Neutrophil NADPH oxidase plays a key role in host defense and in inflammation by releasing large amounts of superoxide and other ROSs. Prolinflammatory cytokines such as GM-CSF and TNF-α prime ROS production by neutrophils through unknown mechanisms. Here we used peptide sequencing by tandem mass spectrometry to show that GM-CSF and TNF-α induce phosphorylation of Ser345 on p47phox, a cytosolic component of NADPH oxidase, in human neutrophils. As Ser345 is located in the MAPK consensus sequence, we tested the effects of MAPK inhibitors. Inhibitors of the ERK1/2 pathway abrogated GM-CSF–induced phosphorylation of Ser345, while p38 MAPK inhibitor abrogated TNF-α–induced phosphorylation of Ser345. Transfection of HL-60 cells with a mutated p47phox (S345A) inhibited GM-CSF– and TNF-α–induced priming of ROS production. This event was also inhibited in neutrophils by a cell-permeable peptide containing a TAT-p47phox-Ser345 sequence. Furthermore, ROS generation, p47phox-Ser345 phosphorylation, and ERK1/2 and p38 MAPK phosphorylation were increased in synovial neutrophils from rheumatoid arthritis (RA) patients, and TAT-Ser345 peptide inhibited ROS production by these primed neutrophils. This study therefore identifies convergent MAPK pathways on Ser345 that are involved in GM-CSF– and TNF-α–induced priming of neutrophils and are activated in RA. Inhibition of the point of convergence of these pathways might serve as a novel antiinflammatory strategy.

Introduction

Neutrophils play a key role in host defenses against invading microorganisms and have a major role in inflammation (1–4). In response to a variety of agents, they release large quantities of superoxide anion (O₂⁻) and other ROSs in a phenomenon known as the respiratory burst (4). Neutrophil production of O₂⁻ is dependent on activation of NADPH oxidase, a multicomponent enzyme system that catalyzes NADPH-dependent reduction of oxygen to O₂⁻ (5–7). In resting cells, NADPH oxidase is inactive, and its components are distributed between the cytosol and membranes. When cells are activated, the cytosolic components of the phagocyte oxidase (phox), such as p47phox, p67phox, p40phox, and Rac2, migrate to the membranes, where they associate with the membrane-bound component (flavocytochrome b558) to assemble the catalytically active oxidase (8–11). Upon NADPH oxidase activation, p47phox, p67phox, p40phox, and p22phox become phosphorylated (12–16). p47phox phosphorylation on several serines plays a pivotal role in oxidase activation in intact cells (17–19). As oxidants produced by NADPH oxidase are highly toxic not only for infectious agents but also for neighboring host tissues, tight regulation of the enzyme complex is necessary to control their production. Phosphorylation/dephosphorylation of the oxidase subunits is one such regulatory mechanism. Various kinases have been shown to phosphorylate p47phox in vitro, but the regulatory pathways involved in the in vivo priming and activation steps are unclear.

Neutrophil O₂⁻ production can be potentiated by prior exposure to “priming” agents such as the proinflammatory cytokines GM-CSF, TNF-α, and IL-8 (20, 21). These cytokines inherently induce a very weak oxidative response by neutrophils, but they strongly enhance neutrophil release of ROS on exposure to a secondary applied stimulus such as bacterial N-formyl peptides (22, 23). The intimate mechanisms involved in the priming process are poorly understood. Some studies have suggested that priming is regulated at the receptor and heterotrimeric G-protein levels (24, 25) or through an increase in cytochrome b558 expression (26–28). We and others have reported that priming of the human neutrophil respiratory burst by GM-CSF, LPS, and TNF-α is associated with partial phosphorylation of the cytosolic NADPH oxidase component p47phox (26, 29–31), but the identity of the phosphorylated serines and the kinases involved in their phosphorylation remained to be discovered. No direct link between the phosphorylation of p47phox and the priming process has yet been established.

The GM-CSF receptor is composed of 2 chains, designated α and β (32, 33). Engagement of GM-CSF with its receptor activates a number of signal transduction pathways in human neutrophils, including protein tyrosine kinase, PI3K (34–36), and the MAPK family members ERK1/2 (37–40). GM-CSF binding to its receptor induces and increases the binding of the protein tyrosine kinase JAK2 and the src-tyrosine kinase lyn to the intracellular domain of the receptor (34, 41, 42). These tyrosine kinases transduce the GM-CSF signal by phosphorylating other

Nonstandard abbreviations used: fMLP, formyl-methionyl-leucyl-phenylalanine; phox, phagocyte oxidase.

