Supplementary Figure 2

A)

- isotype
- OVA-Vehicle-DC
- OVA-iloprost-DC

B) BAL-Macrophages

C) Lung-Macrophages
Supplementary Figure 3

The graph shows the migration index of DCs (dendritic cells) treated with different concentrations of CCL19. The treatments include PBS-DC, OVA-Vehicle DC, and OVA-iloprost-DC. The x-axis represents the concentration of CCL19 (100 ng, 10 ng, 1 ng, 0.1 ng), and the y-axis represents the migration index. The bars indicate the mean migration index with error bars for each treatment group at different CCL19 concentrations.
Supplementary Figure 4

The figure shows the levels of cytokines (IL-4, IL-5, IL-13, IFN-γ) expressed in different conditions: PBS-DC/Do11.10, OVA-DC/Do11.10, iloprost-OVA-DC/Do11.10. The data is represented as mean ± SEM, and statistical significance is indicated by * (p < 0.05), ** (p < 0.01), and *** (p < 0.001).
Supplementary Figure 1: Dose dependency of iloprost effect on asthmatic airway inflammation in vivo. Mice were sensitized by an i.p. injection of OVA/alum on days 0 and 7. On days 19-21, mice were exposed to OVA aerosols. 30 min before each aerosol, mice received an i.t injection of Vehicle or the indicated concentrations of iloprost A) BAL fluid was analyzed by flow cytometry. B) Mediastinal lymph node cell suspensions were restimulated in vitro for four days with OVA, and assayed for cytokine production using ELISA assay. Coding is as immunisation/treatment/challenge; n =6-8 mice per group. Data are shown as mean±SEM. *p<0.05, **p<0.01, ***p<0.001, representative of 3 independent experiments.

Supplementary Figure 2: Effect of iloprost on expression of co-stimulatory molecules in vitro and in vivo. A) BMDC were incubated with vehicle-OVA or iloprost-OVA overnight. CD11c+MHCIIhi DCs were analyzed for their expression of CD40, CD80, CD83 and CD86. Data from one representative experiment out of three is shown. B, C) Experiments were set up as in fig1. Groups are coded as sensitization/treatment/challenge. BAL was collected and single cell suspension was prepared from the lungs. BAL and lung macrophages were analyzed for their expression of CD40, CD80, CD86. Data from one representative experiment out of three is shown.

Supplementary Figure 3 Effect of iloprost treatment on migratory capacity of DCs in vitro

DCs were generated from bone marrow cells cultured in GM-CSF. After 9 days of culture, cells were exposed overnight to iloprost or to vehicle either in the presence or
absence of OVA antigen (containing a trace amount of LPS). Next, cells were placed in the upper chamber of a Transwell migration assay. OVA exposed DCs showed enhanced migration towards the CCR7 agonist CCL19 (MIP3β), and this was severely impaired by iloprost pretreatment.

Supplementary Figure 4: Effect of *in vitro* iloprost treatment on the capacity of DCs to polarize Ag specific T cells *in vitro*.

BMDCs were pulsed or not with 100 μg/ml of OVA overnight. DCs were also treated for 30 min with iloprost (iloprost/OVA-DCs) or vehicle (vehicle/OVA-DCs) before addition of OVA. DCs were collected and co-cultured for 4 days with naive OVA-specific CD4⁺ T cells. Cytokines were measured in supernatants by ELISA. Data are shown as mean±SEM. *p<0.05, **p<0.01, ***p<0.001, representative of 3 independent experiments.