Antigen-specific immune responses to influenza vaccine in utero

Deepa Rastogi,1 Chaodong Wang,2 Xia Mao,3 Cynthia Lendor,3 Paul B. Rothman,4 and Rachel L. Miller3

1Children’s Hospital at Montefiore, Albert Einstein College of Medicine, New York, New York, USA. 2Department of Neurology, Jiangxi Provincial People’s Hospital, Nanchang, People’s Republic of China. 3Department of Medicine, Division of Pulmonary, Allergy, and Critical Care Medicine, Columbia University College of Physicians and Surgeons, New York, New York, USA. 4Department of Internal Medicine, University of Iowa, Iowa City, Iowa, USA.

Initial immune responses to allergens may occur before birth, thereby modulating the subsequent development of atopy. This paradigm remains controversial, however, due to the inability to identify antigen-specific T cells in cord blood. The advent of MHC tetramers has revolutionized the detection of antigen-specific T cells. Tetramer staining of cord blood after CMV infection has demonstrated that effective CD8+ antigen-specific immune responses can follow intrauterine viral infections. We hypothesized that sensitization to antigens occurs in utero in humans. We studied cord blood B and T cell immune responses following vaccination against influenza during pregnancy. Anti-Fluzone and anti-matrix protein IgM antibodies were detected in 38.5% (27 of 70) and 40.0% (28 of 70), respectively, of cord blood specimens. Using MHC tetramers, HA-specific CD4+ T cells were detected among 25.0% (3 of 12) and 42.9% (6 of 14) of cord blood specimens possessing DRB1*0101 and DRB1*0401 HLA types, respectively, and were detected even when the DRB1 HLA type was inherited from the father. Matrix protein–specific CD8+ T cells were detected among 10.0% (2 of 20) of HLA-A*0201+ newborns. These results suggest that B and T cell immune responses occur in the fetus following vaccination against influenza and have important implications for determining when immune responses to environmental exposures begin.

Introduction

Fetal immune responses following exposure of the mother to allergens or other molecules during pregnancy may affect the risk for the subsequent development of atopy and other diseases. Support for this has been based on several observations. Allergen-induced cord blood mononuclear cell (CBMC) proliferative responses have been documented and found to be distinct from the mother's mononuclear cell proliferative responses (1–5). Altered cord blood cytokine levels, such as reduced IFN-γ, appear to be associated with the subsequent development of atopic dermatitis, allergic rhinitis, or asthma (6, 7). In a murine model, prenatal exposure to endotoxin downregulated allergic sensitization and airway inflammation in the offspring (8). Finally, maternal, but not paternal, IgE levels have been associated with infant IgE levels and later development of atopy (9).

Nonetheless, the fundamental paradigm that adaptive antigen-specific T cell and B cell immune responses to environmental exposures occurs prenatally remains controversial (10, 11). The specificity of the cord blood proliferation has been challenged, as T cell epitope mapping of cord blood responses to allergens indicates that neonatal immune cells lack the fine specificity of adult memory cells (12). Also, CBMCs can proliferate following in vitro stimulation with non-recall antigens, to which natural exposure is extremely unlikely (13, 14).

The controversy surrounding in utero sensitization is not only due to the uncertainty regarding the specificity of proliferative immune responses but, importantly, stems from the inability to detect and immunophenotype antigen-specific CD4+ and CD8+ T cells. Conventional techniques for quantifying antigen-specific T cells include limiting dilution cloning, ELISPOT, and intracellular cytokine assays.

Although these assays can be conducted without knowledge of specific antigen epitopes and/or the MHC restriction elements, they are indirect and prone to considerable experimental variability (15).

The advent of MHC class I and II multimer reagents has revolutionized the detection of antigen-specific T cells. The recognition is based on the specific trimolecular interaction between the MHC-peptide and the T cell receptor (16). MHC class I and II tetramers consist of 4 linked HLA molecules loaded with a peptide (epitope) such that the MHC-peptide complex can be recognized by a subset of specific T cells via the TCR. The tetramer molecule is covalently conjugated to a fluorochrome, allowing sensitive, direct, and specific detection of CD4+ or CD8+ T cells by flow cytometry (16, 17). Tetramer-based detection has been applied successfully to the investigation of cell-mediated immunity to a number of pathogens in peripheral blood from children and adults (17–19). Further, tetramers have been able to identify and immunophenotype antigen-specific T cells in cord blood following human CMV infection, demonstrating that effective CD8+ antigen-specific immune responses can occur following intrauterine viral infection (20).

