TB, or not TB: that is the question — does TLR signaling hold the answer?

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Innate immunity critically depends on signaling by Toll-like receptors (TLRs) that rely heavily on an intracellular adapter protein called myeloid differentiation factor 88 (MyD88). Adaptive immune defenses are generally thought to be orchestrated by innate immune responses and so should require intact Toll-like receptor resistance; TLR, Toll-like receptor; TRAM, TRIF-related adapter molecule; TRIF, TIR domain–containing adapter inducing IFN-β.

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In this issue of the JCI, Fremond and coworkers report data that expand our understanding of the host defense against Mycobacterium infection yet raises provocative new questions that will be sure to spark controversy (3). They used mice with a genetic deficiency in myeloid differentiation factor 88 (MyD88), a critical adapter molecule common to signaling by most Toll-like receptors (TLRs). MyD88-/- mice were subject to infectious challenge with Mycobacterium tuberculosis (MTB) and died within 4 weeks, showing evidence of massive, uncontrolled pulmonary growth of the infectious organism. Adaptive immune responses appeared to be unaffected, but iNOS activation and expression of the defensive cytokines TNF-α, IL-6, and IL-12 p40 were strongly reduced in MyD88-null mice infected with Mycobacterium tuberculosis. The authors concluded that MyD88-dependent signaling is not significantly involved in T cell activation but that in the absence of MyD88, T cell–mediated immunity can afford only partial protection from infection. At face value, this work would appear to offer a plausible answer to the question posed — TB, or not TB? — namely, that innate immunity in general and TLR signaling through MyD88 are critical for the host to take arms against the sea of troubles that MTB infection brings. But how does this answer square with what we know about the interaction between innate and adaptive immunity?

Signaling by Toll-like receptors and MyD88 in innate immunity

Defense against invading pathogens comes in 2 major forms. Innate immunity rapidly mounts a multipronged defense that is relatively nonspecific and relies heavily on inflammatory molecules directly or indirectly toxic to pathogens. Adaptive

Every second of every day, Mycobacterium tuberculosis (MTB) infects another human being somewhere in the world (1), yet the global incidence of around 8 million cases of active tuberculosis (TB) per year is only a tiny fraction of the estimated 2 billion people infected. Only around 1 in 10 of those infected will ever develop active disease, which raises a question: why do some individuals develop disease, while others do not? The answer is not trivial: in 2002, TB caused an estimated 2 million deaths, mostly in underdeveloped countries, and over the last century, TB has killed an estimated 100 million people (1, 2).

A solid mechanistic understanding of how MTB infection proceeds and how host defenses are marshaled against it (both successfully and unsuccessfully) has proven elusive and remains an important goal.

Nonstandard abbreviations used: IRF, IFN-regulated factor; MTB, Mycobacterium tuberculosis; MyD88, myeloid differentiation factor 88; NOD, nucleotide-binding oligomerization domain; TLR, toll-like receptor; TIR, Toll/IL-1 receptor resistance; TLR, Toll-like receptor; TRAM, TRIF-related adapter molecule; TRIF, TIR domain–containing adapter inducing IFN-β.
immune defenses are deployed more slowly but involve T cells and antibodies produced by leukocytes that target the pathogen with surgical precision. These 2 arms of host defense interact and cooperate in numerous ways to accomplish their task.

The discovery of the central importance of TLR-mediated signaling to innate immune mechanisms has transformed immunology and answered a fundamental question: how does the host sense the presence of foreign pathogens and how does that generate a signal to rapidly mount a vigorous defense (4, 5)? We now know that there are 13 mammalian TLRs that can sense molecular patterns that are common constituents of a very wide range of pathogens but are rarely found in the host. All TLRs can utilize the adapter protein MyD88 to propagate signals to gene targets and generate a rapid protective response, either by activating NF-κB or via other routes. MyD88 is also used by other inflammatory signaling pathways (e.g., IL-1 and IL-18). However, at least 2 TLRs (TLR3 and TLR4) can use alternative adapters such as Toll/IL-1 receptor resistance (TIR) domain–containing adapter inducing IFN-β (TRIF) and TRIF–related adapter molecule (TRAM; used by TLR4) that can activate responses distinct from those elicited by MyD88. So TLR4, for example, can utilize either the MyD88/TIR domain–containing adapter protein or TRAM/TRIF adapter pairs to generate distinct responses (5); the latter pathway is known to be critical to the induction of DC maturation. While MyD88 typically activates NF-κB, TRAM and TRIF activate IFN-regulated factor–3 (IRF-3) and IRF-7, and can also induce late activation of NF-κB (Figure 1) (reviewed in ref. 5).

MyD88-deficient mice have an impaired ability to signal through TLRs and, consequently, their macrophages do not produce proinflammatory cytokines in response to bacterial components that are known TLR ligands (5), although TLR4 and TLR3 can link with other adapter proteins to mediate signaling in the absence of MyD88 (5, 6). Since TLRs directly recognize a very broad spectrum of foreign pathogens and rapidly generate a defensive response involving numerous proinflammatory cytokines, the innate immune system in MyD88-null mice is severely disabled. Besides its pivotal role in host defense, TLR signaling has also been directly implicated in an expanding list of diseases characterized by chronic inflammation, most recently atherosclerosis (7–9).

