Normal myelopoiesis but abnormal T lymphocyte responses in vitamin D receptor knockout mice

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The vitamin D receptor (VDR) is a transcription factor that mediates the actions of its ligand, 1,25-dihydroxyvitamin D3 [1,25(OH)2D3], which can promote monocyte/macrophage differentiation and inhibit proliferation and cytokine production by activated T lymphocytes. In this study, VDR knockout (KO) mice were used to investigate the possible role of VDR in hematopoiesis. The relative number of red and white peripheral blood cells and the percentage of bone marrow macrophages did not differ between VDR KO and wild-type mice. 12-O-tetradecanoylphorbol-13-acetate, but not 1,25(OH)2D3, induced differentiation of bone marrow-committed myeloid stem cells from VDR KO mice to monocytes/macrophages. Production of IL-18, a Th1-promoting cytokine, was reduced in macrophages from these mice. Antigen-stimulated spleen cells from VDR KO mice showed an impaired Th1 cell response and had decreased expression of STAT4, a Th1 cell transcription factor. These results demonstrate the absolute requirement of VDR for 1,25(OH)2D3-induced monocyte/macrophage differentiation but show that monocyte/macrophage differentiation can occur in the absence of this receptor. The observed reduction in Th1 population in these mutant mice may be explained by a loss of macrophage IL-18 production or a suppression of STAT4 expression by activated splenocytes.


Introduction

The genomic actions of 1,25-dihydroxyvitamin D3 [1,25(OH)2D3] are mediated by the intracellular vitamin D receptor (VDR), a ligand-activated transcription factor belonging to a large family of nuclear receptors (1). VDR forms a heterodimer with the retinoid X receptor; this complex regulates expression of target genes by binding to vitamin D–responsive elements in their promoter regions (2). The classical actions of vitamin D are those concerned with the regulation of calcium homeostasis; however, the widespread distribution of VDR in many cell types clearly shows that the vitamin D endocrine system is involved in the regulation of many cell types and cell lineages (3). Studies using VDR knockout (KO) mice have demonstrated that after weaning, VDR plays an essential role in growth, bone formation, uterine maturation, hair production, and estrogen metabolism (4–7).

Expression of VDR has been detected in various normal and leukemic hematopoietic cells, including macrophages and activated T lymphocytes (8, 9). Evidence from other studies strongly suggests a role for VDR in hematopoiesis: Normal and leukemic myeloid progenitor cells can be induced to differentiate into monocytes and macrophages by 1,25(OH)2D3 (10–14); activated normal macrophages are able to synthesize 1,25(OH)2D3 (15, 16); and 1,25(OH)2D3 inhibits proliferation of activated T lymphocytes (17) and suppresses their synthesis of GM-CSF (18), IFN-γ (19), and IL-2 (17, 20).

The actions of 1,25(OH)2D3 on the immune system are thought to be targeted primarily at Th cells (21, 22), which can be divided into two subsets that direct divergent immune responses through the secretion of distinct cytokine profiles (23, 24). Th1 cells produce IL-2 and IFN-γ and regulate cell-mediated immunity. In contrast, Th2 cells produce IL-4, IL-6, and IL-10, which generate a humoral immune response by enhancing B cell activation and differentiation. The observation that 1,25(OH)2D3 inhibits the Th1-specific cytokines IL-2 and IFN-γ, but has little effect on Th2-specific IL-4 production, has led to the hypothesis that its immunosuppressive properties are achieved by regulation of Th1 cell activity (21). In support of this theory, production by activated macrophages of IL-12, which promotes a Th1 response, is inhibited by 1,25(OH)2D3 (25).

Myeloid and lymphoid cells from patients with vitamin D–dependent rickets type II, in which VDR is functionally defective, are unable to respond to 1,25(OH)2D3 (26, 27). Therefore, normal expression of VDR appears to be a requirement for the wide-ranging effects of
1,25(OH)₂D₃ on hematopoietic cells. The VDR KO mouse provides an excellent model with which to study the importance of VDR in the hematopoietic system. In this study, we explored the consequences of deletion of the VDR on myelopoiesis by examining cell fractions present in the peripheral blood and by comparing the differentiation and function of bone marrow–derived macrophages. Concerning the immune system, activated splenic T cells were demonstrated to be defective in VDR KO mice, revealing a potentially new role for VDR in the regulation of the immune response.

