Syndrome of Apparent Mineralocorticoid Excess
A Defect in the Cortisol–Cortisone Shuttle

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Abstract

The first adult case of 11β-hydroxysteroid dehydrogenase (11β-OHSD) deficiency is described. The impaired conversion of cortisol to cortisone (indicated by urinary cortisol and cortisone metabolites and failure to metabolize 11α-[3H]cortisol to [3H]H2O), was associated with hypertension, hypokalemia, and suppression of the renin-angiotensin-aldosterone system. When established on a fixed Na+/K+ intake, dexamethasone, given orally, produced a natriuresis and potassium retention. Plasma renin activity became detectable. When hydrocortisone (10 mg daily s. c. for 4 d) was added, there was marked Na+ retention, a kaliuresis (urinary Na+/K+ falling from 1.2 to 0.15), with suppression of plasma renin activity and an increase in blood pressure. These changes were also seen with the subject on no treatment. Conversion of cortisone to cortisol was not affected.

These results suggest that cortisol acts as a potent mineralocorticoid in 11β-OHSD deficiency. The major site for the oxidation of cortisol to cortisone is the kidney. In this patient congenital deficiency of 11β-OHSD results in high intrarenal cortisol levels which then act on renal type I mineralocorticoid receptors. This condition can be treated with dexamethasone, which suppresses cortisol secretion and binds to the type II glucocorticoid receptor.

We suggest that 11β-OHSD exerts a critical paracrine role in determining the specificity of the type I receptor. In the normal state cortisol is converted by 11β-OHSD to cortisone which thus allows aldosterone to bind preferentially to the type I receptors in the kidney and gut. In this patient deficiency of 11β-OHSD results in high intrarenal cortisol concentrations that then bind to the type I receptor.

Introduction

Impaired conversion of cortisol (compound F) to cortisone (compound E) has been associated with low renin, low aldosterone hypertension with hypokalemia in children (known as the syndrome of “apparent mineralocorticoid excess”) (1–5).

The interconversion of cortisol and cortisone has been considered to be carried out by 11β-hydroxysteroid dehydrogenase (11β-OHSD), a microsomal enzyme system. Recent evidence suggests that the interconversion is a result of the coordinated action of two independent enzymes, a 11β-hydroxysteroid dehydrogenase and a 11-oxo-steroid reductase (6, 7). For the purpose of this study the familiar term 11β-OHSD will be used to describe the interconversion.

It was the seminal work by Ulick et al. (5) in two hypertensive children, which first demonstrated that there was a defect in the conversion of cortisol to cortisone (detailed further in publications by New and collaborators (2, 8–10)). Since then the syndrome of apparent mineralocorticoid excess has been detected in about 15 children. Five cases have been documented by Shackleton and co-workers (4, 11), with individual cases from Fiselier et al. (12), Honour et al. (3), and Harinck et al. (13), and most recently three further cases by Monder, New, and co-workers (14). Other cases have been documented but not published.

The characteristic urinary steroid metabolite profile is well described (4, 14). Patients have limited ability to metabolize cortisol to cortisone but can carry out the reverse reaction. As a result the urinary 11β-hydroxy metabolites of cortisol are elevated (cortols, tetrahydrocortisol [THF] and C-19 steroids) while the 11-oxo metabolites (cortolones and tetrahydrocortisone [TNE]) are diminished. In addition there appears to be an alteration in the allo-THF (5αTHF) to THF (5βTHF) ratio with 5αTHF formed in preference to 5βTHF in most cases. Whether this reflects a true deficiency of 5β-reductase has not yet been answered (14).

Defective peripheral cortisol metabolism results in a prolonged plasma cortisol half-life, a reduced daily secretion rate, and normal plasma levels. Many of the cases reported have elevated urinary free cortisol levels (4, 8, 13).

Methods

Case report

G.B., a Caucasian male, presented aged 21 with a 2-wk history of altered vision. During the preceding 2 yr he had sustained three episodes of tonsillitis; each time this was associated with carpopedal spasm and perioral paresthesia. He had had polydipsia and nocturia for several years. There was no other past medical history and he had never had his blood pressure recorded previously. He did not smoke, consume alcohol, or take any regular medications. Consumption of exogenous mineralocorticoids such as liquorice was denied. A strong family history of hypertension was noted (see below).

Examination revealed a blood pressure of 200/145 mmHg (right arm), 192/140 mmHg (left arm), sinus rhythm with no radial-femoral
delay or vascular bruits. The apex beat was positioned normally but heaving in quality. The second heart sound was loud. Fundoscopy showed grade III hypertensive changes. Height was 173 cm; weight 53 kg; secondary sexual characteristics were normal. Chest X-ray was normal; ECG showed voltage criteria for left ventricular hypertrophy.

