnostic Systems, Raritan, NJ) and Leu 4 (Becton-Dickinson & Co., Mountain View, CA).

Assays for IL-1, IL-1 inhibitor, IL-2, and IL-2 receptors. The lymphocyte-activating factor assay (22) was used to measure IL-1 and IL-1 inhibitor activity in 24-h supernatants from cells that had been cultured at 2 x 10^6/ml in standard culture medium with and without mitogen. Thymocytes from 4-6-wk-old C3H/HeJ mice were suspended in RPMI 1640 supplemented with 5% heat-inactivated fetal bovine serum, 5 x 10^-5 M 2-mercaptoethanol, 100 U/ml penicillin, 100 μg/ml streptomycin, 2 mM glutamine, and 12.5 μM/m polynoxyn B. Cells were cultured at 10^4/well in 96-well flat-bottom tissue culture plates in the presence of 1 μg/ml PHA and a 25% final concentration of the peripheral blood mononuclear cell (PBMC) and synovial fluid mononuclear cell (SFMC) culture supernatants. Titration of these supernatants resulted in a dose-dependent loss of IL-1 activity. Thymocyte proliferation was measured by [3H]thymidine incorporation after 48 h. To test for specificity of the activity, replicates of the supernatants were preincubated for 2 h at 37°C with a 1:100 dilution of antibody against human IL-1 (18) and then tested for residual IL-1 activity. In these studies, neutralization was accomplished by mixing culture supernates with a 1:100 dilution of antibody for ~2 h at 37°C and then added these to murine thymocytes. Supernatants to be tested for IL-1 inhibitor activity were titrated against 10 U of purified IL-1.

IL-2 was assayed after 48 h on the IL-2-dependent murine cell line CTLL-2 (20). Specificity assays were performed by neutralization (2 h, 37°C) with anti-IL-1 antiserum (1:100) and anti-IL-2 monoclonal antibody (1:10), respectively. All supernatant samples were tested at a 1:4 final dilution which was determined to be an optimal endpoint by titration curves. IL-2 receptors were detected after incubation of extensively washed cells with a fluorescein-conjugated anti-IL-2 receptor monoclonal antibody (Becton, Dickinson & Co., Sunnyvale, CA), followed by cytofluorographic analysis with a FACS IV (Becton, Dickinson & Co.) as previously described (23).

Data analysis. All cultures were performed at least three times, each in triplicate. Mean values were compared by Student's t test (two tails).

Results

Proliferative responses and lymphokine production in cultures of RA and normal PBMC after stimulation with antibody OKT3. Optimal concentrations of antibody OKT3 (1 ng/ml) induced vigorous proliferation of PBMC from normal subjects and RA patients. The ranges of [3H]thymidine uptake in the normal group (26,382–112,123 cpm) and in the RA group (13,638–121,453 cpm) overlapped. The mean values of cultures with normal lymphocytes (70,212±530,470) were not significantly different from those of RA lymphocytes (45,616±64,378 cpm). Cultures from several RA patients, however, showed mean values considerably below those from normal patients. This group of patients had more active disease as characterized by erythrocyte sedimentation rate, rheumatoid factor titer, and joint counts.

In contrast, with suboptimal doses of OKT3, proliferation of lymphocytes from nearly all RA patients was depressed (Table II). Thus, using 0.1 ng of OKT3, the [3H]thymidine uptake was 8,415±2,1036 for the RA patients compared with 35,175±2,432 for the normal subjects (P < 0.01). Stimulation of RA and normal lymphocytes with 0.01 ng of OKT3 yielded a similarly significant difference.

714 Lotz et al.
Table II. Proliferation and Interleukin Production after Stimulation with OKT3

<table>
<thead>
<tr>
<th>OKT3 (ng/ml)</th>
<th>0</th>
<th>0.01</th>
<th>0.1</th>
<th>1.0</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>NL</td>
<td>RA</td>
<td>NL</td>
<td>RA</td>
</tr>
<tr>
<td>cpm</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Prolif.</td>
<td>716±208</td>
<td>193±94</td>
<td>2,179±320</td>
<td>500±182</td>
</tr>
<tr>
<td>IL-2</td>
<td>2,682±312</td>
<td>1,034±54</td>
<td>4,322±538</td>
<td>1,628±304</td>
</tr>
<tr>
<td>IL-1</td>
<td>1,562±296</td>
<td>986±282</td>
<td>20,324±1,832</td>
<td>1,426±256</td>
</tr>
</tbody>
</table>

PBMC from RA patients (n = 6) and normal subjects (NL) (n = 6) were stimulated with 0.01–1 ng/ml OKT3. Proliferation was assayed by [3H]thymidine uptake after 72 h. IL-1 activity was measured on mouse thymocytes (assay background 1,326 cpm) and IL-2 activity on CTLL-2 cells (assay background 1,834 cpm) after 36 h. Each culture was performed in triplicate. Each point represents the mean counts per minute±SEM of six experiments in the various assays.