How does MyD88 affect adaptive immunity?

In 1989 Charles A. Janeway Jr. suggested that the innate immune response initiates the adaptive immune response through pattern-recognition receptors that recognize microbial products (10). Janeway’s widely accepted model posits that the coupling of microbial recognition with the induction of the costimulators is accomplished by TLR signaling and is required for activation of pathogen-specific T cells (11–13). Thus, innate immunity orchestrates the adaptive response through TLR signaling. CD80 and CD86 costimulators “flag” the antigenic peptide presented by MHC molecules on the same DC as being microbe derived. Since T cells can only receive the costimulatory signal in an antigen-specific, cognate interaction with DCs, TLR-induced expression of costimulators translates the nonclonal pattern recognition signals into clonal antigen-specific immune responses (Figure 2). Thus, TLR...
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signaling constitutes the essential link between innate (e.g., APCs) and adaptive (e.g., T cells) immune cells. Once CD4+ T cells are activated, their differentiation into effector cells is controlled by a variety of factors, including cytokines produced by DCs (Figure 3). TLR-induced cytokines, such as IL-12, generally specialize in the induction of Th1 responses. Mice deficient in MyD88, in contrast, are severely compromised in Th1 differentiation and instead generate mostly Th2 responses (11, 14). These data are consistent with the view that TLRs primarily control the induction of Th1 but not Th2 responses (13).

Immune responses are tailored to the threat posed by specific types of microbes (reviewed in ref. 15). For example, intracellular pathogens and mycobacterial infections instigate a robust Th1 response for clearance, but helminthic infestation generates a Th2 response. Therefore, it is likely that distinct combinations of activated TLRs triggered by each microbe might dictate the outcome that occurs, and TLRs are perhaps best thought of as a kind of microbial “barcode reader” (16).

Exactly how the complete response to the barcode occurs is not clear, but it is likely that phagosome and cell surface signaling are differentially involved (Figure 2).

But in MyD88-deficient mice, TLR signaling is severely disabled. How then, in the study by Fremond et al. (3), could adaptive responses to MTB-specific antigens be preserved? This is difficult to answer right now. On one hand, we do not yet know enough about the specific TLR signaling events that mediate the response to MTB infection. That said, it seems likely that MyD88 deficiency critically interferes with one or more steps in the process (17, 18). For example, phagocytosis of the pathogen can be accomplished by a number of different receptors present on professional APCs; the particular receptor involved appears to be an important determinant of intracellular fate and the subsequent immune response. Some TLR signaling originates from phagosomes and might be expected to control specific aspects of the overall response. Separately, recognition of MTB at the cell surface by TLRs is also an important determinant of the effectiveness of the host response. MTB products can be recognized by TLR2 or TLR4, and in addition, TLR9 can sense the presence of MTB DNA (Figure 1). Production of both inflammatory and anti-inflammatory cytokines and chemokines is another key step in the defensive response that may be particularly relevant to the findings of Fremond et al. (3). Molecules that generate TLR2-dependent signals are entirely dependent upon MyD88, but as noted above, TLR4 has MyD88-independent options available for signaling. It is possible, then, that some of these processes could be mediated by TLR4 in a MyD88-independent manner. Alternatively, other innate immune receptors such as nucleotide-binding oligomerization domain (NOD) may play a more important role than presently understood. Obviously, considerable work needs to be done to clarify these issues.

**Immune mechanisms in mycobacterial infection: controversies**

The work of Fremond et al. (3) generates interesting questions and adds to a growing controversy. First of all, is it really true that MyD88 is not involved in regulating the adaptive immune response to MTB?
Scanga et al. (19) first established that host resistance to infection with MTB is dependent on MyD88 and therefore strongly implicated TLR signaling in this response, since mice deficient in the IL-1 or IL-18 signaling pathway show only minor alterations in the normal defense to MTB infection (reviewed in ref. 19). There is evidence to corroborate this and suggest that the principle may be more general. MyD88-null mice have been shown to be highly susceptible to a number of pathogens, including Listeria monocytogenes (20), Staphylococcus aureus (21), Toxoplasma gondii (22), and Mycobacterium avium (23). In the study by Scanga et al. (19), expression of IL-12, TNF-α, IFN-γ, and nitric oxide synthase 2 were markedly decreased but not completely eliminated in MTB-infected MyD88-deficient mice, which indicates a severe impairment in the normal adaptive immune response to this infection. In another study, adaptive immunity and antigen-specific T cell activation were significantly impaired in CD4+ T cells from MyD88−/− mice infected with M. avium (23). Therefore, it is unclear why adaptive immunity would be impaired for M. avium infection but conserved for M. tuberculosis, as reported by Fremond et al. (3).