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proteins such as STAT, PKB, and PI3K. TNF-α exerts its cellular effects by binding to its 2 receptors, p55 and p75. Neutrophils express both receptor types (43), which are linked to different signaling pathways such as p38 MAPK (27, 39). The molecular mechanisms by which GM-CSF and TNF-α prime the respiratory burst in human neutrophils are not fully known.

RA is a systemic inflammatory disorder most commonly targeting the joints. The pathophysiology of RA involves dysregulated cytokine production and neutrophil accumulation in synovial fluid (44). Both excessive production of ROS and release of degradative enzymes by neutrophils have been implicated in rheumatoid tissue damage (4, 45).

The aims of this study were to identify the p47phox site that is phosphorylated during TNF-α- and GM-CSF–induced priming of the oxidative burst; to determine the protein kinase(s) involved in this process; and to determine whether these phosphorylation events occur during an inflammatory disease. We used tandem mass spectrometry (MS/MS) to show that phosphorylation of Ser345 on p47phox is directly related to GM-CSF– and TNF-α–induced priming of ROS production. These results show that Ser345 is a point of convergence used by different MAPK activities to induce priming of ROS production. The selective phosphorylation of Ser345 on p47phox is, to our knowledge, a novel priming mechanism and might serve as a novel antiinflammatory target.

**Results**

**Identification of the GM-CSF– and TNF-α–induced p47phox phosphorylation target site.** In formyl-methionyl-leucyl-phenylalanine– (fMLP–) or PMA-stimulated human neutrophils, p47phox is phosphorylated on several serines located between Ser303 and Ser379 (17, 18). In contrast, in GM-CSF– or TNF-α–primed human neutrophils, p47phox is partially phosphorylated on a common peptide that could contain several serines as potential targets of the GM-CSF– and TNF-α priming cascades (29, 30). In order to identify the p47phox site(s) that are phosphorylated upon GM-CSF or TNF-α exposure, we immunopurified p47phox from GM-CSF– and TNF-α–treated cells (5 × 10⁶) and analyzed it by mass spectrometry. The protein was digested with trypsin and endoproteinase Lys-C, and the resulting peptide mixtures were analyzed by liquid chromatography–elec-
The phosphopeptide (amino acids 338–354) was detected at 26 minutes and was automatically selected for sequencing by MS/MS (Figure 1). (The same result was obtained with GM-CSF and TNF-α, but only the GM-CSF result is shown here). The presence of a phosphoserine at position 345 (QARPGPQ[pS]PGSPLEEER) was revealed by MS/MS, based on the mass difference of 69 Da (dehydroalanine) between the y9 and y10 fragment ions (Figure 1B). Dehydroalanine is generated upon phosphoric acid elimination from phosphoserine and is commonly observed in MS/MS (46, 47). The MS/MS spectrum also demonstrated that Ser348 was not phosphorylated, as dehydroalanine was not detected at this position.

**Figure 2**
Use of an antibody to phospho-Ser345 demonstrates that GM-CSF and TNF-α induce phosphorylation of p47^phox^ on Ser345 in a concentration- and time-dependent manner. (A) Neutrophils (1 x 10^7^ cells/ml) were incubated with various concentrations of GM-CSF for 20 minutes or were incubated with GM-CSF (12.5 ng/ml) for the times indicated. Cells were lysed, and proteins from 4 x 10^5^ cells were analyzed with SDS-PAGE and immunoblotting with anti–phospho-Ser345 antibody (p-Ser345) or anti-p47^phox^ antibody (p47^phox^). (B) Neutrophils (1 x 10^7^ cells/ml) were incubated with various concentrations of TNF-α for 20 minutes or were incubated with TNF-α (10 ng/ml) for the times indicated. Cells were lysed, and proteins from 4 x 10^5^ cells were analyzed with SDS-PAGE and immunoblotting with anti–phospho-Ser345 antibody or anti-p47^phox^ antibody. Western blots from different experiments were scanned; phosphorylated and total p47^phox^ were quantified by densitometry; and the intensity of phosphorylated p47^phox^ was corrected for the amount of p47^phox^. Results are expressed as mean ± SEM (n = 3). *P < 0.05 compared with untreated neutrophils or time 0 minutes.
Use of an antibody against phospho-Ser345 demonstrates that GM-CSF and TNF-α induce Ser345 phosphorylation. To examine Ser345 phosphorylation status in cellular extracts, we prepared a polyclonal antibody directed against phospho-Ser345 by using as antigen the peptide QARPGQ[pS]PGSPLEER (p47phox amino acids 338–354). Western blot analysis of whole-cell lysates from GM-CSF– or TNF-α–treated neutrophils (from as few as 4 × 10^5 cells) showed (Figure 2, A and B; p-Ser345) that this antibody was highly specific for phosphorylated p47phox, as it did not recognize other phosphorylated proteins. Total p47phox protein was detected with an antibody that recognizes the nonphosphorylated as well as the phosphorylated protein (Figure 2, A and B; p47phox). Phosphorylated and total p47phox from several experiments were quantified by densitometry and the amount of phosphorylated p47phox corrected for the amount of p47phox. Together, the results show that GM-CSF– and TNF-α–induced p47phox phosphorylation at Ser345 were both concentration and time dependent and closely matched the priming effects of GM-CSF and TNF-α on O2- production (29, 30) (Figure 2, A and B).