**Methods**

**Animals and cells.** VDR KO mice were generated as described previously (4) and bred under sterile conditions in the animal facility at the Cedars-Sinai Medical Center. Genotypes were determined by Southern blot analysis of genomic DNA from tail clippings, which was digested with the restriction enzyme PvuII. For experiments using VDR KO mice, their wild-type (WT) littermates were used as controls. Mice were killed by cervical neck dislocation. Peripheral blood was obtained by bleeding from the orbital sinus, and bone marrow was flushed out of isolated femurs with Alpha Minimum Essential Medium (α-MEM; Gibco BRL, Grand Island, New York, USA) containing 10% FCS using a 26-gauge needle. Isolated spleens were injected with DMEM (Gibco BRL) plus 10% FCS and crushed with forceps to release cells. Mononuclear cells from bone marrow or spleen were separated by Ficoll-Hypaque density centrifugation (Amersham Pharmacia, Uppsala, Sweden).

**Bone marrow macrophage cultures.** Bone marrow mononuclear bone marrow cells (2 × 10⁶ cells/ml) were added to modified methyl cellulose medium M3234 (StemCell Technologies Inc., Vancouver, British Columbia, Canada) to yield a final concentration of 1% methylcellulose, 30% FCS, 1% BSA, 10⁻⁴ M mercaptoethanol, and 2 mM l-glutamine. Cytokines were used at the concentrations indicated later in this section. 12-O-tetradecanoylphorbol-13-acetate (TPA) and 1,25(OH)₂D₃ were dissolved in ethanol, diluted in media, and used at the concentrations indicated later in this section. Cells were plated in six-well plates in a volume of 1 ml and incubated at 37°C in a humidified atmosphere containing 5% CO₂. Colonies were counted after 2 weeks. Colony type was established by morphology, and to ensure accurate determination, representative colonies were picked from the plates, centrifuged onto slides, and stained with Wright-Giemsa stain.

**Spleen cell cultures.** Twenty-four-well culture plates were coated with anti-CD3 antibody (10 μg/ml; PharMingen, San Diego, California, USA) in PBS, either overnight at 4°C or for 2 hours at 37°C. Cells (10⁶ cells/ml) were incubated with anti-CD28 antibody (2 μg/ml; PharMingen) in DMEM plus 10% FCS and antibiotics at 37°C with 5% CO₂. To stimulate Th1 cell differentiation, IL-12 (100 ng/ml; PharMingen) was added; to stimulate Th2 cell differentiation, anti–IL-12 antibody (50 μg/ml; PharMingen) or IL-4 (10 ng/ml; PharMingen) was added at the start of culture. Cytokine production was assessed using commercial ELISA kits (R&D Systems Inc., Minneapolis, Minnesota, USA). Cells were grown as described above by adding 1 ml of cell culture to 24-well plates and incubating for 3–5 days, and culture supernatants were stored at −20°C until analysis was performed. Proliferation was determined by incubating cells (200 μl) with plate-bound anti-CD3 and soluble anti-CD28 for 3 days in triplicate wells of 96-well plates. [³H]thymidine (0.25 μCi) was added to each well for the last 14 hours of culture. Cells were harvested onto filters, which were assayed for [³H]thymidine content using a plate reader.