The levels of IL-1 and IL-2 activities in the OKT3-stimulated cultures correlated well with the proliferative responses (Table II). Again, lymphocytes from RA patients and normal subjects released markedly different amounts of the two lymphokines after stimulation with 0.1 and 0.01 ng/ml OKT3 antibody.

Decreased proliferative responses in synovial fluid lymphocytes. Mononuclear cells from RA synovial fluids (SFMC) responded very weakly to OKT3 as compared with autologous PBMC (P < 0.001; Fig. 1). The low response occurred throughout a wide dose range (Table III). The spontaneous proliferation (no added OKT3) of synovial fluid cells (1,592±1,188 cpm) was not significantly (P > 0.05) different from that of peripheral blood lymphocytes (1,136±946 cpm). Cell viability as estimated by trypsin blue exclusion exceeded 90% in both SFMC and PBMC after 3 d in culture. Such variables as disease duration, disease activity, or medications did not appear to account specifically for the SFMC-impaired proliferation (cf. Table I).

To exclude the possibility that the low response of SFMC was related to the particular IgG subclass of the anti-CD3 antibody used, we compared the effects of two anti-CD3 antibodies, Leu 4 (IgG1) and OKT3 (IgG2a). The proliferative responses of RA SFMC were severely depressed (Fig. 1) with both antibodies. The Leu 4 monoclonal antibody will not activate the PBMC from ~40% of normal donors (10, 12, 24). The frequency of Leu 4 nonresponders did not differ among RA subjects (45%); the data in Fig. 1 for RA were derived from responders.

We tested the possibility that the low response in SFMC could have been due to lower numbers of T lymphocytes or to modulation of the T3 molecule. The relative proportions of OKT3, or of Leu 4-reactive T cells, to the total numbers of PBMC or SFMC were similar. In addition, FACS analysis revealed that the density of surface molecules reactive with the OKT3 and Leu 4 antibodies was indistinguishable in SFMC and PBMC (data not shown).

Effects of indomethacin and of preincubation on OKT3-induced proliferation. Activated cells in synovial fluid may release prostaglandins which can inhibit lymphocyte responses, particularly the secretion of IL-2 (25). To evaluate whether prostaglandins were involved in the low proliferative responses observed, OKT3-stimulated PBMC and SFMC were cultured with indomethacin (1–10 μg/ml), an inhibitor of prostaglandin synthesis. Under these conditions proliferation of PBMC was augmented by only 34% and that of synovial cells by 28% (data not shown). The difference between the responses of cells from synovial fluid and blood remained significant.

Cells in the synovial fluid are exposed in vivo to immunoglobulin aggregates, which may block Fc receptors on monocytes and thereby prevent OKT3 mitogenesis (26). Therefore, we preincubated SFMC for 24, 48, and 74 h in medium alone before the addition of OKT3 in order to permit shedding of immunoglobulins and clearing of Fc receptors. The preincubation did not alter the low proliferative response of SFMC (data not shown).

Production of interleukins by OKT3-stimulated PBMC and SFMC. Supernatants of PBMC and SFMC cultures stimulated with OKT3 (1 ng/ml) were harvested after 24 and 48 h and assayed for IL-1 and IL-2 activities, respectively. The interleukin results corresponded remarkably with the proliferative responses (Figs. 2 and 3). IL-1 and IL-2 activities were markedly lower in SFMC culture supernatants as compared with PBMC. The supernatants of SFMC did not have toxicity for the cells used in the bioassays. They impaired neither the proliferation of mouse thymocytes stimulated with 10 μg/ml PHA, nor the proliferative responses of CTLL-2 to recombinant human or rat IL-2 (results not shown).