We next examined whether other agents known to activate neutrophils such as the chemoattractant peptide fMLP and the protein kinase C activator PMA could induce Ser345 phosphorylation. Time course and dose-response studies (Figure 3, A and B) showed that fMLP and PMA weakly induced Ser345 phosphorylation as compared with the phosphorylation induced by the proinflammatory cytokines GM-CSF and TNF-α. Specific detection of p47phox with an antibody that recognizes the nonphosphorylated as well as the phosphorylated protein showed that the same amount of p47phox was present in each sample. These results suggest that Ser345 is more strongly phosphorylated during GM-CSF– or TNF-α–induced priming than in stimulatory conditions.

Effect of tyrosine kinase and MAPK inhibitors on GM-CSF– and TNF-α–induced p47phox phosphorylation. Myeloid cells such as neutrophils are terminally differentiated, short-lived cells and are resistant to transfection. An alternative strategy to study the role of specific enzymes is to use cell-permeant pharmacologic inhibitors. In previous studies (29, 30) we showed, by 32P labeling, that the broad-range protein tyrosine kinase inhibitor genistein inhibited GM-CSF– and TNF-α–induced p47phox phosphorylation. To verify that Ser345 phosphorylation detected with the specific antibody is controlled by the same upstream tyrosine kinases, we tested the effects of genistein on this process. First, p47phox from 32P-labeled neutrophils (5 × 10^5 cells) was immunoprecipitated with anti-p47phox and analyzed by SDS-PAGE, Western blotting, and autoradiography (Figure 4, A and B; [32P]p47phox). Second, total cell lysates (4 × 10^5 cells) were analyzed by SDS-PAGE and Western blotting with the anti–phospho-Ser345 antibody (Figure 4, A and B; p-Ser345). Both methods showed that genistein inhibited GM-CSF– and TNF-α–induced p47phox phosphorylation, confirming that the anti–phospho-Ser345 antibody recognizes the phosphorylated site on p47phox induced by GM-CSF and TNF-α and that a genistein-sensitive tyrosine kinase controls this event.