**Western blot analysis and immunoprecipitation.** Cells were washed twice in ice-cold PBS and lysed in radioimmunoprecipitation assay (RIPA) buffer (150 mmol/l NaCl, 50 mmol/l Tris-HCl [pH 8.0], 0.5% deoxycholate, 1% NP-40, and 1% SDS) containing protease inhibitors over a period of 30 minutes on ice. After centrifugation at 15,000 g for 20 minutes at 4°C, the supernatant was collected. Protein concentrations were determined using an assay kit according to the manufacturer’s instructions (Bio-Rad Laboratories Inc., Hercules, California, USA). Lysates were separated by SDS-PAGE on a 4–15% linear gradient Ready Gel (Bio-Rad Laboratories Inc.) and transferred to an Immobilon-P membrane (Millipore Corp., Bedford, Massachusetts, USA). Lysates were then probed with 1 μg/ml anti-p21Waf1/Cip1 rabbit polyclonal antibody, anti-actin goat polyclonal antibody, mouse monoclonal antibody against nuclear factor of activated T cells (NFATc), rabbit polyclonal antibody against signal transducer and activator of transcription 4 (STAT4) (all from Santa Cruz Biotechnology Inc., Santa Cruz, California, USA), and anti-p21Waf1/Cip1 murine monoclonal antibody (Oncogene Science Inc., Uniondale, New York, USA). Bound antibody was detected using horseradish peroxidase–conjugated secondary antibodies and developed using the enhanced chemiluminescence (ECL) substrate from Amersham Corp. (Arlington Heights, Illinois, USA). For immunoprecipitation, cell lysates were incubated with 50 μl of anti-fyn goat polyclonal antibody on ice for 1 hour. The antibody–antigen complex was precipitated with 50 μl of protein A (Calbiochem-Novabiochem Corp., San Diego, California, USA) on ice for 30 minutes, washed twice, resuspended in loading buffer, and subjected to SDS-PAGE as
described above. The membrane was probed with 1 µg/ml anti–tyrosine kinase antibody and detected using the ECL system.

PCR analysis. A twenty-microliter volume of cDNA was prepared from 1 µg of Trizol-extracted RNA. The specific primers were as follows: IL-4, 5′-GAATGTACCAGAGCCATATC-3′ and 5′-CTCAGTACTACGATTAATCCA-3′; IFN-γ, 5′-AACGCTACACACTGCATCTTGG-3′ and 5′-GACTTCAAAGAGTCTGAGG-3′; IL-2, 5′-GAGTCGAAATCCAGAACATGCC-3′ and 5′-TCCACTTCAAGCTCTACAG-3′; GM-CSF, 5′-GCTACCACCTATGCGGATTT-3′ and 5′-CTGTGCCACATCTCTTGTC-3′; 18S, 5′-AAACGGCTACCACATCCAAG-3′ and 5′-CCTCCAATGGATCCTGTTA-3′. Cycling was done with an initial denaturation step of 9 minutes, followed by 30 seconds at 94°C, 30 seconds at annealing temperature (50°C for cytokines and 55°C for 18S), 2 minutes at 72°C, and a final elongation step of 7 minutes at 72°C. The cycle number was 22 for 18S and 30 for cytokines. PCR product was separated on a 2% agarose gel, stained with ethidium bromide, and photographed.

Results

Hematopoiesis. Analysis of peripheral blood revealed no differences in the numbers or percentages of red and white cells between VDR KO and WT mice (Table 1). 1,25(OH)２D₃, acting through the VDR, has previously been shown to stimulate the differentiation of normal bone marrow cells into monocytes/macrophages (10). The percentages of macrophages in the VDR KO bone marrow and monocytes in the peripheral blood were both the same as those found in WT mice (Table 1).

