Transinhibition of C1 Inhibitor Synthesis in Type I Hereditary Angioneurotic Edema

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Abstract

To ascertain the mechanism for decreased synthesis of C1 inhibitor (Cl INH) in certain patients with the autosomal dominant disorder hereditary angioneurotic edema, we studied expression of Cl INH in fibroblasts in which the mutant and wild type mRNA and protein could be distinguished because of deletion of exon 7 (Δex7). In the HANE Δex7 cells, the amount of wild type mRNA (2.1 kb) was expressed at 52±2% (n = 5) of normal, whereas the mutant mRNA was 17±1% (n = 5) of normal. Rates of synthesis of both wild type and mutant proteins (11±3 and 3±1% of normal, respectively) were lower than predicted from the mRNA levels. There was no evidence of increased Cl INH protein catabolism. These data indicate that there are multiple levels of control of Cl INH synthesis in type I hereditary angioneurotic edema. Pretranslational regulation results in <50% of the mutant truncated 1.9-kb mRNA. In addition, translational regulation results in decreased synthesis of both wild type and mutant Cl INH proteins. These data suggest a transinhibition of wild type Cl INH translation by mutant mRNA and/or protein. (J. Clin. Invest. 1993. 91:1258–1262.)

Key words: hereditary angioneurotic edema • C1 inhibitor • synthesis • translation • transinhibition

Introduction

Examples in which the products of a eukaryotic gene effect translational regulation of its expression have been recognized in only a few instances (1, 2). For example, a recent study Chu et al. showed that thymidylate synthase protein inhibits translation of thymidylate synthase mRNA without affecting the translation of other mRNA species (2). While several different mechanisms account for the decreased net synthesis of a specific protein in several genetic deficiencies (3), transinhibition of normal gene expression by products of the mutant allele has not yet been reported.

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1. Abbreviations used in this paper: C1 INH, C1 inhibitor; Δex7, deletion of exon 7; HANE, hereditary angioneurotic edema.

Hereditary angioneurotic edema (HANE) is an autosomal dominant disorder caused by a deficiency of the inhibitor of the first component of complement (C1 INH) (4). Two types of HANE have been recognized: type I, with low serum concentrations of functionally active C1 INH protein, and type II, with normal to increased concentrations of a dysfunctional C1 INH protein (5). The serum of HANE patients contains wild type, functionally active C1 INH protein at concentrations between 5 and 30% of normal, instead of the 50% expected if the single normal allele were fully expressed (4). This discrepancy between the expected and observed levels of C1 INH protein in HANE has been explained from in vivo studies in HANE patients in which an increased fractional catabolic rate of C1 INH was measured, and a decreased rate of synthesis of C1 INH protein was calculated (6, 7). Previous studies of C1 INH biosynthesis in cell culture demonstrated that type I HANE cells synthesized only 20% of normal amounts of C1 INH (8), similar to the rate of synthesis calculated from the in vivo turnover studies (6). These studies, however, were not able to ascertain separately the products of mutant and wild type genes. Ease of identification of wild type and mutant C1 INH mRNAs/proteins in cells from a type I HANE kindred in which exon 7 of the C1 INH gene is deleted (Δex7) (9–11) make it possible to separately estimate rates of translation of the wild type and mutant alleles.

Methods

Patient cells. Normal human adult skin fibroblast lines were obtained from human genetic mutant cells, National Institute of General Medical Sciences (Camden, NJ), or were propagated in our laboratory by standard techniques. A fibroblast line was also started from a skin biopsy obtained from a subject type I HANE caused by deletion of exon 7 of the C1 INH gene (Δex7). Fibroblast lines from the other HANE patients, four with type I HANE (three females and one male from four different kindred) and three with type II (one individual from the Ta kindred and two individuals of We kindred), have been studied previously (8). In these two type II kindreds, the mutant mRNA was the same mass as the wild type mRNA in Northern blot analysis (8). However, in each kindred, the mutant intracellular C1 INH protein is 2–3 kD larger than the wild type protein (8). For one of these mutant proteins, the mutation (deletion of Lys 251) adds a glycosylation site that yields the change in apparent molecular mass, without altering the primary translation product (12). For the other mutant protein, the mutation (a P14 hinge region mutation from Val to Glu) increases the apparent mass of the primary translation product (13). Cells from passages 4–8 were used in all experiments.