During an intravenous urogram (IVU) 24 h after admission he sustained two episodes of ventricular fibrillation treated with D.C. shock. Biochemistry showed a plasma Na⁺ 148 mmol·liter⁻¹ (meq/liter), K⁺ 1.7 mmol·liter⁻¹, bicarbonate 32 mmol·liter⁻¹. Creatinine clearance and 24-h urinary metanephrines were normal. The IVU showed mild bilateral nephrocalcinosis with two renal cysts confirmed on subsequent ultrasonography and computerized tomography. Serum and 24-h urinary calcium were normal. Supine plasma renin activity and aldosterone levels were suppressed at 0.1 ng ml⁻¹ h⁻¹ (reference range 0.5–1.5) and < 100 pmol·liter⁻¹ (reference range 150–500 pmol·liter⁻¹), respectively.

Over the next few months control of his blood pressure and hypokalemia was poor despite a variety of antihypertensive medications (atenolol 300 mg, captopril 150 mg, hydralazine 200 mg, amiloride 10 mg, prazosin 25 mg, nifedipine retard 40 mg, and Sando K 48 mmol per d). He was then referred to this hospital and a diagnosis of 11β-OHSD deficiency made from urinary steroid metabolites as measured by gas chromatography/mass spectrometry (Table 1).

The elevated THF + allo THF/THE ratio is diagnostic of 11β-OHSD deficiency, the elevated 5α THF (allo THF)/5α THF ratio in keeping with other reported cases.

GB was then admitted to the Metabolic Unit for further studies; local Ethical Committee and ARSC approval was obtained for administration of radioisotopes.

**Procedures**

The patient was weighed at 0830 hours each morning after emptying his bladder. Blood pressure was recorded automatically using a commercially available automatic Copal sphygmonanometer (Andrew Stephens, Blackpool, England), which was checked against a random zero sphygmonanometer at weekly intervals.

During all metabolic studies BP was recorded in the supine position at 5-min intervals from 1400 hours to 1500 hours each day. When analyzing blood pressure recordings during the metabolic studies, statistical analysis was performed on consecutive readings from three days, i.e., at least 30 systolic and diastolic recordings, using Student's paired t test.

**Assays**

Plasma and urinary free cortisol were measured by radioimmunoassay, using the commercially available Amerlex kit (Amersham International) (15). Corticosterone (16), urinary aldosterone (17), and plasma renin activity (PRA) (18) were measured by modifications of previously reported methods. Plasma aldosterone was measured by the direct radioimmunoassay of Al-Dujaili and Edwards (19), except that (a) sample volume has been reduced to 50 µl with a resultant fall in the nonspecific binding, improving assay precision and (b) charcoal separation rather than a double antibody separation has considerably improved the sensitivity at lower values. The lowest detection limit for the assay is now 100 pmol l⁻¹. Plasma and urinary electrolyte and creatinine were measured by an ion selective method using an Astra 4.

**Deoxycorticosterone (DOC) assay.** Plasma was extracted with 5 vol of freshly distilled dichloromethane, the extract washed with 0.1 M NaOH and with water before aliquots (equivalent to 100 µl plasma) were evaporated to dryness under nitrogen at 45°C.

The assay used rabbit antiserum (R7-6; St. Bartholomew's Hospital) and tritium-labeled (Amersham International). Because of cross-reactivity with testosterone and progesterone, extracts were purified further by HPLC using a μBondapak C₈ reverse-phase column with 60:40 (vol/vol) methanol/water as mobile phase. The appropriate fraction of eluate was collected, methanol evaporated off under nitrogen and the aqueous solution back extracted with dichloromethane. This extract was evaporated to dryness under nitrogen, dissolved in 100 µl assay diluent (0.05 M phosphate buffer, pH 7, containing 0.1% BSA), [³H]-DOC (~ 2,000 cpm) and antiserum in 100 µl assay diluent added; final antiserum dilution 1:20,000. After overnight incubation at 4°C, bound and unbound DOC were separated using dextran-coated charcoal and supernatant (following centrifugation) counted in Scintran T (BDH Chemicals, Poole, England).

Aliquots from 24 h urine collections were stored at −20°C and the urinary steroid metabolites analyzed using gas chromatography and gas chromatography/mass spectrometry using previously reported methods (4, 20).

| Table 1. Blood Pressures and Urinary Steroid Metabolites in G.B., His Parents, and Siblings as Compared to 22 Normal Controls Aged 18–55 (16 Males) |
|------------------|--------|--------|--------|--------|--------|--------|--------|
|                  | G.B.   | S.B.   | P.B.   | N.D.   | T.B.   | E.B.   |
| Blood pressure (mmHg) |       |        |        |        |        |        |
| Blood pressure reference range (mean±SD) | 235/125 | 166/102 | 165/92 | 125/82 | 134/84 | 135/85 |