Exogenous purified IL-1 at 32 U/ml caused a threefold increase in OKT3-induced IL-2 secretion from SFMC. Under these conditions, therefore, IL-2 levels in SFMC and PBMC super-
Table III. Proliferative Responses of SFMC to OKT3 Stimulation

<table>
<thead>
<tr>
<th>OKT3 (ng/ml)</th>
<th>cpm</th>
<th>cpm</th>
<th>cpm</th>
<th>cpm</th>
<th>cpm</th>
<th>cpm</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>1,592±632</td>
<td>1,786±582</td>
<td>2,268±540</td>
<td>3,424±636</td>
<td>9,582±1,210</td>
<td>15,450±1,352</td>
</tr>
</tbody>
</table>

SFMC were stimulated with varying doses of OKT3 (0–10 ng/ml). Proliferation was quantified after 72 h as [3H]thymidine uptake. Data are shown as mean counts per minute±SEM of six experiments.

Increased responses of the SFMC to OKT3 (1 ng/ml). With both, the increase was dose-dependent. At 30 U/ml of IL-2, the OKT3-induced proliferation of SFMC was close to that of PBMC (Fig. 4 B). Purified IL-1 by itself did not trigger significant levels of proliferation.

IL-1 inhibitor in culture supernatants from RA PBMC and SFMC. Supernatants from RA PBMC and SFMC cultured with or without OKT3 were tested for neutralization of a standard IL-1 preparation. Fig. 5 shows that supernatants from both types of cells contained a factor that neutralizes IL-1. However, much higher concentrations of the IL-1 inhibitor were detected in SFMC cultures. The inhibitor activity decreased after OKT3 stimulation, but still persisted in high titer in SFMC cultures (Fig. 5). It was not detectable in supernatants of normal PBMC cultures. Culture supernatants from RA SFMC did not neutralize the effect of purified or recombinant human IL-2 on CTLL-2 cells (Table V). Furthermore, [3H]thymidine uptake by mouse thymocytes stimulated with optimal doses of PHA was not reduced by RA SFMC culture supernatants (data not shown), excluding nonspecific inhibition of thymocyte proliferation. The IL-1 inhibitor was nondialyzable, and therefore it could not be a smaller molecule such as a prostaglandin. In support of this conclusion, supernatants from cells cultured with indomethacin retained IL-1 inhibitory activity (not shown). More important, SFMC supernatants containing IL-1 inhibitory activity decreased OKT3-induced proliferation of normal PBMC (Table VI). The inhibition could be overcome by adding exogenous IL-1.

To determine the cellular source of the IL-1 inhibitor activity

![Figure 2](image1.png)  
**Figure 2.** IL-1 activity in culture supernatants collected after 24 h from unstimulated (Contr) and OKT3 (1 ng/ml) stimulated PBMC (open bars) and SFMC (hatched bars) of RA patients. IL-1 activity was assayed on mouse thymocytes. Data are shown as mean counts per minute±SEM. Background [3H]thymidine uptake of thymocytes was 1,422±294 cpm. The inserted solid bar indicates residual proliferative activity after preincubation of the supernatant with anti-IL-1 antibody.

![Figure 3](image2.png)  
**Figure 3.** IL-2 activities in culture supernatants of RA PBMC (open bars) and SFMC (hatched bars). The addition of increasing amounts of purified IL-1 to a standard dose of OKT3 (1 ng/ml) increased progressively the responses of the SFMC to the OKT3. Inserted solid bars indicate residual proliferative activity after preincubation of the supernatant with anti-IL-2 antibody. IL-2 was assayed after 48 h of incubation on CTLL-2 cells.
Table IV. IL-2 Receptor Expression