As Ser345 is located in a sequence recognized and phosphorylated by MAPK (–PXSP–), we examined which MAPK was involved in GM-CSF– and TNF-α–induced p47phox phosphorylation at Ser345 by testing the effects of different MAPK inhibitors. As GM-CSF and TNF-α induce the activation of ERK1/2 and p38 MAPK, respectively, in suspended human neutrophils (Figure 5A and refs. 39, 40), we used PD98059 and UO126, which inhibit MEK1/2 (the upstream activator of ERK1/2) (48, 49) and also SB203580, a p38 MAPK inhibitor (50), to analyze the role of these 2 MAPK pathways in p47phox phosphorylation in intact neutrophils. Neutrophils were incubated for 45 minutes with PD98059 (50 μM), UO126 (10 μM), or SB203580 (10 μM), then treated with GM-CSF or TNF-α. p47phox phosphorylation status was analyzed by comparing the results of the 2 methods (32P labeling and anti–phospho-Ser345 antibody). As shown in Figure 5B, PD98059 and UO126 abrogated p47phox phosphorylation on Ser345 in parallel with the inhibition of ERK1/2 phosphorylation (Figure 5B;
p-ERK1/2) induced by GM-CSF. SB203580 had no inhibitory effect; it rather moderately enhanced GM-CSF-induced phosphorylation of p47^phox and ERK1/2, an effect not further examined here. This strongly pointed to ERK1/2 as the kinases involved in p47^phox phosphorylation on Ser345 in GM-CSF–primed human neutrophils. However, with regard to TNF-α–treated neutrophils, the p38 MAPK inhibitor SB203580 inhibited phosphorylation of p47^phox on Ser345, but the MEK1/2 inhibitors (PD98059 and U0126) had no effects (Figure 5C). In these conditions SB203580 inhibited phosphorylation of hsp27, a known substrate of MAPK-activated kinase 2, which is a downstream target of p38 MAPK. In all these experiments, specific detection of p47^phox, ERK1/2, and hsp27 with specific antibodies that recognize the nonphosphorylated as well as the phosphorylated proteins showed that the same amounts of each protein were present in each sample (Figure 5, A–C, and data not shown). All these results suggest that ERK1/2 are the kinases involved in p47^phox phosphorylation on Ser345 in GM-CSF–primed human neutrophils, while p38 MAPK is the kinase involved in p47^phox phosphorylation on Ser345 in TNF-α–primed human neutrophils. These 2 pathways could be convergent on the phosphorylation of p47^phox on this specific site.

**Figure 5** Effect of MAPK inhibitors on GM-CSF– and TNF-α–induced p47^phox and MAPK activation. (A) Neutrophils (1 × 10^6 cells/ml) were incubated with GM-CSF (12.5 ng/ml) or TNF-α (10 ng/ml) for 20 minutes and lysed, and proteins from 4 × 10^6 cells were analyzed with SDS-PAGE and immunoblotting with anti–phospho-ERK1/2 or anti–phospho-p38 antibody (p-p38MAPK). (B and C) Neutrophils were incubated with SB203580 (10 μM), PD98059 (50 μM), or UO126 (10 μM) for 30 minutes, then treated with GM-CSF (B) or TNF-α (C) for 20 minutes. p47^phox from 32P-labeled neutrophils (5 × 10^6 cells) was immunoprecipitated with anti-p47^phox antibody and analyzed by SDS-PAGE, Western blotting, and autoradiography. Total cell lysates (4 × 10^6 cells) from unlabeled cells were also analyzed by SDS-PAGE and Western blotting with anti–phospho-Ser345 antibody or anti–phospho-ERK1/2 antibody (p-ERK1/2), which reflects MEK1/2 activity. p38 MAPK activity was assessed by in vitro phosphorylation of hsp27, as described in Methods. Data are representative of 3 experiments.

**Figure 6** Mutation of Ser345 to Ala inhibits the priming process in HL-60 cells. To test whether phosphorylation of Ser345 of p47^phox is required for NADPH oxidase priming in intact cells, we used a plasmid that encodes p47^phox in which Ser345 was mutated to alanine (S345A), as compared with a plasmid that encodes WT p47^phox. Because neutrophils are terminally differentiated, short-lived cells that are resistant to transfection, we used HL-60 cells, which can be transfected and differentiated into neutrophil-like cells. Results showed (Figure 6A) that nondifferentiated transfected HL-60 cells expressed the same amount of WT and mutant p47^phox but not p67^phox, while after differentiation an increase in p47^phox expression and in p67^phox expression occurred. ROS production by differentiated HL-60 cells transfected with WT p47^phox was primed by TNF-α or GM-CSF. In contrast, ROS production by differentiated HL-60 cells transfected with the S345A-p47^phox mutant was not primed by TNF-α or GM-CSF, as the level of ROS production after TNF-α or GM-CSF
massive ROS release are believed to contribute to tissue injury in inflammatory diseases. ROS release can be potentiated by the presence of proinflammatory cytokines such as GM-CSF and TNF-α at the inflammatory site. Because proinflammatory cytokines are involved in several inflammatory diseases, such as RA, and anti-TNF-α therapies are very beneficial in this disease, we examined NADPH oxidase priming, p47^phox-Ser345 phosphorylation, and ERK1/2 or p38 MAPK phosphorylation status in neutrophils from patients with RA. The results showed (Figure 8A) that neutrophils from synovial fluid of RA patients had higher basal NADPH oxidase activity than neutrophils isolated from blood of the same patient. When stimulated with fMLP, neutrophils from synovial fluid of RA patients had higher ROS production than circulating neutrophils (Figure 8B), suggesting that they were primed. Results obtained from 10 patients are summarized in Figure 8C. Furthermore, phosphorylation of p47^phox on Ser345, phosphorylation of ERK1/2, and phosphorylation of p38 MAPK were all increased in RA patients’ synovial neutrophils as compared with their blood neutrophils (Figure 9). This result was obtained from 10 different RA patients, but data for only 3 are shown on this figure. In all these experiments, specific detection of p47^phox, ERK1/2, and p38 MAPK with specific antibodies that recognize the nonphosphorylated as well as the phosphorylated proteins showed that the same amounts of each protein were present in each sample. Taken together, these results suggest that resident synovial neutrophils from RA patients are in a primed state and that phosphorylation of p47^phox on Ser345 by convergent kinases such as ERK1/2 and p38 MAPK could be the molecular basis of this process.