Myeloid differentiation. We used a soft-gel clonogenic assay to determine the number and percentage of granulocyte, macrophage, and granulocyte/macrophage mixed colonies derived from committed myeloid stem cells from the mononuclear bone marrow cells. Addition of 1,25(OH)₂D₃ to these soft-gel cultures markedly altered the differentiation of committed myeloid stem cells from WT mice, but not those from VDR KO mice (Figure 1). For example, in the WT mice, 1,25(OH)₂D₃ (10⁻⁷ M) dramatically increased the percentage of macrophage colonies [control dishes, 28% ± 3%; 1,25(OH)₂D₃-containing dishes, 67% ± 9%; mean ± SD] and decreased the percentage of mixed colonies [control dishes, 55% ± 8%; 1,25(OH)₂D₃-containing dishes, 30% ± 1%; mean ± SD] (Figure 1). By contrast, in the KO mice, 1,25(OH)₂D₃ (10⁻⁷ M) did not affect the percentage of macrophage colonies [control dishes, 25% ± 9%; 1,25(OH)₂D₃-containing dishes, 23% ± 5%] or the percentage of mixed colonies [control dishes, 61% ± 9%; 1,25(OH)₂D₃-containing dishes, 66% ± 9%; mean ± SD] (Figure 1). One possible explanation for these observations is that bone marrow cells from VDR KO mice were unable to respond to a differentiation stimulus. Therefore, cultures were treated with TPA, as this stimulator of phosphokinase C is known to induce macrophage colony formation (28). TPA (5 × 10⁻¹⁰M) increased the proportion of macrophage colonies from bone marrow of both WT and KO mice.

Table 1

<table>
<thead>
<tr>
<th></th>
<th>WT mice</th>
<th>VDR KO mice</th>
</tr>
</thead>
<tbody>
<tr>
<td>Peripheral blood</td>
<td></td>
<td></td>
</tr>
<tr>
<td>WBCs (10³/mm³)</td>
<td>7.0 ± 0.3</td>
<td>6.4 ± 0.7</td>
</tr>
<tr>
<td>RBCs (10³/mm³)</td>
<td>9.1 ± 0.2</td>
<td>9.1 ± 0.5</td>
</tr>
<tr>
<td>Hemoglobin (g/dl)</td>
<td>15.1 ± 1.3</td>
<td>13.9 ± 1.0</td>
</tr>
<tr>
<td>Hematocrit (%)</td>
<td>43.7 ± 0.7</td>
<td>43.1 ± 2.5</td>
</tr>
<tr>
<td>Lymphocytes (%)</td>
<td>82.3 ± 12.3</td>
<td>84.3 ± 12.1</td>
</tr>
<tr>
<td>Granulocytes (%)</td>
<td>8.3 ± 5.4</td>
<td>7.0 ± 5.7</td>
</tr>
<tr>
<td>Bone marrow</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Macrophages (%)</td>
<td>6.5 ± 1.0</td>
<td>7.6 ± 1.6</td>
</tr>
</tbody>
</table>

Peripheral blood was drawn from mice at the time of their sacrifice (4–7 weeks of age), and bone marrow macrophages were counted by α-naphthyl esterase staining of bone marrow cytopreparations. Data are presented as mean ± SD of measurements from three mice using light microscopy.

Figure 1

Colony formation by mononuclear bone marrow cells from VDR KO and WT mice. Mononuclear cells were obtained from femoral bone marrow plugs and grown in methylcellulose media containing cytokines and various concentrations of 1,25(OH)₂D₃. Colonies were counted on day 10 of culture. Control cultures from WT and KO mice contained a mean of 96 ± 37 and 98 ± 15 myeloid colonies, respectively. Numbers represent the mean ± SD of three experiments performed on triplicate wells using cells from three KO and three WT mice. Triangles, granulocyte colonies; circles, macrophage colonies; squares, mixed granulocyte/macrophage colonies.
Mononuclear cells were obtained from femoral bone marrow and grown in methylcellulose media containing cytokines and various concentrations of TPA. Colonies were counted on day 10 of culture. Control cultures from WT and KO mice contained a mean of 93 ± 22 and 117 ± 15 myeloid colonies, respectively.

### Table 2

<table>
<thead>
<tr>
<th>TPA</th>
<th>WT mice</th>
<th>KO mice</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Granulocyte</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mixed</td>
<td>20 ± 4</td>
<td>64 ± 5</td>
</tr>
<tr>
<td>Macrophage</td>
<td>58 ± 2</td>
<td>22 ± 1</td>
</tr>
<tr>
<td>Granulocyte</td>
<td>16 ± 4</td>
<td>43 ± 3</td>
</tr>
<tr>
<td>Macrophage</td>
<td>43 ± 7</td>
<td>3 ± 0</td>
</tr>
</tbody>
</table>

Mononuclear cells were counted on day 10 of culture. Control cultures from WT and KO mice contained a mean of 93 ± 22 and 117 ± 15 myeloid colonies, respectively. Numbers represent the mean ± SD of separate experiments performed on three pairs of mice, using triplicate wells per experimental point.