<table>
<thead>
<tr>
<th>Steroid</th>
<th>G.B.</th>
<th>S.B.</th>
<th>P.B.</th>
<th>N.D.</th>
<th>T.B.</th>
<th>E.B.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Androsterone</td>
<td>2,068±892</td>
<td>10,176</td>
<td>700</td>
<td>470</td>
<td>1,966</td>
<td>7,018</td>
</tr>
<tr>
<td>Etocholanolone</td>
<td>1,690±874</td>
<td>4,427</td>
<td>1,137</td>
<td>767</td>
<td>2,379</td>
<td>6,606</td>
</tr>
<tr>
<td>11-oxo-etiocholanolone</td>
<td>426±461</td>
<td>537</td>
<td>680</td>
<td>534</td>
<td>1,059</td>
<td>1,257</td>
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<tr>
<td>11β-hydroxyandrostenedone</td>
<td>1,058±875</td>
<td>1,555</td>
<td>852</td>
<td>799</td>
<td>1,821</td>
<td>3,510</td>
</tr>
<tr>
<td>11β-hydroxyetiocholanolone</td>
<td>396±272</td>
<td>1,075</td>
<td>309</td>
<td>686</td>
<td>1,037</td>
<td>1,270</td>
</tr>
<tr>
<td>(THE)</td>
<td>2,589±1,292</td>
<td>537</td>
<td>2,435</td>
<td>4,414</td>
<td>2,545</td>
<td>4,619</td>
</tr>
<tr>
<td>(THF)</td>
<td>1,469±585</td>
<td>2,472</td>
<td>2,076</td>
<td>3,019</td>
<td>1,380</td>
<td>3,679</td>
</tr>
<tr>
<td>Allo tetrahydrocortisol (allo-THF)</td>
<td>1,033±525</td>
<td>4,819</td>
<td>1,019</td>
<td>1,860</td>
<td>839</td>
<td>3,176</td>
</tr>
<tr>
<td>α-cortolone</td>
<td>913±386</td>
<td>182</td>
<td>513</td>
<td>778</td>
<td>622</td>
<td>874</td>
</tr>
<tr>
<td>α-cortol</td>
<td>303±100</td>
<td>422</td>
<td>342</td>
<td>338</td>
<td>310</td>
<td>690</td>
</tr>
<tr>
<td>δ-cortol</td>
<td>920±414</td>
<td>537</td>
<td>241</td>
<td>916</td>
<td>310</td>
<td>765</td>
</tr>
<tr>
<td>THF + allo THF/THE ratio</td>
<td>0.97±0.30</td>
<td>13.5</td>
<td>1.27</td>
<td>1.10</td>
<td>0.87</td>
<td>1.60</td>
</tr>
</tbody>
</table>

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Synthesis and metabolism of 11α-[3H]cortisol. 11α-[3H]Cortisol was prepared as described by Hellman et al. (21) but with modifications to improve the radiochemical yield and define the specific activity. The main changes were a threefold increase in the ratio of cortisol-3, 20-bis-butylenyl ester to sodium [3H]borohydride and purification of the final product by thin layer chromatography on silica gel GF254 (20 × 20 cm × 0.25 mm) in chloroform-methanol (9:1 by vol). The cortisol band was located under ultraviolet light, scraped from the plate and eluted with ethanol (1 × 5 ml, 6 × 1 ml). The cortisol concentration in recovered solution (total volume 9 ml) determined by quantitative high pressure liquid chromatography (μBondapak C18; mobile phase methanol-water 1:1 by volume; calibrated against a standard cortisol solution) was 1.48 mg/ml, and the radioactive concentration determined by scintillation counting was 5.91 MBq/ml. The specific activity was therefore 1.44 GBq/mmol. No cortisol or 11-epicortisol was detected in the purified product (lower limit of detection of either compound was 0.5%). The ethanol solution was passed through a 0.2-μm filter, aliquoted into sterile vials (1.19 MBq/vial) and stored at 4°C.

The measured specific activity was substantially lower than that of the sodium borohydride (2.6 GBq per hydride) due to the well-known primary isotope effect.

With G.B. fasted overnight diuresis was established by an oral water load. When a constant urinary flow rate was achieved, 1.19 MBq 11α-[3H]cortisol in 0.36 ml ethanol i.v. was injected as a bolus in 15 ml water. Urine was collected at 15-min intervals, blood at 15-min intervals during the first 60 min, then every 30 min for 120 min. The samples were counted for total 3H and for [3H]H2O following sublimation using a (Tricarb 4330 series; Packard Instruments, Inc., Downers Grove, IL) liquid scintillation beta counter after correction for quench. Cocktail T (BDH Chemicals) was used as scintillant. Subtraction of [3H]H2O from total 3H gave a value in counts/min representing activity of the titrated steroid. Separate experiments were performed with the HPLC system outlined above to demonstrate that the residual radioactivity after sublimation resided in 11α-[3H]cortisol. Plots of log 10 11α-[3H]cortisol versus time were made and the slope analyzed by linear regression. Only the elimination phase (time points 45–120 min) was studied and expressed as a half-life.

