Degeneration of brain tissue following stroke leads to functional impairment with limited brain self-repair. New evidence suggests that delivery of circulating CD34+ human umbilical cord blood cells can produce functional recovery in an animal stroke model with concurrent angiogenesis and neurogenesis leading to some restoration of cortical tissue (see the related article beginning on page 330). While some alternative interpretations of this data are offered herein, the study provides encouraging evidence of functional recovery from stroke in an animal model using stem cell therapy.

Focal brain ischemia results from blockage of the blood supply to the region supplied by the affected vessel with subsequent degeneration of that tissue. The severity of this degeneration is a function of the extent and location of the injury. There is little spontaneous repair of the injured region, and most post-stroke improvement appears to reflect the recruitment of intact circuitry to perform the function of the degenerated region. Because of the public health importance of developing an effective treatment leading to recovery from stroke, various animal models have been developed to simulate the mechanism of injury and to evaluate intervention strategies.

The observation that new neurons are generated in specific regions of the adult brain has raised the possibility that this process of neurogenesis may be harnessed to repopulate degenerated regions (1). Neurogenesis in the adult brain occurs in the hippocampal dentate gyrus and the forebrain subventricular zone (SVZ), it has been reported that new neurons can be generated either transiently or at a low rate in the cerebral cortex, substantia nigra, and hypothalamus (2–4), although the existence of cortical neurogenesis remains controversial (5). As a result, the adult brain can be divided into regions designated as neurogenic (dentate gyrus and forebrain SVZ) or nonneurogenic (the rest of the brain). However, recent evidence indicates that the forebrain SVZ of humans does not give rise to new neurons, suggesting that in the human brain, only the dentate gyrus may support substantial amounts of neurogenesis (6). Such restriction in the human brain may have implications for the therapeutic use of neural stem cells. Nevertheless, in animal models, neural stem cells can be extracted from various brain regions and expanded in vitro (7). Recent studies suggest that cells expressing the chondroitin sulphate proteoglycan NG2, thought to serve as a marker for neural progenitors committed to an oligodendrocyte lineage, may exhibit multipotentiality in vitro (8, 9). As NG2-positive cells are widespread throughout the mature brain, they may represent a local reserve of stem cells that may be recruited for repair of focal injury.

Within neurogenic regions, the rate of neurogenesis appears to be modulated by local environmental changes (10). Neurogenesis can also be promoted by the delivery to neurogenic regions of various growth factors, including brain-derived neurotrophic factor, FGF-2, IGF, and erythropoietin (11–13). Following experimental injury, there is an upregulation of neurogenesis, and delivery of growth factors in injury models can augment this neurogenic response as well as contribute to neuroprotection, which suggests that this may be a suitable strategy to pursue for development of therapeutic intervention (14, 15). Specific enhancement of neurogenesis has been reported as a result of ischemic injury with migration of neuroblasts to the site of degeneration, although only a small fraction of these new neurons survive (reviewed in ref. 16).

It is not yet understood what environmental differences between neurogenic and nonneurogenic brain regions support the generation of new neurons, but this process may be regulated by the availability of factors to enhance neurogenesis, the presence of factors that suppress neurogenesis, or a combination of the two. The situation is made more complex by the progression of cells from proliferation through lineage commitment and functional differentiation, all of which are sequential components in the process of neurogenesis. Furthermore, cell progression through each of these components of neurogenesis is likely to be regulated by different factors at each step. Thus, the development of successful therapeutic strategies to use neural stem cells for brain repair will require that appropriate regulatory factors be presented in the proper spatial and temporal sequence. This is a daunting prospect, and progress will come as individual pieces of this puzzle are assembled.

Commentaries

Umbilical cord blood cells and brain stroke injury: bringing in fresh blood to address an old problem

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Degeneration of brain tissue following stroke leads to functional impairment with limited brain self-repair. New evidence suggests that delivery of circulating CD34+ human umbilical cord blood cells can produce functional recovery in an animal stroke model with concurrent angiogenesis and neurogenesis leading to some restoration of cortical tissue (see the related article beginning on page 330). While some alternative interpretations of this data are offered herein, the study provides encouraging evidence of functional recovery from stroke in an animal model using stem cell therapy.
Figure 1
Morphological response of stroke-injured brain to delivery of umbilical cord blood cells. (A) Occlusive stroke results in the degeneration of brain tissue supplied by the occluded vessel, which produces a lesion cavity. The lesion cavity compromises the cerebral cortex and lies some distance from a neurogenic region, the SVZ. The major fiber tracts forming the corpus callosum lie between the lesion cavity and the SVZ, a putative source of newly generated neurons observed in response to stem cell therapy. (B) Delivery of umbilical cord blood cells enriched with the CD34+ cell fraction to the tail vein results in circulation of these primitive stem cells throughout the body’s vascular system. (C) The circulating CD34+ cord blood cells in vessels adjacent to regions of neurodegeneration provide a possible supply of a number of cytokines and growth factors, including FGF-2 and IGF, that are proposed to cross into brain parenchyma directly or to stimulate local environmental production of growth factors. (D) Factors released by the circulating CD34+ cells also serve to recruit angioblasts, leading to angiogenesis. (E) Following CD34+ cord blood cell delivery, new cells are born and express neuronal lineage markers in a process called neurogenesis. In addition to being present in tissue remaining after the lesion, these new neurons contribute to restored tissue in the margin of the lesion cavity, which results in a small lesion cavity. It remains to be determined whether the newly generated neurons migrated from the neurogenic SVZ or were recruited from quiescent neural progenitor or stem cells residing in the parenchyma closer to the lesion cavity. Figure adapted with permission from Current Opinion in Neurobiology (16).