### Table 3

<table>
<thead>
<tr>
<th>Cytokine</th>
<th>CSF (ng/ml)</th>
<th>WT mice</th>
<th>KO mice</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Control</td>
<td>10</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>GM-CSF</td>
<td>10</td>
<td>115 ± 23</td>
<td>116 ± 27</td>
</tr>
<tr>
<td></td>
<td>100</td>
<td>132 ± 30</td>
<td>134 ± 29</td>
</tr>
<tr>
<td>GM-CSF + IL-3</td>
<td>10</td>
<td>149 ± 29</td>
<td>172 ± 49</td>
</tr>
<tr>
<td></td>
<td>100</td>
<td>150 ± 33</td>
<td>164 ± 45</td>
</tr>
<tr>
<td>G-CSF</td>
<td>10</td>
<td>66 ± 28</td>
<td>100 ± 66</td>
</tr>
<tr>
<td></td>
<td>100</td>
<td>131 ± 24</td>
<td>164 ± 41</td>
</tr>
<tr>
<td>G-CSF + IL-3</td>
<td>10</td>
<td>167 ± 38</td>
<td>160 ± 41</td>
</tr>
<tr>
<td></td>
<td>100</td>
<td>172 ± 34</td>
<td>187 ± 32</td>
</tr>
<tr>
<td>M-CSF</td>
<td>10</td>
<td>206 ± 57</td>
<td>171 ± 41</td>
</tr>
<tr>
<td></td>
<td>100</td>
<td>216 ± 51</td>
<td>180 ± 44</td>
</tr>
</tbody>
</table>

Mononuclear cells were obtained from femoral bone marrow and grown in methylcellulose media containing cytokines. When present, IL-3 was at a concentration of 10 ng/ml. Colonies were counted on day 10 of culture. Numbers represent the mean of two experiments performed on pairs of mice in triplicate wells per experimental point.
cells compared with WT cells. These data suggest that VDR expression is required for the development of a normal Th1 cell response.

Differentiation of Th cells toward a Th1 or Th2 phenotype can be achieved by costimulation with a polarizing cytokine or antibody. Antigen-stimulated splenocyte cultures were supplemented with IL-12 to induce Th1 cell differentiation, or with either IL-4 or anti-IL-12 antibody to induce Th2 cell differentiation. Exogenous IL-12 increased the amount of IFN-γ secreted by Th1 cells, but production by VDR KO cells remained below normal levels (Figure 5a). Production of IL-4 by Th2 cells was also increased by addition of anti-IL-12 compared with antigen stimulation alone (Figure 5b); and although slightly less IL-4 production was observed in WT cultures, the difference was not statistically significant.

An altered proliferative response to polarizing cytokines could contribute to the differences in the cytokine levels. Splenic cellular proliferation over 3 days showed a dose-dependent response to IL-12 and IL-4, as determined by [3H]thymidine incorporation (Figure 6). Proliferation of VDR KO splenocytes in response to IL-12 was half that of WT cells (Figure 6a), but was up to 1.4-fold greater than that of WT cells in response to IL-4 (Figure 6b). Cell cycl analysis of splenocytes stimulated with 100 ng/ml IL-12 for 3 days revealed that a smaller proportion of the VDR KO cells were in the G1 phase (WT, 68% ± 9%; KO, 53% ± 5%; mean ± SD). These data suggest that the decreased levels of IFN-γ and increased levels of IL-4 produced by activated VDR KO spleen cells in culture are due at least partially to reduced Th1 cells and increased Th2 proliferation.