Subtraction potential difference (p.d.). As an index of in vivo mineralocorticoid activity subtraction p.d. was measured using an Adrenosonde (Kontron, Vienna, Austria) measuring the p.d. across a steroid insensitive mucosa (buccal mucosa) and subtracting this from a steroid sensitive mucosa (rectum) (22). Subtraction of buccal from rectal p.d. eliminates the nonsteroidal influences of rectal p.d.

The reference electrode was placed on the forehead over an intracutaneous injection of 0.3 ml 0.9% saline to eliminate skin p.d. The potential difference was read with the probe electrode placed on either the buccal mucosa 3 cm from the oral orifice or on the rectal mucosa 8 cm from the anal margin, 20 ml of 0.9% saline having been injected into the rectum. This was performed on every second day during the metabolic studies at 1000 hours. Two recordings (which never differed by more than 4 mV) were made at 20-min intervals and the mean recorded. The normal subtraction p.d. is −5 to +25 mV, with values of 40–80 mV occurring in states of mineralocorticoid excess such as primary aldosteronism (23).

Metabolic balance studies

To investigate the role of cortisol in producing the hypertension and hypokalemia in our patient, metabolic balance studies were performed on a fixed Na+ "K+" diet (118 mmol Na+, 78 mmol K+/day). In balance it was possible to withdraw all drugs except for nifedipine, amiloride, and Sando K (48 mmol) with no ill effects apart from mild elevation of blood pressure. The metabolic balance was divided into several studies.

Study 1 involved baseline investigation of the circadian rhythm of cortisol, measurement of other adrenal and gonadal steroids, ACTH and the oral administration of dexamethasone, initially 0.5 mg 6 hourly for 48 h, then 1.5 mg/d.

Study 2 was performed with the patient in balance taking 1.5 mg dexamethasone daily. Hydrocortisone (10 mg) was then given subcutaneously via a pulsatile NIMR Millhill pump delivering steroid at 15 min pulses. 10 mg was chosen as this was the sum of free and conjugated cortisol metabolites excreted in the urine in a 24-h period as determined by GC/MS.

Study 3 was performed with the patient on no treatment for 8 d to reproduce the syndrome.

Study 4. Finally cortisone acetate was given orally (25 mg at 0900 hours, 12.5 mg at 1700 hours).

In study 5 we examined the metabolism of 11α-[3H]cortisol given intravenously as a bolus.

**Hemodynamic assessment**

With the patient supine on no treatment and fasted overnight, a thermodilution flow directed balloon catheter was sited in the pulmonary artery via the femoral vein and its position confirmed by fluoroscopy. Right atrial pressure (RAP) and cardiac output (CO) were measured, the latter by using a thermodilution technique. Blood pressure (BP) was recorded in the right arm during the procedure using a mercury sphygmomanometer. Systemic vascular resistance (SVR) was then calculated using the formula SVR = BP-RA P × 80/CO (dyn s-1 cm-5).

**Results**

**Family data**

Blood pressure recordings were made in the supine position between 0830–0900 hours using a Copal automatic sphygmomanometer in G.B.'s parents and siblings and 24 h urinary steroid metabolites measured (Table I).

G.B.'s father, S.B., had a 20-yr history of hypertension controlled for the last four years on hydralazine and a beta blocker/diuretic combination. PB, his mother, had hypertension diagnosed at the age of 53 and had been treated with diuretics only. Under supervision all medication was withdrawn from S.B. and P.B. for 4 wk, and they were investigated further. S.B. had normal plasma electrolytes (Na+ 137 mmol·liter-1, K+ 3.9 mmol·liter-1, bicarbonate 28 mmol·liter-1), normal urinary free cortisol (155 nmol·liter-1), supine PRA (0.7 ng ml-1 h-1) and urinary aldosterone (23 nmol/24 h). However G.B.'s mother, PB, had a mild hypokalemic alkalosis (Na+ 137 mmol·liter-1, K+ 3.3 mmol·liter-1, bicarbonate 33 mmol·liter-1) associated with suppression of the renin angiotensin aldosterone system (supine PRA 0.3 ng

**Table II. Plasma Steroid, ACTH, and Catecholamine Levels**

<table>
<thead>
<tr>
<th>Day</th>
<th>0900 hours (n range 180-700)</th>
<th>2400 hours (n range &lt;180)</th>
<th>0900 hours (n range 10-80)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>513</td>
<td>156</td>
<td>35</td>
</tr>
<tr>
<td>2</td>
<td>519</td>
<td>290</td>
<td>32</td>
</tr>
<tr>
<td>3</td>
<td>601</td>
<td>217</td>
<td>45</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th></th>
<th>Cortisol nmol·liter-1</th>
<th>ACTH ng·liter-1</th>
</tr>
</thead>
<tbody>
<tr>
<td>Norepinephrine*</td>
<td>1.3 nmol·liter-1</td>
<td>(normal &lt;5)</td>
</tr>
<tr>
<td>Epinephrine*</td>
<td>0.06 nmol·liter-1</td>
<td>(normal &lt;1)</td>
</tr>
<tr>
<td>Testosterone*</td>
<td>24 nmol·liter-1</td>
<td>(normal 10-30)</td>
</tr>
<tr>
<td>Androstenedione*</td>
<td>3.5 nmol·liter-1</td>
<td>(normal 2-11)</td>
</tr>
<tr>
<td>Dehydroepiandrosterone sulfate*</td>
<td>11 μmol·liter-1</td>
<td>(normal 2-9)</td>
</tr>
</tbody>
</table>