The cell-permeable peptide TAT-Ser345 inhibits basal and primed ROS production by neutrophils from RA patients. The effect of the TAT-Ser345 peptide was investigated in this inflammatory disease situation in which neutrophils from synovial fluid are primed and produce high level of ROS. Results showed (Figure 10) that TAT-Ser345 peptide has a strong inhibitory effect on basal ROS production by synovial neutrophils from RA patients even at a low concentration such as 10 μM. fMLP-stimulated ROS generation of these primed synovial neutrophils was also inhibited by TAT-Ser345 peptide. In contrast, TAT-scramble peptide did not inhibit basal or fMLP-stimulated ROS production. This result suggests that TAT-Ser345 peptide could have antiinflammatory properties.

Discussion
The molecular mechanisms underlying human neutrophil respiratory burst priming are unknown. Phosphorylation of the cytosolic subunit p47^phox is necessary for NADPH oxidase activation and regulation. The proinflammatory cytokines GM-CSF and TNF-α induce partial p47^phox phosphorylation in human neutrophils, but the phosphorylated serine remained to be identi-
Primed NADPH oxidase activity in neutrophils isolated from synovial fluid of patients with RA. (A) Resting neutrophils (5 × 10⁶ cells) isolated from blood or from synovial fluid of RA patients were incubated in HBSS in the presence of luminol (10 μM), and spontaneous (without stimulation) chemiluminescence was measured over time. (B) The same preparation was then stimulated with fMLP (10⁻⁷ M), and chemiluminescence was measured over time. (C) Ox• production was quantified by cytochrome c reduction assay; control (100%) basal O₂•⁻ production was 0.14 ± 0.01 nmol/min/10⁶ cells, and control (100%) fMLP-induced O₂•⁻ production was 3.6 ± 0.5 nmol/min/10⁶ cells. Data are presented as mean ± SEM (n = 10). *P < 0.05 compared with control.

This is the first report showing a pivotal role of a specific serine phosphorylated on p47phox by convergent MAPKs in the respiratory burst priming of neutrophils in an inflammatory environment. This phosphorylated site could potentially serve as a pharmacological target in RA and other inflammatory diseases.

In fMLP- or PMA-activated neutrophils, p47phox is phosphorylated on several serines (17, 18). Tryptic peptide mapping of p47phox showed that these phosphorylated serines were located on several peptides on the phosphopeptide map. We have shown that GM-CSF induces the phosphorylation of a single major p47phox peptide (29), suggesting that GM-CSF induces partial phosphorylation of p47phox only on certain serines; this might potentiate the phosphorylation of the other serines after secondary fMLP stimulation. The position of the phosphorylated peptides on the p47phox phosphopeptide map identified serines 345, 348, 359, and 370 as candidate targets for GM-CSF–induced phosphorylation. In this study, MS/MS analysis identified Ser345 as the phosphorylated serine. It is noteworthy that 2-dimensional phosphopeptide mapping of phosphorylated p47phox in TNF-α–primed neutrophils revealed partial phosphorylation of p47phox on the same peptide as in GM-CSF–exposed neutrophils (30), suggesting that the same phosphorylated site is targeted during priming. Indeed, our results show that phosphorylation of Ser345 is also induced by TNF-α.