We used Western analysis to determine expression of the transcription factors STAT4 and NFATc, which are involved in directing the differentiation of naive Th cells toward Th1 cells or Th2 cells, respectively. Decreased expression of STAT4 was observed in antibody-stimulated splenocytes from VDR KO mice compared with those from WT mice (Figure 7). Spleen cells from KO mice given additional stimulation with IL-12 or IL-4 also expressed reduced levels of STAT4 compared with those from WT mice. In contrast, levels of NFATc were the same in activated splenocytes from VDR KO and WT mice (Figure 7). We tested the expression of the NFATc target genes GM-CSF and IL-2 by RT-PCR. No difference was found in the expression of GM-CSF or IL-2 in activated splenocytes from VDR KO and WT mice (Figure 7). It is possible that the T cells lacking VDR were unable to properly transduce the signal from the T cell receptor in response to CD3 ligation. Therefore, we examined the phosphorylation status of fyn, whose expression and activation are increased in T cells following anti-CD3 stimulation (32). Protein lysates from stimulated spleen cells were immunoprecipitated with an anti-fyn antibody, and phosphorylation of fyn was detected by Western blot using a tyrosine kinase–specific antibody. We could not detect any differences in the tyrosine phosphorylation of fyn between the VDR KO and WT splenocytes, regardless of the stimulus received (Figure 7).

Discussion

The seco-steroid hormone 1,25(OH)2D3 is widely believed to play a role in normal hematopoiesis by enhancing the differentiation of monocytes/macrophages and inhibiting the proliferation and cytokine production of activated lymphocytes (21, 33, 34). These...
actions of 1,25(OH)2D3 are thought to be mediated through the VDR, which interacts with a broad range of genes to regulate their expression (35). We used a colony assay that allows committed myeloid stem cells to differentiate to either granulocytes or monocytes, or a combination of both. Committed myeloid stem cells from VDR KO mice and WT mice were equally responsive to all combinations of the tested cytokines, as determined by the number of colonies formed in culture. Very low concentrations of 1,25(OH)2D3 (10^{-10} to 10^{-9} M) added to the culture selectively induced WT myeloid stem cells to differentiate into macrophages. This is consistent with our previous observations (10). In stark contrast, these committed stem cells from the VDR KO bone marrow did not preferentially differentiate into macrophages in the presence of increasing concentrations of 1,25(OH)2D3, and even 10^{-7} M 1,25(OH)2D3 was unable to increase the percentage of stem cells differentiating into macrophage colonies. Nevertheless, the myeloid stem cells of the VDR KO mice were able, in soft agar, to differentiate into macrophages with the addition of another macrophage inducer, TPA. 1,25(OH)2D3 induces committed myeloid stem cells to differentiate into monocytes/macrophages by regulating expression of genes, including p21Waf1/Cip1 and c-jun (29, 36, 37). This regulation can occur directly, via interaction of VDR with the promoter regions of target genes (35). For example, p21Waf1/Cip1 has a vitamin D response element in its promoter that responds to activated VDR and transactivates the gene (29, 36). Studies have shown that the c-jun terminal kinase (JNK) pathway is upregulated during monocyte/macrophage differentiation (38, 39). The results presented here demonstrate that 1,25(OH)2D3-activated intracellular signaling pathways require the presence of VDR to stimulate monocyte/macrophage differentiation. The signaling pathways for macrophage differentiation activated by TPA do not require VDR. Therefore, by comparing the regulation of different intracellular pathways in myeloid progenitor cells from VDR KO mice in response to 1,25(OH)2D3 or TPA, it may be possible to determine which pathways are used by the two agents.

Thus, the paradox is that the bone marrow cellularity, the absolute peripheral blood white cell count, and the percentage of monocytes/macrophages in the marrow and blood in the VDR KO and WT mice in our study were identical. This parallels the findings from

**Figure 4**
Cytokine production by splenocytes from VDR KO and WT mice. Mononuclear cells (10^6 cells/ml) from murine spleens were stimulated with 10 µg/ml anti-CD3 and 2 µg/ml anti-CD28. (a) Conditioned media were analyzed for cytokine content by ELISA. The results represent the mean ± SD of experiments performed on triplicate wells using cells from three KO and three WT mice. (b) Cells were stimulated for the number of days indicated, and expression of IFN-γ and 18S was analyzed by RT-PCR.