* Samples taken at 0900 hours on day 1.
Figure 1. Metabolic balance study 1. Urinary free cortisol, urine and plasma electrolytes, blood pressure, plasma aldosterone, renin activity, subtraction p.d. and weight are shown during acute (2 mg/d for 48 h) and chronic (1.5 mg/d) dexamethasone therapy. The reference ranges for plasma Na⁺, K⁺ and renin activity are indicated. The mean±SD of 10 readings for systolic and diastolic blood pressure is charted.
ml\(^{-1}\) h\(^{-1}\), urinary aldosterone 5 nmol/24 h). Despite this metabolic disturbance, urinary free cortisol was normal (272 nmol/24 h) as were her urinary steroid metabolites.

As shown, all the siblings had normal blood pressure but their steroid excretion patterns were not entirely normal. Both brothers in addition to G.B. showed high urinary levels of androstosterone, etiocholanolone, and its \(11\beta\)- and \(\alpha\)-o xo metabolites (Table I). In the 16 control males (mean age 30.7 yr) the mean androstosterone and etiocholanolone excretions were 2,292±1131 (SD) (range 1,016-4,856) and 2,430±744 (range 1,265-3,772) \(\mu\)g/24 h, respectively. One brother, T.B., had a marginally elevated THF + allo THF/THF ratio.

**Study 1**

**Baseline investigations and dexamethasone suppression.** Table II shows that G.B. had a normal cortisol circadian rhythm, normal ACTH levels, plasma catecholamines, testosterone, and androstenedione. Plasma dehydroepiandrosterone sulphate was slightly elevated.

When in balance on his fixed Na\(^+\)/K\(^+\) diet and treated with nifedipine retard 20 mg twice daily, amiloride 5 mg twice daily, and potassium supplements (48 mmol/24 h), dexamethasone 0.5 mg 6 hourly was given for 48 h. As shown in Fig. 1 urinary free cortisol suppressed from 628 nmol/24 h to <30 nmol/24 h (reference range 80-450 nmol/24 h). There was a marked natriuresis with potassium retention, urinary Na\(^+\)/K\(^+\) ratio rising from 1.2 to 3.3. Plasma electrolytes showed a corresponding change, Na\(^+\) falling from 143 to 136 mmol·liter\(^{-1}\), K\(^+\) rising from 3.5 to 4.5 mmol·liter\(^{-1}\). Table III shows the response of plasma cortisol, renin activity, deoxycorticosterone, aldosterone, corticosterone, weight and subtraction potential difference during this and subsequent studies. Baseline values were obtained from the day immediately prior to the first dose of dexamethasone. As shown, PRA initially suppressed despite nifedipine and a low dose of amiloride became detectable. Plasma aldosterone over this short period of time remained low as expected following prolongation of suppression of the zona glomerulosa. When the urinary free cortisol levels reverted to predexamethasone values maintenance dexamethasone was recommenced (1.0 mg 2300 hours and 0.5 mg 0900 hours). Once again this produced a natriuresis and potassium retention, indeed the combination of dexamethasone and amiloride resulted in marked hyperkalemia (K\(^+\) peak 6.8 mmol·liter\(^{-1}\)) and this coincided with and was possibly the cause of the rise in plasma aldosterone. The hyperkalemia was treated by parenteral hydrocortisone (100 mg i.m.), frusemide (40 mg i.v.) and withdrawal of K\(^+\) and amiloride (Fig. 1). Nifedipine was also stopped. When reestablished in balance on dexamethasone 1.5 mg daily and off all other drugs, G.B. maintained a normal plasma K\(^+\), with only a slight rise in blood pressure. On dexamethasone alone, subtraction p.d. remained in the normal range at 3-10 \(\mu\)V, as did PRA (0.5-1.5 ng ml\(^{-1}\) h\(^{-1}\)). The zona glomerulosa showed some signs of recovery immediately prior to study 2 (plasma aldosterone 140 pmol·liter\(^{-1}\)).