<table>
<thead>
<tr>
<th></th>
<th>NL PBMC</th>
<th>RA PBMC</th>
<th>RA SFMC</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>%</td>
<td>%</td>
<td>%</td>
</tr>
<tr>
<td>Control</td>
<td>2.6±1.8</td>
<td>3.4±2.3</td>
<td>19.4±4.6</td>
</tr>
<tr>
<td>OKT3</td>
<td>47.3±6.2</td>
<td>45.6±5.2</td>
<td>22.6±3.9</td>
</tr>
<tr>
<td>IL-1</td>
<td>8</td>
<td>2.3±1.4</td>
<td>2.6±1.2</td>
</tr>
<tr>
<td></td>
<td>16</td>
<td>2.8±2.2</td>
<td>3.2±1.6</td>
</tr>
<tr>
<td></td>
<td>32</td>
<td>3.2±2.1</td>
<td>2.8±1.2</td>
</tr>
<tr>
<td>OKT3+IL-1</td>
<td>8</td>
<td>49.7±5.2</td>
<td>51.2±4.0</td>
</tr>
<tr>
<td></td>
<td>16</td>
<td>52.0±6.7</td>
<td>52.4±5.2</td>
</tr>
<tr>
<td></td>
<td>32</td>
<td>57.3±5.4</td>
<td>50.2±4.6</td>
</tr>
<tr>
<td>IL-2</td>
<td>7.5</td>
<td>3.2±1.4</td>
<td>2.6±1.8</td>
</tr>
<tr>
<td></td>
<td>15</td>
<td>2.8±1.2</td>
<td>2.9±1.2</td>
</tr>
<tr>
<td></td>
<td>30</td>
<td>2.5±1.6</td>
<td>1.8±1.2</td>
</tr>
<tr>
<td>OKT3+IL-2</td>
<td>7.5</td>
<td>51.2±4.9</td>
<td>52.6±4.8</td>
</tr>
<tr>
<td></td>
<td>15</td>
<td>54.4±5.3</td>
<td>56.4±5.2</td>
</tr>
<tr>
<td></td>
<td>30</td>
<td>56.2±6.4</td>
<td>55.3±4.6</td>
</tr>
</tbody>
</table>

Mononuclear cells from normal donors (NL) and paired samples of RA peripheral blood and synovial fluid were cultured in medium alone, in the presence of OKT3 (1 ng/ml) alone, or in the presence of varying concentrations (U/ml) of IL-1 or IL-2 with and without OKT3. Cells were harvested after 48 h, stained with fluorescein-conjugated anti–IL-2 receptor monoclonal antibody and analyzed in the FACS. Data are shown as percent IL-2 receptor-positive cells±SEM of seven experiments.

When we depleted SFMC of monocytes by plastic adherence and subsequent carbonyl-iron treatment, the adherent cells were depleted from the plastic surfaces with EDTA and cultured in parallel with SFMC and monocyte-depleted SFMC. Table VII shows that the IL-1 inhibitor activity is predominantly derived from the monocyte-enriched population.

In an attempt to dissociate IL-1 from the IL-1 inhibitor(s), SFMC culture supernatants were chromatographed on phosphocellulose columns. After elution with a linear salt gradient (0.1–1 M), fractions were dialyzed against RPMI and tested on mouse thymocytes for IL-1 activity. The IL-1 activity eluted as a single peak at 0.3 M NaCl (Fig. 6) and was neutralized with the anti–IL-1 antibody. The IL-1 inhibitory activity did not bind to phosphocellulose and eluted with the void volume of the column (Fig. 6). Thus, SFMC supernatants do contain biologically active IL-1, which can be detected after separation from the IL-1 inhibitor(s).

Mixing of lymphocytes and monocytes from blood and synovial fluid. Having demonstrated that synovial fluid monocytes release much higher levels of IL-1 inhibitor activity than blood monocytes, we tested various combinations of purified lymphocytes and monocytes. PBMC and SFMC from the same patient were partially depleted of monocytes by plastic adherence. Residual monocytes were removed by carbonyl-iron treatment. The adherent cells were dislodged with trypsin-EDTA and recombined with lymphocytes. Table VIII shows that both monocyte-depleted PBMC and SFMC, as expected, did not respond to OKT3. When recombined with 10% blood monocytes, blood lymphocytes (two of three experiments) gave a high proliferative response, but with monocytes from synovial fluid, they uniformly showed low responses. With synovial fluid monocytes, synovial fluid lymphocytes gave a characteristically low proliferative response, but with blood monocytes they gave much higher responses. These results thus demonstrate that the poor mitogen

\[ \text{Figure 4. Proliferative responses of RA PBMC (open bars) and SFMC (hatched bars) in response to OKT3 (1 ng/ml) in the presence of exogenous IL-1 (8–32 U/ml) (B) or IL-2 (7.5–30 U/ml) (A). Proliferation was measured as [\text{H}]thymidine incorporation after 72 h. Data are shown as mean counts per minute±SEM.} \]

\[ \text{Figure 5. Inhibition of purified IL-1 activity by supernatants from RA PBMC (triangles) and SFMC (circles). Open symbols represent supernatants from unstimulated cultures and closed symbols represent supernatants from OKT3 (1 ng/ml)-stimulated cultures. Various dilutions of culture supernatants were mixed with 10 U/ml of purified IL-1, incubated for 2 h at 37°C, and then assayed on mouse thymocytes. Results are expressed as percent inhibition of the activity of 10 U/ml IL-1 on mouse thymocytes.} \]
Table V. RA Culture Supernatants Inhibit IL-1, but not IL-2 Activity