DME (low endotoxin) FBS (endotoxin < 0.01 ng/ml), and DME

without methionine were purchased from Gibco Laboratories (Grand Island, NY) and penicillin-streptomycin solution and L-glutamine from Flow Laboratories, Inc. (McLean, VA). BSA, fraction V (low endotoxin), was purchased from Sigma Chemical Co. (St. Louis, MO). [35S]methionine (specific activity ~1,000 Ci/mmol) was purchased from ICN Radiochemicals (Irvine, CA) and ENHANCE was purchased from New England Nuclear (Boston, MA). Goat antibodies to human C1 INH, CII, factor B, factor H, and C3 were purchased from Atlantic Antibodies (Scarborough, ME) and to human C1s were purchased from Cytotech (now part of Quielil, San Diego, CA).

**Cell culture and analysis of protein synthesis.** Human fibroblasts were grown to confluency in multiwell tissue culture plates in medium containing 10% FBS and then washed to remove serum and spent medium. The cells were labeled biosynthetically immediately after removal of the FBS. Biosynthetic labeling was performed by incubating the cells in methionine-free DME containing 25 μCi/ml [35S]-methionine (14) for 60 min, since there was no or only minimal secretion of newly synthesized proteins during this period. The number of cells adherent at confluency in each 16-mm diameter well was 9.78 × 10^4±0.38 (mean±SD) and did not vary with the type of cells. At the end of the pulse period, cells were rinsed and either lysed by freeze thawing as previously described (14), or in pulse-chase experiments, medium containing ~1,000-fold excess of cold methionine (DME containing 10% FBS) was added and the cells were incubated for varying time periods (chase periods). After chase periods, medium was removed, and the cells were washed with ice-cold HBSS before lysis. Cell lysates and extracellular media were prepared for immunoprecipitation by adding PMSF and leupeptin as enzyme inhibitors and detergents, preabsorbed and then immunoprecipitated with monospecific antibodies using formalin-fixed *S. aureus* (14). Immunoprecipitates were subjected to SDS-PAGE under reducing conditions, and the gels were fixed, impregnated with ENHANCE, dried, and exposed at ~70°C to film (XAR-5; Kodak, Rochester, NY). Total protein synthesis was estimated by incorporation of [35S]methionine into TCA-insoluble protein (14). Incorporation of [35S]methionine into individually immunoprecipitated proteins was determined by subtracting the appropriate background from the counts in the portion of the gel slices containing the protein after digestion with 15% hydrogen peroxide for 16 h at 65°C and addition of Bio-Safe II (Research Products International Corp., Mount Prospect, IL). Background counts were obtained in another portion of the appropriate lane of the gel not containing protein. If necessary, the amounts of protein were corrected on the basis of TCA-insoluble protein, as described previously (14).

**RNA analysis.** Total cellular RNA was extracted from fibroblast monolayers by lysis with guanidinium thiocyanate and isolated by cesium chloride density gradient ultracentrifugation, as described previously (8). The amount of total cellular RNA recovered was quantitated by absorbance at 260 nm, and was similar for the normal and HANE ΔEx7 cell. 10 μg total cellular RNA was subjected to RNA blot analysis using a riboprobe for C1 INH (368 bp representing the Bgl II to Pvu II portion of the cDNA containing the 3’ end of exon IV and all of exons V and VI inserted into the pSp72 vector). The riboprobe was 32P-radiolabeled by using a transcription kit (SP6/T7; Boehringer Mannheim Biochemicals, Indianapolis, IN). C3 and factor B cDNA probes were radiolabeled by random priming (Random Priming Kit; Boehringer Mannheim). The filters were washed and exposed to x-ray film for autoradiography.