**Study 2**

**Hydrocortisone administration.** Administration of hydrocortisone (10 mg/24 h for 4 d) when G.B. was in metabolic balance and only on dexamethasone 1.5 mg/24 h produced urinary free cortisol levels similar to those on no treatment. As shown in Fig. 2 this caused marked Na\(^+\) retention and a kaliuresis with a fall in urinary Na\(^+\)/K\(^+\) ratio from 1.2 to 0.15. Plasma electrolytes showed corresponding changes, Na\(^+\) rising from 136 to 145 mmol·liter\(^{-1}\), K\(^+\) falling from 4.5 to 3.3 mmol·liter\(^{-1}\). As expected there was weight gain and suppression of PRA and plasma aldosterone (Table III). Subtraction p.d. rose to 59 \(\mu\)V, indicative of mineralocorticoid excess. In addition there was a highly significant rise in both systolic and diastolic blood pressure (Table IV).

**Study 3**

**Withdrawal of dexamethasone.** When back in Na\(^+\)/K\(^+\) balance on dexamethasone alone, dexamethasone was discontinued for 8 d (Fig. 3). Urinary free cortisol, which on dexamethasone was <30 nmol/24 h rose to 450 nmol/24 h. There was a kaliuresis and Na\(^+\) retention with urinary Na\(^+\)/K\(^+\) ratio falling to 0.15. Plasma K\(^+\) fell to 2.9 mmol·liter\(^{-1}\) and Na\(^+\) rose to 146 mmol·liter\(^{-1}\). PRA suppressed once more as did plasma aldosterone (which had risen to 210 pmol·liter\(^{-1}\) prior to withdrawal of treatment). Similarly subtraction p.d. was elevated at 48 \(\mu\)V, a pattern identical to that seen following cortisol administration (Table III). In addition there was a significant rise in diastolic blood pressure (Table IV).

These changes were then reversed on restarting dexamethasone; 1.25 mg dexamethasone/d (0.75 mg 2300 hours, 0.5 mg 0900 hours) was sufficient to suppress urinary free cortisol to <30 nmol/24 h and prevent hypokalemia.

**Table III. Plasma Cortisol (F), Renin Activity (PRA), DOC, Corticosterone (B), Aldosterone (Aldo).**

<table>
<thead>
<tr>
<th>Reference range</th>
<th>Baseline</th>
<th>Acute Dexa suppression (2 mg/d for 48 h)</th>
<th>Dexa 1.5 mg/d</th>
<th>Dexamethasone 10 mg/d</th>
<th>No treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td>09.00 F</td>
<td>300-700 nmol liter(^{-1})</td>
<td>601</td>
<td>39</td>
<td>30</td>
<td>239</td>
</tr>
<tr>
<td>09.00 B</td>
<td>1.4-15.6 nmol liter(^{-1})</td>
<td>1.2</td>
<td>1.0</td>
<td>0.4</td>
<td>0.5</td>
</tr>
<tr>
<td>09.00 DOC</td>
<td>150-450 pmol liter(^{-1})</td>
<td>214</td>
<td>223</td>
<td>181</td>
<td>172</td>
</tr>
<tr>
<td>09.00 PRA (supine)</td>
<td>0.5-1.5 mg AI (\mu)mol h(^{-1})</td>
<td>&lt;0.05</td>
<td>0.39</td>
<td>0.50</td>
<td>&lt;0.05</td>
</tr>
<tr>
<td>09.00 Aldo (supine)</td>
<td>135-500 pmol liter(^{-1})</td>
<td>120</td>
<td>&lt;100</td>
<td>140</td>
<td>&lt;100</td>
</tr>
<tr>
<td>Subtraction p.d.</td>
<td>-5±25 mV</td>
<td>—</td>
<td>8</td>
<td>3</td>
<td>59</td>
</tr>
<tr>
<td>Weight, kg</td>
<td>57.0</td>
<td>56.7</td>
<td>56.6</td>
<td>57.2</td>
<td>57.1</td>
</tr>
</tbody>
</table>

Baseline values were obtained on the day immediately before the first dose of dexamethasone when in metabolic balance. Other values reported refer to the last day of the treatment option described.
Table 2. Metabolic balance data, blood pressure, renin activity and subtraction p.d. for study 2. With GB on dexamethasone 1.5 mg/d, the response to hydrocortisone 10 mg/d for 4 d is shown.

Figure 3. Metabolic balance study 3. Dexamethasone was discontinued for 8 d.
Table IV. Mean Blood Pressure Readings during Metabolic Studies

<table>
<thead>
<tr>
<th>Treatment option</th>
<th>Systolic B.P. mmHg</th>
<th>Diastolic B.P. mmHg</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>mean±SD</td>
<td>mean±SD</td>
</tr>
<tr>
<td>A Dexamethasone</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(1.5 mg/d)</td>
<td>161±7</td>
<td>105±7</td>
</tr>
<tr>
<td>B Dexamethasone</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(1.5 mg/d)</td>
<td>177±8</td>
<td>114±12</td>
</tr>
<tr>
<td>Hydrocortisone</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(10 mg/d)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C No treatment</td>
<td>163±6</td>
<td>112±8</td>
</tr>
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</table>

P value

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<tr>
<th></th>
<th>Systolic</th>
<th>Diastolic</th>
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</thead>
<tbody>
<tr>
<td>A vs. B</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>A vs. C</td>
<td>NS</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>

Study 4
Administration of cortisone acetate. This study was performed 12 m after study 3. During this period blood pressure had been measured twice weekly by G.B. himself and averaged 110/70 mmHg. Blood pressure had been controlled in the interim on dexamethasone 1.25 mg/d, captopril 25 mg b.d., frusemide 40 mg/d. When reassessed at this visit, he had a blood pressure of 115/82 mmHg supine and 102/66 mmHg erect suggesting over treatment with frusemide and captopril. These were discontinued 11 d before the administration of cortisone acetate.

