IL-23 produced by CNS-resident cells controls T cell encephalitogenicity during the effector phase of experimental autoimmune encephalomyelitis

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CNS-resident cells, in particular microglia and macrophages, are a source of inflammatory cytokines during inflammation within the CNS. Expression of IL-23, a recently discovered cytokine, has been shown to be critical for the development of experimental autoimmune encephalomyelitis (EAE) in mice. Expression of the p40 subunit of IL-12 and IL-23 by microglia has been shown to be critical for the development of EAE in mice. Expression of the p40 subunit of IL-12 and IL-23 by microglia has been shown in situ and in vitro, but direct evidence for a functional significance of p40 expression by CNS cells during an immune response in vivo is still lacking. Here we report that p40 plays a critical role in maintaining encephalitogenicity during the disease course. By using irradiation bone marrow chimeras, we have generated mice in which p40 is deleted from the CNS parenchyma but not the systemic immune compartment. Our studies show that p40 expressed by CNS-endogenous cells is critical for the development of myelin oligodendrocyte glycoprotein (MOG) peptide–induced EAE. In spite of the reduced clinical disease, the absence of p40 from the CNS has little impact on the degree of inflammation. Expression profiles of the CNS lesions show an increase in Th2 cytokines when compared with mice that develop EAE in the presence of CNS IL-12 and/or IL-23. Taken together, our data demonstrate that p40 expression by CNS-resident cells forms the basis for the Th1 bias of the CNS.


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Nonstandard abbreviations used: experimental autoimmune encephalomyelitis (EAE); myelin oligodendrocyte glycoprotein (MOG); phycoerythrin (Pe).
in a population of MOG peptide–reactive lymphocytes. By using irradiation BM chimeras, we have generated mice in which p40 is deleted from the CNS parenchyma but not the systemic immune compartment. Our studies show that p40 expressed by CNS-endogenous cells is critical for the development of MOG-induced EAE.

**Methods**

**Mice.** Female C57BL/6 and CD45 congenic C56BL/6-Ly5.2 mice were obtained from the National Cancer Institute (Frederick, Maryland, USA). Homozygous IL-12 p35−/− and IL-12 p40−/− C57BL/6 mice were originally purchased from The Jackson Laboratory (Bar Harbor, Maine, USA) and bred in-house under pathogen-free conditions.

**Irradiation BM chimeric mice.** BM-donor mice were euthanized using CO2 and BM cells were isolated by flushing femur and tibia bones with HBSS. BM was filtered through a 100-µm cell strainer and cells were washed with BSS. CD45 congenic recipient mice were lethally irradiated (12 Gy in a split dose) and intravenously injected with 2 × 107 BM cells. Engraftment took place over 6–8 weeks of recovery. Mice were bled retro-orbitally to ensure more than 95% engraftment of blood leukocytes. We have previously reported that this protocol allows us to generate mice in which the APC compartment of the CNS and of the immune system can be completely separated (11).

**Peptides and Ab’s.** MOG35–55 peptide (MEVGWYRSPF-SRVVHYLRNNGK) was obtained from Research Genetics (Huntsville, Alabama, USA). For cytofluorometric analysis, the antibodies against the following were used: CD45-phycocerythrin (CD45-PE), CD11b-FITC (Mac1), CD4-biotin, GR-1-biotin, CD8-biotin, and NK1.1-biotin. Biotin labels were visualized using streptavidin-allophycocyanin (Pharmingen, La Jolla, California, USA). All Ab’s were obtained from Pharmingen.

**Induction of EAE.** For active immunization, 5- to 8-week-old female C57BL/6 or p40−/− mice (13- to 16-week-old BM chimeric mice) were immunized subcutaneously with 200 µg of MOG35–55 peptide emulsified in CFA supplemented with 2 mg/ml of Mycobacterium tuberculosis (Difco Laboratories, Detroit, Michigan, USA). The mice received intraperitoneal injections with 250 ng pertussis toxin (Sigma-Aldrich, St. Louis, Missouri, USA) at the time of immunization and 48 hours later. After 7 days, the mice received an identical booster immunization with MOG peptide in CFA without pertussis toxin. Clinical disease usually commences between day 12 and day 18 after immunization.