Surprisingly, mice deficient in TLR2 (24, 25), TLR4 (24, 26, 27), or TLR6 (25), all of which rely heavily (but not exclusively) in the case of TLR4) on MyD88, show no defects in acute resistance to aerogenic MTB infection. Another study reported that MyD88-null mice may not always succumb to MTB infection (28). Furthermore, a recent study determined that gene expression in macrophages infected with MTB in vitro is largely MyD88 independent (29). On the other hand, mice deficient in MyD88 could mount an apparently normal adaptive response against the intracellular pathogen Listeria monocytogenes (30) and Borrelia burgdorferi (31), which suggests (like Fremond et al.; ref. 3) that MyD88 may not always be needed for an effective adaptive defense. Other data are consistent with this conclusion as well: mice deficient in both MyD88 and TRIF may be tentatively suggested that MyD88 regulates resistance to at least some strains of MTB, but if this is true, how is it accomplished? Does MyD88 control genetic programs only within the jurisdiction of innate immunity, or does it have a more general function in modulating crucial interactions between innate and adaptive immunity? In this regard, Fremond’s conclusion that there is no significant impact of MyD88 on the adaptive response is at odds with the generally accepted model that adaptive immunity is controlled by innate mechanisms and therefore, by extension, MyD88 (13). That said, the contribution of other innate signaling mechanisms besides TLRs to regulation of the adaptive response to MTB infection has not been fully defined. For example, the intracellular NOD proteins, which recognize pathogen-associated molecular patterns independently of TLR signaling, can activate NF-κB and increase production of IFN-γ (32). In addition, double-stranded RNA can elicit an innate response independently of TLR3 and TRIF (33).

**TB or not TB: the unanswered questions**

For now, these data and the current study by Fremond et al. (3) should make one thing very clear: we do not yet understand the signaling mechanisms, transcriptional responses, subcellular processes, or cell-cell interactions that follow MTB infection. This leads us to a series of provocative questions directly relevant to the work of Fremond et al. Is adaptive immunity really dependent upon innate immunity? If MyD88 is not involved in regulating the adaptive response to MTB infection, then what is? What are the MTB-derived ligands and the TLRs that instigate signaling? TLR signaling involves complex and variable assemblies of cofactors and adapters for efficient transduction; how are expression and availability of these accessory molecules regulated, and how important might that be to the response? Does interaction among TLRs or downstream crosstalk play a key role in the response? What are the roles for specific components are dependent both on the nature of the microbe and perhaps also the site of infection.

Reconciling these discordant results is not straightforward. Possible explanations could include differences in MTB strains used, differences in the nature and the dose of the microbe, and differences in genetic backgrounds of MyD88-null mice. But it can be tentatively suggested that MyD88 regulates resistance to at least some strains of MTB. But if this is true, how is it accomplished? Does MyD88 control genetic programs only within the jurisdiction of innate immunity, or does it have a more general function in modulating crucial interactions between innate and adaptive immunity? In this regard, Fremond’s conclusion that there is no significant impact of MyD88 on the adaptive response is at odds with the generally accepted model that adaptive immunity is controlled by innate mechanisms and therefore, by extension, MyD88 (13). That said, the contribution of other innate signaling mechanisms besides TLRs to regulation of the adaptive response to MTB infection has not been fully defined. For example, the intracellular NOD proteins, which recognize pathogen-associated molecular patterns independently of TLR signaling, can activate NF-κB and increase production of IFN-γ (32). In addition, double-stranded RNA can elicit an innate response independently of TLR3 and TRIF (33).
functions of cell surface signaling by TLRs in MTB infection compared with signaling that originates from intracellular sources such as phagosomes and endosomes? Are there different genetic programs controlled by these? How is the entire machinery fine tuned to prevent an inappropriate degree of inflammation, and how is it turned off when it is no longer necessary?

An intriguing model that may at least partly resolve some of the discrepancies outlined above suggests that the TLR-dependent response of APCs is defined by the cell type and the niche in which the cell resides. Closely related lineages that express similar repertoires of TLRs may nevertheless respond very differently to the presence of TLR ligands (13). And finally, McCaffrey and colleagues recently reported that distinct gene clusters may be induced by particular pathogens depending on their subcellular localization in a manner that is independent of TLRs and MyD88 (34).

What dreams may come?

The questions posed outline fertile ground for future investigations that will doubtlessly provide insights that we, like Hamlet, can only wonder about. And the same dilemma that beset Hamlet—‘To be, or not to be: that is the question’—cousin earlier also confronted evolution: how best to respond to the slings and arrows of pathogenic invasion by MTB? Be it coincidence or sublime irony, the resolution arrived at by evolution is the same as Hamlet’s:

But that the dread of something after death,
The undiscover’d country from whose bourn
No traveller returns, puzzles the will . . .
Thus conscience does make cowards of us all.

— William Shakespeare, Hamlet, act 3, scene 1

The internal drives of Hamlet and evolution continue to fuel efforts to take arms against a sea of troubles, to understand how immune defenses protect us from MTB, and so to develop more effective ways to counter the ability of pathogens to cause us to shuffle off this mortal coil. The work of Fremond et al. (3) now takes us a step further along this path.

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