G.B. was once more established on the same Na+/K+ diet, and dexamethasone 1.25 mg/d was continued. When in metabolic balance cortisone acetate 37.5 mg/d (25 mg 0900 hours, 12.5 mg 1700 hours) was given orally for 3 d. This resulted in high urinary free cortisol levels with associated Na+ retention though only a moderate kaliuresis (Fig. 4). Before administration of cortisone there was activation of the renin-angiotensin-aldosterone system (PRA 13.4 ng ml⁻¹ h⁻¹, urinary aldosterone 74.3 nmol/24 h) which was in keeping clinically with over treatment. After 3 d of cortisone acetate PRA fell to 3.4 ng ml⁻¹ h⁻¹, urinary aldosterone to 4 nmol/24 h. Both systolic and diastolic blood pressure rose during this period as shown in Table V.

During the first 24 h of cortisone acetate administration, plasma cortisol, measured throughout the day showed two distinct peaks (720 nmol liter⁻¹ at 1000 hours and 680 nmol liter⁻¹ at 1800 hours) indicating that G.B. is able to convert cortisone to cortisol.

Study 5
Metabolism of 11α-[³H]cortisol. This was performed with G.B. taking dexamethasone 1.25 mg/d only, and off all other medication for over 3 wk. A preliminary study in a normal volunteer showed that dexamethasone therapy did not alter plasma and urinary total ³H and [³H]H₂O levels (data not shown). Similarly in vitro dexamethasone was shown by Lugg et al. to have no effect on 11β-OHSD activity (24).

The half-life of 11α-[³H]cortisol in G.B. is prolonged at 131 min. Eight normal volunteers the mean plasma half-life was 40±5.3 min, which is in agreement with that found by Hellman et al. (21). When 11α-[³H]cortisol is oxidized by 11β-

OHSD equimolar amounts of [³H]H₂O and cortisone are produced. G.B. is unable to carry out this reaction as shown by the failure to produce [³H]H₂O in both his plasma and urine (Figs. 5 and 6).

Hemodynamic results. Heart rate, blood pressure, cardiac output (CO) and systemic vascular resistance (SVR) were measured in the erect and supine position. The mean of three readings recorded at 10-min intervals was taken. In the supine position heart rate was 116 beats/min, mean arterial pressure 149 mmHg. CO elevated at 8.8 l/min (references 4–6), and SVR 1,341 dyn s⁻¹cm⁻² (references 800–1,400). In the erect posture, heart rate rose to 154 beats/min, MAP was 136 mmHg, CO was 4.3 liters/min and SVR elevated at 2,578 dyn s⁻¹cm⁻².
Discussion

It is not surprising that patients with 11β-hydroxysteroid dehydrogenase deficiency with hypertension, hypokalemic alkalosis, and suppressed plasma renin and aldosterone levels should have been described as having the syndrome of apparent mineralocorticoid excess. Extensive studies however, using a variety of mineralocorticoid bioassays have failed to demonstrate any evidence of a circulating mineralocorticoid excess. The cause of the syndrome, which has previously only been described in children, and which is occasionally fatal, remains unclear (1, 3, 5, 12).

Marver reported in 1978 (25) that 5α-dihydrocortisol potentiated the action of aldosterone in an animal model, but although 5α-dihydrcortisol levels were elevated in reported cases these fell at puberty despite continuation of the syndrome (5). When 5α-dihydrcortisol was infused in such patients the syndrome could not be reproduced (10).

Detailed metabolic balance studies by New’s group have indicated the important role of cortisol in 11β-OHSD deficiency. Their work has suggested that in these patients cortisol was acting as a mineralocorticoid and this effect could be blocked by spironolactone (9, 10). They postulated a receptor defect with an abnormal receptor recognizing cortisol as a mineralocorticoid. This syndrome is familial and this hypothesis would presumably require two genetic defects, one relating to the enzyme and the other to the receptor abnormality (assuming that these are not linked).