**Results**

HANE ΔEx7 fibroblasts synthesize reduced amounts of wild type, as well as mutant C1 INH proteins. C1 INH protein synthesis in skin fibroblasts from normal and HANE ΔEx7 was assessed by SDS-PAGE of [35S]methionine labeled protein. Intracellular lysates of normal cells contained C1 INH proteins of 78 and 104 kD, representing the partially glycosylated primary translation product and the fully glycosylated protein, respectively (Fig. 1). In HANE ΔEx7 cells, an additional 66 kD C1 INH protein was detected (Fig. 1). Amounts of mutant (66 kD, ΔEx7) and wild type (78 kD) C1 INH proteins were 3±1% and 11±3% of normal, respectively (n = 6). Synthesis of other complement proteins, C1r, C1s, factor B, C3, and factor H, was comparable or greater in the HANE ΔEx7 than in the normal cells, indicating that reduction in synthesis of the C1 INH proteins was specific. Extracellular media of both normal and HANE ΔEx7 cells contained only a C1 INH protein of 104 kD (Fig. 1). No protein compared to the expected mass of the glycosylated form of the mutant protein (~92 kD) was identified in the extracellular medium, even with prolonged exposure of the SDS-PAGE gel to the x-ray film. In a pulse-chase experiment (Fig. 2), the rate of disappearance of wild type C1 INH protein was identical in normal and HANE ΔEx7 cells (Fig. 2 B); for each cell type, the amount of wild type C1 INH protein recovered in the extracellular medium was ~100% of the amount of wild type protein synthesized during the 1-h pulse period (Fig. 2 C).

To further investigate the fate of the mutant protein, cells were treated with tunicamycin, 5 μg/ml, during a 2-h pulse period (Fig. 3). These conditions allowed detection of four forms of C1 INH in the normal cell lysates, the 53-kD primary translation product, and partially glycosylated forms of 60, 66, and 78 kD. HANE ΔEx7 cells contained these four forms, but also had a 45-kD form representing the primary translation product of the mutant protein. The apparent molecular masses of these primary translation products were confirmed by SDS-PAGE analyses of proteins translated from fibroblast RNA under cell-free conditions (data not shown); the mutant protein is truncated both because of deletion of exon 7 (nucleotides 1065–1284) and a premature stop codon at nucleotide 1323 (9–11). In the extracellular media, tunicamycin treatment yielded the normal 104-kD protein and a 78-kD form, similar to the major form of the protein in the intracellular lysates, for both the normal and HANE ΔEx7 cells. Taken together, these data suggest that the truncated protein is not secreted from the cell in a stable form.
Analysis of C1 INH mRNA suggests that regulation of synthesis of the wild type C1 INH protein in HANE ΔEx7 occurs at the level of translation. Normal cells contain a single C1 INH mRNA species of 2.1 kb, while HANE ΔEx7 cells contain two C1 INH mRNA species (2.1 and 1.9 kb) (Fig. 4). The content of the wild type (2.1 kb) C1 INH mRNA in the HANE ΔEx7 cells was 52±2% (n = 5) of normal, similar to the amount expected from a single allele. The contrast between the level of mRNA (52% of normal) and rate of protein synthesis (11% of normal) suggests that the decreased rate of synthesis of wild type C1 INH protein in the HANE ΔEx7 cells is caused by a specific deficit in translation of the wild type C1 INH mRNA.