Stem cell therapy with cord blood: how might it work?
These are exciting findings with direct therapeutic possibilities, but there are alternative interpretations for some aspects of the study by Taguchi et al. (17). The authors attribute the behavioral recovery of the mice to the functioning of newly generated neurons. However, the improvement in cortical thickness was quite modest. By 90 days, animals receiving CD34+ cells had a 25% thicker cortex than non-CD34+-treated animals or had 15% less loss of cortical thickness, depending on the context in which the results are viewed. It is difficult to see how such a limited morphological increase could account for functional restoration, particularly in the absence of data revealing the morphological and functional connectivity of new cells. It is also possible that the neovascularization the authors observed may have benefited uncompromised tissue, leading to enhanced behavioral performance. Further studies, in which neurogenesis is specifically blocked, will be needed to provide insight into the role of neurogenesis and to validate the authors’ conclusion that behavioral improvement was due to functional neurogenesis.

Taguchi et al. (17) also assume that the new neurons have migrated from the forebrain SVZ because of their expression of the migratory markers polysialylated neuronal cell adhesion molecule (PSA-NCAM) and doublecortin. While these are both markers of migrating neuroblasts, their expression does not provide information about the extent of their migration. For example, expression of PSA-NCAM and doublecortin are also found in the new neurons in the dentate gyrus where the total migratory movement is only a few cell diameters. These cells may be migrating, but they need not have come all the way from the forebrain SVZ. It may be that these cells were recruited from local progenitor cells, such as the NG2-positive cell population described earlier. For example, in the immediate post-stroke interval, substantial cell proliferation was observed near the site of injury and 25% of these cells were identified as adopting an endothelial cell fate. However, the ultimate identity of the remaining BrdU-positive cells was not pursued, and it is possible that these may have eventually expressed neuronal lineage markers in response to signals associated with CD34+ cells and neovascularization.

It would be interesting to know what effect the stimulation provided by circulating CD34+ umbilical cord blood cells would have on neurogenesis in the forebrain SVZ and dentate gyrus regions of naïve and stroke-lesioned animals. Such information...
would be useful for discriminating between the influence of CD34+ cells and that of neovascularization on the overall neurogenic capacity of the dentate gyrus and forebrain SVZ. It may also provide insight into the possibility of recruiting neural stem cells from these regions for brain repair. This information would be particularly relevant in regard to the experimental group that also received erythropoietin, as this treatment has been shown to enhance both angiogenesis and neurogenesis (13). The current study suggests that one mechanism accounting for the functional recovery resulting from cord blood cell delivery works through the enhancement of angiogenesis around the site of degeneration. This finding provides a note of optimism for developing therapeutic strategies for stroke. Taguchi et al. provide an important piece of the puzzle, but there remains much to be determined about the mechanisms involved and the specific role of neurogenesis in brain recovery from stroke before a comprehensive picture will emerge on how to treat this old problem (17).

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Unlocking the secrets of the pancreatic β cell: man and mouse provide the key

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Failure of the pancreas to secrete sufficient insulin results in type 2 diabetes, but the pathogenesis of pancreatic β cell dysfunction is still poorly understood. New insights into β cell failure come from defining the genes involved in rare genetic subtypes of diabetes and creating appropriate animal models. A new mouse model of transient neonatal diabetes mellitus emphasizes that both the number of β cells and their function are critical for insulin secretion and may be regulated by imprinted genes (see the related article beginning on page 339).

The regulated secretion of insulin by the pancreatic β cell maintains blood sugar concentrations within a narrow physiological range. In over 150 million people worldwide, however, pancreatic β cells fail to secrete adequate insulin, usually in the presence of increased insulin resistance, which results in type 2 diabetes (T2D). Understanding the pathways that result in β cell dysfunction at a physiological and molecular level is critical for improved understanding and treatment of T2D.

Learning from rare genetic subtypes of diabetes
How can we study the pancreatic β cell in humans when these cells are not readily accessible? Accidents of nature in which a single gene defect results in severe β cell dysfunction, causing diabetes, offer the chance that the responsible gene can be defined. The best example of such research has been the use of positional cloning to demonstrate that heterozygous mutations of the genes encoding the hepatic transcription factors

Nonstandard abbreviations used: Ig, immunoglobulin; mi, microarray; P0, primitive embryonic; pl, primary lymphocytic; PAC, P1-derived artificial chromosome; TNDM, transient neonatal diabetes mellitus; T2D, type 2 diabetes.

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