<table>
<thead>
<tr>
<th>Dilution of RA culture supernatant</th>
<th>1/4 cpm</th>
<th>1/16 cpm</th>
<th>1/64 cpm</th>
<th>1/206 cpm</th>
<th>1/1,024 cpm</th>
<th>1/4,096 cpm</th>
</tr>
</thead>
<tbody>
<tr>
<td>IL-1</td>
<td>16,384±1,592</td>
<td>1,386±283</td>
<td>1,945±342</td>
<td>2,468±382</td>
<td>7,452±688</td>
<td>15,384±1,293</td>
</tr>
<tr>
<td>IL-2</td>
<td>24,586±2,134</td>
<td>18,934±1,731</td>
<td>22,834±1,926</td>
<td>26,879±2,031</td>
<td>23,452±2,186</td>
<td>23,987±1,824</td>
</tr>
</tbody>
</table>

Culture supernatants from RA SFMC were added to mouse thymocytes stimulated with 10 U/ml IL-1 and to CTLL-2 cells stimulated with 7.5 U/ml IL-2. Data shown are mean counts per minute±SEM of triplicates from four experiments.

Table VI. Inhibition of Proliferation in Normal PBMC by RA SFMC Supernatants Can Be Overcome by IL-1

<table>
<thead>
<tr>
<th>Percent (vol/vol) of RA supernate</th>
<th>OKT3 IL-1 (U/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Control</td>
</tr>
<tr>
<td></td>
<td>cpm</td>
</tr>
<tr>
<td>0</td>
<td>1,438±312</td>
</tr>
<tr>
<td>25</td>
<td>1,234±286</td>
</tr>
<tr>
<td>6.2</td>
<td>1,380±344</td>
</tr>
<tr>
<td>1.6</td>
<td>1,552±368</td>
</tr>
<tr>
<td>0.4</td>
<td>1,636±402</td>
</tr>
</tbody>
</table>

PBMC from normal donors were cultured in medium alone, or were stimulated with OKT3 (1 ng/ml) with simultaneous addition of 0.4–25% of RA SFMC culture supernatants and/or 0–32 U/ml of purified IL-1. Proliferation was assessed by [3H]thymidine incorporated after 72 h. Data are shown as mean counts per minute±SEM of two experiments.

Table VII. Cellular Origin of IL-1 Inhibitor in SFMC

<table>
<thead>
<tr>
<th>Dilution of culture supernatant</th>
<th>SFMC cpm</th>
<th>1/4 1,068±264</th>
<th>1/16 1,820±318</th>
<th>1/64 3,644±428</th>
<th>1/206 8,937±756</th>
<th>1/1,024 15,438±1,036</th>
<th>1/4,096 17,994±1,628</th>
</tr>
</thead>
<tbody>
<tr>
<td>Monocyte-depleted SFMC (Mφ*)</td>
<td>16,386±1,384</td>
<td>18,243±2,044</td>
<td>17,483±1,686</td>
<td>20,344±1,932</td>
<td>19,328±1,757</td>
<td>19,892±1,863</td>
<td></td>
</tr>
<tr>
<td>Monocytes isolated by adherence</td>
<td>1,216±304</td>
<td>1,436±296</td>
<td>2,038±343</td>
<td>5,489±492</td>
<td>11,398±946</td>
<td>18,368±1,456</td>
<td></td>
</tr>
</tbody>
</table>

SFMC, monocyte-depleted SFMC (Mφ*), 2 × 10⁶/ml, and monocytes isolated by adherence from SFMC (Mφ*), 2 × 10⁶/ml were cultured for 48 h and supernatants were tested for IL-1-inhibitor activity against 10 U of a standard IL-1 preparation which gave 18,324±1,626 cpm in the mouse thymocyte assay over a background of 1,268±242 cpm. Data are shown as mean counts per minute±SEM of [3H]thymidine incorporation after 48 h.

Response in SFMC is predominantly a monocyte function. Furthermore, a simple numerical difference in monocytes does not appear to account for the poor mitogen response in SFMC.