Unlike the wild type mRNA, the content of the mutant (1.9 kb) C1 INH mRNA was 17±1% (n = 5) of normal. Factor B (2.6 kb) and C3 (5.1 kb) mRNA content was equal in normal and HANE ΔEx7 cells, indicating that the reduction in C1 INH mutant mRNA is specific. Inhibition of transcription with actinomycin D and timed measurement of levels of the C1 INH mRNAs in HANE ΔEx7 and normal cells indicate that catabolism of the wild type and mutant mRNAs is similar (Fig. 5).

Expression of wild type C1 INH protein in HANE ΔEx7 is unique amongst HANE variants. C1 INH mRNA levels and synthesis rates of mutant and wild type C1 INH proteins in HANE ΔEx7 cells were compared to the findings in type II and

Figure 2. Synthesis and secretion of C1 INH in normal and HANE ΔEx7 fibroblasts: pulse-chase. (A) Confluent fibroblasts were pulse-labeled for 1 h, and then washed and incubated for varying time (chase) periods in medium containing ~ 1,000-fold excess of unlabeled methionine. At various time points ranging from 30 to 240 min, C1 INH proteins in intracellular lysates and extracellular media were analyzed as described in the legend for Fig. 1. (B and C) Incorporation of [35S]-methionine into individually immunoprecipitated wild type proteins in the intracellular lysates (B) and extracellular media (C) from normal (closed symbols) and HANE ΔEx7 (open symbols) cells was determined as described in the Methods. The results represent the mean±SD for three experiments.

Figure 3. Effect of tunicamycin on molecular masses of C1 INH proteins synthesized and secreted from normal and HANE ΔEx7 fibroblasts. Newly synthesized C1 INH proteins were radiolabeled with [35S]-methionine as described in the legend for Fig. 1. For the lanes indicated, tunicamycin (Tu), 5 μg/ml, was present during the labeling period.

Figure 4. C1 INH mRNA content in normal and HANE ΔEx7 fibroblasts. Fibroblasts were grown to confluence and total cellular RNA was extracted by lysis with guanidinium isothiocyanate and isolated by cesium chloride density gradient ultracentrifugation. Equal amounts (10 μg) were electrophoresed on agarose-formaldehyde and transferred to nitrocellulose filters (Bio-Rad Laboratories, Richmond, CA) for RNA blot analysis using a riboprobe for C1 INH labeled with 32P.
other type I HANE patients (Table 1). Fibroblasts from four unrelated patients with type I HANE contained C1 INH mRNA at 27% of normal and synthesized C1 INH protein at 20% of normal. In contrast to the findings in these type I patients, three patients with type II HANE contained C1 INH mRNA at 122% of normal, and synthesized wild type and mutant C1 INH proteins at 43 and 57% of normal, respectively. HANE ΔEx7 is the only variant studied for which synthesis of C1 INH is less than predicted from the content of corresponding C1 INH mRNA. In HANE ΔEx7, decreased synthesis relative to mRNA content occurs for both wild type and mutant C1 INH proteins.

Discussion

In most type I HANE kindred, the mutant allele and its primary transcript and protein product cannot be distinguished from the wild type forms (8). Recognition of type I HANE kindred in which one of several restriction length fragment polymorphisms is present allows the mutant allele to be identified separately from the wild type (15). For one of these kindred, HANE ΔEx7, Cicardi et al. reported the presence of a mutant ΔEx7 mRNA in addition to the normal transcript in peripheral blood monocytes (9). We report that fibroblasts from a patient from this kindred contain this mutant mRNA, and they also synthesize a mutant protein of the molecular mass predicted from the structure of the mutant mRNA. ΔEx7 yields type I HANE because the amount of wild type C1 INH protein synthesized is much less than the 50% expected from the single normal allele, similar to the findings of amounts of C1 INH protein synthesized in other type I HANE cells (8). In HANE ΔEx7, the net amount of wild type C1 INH protein synthesized was approximately fivefold less than the amount that would have been predicted from the wild type mRNA content (Table 1). Synthesis of less wild type protein than predicted could be caused by either a decreased rate of wild type C1 INH translation and/or increased rate of wild type protein catabolism in HANE ΔEx7 cells. Since the intracellular catabolism of the wild type protein was not measurably increased, these data suggest that translation of the wild type mRNA in HANE ΔEx7 cells is inhibited by a transfectant of either mutant mRNA or protein. Inhibition of translation by protein product has been proposed as a mechanism for regulating synthesis of type I procollagen and thymidylate synthase (1, 2). The decreased synthesis of wild type C1 INH protein in the HANE ΔEx7 cells represents the first example of transinhibition of translation of a normal gene product by the products of a mutant allele. It should also be noted that inhibition of translation of the mutant C1 INH mRNA was also observed.