11β-OHSD occurs in many tissues in man, not only the liver (26) and kidney (27) but also placenta (28), gastrointestinal tract (29), prostate, muscle, lung, and thyroid (27). The “enzyme” has recently been shown to exist as two distinct hydroxy-dehydrogenase and reductase enzymes (6). As glucocorticoids and mineralocorticoids are dependent upon an 11β-hydroxyl group at the C-11 position then 11β-OHSD exerts a critical role in the above tissues in regulating the amount of “active steroid” exposed to that tissue by shuttling between the hydroxyl and keto groups. Activities differ markedly in different tissues; the liver, for instance, is strongly reductive (cortisone-cortisol) and dependent upon NADPH (30), the kidney oxidative (cortisol-cortisone) and dependent upon NADP (27). Alteration in enzyme activity within different tissues may reflect a different enzyme or altered redox potential (hence NADPH/NADP ratio) therein.

This study describes the first adult case of 11β-OHSD deficiency. The metabolic balance studies show that cortisol acted as a potent mineralocorticoid on the kidney (and gastrointestinat tract) causing Na⁺ retention and hypertension with hypokalemia. The profound hypokalemia in addition to causing tetany (as previously seen in other enzymatic forms of mineralocorticoid hypertension, e.g., 17α-hydroxylase deficiency [31]), proved to be life-threatening. Nephrocalcinosis and renal cysts were also seen in the first reported case of Liddle’s syndrome (32), and could possibly also relate to chronic hypokalemia. Dexamethasone was able to reverse the hypokalemia, by suppressing the hypotalamico-pituitary adrenal axis, caused a natriuresis and was associated with a lower, but not normal blood pressure, providing a new therapeutic approach in this patient. Only the conversion of cortisol to cortisone was impaired as shown by his failure to metabolize 11α-[3H]cortisol, and normal production of cortisol following oral cortisone acetate.

Several studies indicate that there is a genetic basis for this syndrome (4, 14). Urinary steroid metabolites in G.B.’s parents on no treatment were normal. The finding of a mild hypokalemic alkalosis with suppression of the renin-angiotensin-aldosterone axis in G.B.’s mother is of considerable interest, and raises the question whether a very subtle abnormality in 11β-OHSD could be responsible for her condition. The urinary steroid excretion values for G.B.’s brothers were notable but we hesitate to term them abnormal. Like G.B., the brothers had evidence of high androgen production through elevated excretions of androstenedione and etiocholanolone. Most notable was the high excretion of 11β-hydroxyandrostenedione, which could reflect increased activity of side-chain cleavage in cortisol metabolism or substantial 11β-hydroxylation of testosterone and androstenedione. Apart from T.B., the THF + alloTHF/THF ratios were normal.

The hemodynamic data indicate a “hyperdynamic circulation” with a high resting cardiac output and normal systemic vascular resistance. This has been described in other patients with “mineralocorticoid excess” states such as primary aldosteronism (33). A sinus tachycardia had been noted since discontinuing beta blockade 12 mo earlier. On adopting the erect
posture the heart rate increased and cardiac output fell by 50%, suggesting a noncompliant left ventricle as seen in ventricular hypertrophy.

Although Srivastava et al., measuring cortisol/cortisone ratios in severe liver and renal disease, suggest that the kidney is a major source of cortisone (34), the only direct evidence as to the contribution made by the kidney comes from Hellman et al. (21). After isotopic studies using [14C]cortisol and 11α-[3H]cortisol they concluded that the kidney was responsible for 10% of the total oxidation of cortisol to cortisone.

The recent cloning of the human mineralocorticoid receptor by Arriza et al. (35) has shown remarkable homology between the human glucocorticoid receptor and the human mineralocorticoid receptor. This receptor appears to have similar affinities in vitro for aldosterone, corticosterone, and cortisol, suggesting that something other than receptor structure is important in determining its specificity. Funder had suggested that this may be renal extravascular cortisol binding globulin (CBG) (36). However in recent studies using 10-d-old rats (which have very low levels of CBG), the in vivo specificity for the type I receptor was maintained despite much higher levels of corticosterone than aldosterone (37). We postulate that the oxidation of cortisol to cortisone (corticosterone to 11-dehydrocortisone in the rat) by renal 11β-OHSD is critical in determining the intrarenal concentration of active glucocorticoid, and hence the specificity of the type I receptor. Deficiency of renal 11β-OHSD allows cortisol to act in preference to aldosterone on the mineralocorticoid receptor causing hypertension and hypokalemia. An indication of the abnormal renal handling of cortisol is the moderate elevation of urinary free cortisol in proportion to total cortisol metabolite excretion in the condition, despite normal plasma cortisol levels and a prolonged cortisol half-life. This, through negative feedback control, lowers ACTH and hence cortisol production. If our hypothesis is true this would suggest that this unique biological experiment has disclosed a previously unknown paracrine mechanism by which the kidney protects itself from the mineralocorticoid action of cortisol.

This syndrome indicates the quite striking results of a congenital disturbance of this cortisol–cortisone shuttle. Our recent studies have shown that the sodium retaining effects of liquorice are due to an acquired inhibition of this mechanism and not to a direct effect on renal type I receptors (38). It remains to be seen if more subtle defects in this mechanism are important in other types of hypertension.

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