Literature demonstrating the development of antigen-specific B cells in utero has been scarce. In 2 small case series, maternal vaccination against tetanus was associated with the development of antitetanus IgM in the cord blood (21, 22). In another case series, influenza-specific IgM was measured in the cord blood of 1 of 8 babies born to vaccinated mothers (23). Details concerning the frequency of neonatal humoral immunity following vaccination and exposure to other antigens and the relationship between maternal and cord blood immunoglobulin responses have yet to be addressed.

We hypothesized that in utero sensitization occurs in response to antigens to which the mother is exposed during pregnancy. To address this hypothesis, and to study systematically the mechanism underlying in utero T cell and B cell immune responses in humans, we chose a model in which the pregnant mother’s exposure to anti-
gen is controllable, safe, and recommended by the Centers for Disease Control and Prevention (CDC) (24), as well as a model in which the detection of the fetal immune response to antigen is sensitive and specific. The strategy was to study cord blood B cell and T cell immune responses following vaccination of the pregnant mothers against influenza (Fluzone vaccine, zonal purified, subvirion). We found that the fetus readily produces anti-Fluzone and anti–matrix protein (anti-MP) IgM following vaccination of the mothers against influenza during pregnancy. HA-specific CD4+ T cells were detected in cord blood using tetramer staining of specimens from newborns who inherited the DRB1 HLA types from either the mother or father. Combined, these results suggest that intrauterine B and T cell immune responses occur following vaccination of the mother and support the paradigm that specific fetal immune response to environmental exposures occurs prenatally.

Results

The cohort consisted predominantly of Hispanic women, at least 52% Dominican American, receiving the influenza vaccine at 29.53 ± 0.34 weeks gestation (Table 1). The frequency of HLA-A*0201 in the cohort is within a range reported by other groups (25–27), but the frequencies of DRB1*0101 and DRB1*0401 may be higher when compared with those in other Hispanic cohorts (27–29). Among newborns who inherited 1 of the 3 HLA types of interest, 57.6% (19 of 33), 15.4% (2 of 13), and 44.4% (8 of 18) of them shared the A*0201, DRB1*0401, and DRB1*0101 HLA types, respectively, with their mothers.

B cell responses to influenza vaccine among pregnant women and in cord blood. Maternal B cell immune responses to the influenza vaccine during pregnancy were determined by comparing prevaccination with postvaccination levels of anti-Fluzone and anti-MP IgM and IgG antibodies. Among the 89 mothers for whom both prevaccination and postvaccination specimens were available, 43.8% (39 of 89) produced greater than 2-fold increases in IgM antibodies against Fluzone, a seroconversion rate modestly lower than that in previous reports following vaccination of pregnant women against influenza (30) and group B streptococcus (31). In contrast, 20.2% (18 of 89) of mothers produced greater than 2-fold increases in levels of IgM antibodies against MP (Figure 1). Further analysis demonstrated that the mothers’ overall mean postvaccination anti-Fluzone IgM antibody was significantly greater than the prevaccination level (0.35 ± 0.04 versus 0.17 ± 0.03; P < 0.0001; antibody levels expressed in OD units). Such overall increases following vaccination were not significant against MP (0.12 ± 0.02 versus 0.06 ± 0.01; P = 0.11). The likelihood of seroconversion was not affected by the time interval between pre- and postvaccination measurements, when groups of women vaccinated at 24–28, 28–32, and more than 32 weeks gestation were compared (36%, 31.8%, and 31.8%; P = 0.95). These results also indicate that anti-Fluzone IgM antibodies can be maintained for up to 16 weeks. In comparison, measured postvaccination anti-Fluzone and anti-MP IgG antibodies were increased among 13.5% (12 of 89) and 27% (24 of 89) of the pregnant mothers, respectively. The mean postvaccination anti-Fluzone, but not anti-MP, IgG antibody was significantly greater than the prevaccination level (Fluzone, 1.85 ± 0.09 versus 1.32 ± 0.07, P < 0.0001; MP, 0.27 ± 0.07 versus 0.19 ± 0.05, P = 0.14).