Discussion

The present investigation has analyzed the basis for the defective mitogen responses of RA SFMC with anti-CD3 antibodies. In nearly all cases, the OKT3-induced proliferation of SFMC was depressed severely and indeed was significantly lower than that of PBMC in paired samples. To analyze the hyporesponsiveness of SFMC, the sequence of activation events leading to T cell mitogenesis with anti-CD3 antibodies was dissected. OKT3-stimulated SFMC released minimal IL-1 and IL-2 activity. The addition of purified IL-1 to OKT3-stimulated SFMC markedly increased IL-2 secretion, suggesting that the lack of IL-2 was the consequence of deficient IL-1. IL-2 receptors were expressed on a large proportion of SFMC without prior in vitro stimulation, consistent with their activated state (27–30). The addition of recombinant IL-2 to SFMC triggered modest proliferation demonstrating that the IL-2 receptors were functional. Little increase in IL-2 receptors occurred after stimulation of SFMC with OKT3 alone. Only the simultaneous addition of either IL-1 or IL-2 together with OKT3 induced a major increase in IL-2 receptors in SFMC; both also increased proliferation. The recent finding that purified IL-1 induces IL-2 receptor expression in PBMC
incubated with immobilized anti-T3 antibody (64.1) is in accord with these results (13).

In control experiments with PBMC, IL-2 receptor expression and/or proliferation increased very little after addition of exogenous IL-1 or IL-2, although they did significantly so with SFMC. Mixing experiments with purified monocytes and lymphocytes from blood and synovial fluid showed that synovial fluid lymphocytes gave a much higher proliferative response in the presence of autologous blood monocytes. Conversely, blood lymphocytes gave a low response in the presence of autologous synovial monocytes. In other experiments, synovial fluid monocytes were shown to be a rich source and blood monocytes a weak source of an IL-1 inhibitor. Collectively, these results strongly suggest that the defective proliferative and IL-2 secretory responses of OKT3-stimulated SFMC from RA patients are largely attributable to a deficiency in functional IL-1 activity.

The importance of IL-1 in anti-CD3 mitogenesis is demonstrated by several experiments that will be reported separately (Lotz, M., J. Bentine, and C. D. Tsoukas, unpublished data). Thus, the addition of antibody against IL-1 to OKT3 stimulated normal PBMC reduces the proliferative response. Furthermore, IL-1 is not detectable in Leu 4-stimulated PBMC of Leu 4 non-responders. Finally, soluble OKT3 did not induce IL-1 from isolated monocytes, but did so when both monocytes and T cells were present, indicating that IL-1 release after OKT3 is the result of a macrophage-T cell interaction and not due simply to a contaminant in the antibody preparation.

We were able to explain the low IL-1 activity in RA cultures on the basis of a specific IL-1 inhibitor. The IL-1 inhibitor was detected in large quantities in SFMC cultures. Its action was selective for IL-1, in that it neutralized neither purified nor recombinant human IL-2. The inhibitor(s) was not a prostaglandin, nor were prostaglandins apparently involved in its induction. The inhibitor was not nonspecifically toxic to mouse thymocytes. SFMC culture supernatants with IL-1 inhibitory activity decreased OKT3-induced proliferation of normal PBMC. This effect was overcome by purified exogenous IL-1.

Fractionation of SFMC culture supernatants on phosphocellulose allowed separation of biologically active IL-1 from the IL-1 inhibitor. Several factors that neutralize IL-1 have been described. Epstein-Barr virus-transformed B cell lines released a 95-kD protein that blocks IL-1 effects on mature T lymphocytes and thymocytes (31). We have also seen an IL-1 inhibitor in the supernatants of RA PBMC infected in vitro with Epstein-Barr virus (32). During endotoxin fever, human serum contains a factor that inhibits IL-1 effects on mouse thymocytes (33). Furthermore, in the urine of febrile patients a 20-40-kD glycoprotein was identified which specifically blocks IL-1-induced thymocyte proliferation (34). Synovial fluid also contains a factor that masks IL-1 activity, but this has not been characterized (35). Arend et al. (36) induced the production of a 22-kD IL-1 inhibitor by culturing human monocytes on adherent immune complexes or antibodies. Cytomegalovirus infection of monocytes and a monocytic cell line caused release of an IL-1-specific inhibitor (37). Thus, a variety of IL-1 inhibitors have been described in vivo and in vitro. To ascertain the biochemical and immunologic relationships of the different IL-1 inhibitors will require their purification, the preparation of monospecific antibodies to them, and characterization of their genetic sources.