For adoptive transfer, donor mice were immunized subcutaneously with 200 µg MOG35–55 in CFA supplemented with 500 µg Mycobacterium tuberculosis. Immediately after immunization and 2 days later, the mice received 250 ng of pertussis toxin. Eleven days after immunization, the mice were sacrificed, spleens were removed and homogenized, and red blood cells were lysed. The cells were cultured for 4 days in RPMI 1640 supplemented with 10% FCS (both obtained from BioWhittaker Inc., Walkersville, Maryland, USA), 10 µg/ml of MOG peptide, and 2.5 ng/ml of recombinant IL-12 (PeproTech Inc., Rocky Hill, New Jersey, USA). The cells were harvested, and dead cells were removed by Ficoll (Sigma-Aldrich) centrifugation. Cells were then washed and injected into recipient mice (5 × 106 to 2.5 × 107 cells/mouse). Animals received 250 ng/mouse pertussis toxin on day 0 and day 2 after transfer. Clinical disease usually commences 6–10 days after cell transfer.

**Clinical evaluation.** The mice were scored four times per week as follows: 0, no detectable signs of EAE; 0.5, distal limp tail; 1, complete limp tail; 1.5, limp tail and hind limb weakness; 2, unilateral partial hind-limb paralysis; 2.5, bilateral partial hind-limb paralysis; 3, complete bilateral hind-limb paralysis; 3.5, complete

**Table 1**

<table>
<thead>
<tr>
<th>Group</th>
<th>Incidence</th>
<th>Mean day of disease onset</th>
<th>Mean maximal clinical score</th>
</tr>
</thead>
<tbody>
<tr>
<td>WT + 2.5 × 10^7 MOG-reactive cells</td>
<td>6/6</td>
<td>100%</td>
<td>8.3</td>
</tr>
<tr>
<td>WT + 1.25 × 10^7 MOG-reactive cells</td>
<td>5/6</td>
<td>83.33%</td>
<td>11.2</td>
</tr>
<tr>
<td>p40−/− + 2.5 × 10^7 MOG-reactive cells</td>
<td>2/5</td>
<td>40.00%</td>
<td>11.5</td>
</tr>
<tr>
<td>p40−/− + 1.25 × 10^7 MOG-reactive cells</td>
<td>2/5</td>
<td>40.00%</td>
<td>11.5</td>
</tr>
</tbody>
</table>

* Diseased animals only.

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hind-limb paralysis and unilateral forelimb paralysis; 4, total paralysis of fore and hind limbs (score greater than 4 to be euthanized); 5, death.

**Histology.** Mice were euthanized with CO2. They were then perfused with PBS and the spinal column of each mouse was removed and fixed in 10% buffered formalin. The spinal cord was dissected and paraffin embedded prior to staining with H&E to assess infiltration.

**Flow cytometry.** Mice were euthanized with CO2 and spinal cords were removed by flushing the spinal column with sterile BSS. The brain was dissected to isolate the brain stem. Both tissues were homogenized and strained through a 100-µm nylon filter (Fisher Scientific Co., Pittsburgh, Pennsylvania, USA). After centrifugation, the cell suspension was resuspended in 37% isotonic Percoll and underlaid with 70% isotonic Percoll. The gradient was centrifuged at 600 g for 25 minutes at room temperature. The interphase cells were collected and extensively washed prior to staining. For flow cytometry, the cells were stained with primary Ab’s for 30 minutes at 4°C, washed, and incubated with streptavidin-allophycocyanin (Pharmingen) for 15 minutes. The cells were washed and analyzed using a FACS Calibur system and CellQuest software (Becton, Dickinson and Co., San Jose, California, USA). Data analysis was performed using WinMDI 2.8 software (Scripps Research Institute, La Jolla, California, USA).