To our surprise, Ser345 was only moderately phosphorylated in neutrophils stimulated with fMLP or PMA, suggesting that phosphorylation of this site is selectively targeted by priming conditions. A cell-permeable peptide containing the Ser345 sequence specifically inhibited the priming effect of GM-CSF and TNF-α on neutrophil O₂•⁻ production, suggesting that Ser345 phosphorylation plays a functional role by potentiating NADPH oxidase activation. Mutation studies of p47phox serines in EBV-transformed lymphoblasts (19) showed that mutation of Ser345 to Ala had only a moderate effect on PMA-induced O₂•⁻ production. Our results are in keeping with these data, showing that phosphorylation of Ser345 was weakly induced by PMA, and also lend further weight to the hypothesis that Ser345 is specifically induced and has a role in the priming process.

As Ser345 is located in the MAPK phosphorylation consensus sequence (-PXSP-), we tested the effects of MAPK inhibitors on this phosphorylation. We found that PD98059 and UO126, which inhibit MEK1/2 (the upstream activator of ERK1/2), abrogated p47phox Ser345 phosphorylation induced by GM-CSF, while SB203580, an inhibitor of p38 MAPK, had no inhibitory effect. On the other hand, in TNF-α–treated neutrophils, SB203580 inhibited p47phox Ser345 phosphorylation, but PD98059 and UO126 had no effect. As p47phox is a good in

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fied, and a direct link to the priming of ROS production remained to be determined. In this study we show that GM-CSF and TNF-α induce p47phox phosphorylation on Ser345. In the respiratory burst priming conditions induced by GM-CSF and TNF-α, phosphorylation of this site was markedly higher than in stimulatory conditions created by the chemoattractant peptide fMLP or the protein kinase C activator PMA. We also show that ERK1/2 are the kinases involved in GM-CSF–induced phosphorylation of Ser345 and that p38 MAPK is the kinase involved in TNF-α–induced phosphorylation of Ser345. Mutation of this serine specifically inhibited the priming effect of GM-CSF and TNF-α on O₂•⁻ production by neutrophils in response to fMLP. Furthermore, the TAT-Ser345 peptide inhibited ROS production by synovial neutrophils of these RA patients. To our knowledge,
vitro substrate for ERK2 and P38 MAPK (53, 54), these 2 kinases could target p47phox in intact neutrophils when recruited by specific agonists (GM-CSF and TNF-α, respectively). In addition, inhibitors of ERK1/2 and p38 MAPK inhibited neutrophil respiratory burst priming by GM-CSF and TNF-α (data not shown and refs. 27, 31, 39, 40). These results strongly suggest that ERK1/2 are the kinases that mediate p47phox phosphorylation on Ser345 in GM-CSF–treated neutrophils and that p38 MAPK is the kinase that mediates p47phox phosphorylation on Ser345 in TNF-α–treated neutrophils. Ser345 serves as a point of convergence of different MAPKs to prime ROS production. Several reports suggest that phosphorylation of p47phox induces conformational changes (55–57) that initiate assembly of the active enzyme via interaction of the SH3 domains with the proline-rich region of p22phox. p47phox contains 2 SH3 domains, one of which interacts intramolecularly with a p47phox-polypoline sequence in the nonphosphorylated protein. This interaction switches to the p22phox-polypoline sequence when p47phox is phosphorylated. Pre-phosphorylation of p47phox on Ser345 by ERK1/2 or p38 MAPK could facilitate conformational changes induced by a secondary phosphorylation step.

Figure 9
Phosphorylation of p47phox on Ser345 and phosphorylation of ERK1/2 and p38 MAPK are increased in neutrophils isolated from synovial fluid of patients with RA. Resting neutrophils (5 × 10^5 cells) isolated from blood or from synovial fluid of RA patients were lysed, and proteins were analyzed with SDS-PAGE and immunoblotting with anti–phospho-Ser345 antibody, anti–phospho-ERK1/2, or anti–phospho-p38 antibody. Total p47phox, ERK1/2, and p38 MAPK were detected using specific antibody. Results from 3 of 10 RA patients are shown.