To determine whether antigen-specific B cell responses to influenza antigens occur in utero, and to describe its frequency, anti-influenza IgM and IgG antibodies in cord blood were measured. Increased (i.e., ODs 2-fold greater than those among the “nonvaccinated” negative control cohort) anti-Fluzone and anti-MP IgM antibodies were measured among 38.6% (27 of 70) and 40% (28 of 70), respectively, of the cord blood specimens following vaccination of the mothers. Also, anti-Fluzone and anti-MP IgM mean antibodies were significantly greater in cord blood from mothers who received the influenza vaccine when compared with cord blood from newborns of unvaccinated mothers (Fluzone, 0.05 ± 0.01 versus 0.01 ± 0.005; MP, 0.03 ± 0.01 versus 0.006 ± 0.003, respectively; P < 0.05). Moreover, mean levels of cord blood anti-Fluzone and MP IgM antibodies were significantly higher than those of antibodies against an irrelevant antigen, Japanese encephalitis (JE) (Fluzone, 0.09 ± 0.02 and MP, 0.07 ± 0.01 versus JE, 0.01 ± 0.004; P < 0.001) (Figure 1). As an additional negative control, cord blood anti-Fluzone and anti-MP IgM levels of newborns of unvaccinated mothers were found to be similar to cord blood anti-JE IgM levels of newborns.

![Figure 1](http://www.jci.org/Volume117/Number6/June2007)

**Table 1.** Characteristics of the fully enrolled study cohort (n = 126)

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ethnicity</td>
<td>Hispanic: 111, African American: 13, White: 2</td>
</tr>
<tr>
<td>Age at enrollment (yr, mean ± SEM)</td>
<td>25.41 ± 0.42</td>
</tr>
<tr>
<td>Gestation at vaccination (wk, mean ± SEM)</td>
<td>29.53 ± 0.34</td>
</tr>
<tr>
<td>HLA-A*0201+</td>
<td>43.7%</td>
</tr>
<tr>
<td>DRB1*0101+</td>
<td>17.5%</td>
</tr>
<tr>
<td>DRB1*0401+</td>
<td>18.2%</td>
</tr>
</tbody>
</table>

![IgM response](http://www.jci.org/Volume117/Number6/June2007)
of mothers vaccinated against influenza. Cord blood anti-Fluzone and anti-MP IgG antibodies also tended to increase following vaccination of the mother during pregnancy (Fluzone, 2.09 ± 0.1 versus 1.71 ± 0.12, P = 0.05; MP, 0.37 ± 0.10 versus 0.07 ± 0.02, P < 0.005). But cord blood anti-Fluzone and anti-MP IgM antibodies did not correlate with their respective IgG antibodies (Fluzone, r = −0.23, P = 0.06; MP, r = −0.02, P = 0.85). Combined, these results suggest that both anti-influenza IgM and IgG antibodies can be detected in the cord blood following vaccination of the mother during pregnancy, with the latter presumably developing in the cord blood as a result of receptor-mediated maternofetal transfer (32).

The specificity of anti-Fluzone IgM in maternal and cord blood was confirmed further by Fluzone inhibition assays. As shown in Figure 2, the percentage of Fluzone inhibition increased following preincubation with increasing concentrations of Fluzone vaccine (from −3.6% ± 4.1% to 86.2% ± 7.6% maternal postvaccination positive samples). Such inhibition was not found among the maternal prevaccination levels. In comparison, substantial tetramer staining ranging 0.03% to 0.18% and 0.01% to 0.10%, respectively.

To determine whether the simultaneous detection of antigen-specific T cells in both maternal peripheral blood and cord blood represents contamination of maternal cells during the collection of cord blood, cord blood specimens from newborns who inherited the DRB1*0101 or DRB1*0401 HLA type from the father were examined. Among the 9 newborns with evidence of increased DR1/HA or DR4/HA tetramer staining, 5 inherited the HLA type from the father (Table 2). These data argue against the possibilit
ity that tetramer positivity of cord blood specimens during flow cytometry represents contamination by maternal cells.

Identification of maternal and cord blood MP-specific CD8+ T cells by direct staining with MHC class I tetramer. Relatively little is known about the CD8+ T cell response to influenza vaccination during pregnancy as measured in either maternal or cord blood. Notably, neonatal CD8+ T cell responses have been described as weak compared with those of adults (33) but perhaps more competent than previously recognized (20, 34). To help characterize these responses, we determined both maternal and cord blood influenza-specific CD8+ T cell responses following vaccination of the mother during pregnancy by immunostaining with MHC class I tetramer (A2.1/MP tetramer). Tetramer staining was detected only among 25% (4 of 16) of HLA-A*0201+ mothers following vaccination and was not detected among any of the HLA-A*0201- (n = 14) specimens. Positive tetramer staining was present among only 10% (2 of 20) of HLA-A*0201+ newborns tested and none (n = 13) of the HLA-A*0201- newborns tested (Figure 6). These results suggest that CD8+ MP tetramer positivity following vaccination during pregnancy is infrequent among vaccinated mothers and even less frequent among their newborns.