**Recall responses.** Mice were primed by flank injections of MOG peptide in CFA. After 5 days, the axillary and inguinal lymph nodes were removed and homogenized. Lymph node cells (5 x 10⁶) were placed in triplicate in a 96-well plate and pulsed with different amounts of MOG peptide or the irrelevant peptide myelin proteolipid protein (PLP139-151; Research Genetics) as a control. After 48 hours, cells were pulsed with [³H]thymidine (NEN Life Science Products, Boston, Massachusetts, USA) and incubated for an additional 15 hours before cells were harvested. Thymidine incorporation was assessed using a Filtermate harvester and a TopCount NXT microplate scintillation and luminescence counter (both from Packard Bioscience Co., Meriden, Connecticut, USA). For cytokine analysis, sister cultures were harvested 48 hours later, and culture supernatants were analyzed by ELISA for IFN-γ and IL-2 (Pharmingen).

**Real-time PCR.** RNA was extracted from spinal cords of mice with EAE as described (11). Briefly, the spinal column was flushed with ice-cold HBSS and the cords were homogenized in Trizol reagent (Invitrogen Corp., Carlsbad, California, USA). RNA was extracted following the manufacturer’s instructions. The samples were...
Table 2
Phenotype of BM chimeras

<table>
<thead>
<tr>
<th>BM donor</th>
<th>Recipient mouse</th>
<th>p40 expression</th>
</tr>
</thead>
<tbody>
<tr>
<td>WT</td>
<td>WT</td>
<td>Whole body</td>
</tr>
<tr>
<td>WT</td>
<td>p40–/–</td>
<td>Immune compartiment</td>
</tr>
<tr>
<td>p40–/–</td>
<td>WT</td>
<td>CNS parenchyma</td>
</tr>
<tr>
<td>p40–/–</td>
<td>p40–/–</td>
<td>Null</td>
</tr>
</tbody>
</table>

Therefore, whether one examines inflammation with myelin antigen (1–3).

Therefore, whether one examines inflammation with myelin antigen (1–3). The induction of EAE upon active immunization in p40–/– or p19–/– mice, the phenotype is exclusively due to the lack of IL-23. We wanted to extend these studies to assess the role and function of p40 during the effector phase of EAE. In order to do so, we adoptively transferred MOG-reactive lymphocytes (generated in WT mice) into either p40–/– or WT mice. Figure 1 and Table 1 show that p40–/– mice develop EAE with significantly delayed kinetics and decreased severity (P < 0.01), indicating that host-expressed p40 is involved in the maintenance of encephalitogenicity during the effector phase of EAE. These data are consistent with the observations of Cua and colleagues have shown that intracerebral administration of IL-23 after disease onset can restore disease in p19–/– mice (3).

**p40 drives encephalitogenicity but not inflammation.** In spite of the reduced clinical phenotype of p40–/– mice upon adoptive transfer of MOG-reactive lymphocytes, histological analysis (Figure 2) revealed significant inflammation in the spinal cords of p40–/– mice that was comparable to that of WT mice with significantly more severe EAE. This finding supports the notion that p40 is not absolutely required for the infiltration of inflammatory cells into the CNS, but rather alters the degree of encephalitogenicity upon entry into the CNS. For a more quantitative analysis.