In addition to the markedly decreased rate of synthesis of wild type protein that yields type I HANE in ΔEx7, the mutant protein is not present outside the cells. The rate of disappearance of the mutant protein from the HANE ΔEx7 cells was similar to disappearance of the wild type protein, but no evidence for secretion was obtained. The possibility that the truncated protein is secreted but unstable in the extracellular medium cannot be eliminated.

The level of mutant mRNA in HANE ΔEx7 cells is decreased below the 50% expected for the product of a single allele. Since the catabolism of this mRNA in the cytoplasm is normal, the reduced steady state mutant mRNA content is probably caused by either decreased transcription or intranuclear degradation of mutant C1 INH heteronuclear RNA. HANE ΔEx7 is unique amongst HANE variants studied (Table 1). HANE ΔEx7 is the only one in which the rate of wild type

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N, number of fibroblast cell lines studied. * For mRNA studies, RNA from a single normal fibroblast line, GM8399 (Human Genetic Mutant Cells, National Institute of General Medical Sciences), was included on each RNA blot. For quantitation, density of the normal signal on each blot was defined as 100%. 1 Specific protein synthesis was determined as described in the legends for Figs. 1 and 2. B and C, and in reference 8. 4 Values are mean±SD for five experiments for RNA and six experiments for protein synthesis. 8 Data from previous studies with fibroblast lines developed from four patients with type I and three patients with type II HANE (8). 1 For the typical type I cells, products of the mutant allele, if present, could not be distinguished from normal either structurally or immunologically. ** For the type II cells, structural differences in the mRNAs for the two alleles were not detected using Northern blot analysis.

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C1 INH protein synthesis (11% of normal) is markedly less than the level of wild type C1 INH mRNA (51% of normal). Hence, the structure of the ΔEx7 mRNA and/or protein is specific in its capacity to inhibit translation of C1 INH. In the four other HANE type I variants studied, only single species of C1 INH mRNA and protein could be identified. The similarities in reductions of mRNA content and protein synthesis indicate that C1 INH synthesis in these cells is reduced by alterations in mRNA content that occur before translation. Preliminary data indicate that the stability of C1 INH mRNA in these HANE type I variants is similar to normal, suggesting that regulation occurs during transcription or during processing of RNA in the nucleus. In the HANE type II variants studied, products of the wild type and mutant genes can be distinguished, similar to the situation in HANE ΔEx7. However, unlike HANE ΔEx7, type II HANE cells contain approximately the expected amount of C1 INH mRNA and synthesize ~50% of normal amounts of each of the wild type and mutant proteins (8). Also unlike HANE ΔEx7, the mutant protein synthesized in type II cells is secreted normally and is stable in extracellular medium (8), where abnormalities in structures of the mutant proteins alter function without appreciably affecting mass (16). The ΔEx7 mRNA and protein have substantial differences from normal in molecular mass and probably in tertiary structure. Which of these or other abnormalities result in the differences for the processing and molecular regulation between normal (or type II HANE) and the HANE ΔEx7 is not known.

Analysis of the mechanism of the transinhibition of wild type protein synthesis that occurs in HANE ΔEx7 should provide important information about regulation of protein synthesis in other genetic deficiencies, as well as translation of mRNA in normal cells.

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References