We compared NADPH oxidase activation in circulating and synovial neutrophils from RA patients. Neutrophils isolated from synovial fluid showed a stronger oxidative burst than neutrophils isolated from peripheral blood of the same patients and of healthy controls. These results showed that the patients’ synovial neutrophils were primed. In addition, p47phox-Ser345 phosphorylation and the MAPKs (ERK1/2 and p38) that mediate this process were upregulated. These results suggest that increased NADPH oxidase activity and excessive ROS generation could be involved in arthritic joint inflammation and destruction, through increased p47phox phosphorylation. This would be related to high levels of proinflammatory cytokines in synovial fluid, such as TNF-α, GM-CSF, IL-8, or other agents that activate ERK1/2 or p38 MAPK. Additionally, the high levels of TNF-α in synovial fluid from RA patients, along with the beneficial effect of anti-TNF-α therapies in this rheumatic disease, strongly support a pivotal action of this cytokine in the pathogenesis of the joint lesions. The effect of TAT-Ser345 peptide was tested on neutrophils from synovial fluid of RA patients. It is noteworthy that TAT-Ser345 peptide has a strong inhibitory effect on basal ROS production by neutrophils from RA patients even at low concentration. fMLP-stimulated

Figure 10
The cell-permeable peptide TAT-Ser345 inhibits the basal and primed ROS production by neutrophils from patients with RA. Neutrophils isolated from RA patients (5 × 10^5) were incubated with a cell-permeable peptide corresponding to TAT-peptide-Ser345 sequence or with a scramble peptide for 30 minutes, then ROS production was measured with the luminol-amplified chemiluminescence technique in the absence (A) or presence (B) of fMLP (10^-7 M). Data are presented as mean ± SEM (n = 3). *P < 0.05.
ROS generation by primed neutrophils from RA patients was also inhibited. This result suggests that Ser345 could be continuously phosphorylated/dephosphorylated in inflammatory conditions and this inhibitory peptide could have anti-inflammatory properties involving inhibition of the phosphorylation process.

In conclusion, this study shows that, in respiratory burst priming conditions, GM-CSF and TNF-α induce p47phox phosphorylation on Ser345, while stimulation with the bacterial N-formyl peptide fMLP has much weaker effects. Phosphorylation of this site can be mediated by ERK1/2 (in GM-CSF–primed cells) or by p38 MAPK (in TNF-α-primed cells). A cell-permeable peptide containing the Ser345 sequence specifically inhibited the priming effect of GM-CSF and TNF-α on ROS production. Furthermore, neutrophils isolated from synovial fluid of RA patients were found to be primed for ROS production and exhibited specific phosphorylation of p47phox on Ser345, as well as upregulation of both ERK1/2 and p38 MAPK phosphorylation. The TAT-Ser345 peptide inhibited ROS production by synovial neutrophils of these patients. These results strongly suggest that Ser345 serves as a point of convergence of different MAPKs to induce priming of ROS production. Pharmacological inhibition of this common phosphorylation site on p47phox might avoid exaggerated neutrophil responses in inflammatory environments while preserving the physiological ability of the bacterial N-formyl peptides to activate neutrophils.

Methods
Reagents and antibodies. Recombinant human GM-CSF and TNF-α were from R&D Systems and PeproTech. The fMLP, PMA, and proteinase Gphosphatase inhibitors were from Sigma-Aldrich. [32P]orthophosphoric acid was from PerkinElmer. Kinase inhibitors were from Calbiochem. EDTA, 15 mM NaF, 5 mM NaVO4, 1.5 mM PMSF, 1 mM diisopropyl fluorophosphate, 0.5 mM Triton X-100, 25 mM NaF, 5 mM NaVO4, 5 mM β-glycerocephosphate, 1 mM p-nitrophenol phosphate, 1 mg/ml DNase I) and sonication on ice (3 × 15 s); the lysate was then centrifuged at 100,000 × g for 30 minutes at 4°C in a TL100 ultracentrifuge (Beckman Coulter). The cleared supernatant was incubated overnight with anti-p47phox (1:200), and protein was immunoprecipitated with GammaBind G Sepharose beads (GE Healthcare), washed 4 times in lysis buffer, and denatured in Laemmli sample buffer. The samples were subjected to SDSPAGE in 10% polyacrylamide gels, using standard techniques (58). The separated proteins were stained with Coomassie blue, and the protein band corresponding to p47phox was cut out, treated with protease, and analyzed by mass spectrometry as described below.

Preparation of p47phox samples and automated nanoflow LC-MS/MS analysis. P47phox was isolated by SDS PAGE, stained with Coomassie blue, and digested in situ with trypsin and LysC. Extracted peptide mixtures were analyzed with a nanoflow capillary high-pressure liquid chromatography system (UltiMate; LC Packings) directly coupled to a Q-Tof tandem mass spectrometer (Q-Tof Ultima API, Waters) as described previously (46). Peptide mixtures were separated on a 9 cm custom-packed C18 reverse-phase column (75 μm ID ZORBAX SB-C18; Agilent Technologies, 3.5 μm particle size) during a 35-minute gradient of 0–90% acetonitrile (vol/vol) containing 1% (vol/vol) formic acid, 0.6% (vol/vol) acetic acid, and 0.005% (vol/vol) heptfluorobutryic acid at a flow rate of 175 nl/min. The mass spectrometer was calibrated with NaCl.