**Figure 4**
BM transfer fully reconstitutes the host’s ability to drive Th1 immunity. After 2 months of recovery, BM chimeric mice were immunized with MOG peptide in CFA and lymph nodes were removed 5 days later. Recall proliferation (a), IL-2 (b), and IFN-γ secretion (c) in response to either 100 µg MOG peptide or irrelevant myelin proteolipid protein peptide was assessed in vitro. White bars, medium alone; black bars, 100 µg MOG peptide; gray bars, 100 µg PLP peptide. *, WT; –, p40–/–; arrows indicate the transfer of BM. (d) After BM transplant, mice (diamonds, p40+/+ → p40–/–; squares, p40–/– → p40+/–; x, p40–/– → p40–/–; triangles, p40–/– → p40–/–) were allowed to recover for 2 months, then immunized with MOG35,55 in CFA as described and scored for clinical disease. In all experiments, the disease score between the susceptible p40+/+ → p40–/– and p40–/– → p40–/– mice was significantly different as assessed using a nonpaired Student’s t test (P < 0.01). Shown is a representative of four individual experiments.
of CNS infiltrates, 5 days after disease onset, the recipient mice were euthanized, and infiltration into the spinal cord was measured by flow cytometry. Figure 3a shows that the absence of p40 in the recipient does not significantly alter the degree of inflammation. Both WT and p40–/– mice immunized with MOG35–55 peptide display a high number of CD45 hi infiltrating cells compared with nonimmunized mice. We did not observe an overt change in the cellular profile of infiltrating cells with regard to the percentage of CD4+ or CD8+ T cells, macrophages, or polymorphonuclear cells as assessed by flow cytometry (Figure 3b). The number of NK cells or NK T cells was negligible as assessed by NK1.1 staining (not shown). The data indicate that the quality of the CNS immune response is different in the WT than in the p40–/– CNS, but that the differences do not lie in the composition of the infiltrating cells.

Production of p40 CNS-endogenous cells is critical for the induction of EAE upon active immunization. We have previously shown that CNS-endogenous cells, most likely microglia, control the extent of inflammation in EAE via CD40 (11). Mice in which CD40 is not expressed within the CNS show a decrease in inflammatory infiltrates and a paucity in early chemokine expression by CNS-resident cells. In order to assess the role of p40 expressed by cells within the CNS during the effector phase of EAE, we selectively expressed p40 within the CNS using irradiation BM chimeras. Following irradiation and BM reconstitution, the peripheral APC compartment is comprised entirely of BM-donor–derived cells, whereas 100% of CNS-resident microglia remain of host origin (11). We generated BM chimeras by transferring p40–/– or WT BM into irradiated p40–/– and WT recipients (Table 2). To confirm the phenotype of the reconstituted peripheral APC compartment, we immunized these mice and determined the capacity of APCs in the periphery to support Th1 responses in a recall assay. Figure 4, a and b, shows that the antigen-induced T cell proliferation and production of IL-2 and IFN-γ are intact in WT → WT and WT → p40–/– mice. In contrast, in p40–/– → WT and p40–/– → p40–/– mice, T cells were incapable of inducing IFN-γ secretion.

Using this panel of mixed BM chimeric mice, mice were immunized with MOG35–55 in CFA and disease development was assessed as described. Figure 4d and Table 3 show that p40 produced by cells resident to the CNS is essential for full disease development. In the absence of CNS-derived p40 (WT → p40–/–), we saw a marked delay in disease onset and a decrease in disease severity. It is important to emphasize that while both WT → p40–/– and WT → p40–/– chimeras have an equivalent capacity to generate peripheral Th1-type T cells, the disease in the latter is compromised. Th1 polarization by CNS-derived IL-12 p40. We wanted to determine the impact of p40 expressed within the CNS on the cytokine profile of the infiltrating lymphocytes. We immunized p40–/– BM chimeras with MOG35–55 in CFA as described. The mice were sacrificed at the first sign of clinical disease to extract RNA from the spinal cords. Expression levels of Th1 and Th2 markers were assessed by real-time PCR using hydrolysis probes (TaqMan) (Figure 5). The data reveal that the infiltrates in p40–/– mice display a Th2 polarized phenotype, whereas the WT mice appear more Th1 biased. A similar trend could be observed in p40–/– mice upon adoptive transfer (data not shown). The data indicate that the encephalitogenic potential of infiltrating lymphocytes is nurtured by p40 expressed within the CNS.