**Figure 5**
Cytokine production by splenocytes stimulated to differentiate toward either Th1 or Th2 cells. Spleen cells were stimulated as described in Figure 4 legend, but with the addition of a cytokine or antibody that enhances Th1 or Th2 differentiation. (a) Exogenous IL-12 (50 µg/ml) was added to induce Th1 cell differentiation. (b) Anti-IL-12 (100 µg/ml) was added to induce Th2 cell differentiation. Conditioned media were analyzed for cytokine content by ELISA. The results represent the mean ± SD of three experiments performed on triplicate wells using cells from three KO and three WT mice.
patients with vitamin D–dependent rickets type II, a disease associated with germline mutations of the VDR gene that either partially or completely inactivate the function of VDR. Their hematopoiesis, as reflected by their peripheral blood counts, bone marrow morphology, and myeloid colony formation, appear normal (27). These observations indicate that normal hematopoiesis, including development of monocytes/macrophages, does not require the nuclear VDR in vivo. Perhaps other mechanisms assure normal differentiation along the monocytic pathway.

Differential screening of immediate-early-response genes induced by 1,25(OH)\(_2\)D\(_3\) in the myelomonocytic cell line U937 has revealed that the cyclin-dependent kinase inhibitor p21\(^{\text{Waf1/Cip1}}\) is important in VDR-directed myeloid differentiation into monocytes/macrophages (29, 36). Forced overexpression of this gene enhanced the differentiation of U937 cells into monocytes/macrophages. Our finding of normal expression of p21\(^{\text{Waf1/Cip1}}\) protein in mature macrophages from VDR KO mice indicates that p21\(^{\text{Waf1/Cip1}}\) expression is controlled by a variety of transcriptional regulators during macrophage differentiation.

Expression of VDR and retinoid X receptor is required for the 1,25(OH)\(_2\)D\(_3\)–mediated repression of transcription and production of IL-12 in macrophages (25). However, we found that the absence of VDR in murine macrophages did not affect their production of IL-12 in response to activation, suggesting that multiple regulatory elements affect its expression. IL-18 (also known as IFN-γ-inducing factor) is a novel cytokine produced by macrophages that has recently been shown to induce a Th1 immune response (30, 31, 40). Interestingly, production of IL-18 was less in macrophages from VDR KO animals than in WT animals, and 1,25(OH)\(_2\)D\(_3\) increased the IL-18 levels in WT macrophage cultures (data not shown). This is the first report demonstrating the regulation of IL-18 by VDR or its ligand, 1,25(OH)\(_2\)D\(_3\), and may account for some of the observations from the spleen cell cultures.

Generation of a coordinated Th1 and Th2 immune response results from the resolution of conflicting signals received by activated Th cells (41). The relationship between the two cell types is antagonistic, as these cells produce cytokines that both enhance their own differentiation and inhibit the development of the opposing subtype. Inhibition of the Th1 cytokines IFN-γ and IL-12 by 1,25(OH)\(_2\)D\(_3\) contributes to its anti-inflammatory properties (21). Therefore, splenocyte cultures...
from VDR KO mice might be expected to be relieved of that repression and generate a greater Th1 response by producing more IFN-γ and less IL-4. In fact, the opposite result was observed: less IFN-γ and more IL-4 was produced by murine VDR KO spleen cells than by WT spleen cells. This effect endured when either a cytokine or an antibody was added as a differentiation stimulus, suggesting that VDR is required to generate a normal Th1 immune response. The regulation of IFN-γ did not occur at the transcriptional level, as shown in Figure 4b. Other cytokines produced by lymphocytes have been reported to regulate IFN-γ by posttranscriptional mechanisms (42). Therefore, higher levels of Th2 cytokines produced by differentiating VDR KO spleen cells may act to suppress production of IFN-γ. Examination of transcription factors involved in Th cell differentiation revealed that protein levels of STAT4 were reduced in activated spleen cells from VDR KO mice compared with those from WT mice. Activation of STAT4 occurs after T cell stimulation in the presence of IL-12 and is required for Th1 cell differentiation (43, 44). Additionally, T cells from STAT4 KO mice have been demonstrated to have a reduced ability to mount a Th1 immune response (45). Our results suggest that the reduced amounts of IFN-γ produced by Th1 cells from VDR KO mice are due to their lower levels of STAT4 expression.