Mass spectra obtained by automated nanoflow LC-MS/MS, and data-dependent acquisition modes were analyzed with MassLynx 3.5 software (Waters) and Mascot MS/MS Ions Search software (Matrix Science). Tandem mass spectra obtained with phosphopeptides were manually inspected and compared with theoretical data obtained using the GPMW 6.20 program (Lighthouse data).

Detection of Ser345 phosphorylation in neutrophils by a specific antibody. Neutrophils (1 × 10⁷ cells/ml) were incubated in phosphate-free buffer (20 mM Hepes pH 7.4, 140 mM NaCl, 5.7 mM KCl, 0.8 mM MgCl₂, and 0.025% BSA) containing 0.5 μCi [32P]orthophosphoric acid/10⁶ cells/ml for 60 minutes at 30°C, as previously reported (29). Cells were then treated with GM-CSF or TNF-α. The reaction was stopped by adding ice-cold buffer and centrifugation at 400 g for 6 minutes at 4°C. The cells were lysed, and p47phox was immunoprecipitated as described above. The samples were subjected to SDS-PAGE in 10% polyacrylamide gels, using standard techniques. The separated proteins were transferred to nitrocellulose and detected as described previously (29).

Detection of Ser345 phosphorylation in neutrophils by a specific antibody. Neutrophils (1 × 10⁷ cells/ml) were incubated in various concentrations of GM-CSF, TNF-α, fMLP, or PMA at different incubation times. Cells were lysed in the lysis buffer as described above, and proteins from 4 × 10⁶ cells were analyzed with SDS-PAGE and immunoblotting with anti–phospho-Ser345 antibody (1:10,000 dilution) or anti-p47phox antibody (1:5,000 dilution) and HRP-labeled goat anti-rabbit antibody. The reaction was detected using ECL reagents.

HL-60 cell culture, transfection, and differentiation. HL-60 cells were purchased from ATCC and cultured in RPMI-1640 containing 10% heat-inactivated fetal bovine serum, 100 U/ml penicillin, 100 μg/ml streptomycin at 37°C in humidified atmosphere, 5% CO₂ in air. Plasmids (pEBO) that encode WT p47phox or p47phox in which Ser345 was mutated to alanine (S345A) were constructed as we have described (18, 19) (gift from B. Babior). HL-60 cells were transfected by electroporation using the same protocol as for B cells (59). After 48 hours, hygromycin B was added to a final concentration of 200 μg/ml, and the cells were grown under hygromycin B selection pressure for 2 weeks. p47phox protein expression in transfected HL-60 cells was monitored by Western blotting. For differentiation, exponentially growing cells were harvested and resuspended at a density of 5 × 10⁶ cells/ml in complete medium with hygromycin B in the presence of 1.3% DMSO. After 5 days, the cells were washed and incubated for 5 hours in RPMI alone, then assessed for NADPH oxidase activity using chemiluminescence.

Preparation of neutrophils from synovial fluid and peripheral blood from RA patients. With the subjects’ informed consent, knee-joint synovial fluid and peripheral blood were collected from patients who met the American College of Rheumatology criteria for RA (60, 61) and who did not receive any...
anti-cytokine biotherapy. Samples were collected in sterile lithium hepa-
rinate tubes. To make sure that neutrophil preparation was performed in 1-step centrifugation using a Polymorphprep gradient (Abcys SA; Axis-Shift). After centrifugation at 500 g for 30 minutes at 22 °C, the neutrophil band was collected and cells washed in PBS and counted, then used for ROS production and Western blots.

Measurement of O2− and ROS production. O2− production was measured in response to FMLP (10−7 M) in terms of O2−-dismutase-inhibitable ferricy-
toehrome c reduction as described previously (29) or the chemilumines-
cence method: cells (2.5 × 107) were suspended in 0.5 ml HBSS containing 10 μM luminal preheated to 37 °C in the thermostatted chamber of the luminometer (Biolumat LB937; Berthold). After a baseline reading, cells were stimulated with 10−7 M FMLP, and chemiluminescence was recorded.