Table 3
EAE in BM chimeras induced by active immunization with MOG peptide (see Figure 4d)

<table>
<thead>
<tr>
<th>Group</th>
<th>Incidence</th>
<th>Mean day of disease onsetA</th>
<th>Mean maximal clinical scoreA</th>
</tr>
</thead>
<tbody>
<tr>
<td>WT → WT</td>
<td>4/5 80%</td>
<td>17.2</td>
<td>4</td>
</tr>
<tr>
<td>WT → IL-12 p40–/–</td>
<td>4/6 67%</td>
<td>23.3</td>
<td>2.1</td>
</tr>
<tr>
<td>IL-12 p40–/– → IL-12 p40–/–</td>
<td>0/3 0%</td>
<td></td>
<td></td>
</tr>
<tr>
<td>IL-12 p40–/– → WT</td>
<td>0/4 0%</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

A Diseased animals only.

**Figure 5**
Expression profile of inflamed spinal cords. RNA was isolated as described from inflamed spinal cords 24 days after immunization. Real-time RT-PCR analysis was performed by measuring amplicon accumulation using TaqMan probes (Qiagen Inc.) (black bars, WT; white bars, p40–/–). Data are expressed as arbitrary units based on the standard curve of triplicate wells ± SD. Shown is a representative of at least three individual experiments. Levels of β-actin mRNA were similar in both samples, and other mRNA species levels were normalized to β-actin.
Discussion
The goal of our study was to determine the role of CNS-expressed p40 during the effector phase of EAE. Upon adoptive transfer of fully primed and activated WT MOG-reactive lymphocytes into p40−/− mice, we observed a marked delay in disease onset and a significant decrease of disease severity when compared with WT mice. Interestingly, although the degree of clinical symptoms was modest, the spinal cords of these mice still showed a large number of inflammatory infiltrates. Upon entry of leukocytes into the CNS, resident cells, in particular microglial cells, become activated and secrete a variety of proinflammatory factors including IL-12 and IL-23 (3, 9). We hypothesized that it is the CNS microenvironment that regulates T cell behavior during CNS inflammation in both EAE and MS. We generated BM chimeras in order to assess the role of p40 expressed by CNS-endogenous cells (radioresistant compartment), while having an immune compartment fully capable of supporting T cell activation, Th1 polarization, and encephalitogenicity. After active immunization of these mice, in the absence of p40 expressed within the CNS, we observed only subclinical EAE. In the absence of p40 from the CNS, autoreactive T cells still infiltrate the CNS, but do not fully develop their encephalitogenic potential. This finding is consistent with a recent report showing that mice deficient in the p19 subunit of IL-23 develop clinical EAE only upon delivery of IL-23 into the CNS. Peripheral complementation was insufficient to break EAE resistance (3). Furthermore, CNS-infiltrating inflammatory macrophages have been shown to express p19 and thus IL-23 at levels comparable to those expressed by CNS-resident microglia (3). However, the amount of IL-23 produced by CNS-invading cells alone is insufficient to drive a fulminant encephalitogenic Th1 response. It appears that the CNS residents participate in Th polarization and that this participation is a critical determinant in the disease pathogenesis.

To assess the impact of p40 secretion by CNS-resident cells on the inflammatory response, we evaluated the expression profile of Th1/2-associated genes expressed by CNS-infiltrating T cells. Expression analysis of spinal cords for Th1 and Th2 markers revealed that in the absence of p40 from the CNS, a shift from a Th1 profile toward a Th2/0 polarized phenotype was induced.

The Th1/2 paradigm of inflammation in EAE and MS is challenged by gene-deletion studies showing that EAE can develop irrespective of Th1 cytokines (12–16) and Th1-inducing cytokines (e.g., IL-12). Thus, one cannot assume that the expression of Th1 markers correlates with encephalitogenicity. Nonetheless, it appears that continued exposure of lymphocytes to IL-23 at the site of inflammation is critical for the sustained expression of Th1-associated genes. As for MS, therapeutic intervention by targeting the IL-12/23 pathway may provide a powerful inhibitor of disease progression without the side effects of broad immune suppression.

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