**Table VIII. Mixing of Lymphocytes and Monocytes from Blood and Synovial Fluid**

<table>
<thead>
<tr>
<th></th>
<th>PBMC</th>
<th>SFMC</th>
<th>M⁺⁺PBMC</th>
<th>M⁺⁺SFMC</th>
<th>M⁺⁺PBMC +10% PBMO</th>
<th>M⁺⁺PBMC +10% SFMO</th>
<th>M⁺⁺SFMC +10% PBMO</th>
<th>M⁺⁺SFMC +10% SFMO</th>
</tr>
</thead>
<tbody>
<tr>
<td>cpm</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Exp. 1</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Contr.</td>
<td>1,154</td>
<td>842</td>
<td>1,892</td>
<td>2,689</td>
<td>1,783</td>
<td>1,139</td>
<td>1,832</td>
<td>3,044</td>
</tr>
<tr>
<td>OKT3</td>
<td>50,384</td>
<td>6,843</td>
<td>2,684</td>
<td>3,246</td>
<td>44,892</td>
<td>8,364</td>
<td>9,024</td>
<td>42,822</td>
</tr>
<tr>
<td>Exp. 2</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Contr.</td>
<td>1,682</td>
<td>1,234</td>
<td>1,434</td>
<td>2,382</td>
<td>1,434</td>
<td>1,580</td>
<td>2,034</td>
<td>2,643</td>
</tr>
<tr>
<td>OKT3</td>
<td>66,493</td>
<td>10,493</td>
<td>2,281</td>
<td>2,883</td>
<td>57,983</td>
<td>14,394</td>
<td>11,386</td>
<td>36,587</td>
</tr>
<tr>
<td>Exp. 3</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Contr.</td>
<td>1,904</td>
<td>2,638</td>
<td>1,428</td>
<td>3,022</td>
<td>1,228</td>
<td>1,779</td>
<td>2,976</td>
<td>2,588</td>
</tr>
<tr>
<td>OKT3</td>
<td>83,685</td>
<td>13,892</td>
<td>1,139</td>
<td>2,756</td>
<td>5,364</td>
<td>6,824</td>
<td>4,582</td>
<td>52,893</td>
</tr>
</tbody>
</table>

PBMC or SFMC from three RA patients were depleted of monocytes (M⁺⁺) and recombined with 10% autologous monocytes from the same source or mixed in a crisscross manner. Proliferation was measured as [³H]thymidine uptake after 72 h. Data are shown as mean counts per minute from triplicates.

Response to Anti-CD3 Antibodies in Rheumatoid Arthritis 719
The prominent feature of the IL-1 inhibitors is that they appear regularly during inflammatory responses induced by diverse stimuli. Perhaps immune complexes or a viral agent represent the eventual stimulus for IL-1 inhibitor production in all cases. In this fashion, products of an inflammatory response may control T lymphocyte activation and lymphokine production via a normal feedback mechanism.

In RA patients, lymphocyte proliferation and lymphokine secretion in the synovial fluid may be dampened by the IL-1 inhibitor(s). However, it remains unclear whether this is a deleterious or beneficial phenomenon. Certainly the inhibitor(s) should impair T cell-mediated inflammatory responses. At the same time the resultant T cell immunodeficiency might facilitate the persistence of inciting agents and release autoantibody-secreting B cells from regulatory control. An analysis of the effects of the IL-1 inhibitor(s) on T cell-regulated autoantibody responses, and on the immune response to environmental pathogens, warrants further detailed study.

We do not know whether there is anything specific about the IL-1 inhibitor for RA synovitis as compared with other synovitic disease. We do know, however, that the appearance of an IL-1 inhibitor in the supernatants of Epstein-Barr virus-infected PBMC is not limited to RA, but may occur in other autoimmune diseases, although not in normal controls (38).

Acknowledgments

We gratefully acknowledge the BCR Word Processing Center for preparing the manuscript for this article.

Funding for this research was provided in part by grants AM-21175, AM-02144, AM-25443, and RR08333 AI-15614 from the National Institutes of Health and grants from the Rheumatic Diseases Research Foundation, the Lilly Research Laboratories, and the Deutsche Forschungsgemeinschaft (Lo 3181/1).

References