Among cytokines directly repressed by 1,25(OH)₂D₃, IL-2 and GM-CSF have been shown to have their expression inhibited by blockade of NFATc-driven transcription (46–48). When we examined NFATc protein levels, no differences were detected between splenocytes from WT and KO mice. However, the presence of VDR-Retinoid X receptor heterodimers has been shown to block binding of NFATc to the promoter regions of its target genes (46–48), so a lack of change in protein expression may not fully reflect events occurring in the promoter regions of genes targeted by this transcription factor. Therefore, we examined mRNA levels of IL-2 and GM-CSF. We found no differences in expression between VDR KO and WT cells, suggesting that the absence of VDR does not affect NFATc signaling in activated splenocytes.

The decreased proliferation of VDR KO splenocytes compared with WT splenocytes in response to IL-12 would be expected to contribute to the lower levels of IFN-γ and higher levels of IL-4 seen in these cultures. The mechanisms behind the differences in proliferation in response to stimulation are unknown, but they do not appear to be due to defects in signaling by the T cell receptor. This was demonstrated by the normal phosphorylation of the signaling molecule fyn, which had previously been shown to be activated in Th1 cell clones following anti-CD3 stimulation (49).

The weaker Th1 cell response we observed in the VDR KO murine splenocyte cultures is difficult to reconcile with the numerous studies showing that 1,25(OH)₂D₃ is able to suppress this pathway in vivo and in vitro. A study using oligonucleotide array analysis to compare gene expression profiles between Th1 and Th2 cells showed that levels of VDR were elevated in Th1 cells when they were treated with IL-12, in a pattern similar to that of known Th1-specific genes (50). In this context, the data suggest that expression of the VDR itself may be important in Th1 cell development. In support of this concept, one study in which U937 cells were transfected with cadmium-inducible antisense VDR showed increased levels of apoptosis and a decreased proportion of cells in the cell cycle when VDR was inhibited (51). This pattern appears to also occur in Th1 cells, where in the absence of VDR, exogenous IL-12 is unable to induce normal levels of splenic cell proliferation.

The reduced ability of VDR KO macrophages to produce IL-18 may contribute to the weaker Th1 response by the activated splenocytes. IL-18 has been shown to directly enhance cytokine production by activated Th1 cells, an effect partially dependent on IL-2, but independent of IL-12 (30, 31). In addition, spleens from IL-18-deficient mice infected with either Mycobacterium tuberculosis or Mycobacterium bovis BCG Pasteur produced less IFN-γ than WT mice did (52). Therefore, IL-18 synthesized by splenic macrophages may be important in the generation of cytokines by Th1 cells. In our experiments, insufficient IL-18 secretion by macrophages appears to contribute to reduced IFN-γ production by VDR KO splenocyte cultures, which leads to greater production of IL-4 by Th2 cells.

In summary, we have demonstrated that VDR is absolutely required for 1,25(OH)₂D₃ to induce the differentiation of bone marrow progenitors into monocytes/macrophages, but monocyte/macrophage differentiation can occur in the absence of VDR. Expression of VDR was shown to be important for the generation of a Th1-type immune response by spleen cells. Our results suggest that fewer Th1 cells are generated in the absence of VDR in response to antibody stimulation, and that VDR regulation of IL-18 production by macrophages, and STAT4 expression by activated splenocytes may be involved in this process.

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