**HA-specific T cell responders may be effector memory T cells.** The demonstration of both humoral and cellular responses to influenza antigens in cord blood lends support to the notion that the fetus is capable of developing adaptive immunity to antigens encountered in utero. Such immunity would be expected to result in immunological memory in the newborn. To determine whether the HA tetramer-positive CD4+ T cells were of an effector memory phenotype, we performed additional costaining with CD45RO. As shown in Figure 7, the majority of maternal DR1/HA, DR4/HA tetramer-positive CD4+ T cells were CD45RO+, an immunophenotype consistent with effector memory. To determine whether tetramer positivity in cord blood is associated with a memory phenotype, cord blood samples were stained for CD45RO cell surface expression. Convincing evidence of CD45RO expression (beyond that stained with the isotype staining control) was present among CD4+ DR4/HA+ cord blood cells (66.7% and 14.1% of CD4+DR4/HA+ cells versus 6% and 0% of CD4+DR4/glutamic acid decarboxylase (CD4+DR4/GAD+) cells in 2 consecutive experiments), consistent with an effector memory phenotype.

**In vitro expansion of tetramer-positive T cells.** To determine whether tetramer-positive T cells expand following in vitro stimulation with influenza vaccine, in vitro proliferation in thawed aliquots of available DR1, DR4/HA tetramer-positive and -negative cord blood specimens were tested using the cell tracking dye CSFE (35, 36). In vitro proliferation following by positive tetramer staining of CD4+ cells was limited and nonsustainable beyond 2–3 generations of divisions. Enrichment of tetramer staining was minimal (data not shown).

Influenza antigen-specific CBMC proliferation was infrequent and discordant with tetramer staining. To assess the in vitro proliferative response against influenza in maternal versus cord blood, and to compare influenza-specific T cell proliferation with tetramer staining and B cell responses, Fluzone-specific PBMC and CBMC proliferative responses were measured. Among the vaccinated mothers examined, 78% (32 of 46) exhibited 2-fold increases in proliferation following in vitro stimulation with Fluzone, compared with proliferation in the absence of antigens. In comparison, 13.6% (6 of 44) of CBMC specimens exhibited 2-fold increases in proliferation following stimulation with Fluzone, compared with background proliferation. Notably, no association between the presence of influenza vaccine-induced proliferation and detection of CD4+HA-specific cells by tetramer staining was found (Table 3). Likewise, positive results from either T cell assay were not associated with greater levels of IgM antibodies (Table 3).

**Discussion**

The determination that the fetus can initiate B and T cell responses following exposure of the mother during pregnancy has wide implications for multiple diseases, including atopy. This study provides the first direct evidence of antigen-specific T cell fetal immune responses to prenatal antigen exposure via vaccination. Without in vitro manipulation, MHC class II HA-specific CD4+ T cells were detected in 34.6% (9 of 26) of the cord blood samples. MHC class I
MP-specific CD8+ T cells were detected less frequently. In addition, influenza-specific B cell responses in cord blood, as assessed by IgM production following vaccination of the mother were relatively common (38.5% to Fluzone; 40% to MP). Combined, these results suggest that T and B cell immune responses occur in the fetus following maternal vaccination against influenza during pregnancy.

MHC tetramers utilize the MHC-peptide complex to detect antigen-specific T cells at the single-cell level, thus surpassing the detection limits of conventional cell-based methods. Lower detection limits for the tetramers utilized here have been reported. For example, Dunbar and colleagues reported that the A2.1/MP tetramer stained 0.0017%–0.067% of PBMCs from HLA A*0201+ individuals with no known history of influenza infection to demonstrate that tetramers could be used to analyze low-frequency CD8+ populations (17). Hoffmann and colleagues reported that 0.11% of HLA A*0201+ CD8+ cells collected from healthy donors stain with the A2.1/MP tetramer (37). The latter group also calculated a geometric mean background staining among HLA A*0201– subjects of 0.0045%. They also established a cutoff for the lower detection limit of the assay at the upper 99th percentile of tetramer-positive CD8+ T cells in HLA A*0201 individuals at 0.01% (37), providing guidelines for discriminating true positive events from false negative ones. Tetramer staining also detected 0.13%–7.5% CD8+ cells from CMV-infected cord blood but repeatedly less than 0.01% of CD8+ cells from noninfected cord blood (20). When examining CD4+ immune responses following vaccination against influenza, Danke and Kwok reported that 0.0033%–0.167% of CD4+ cells stain with the DR1/HA or DR4/HA tetramer (38), though expected background tetramer staining among DRB1*0101– or DRB1*0401– nonvaccinated subjects has not been well described. While potential overlap tetramer staining between true antigen-specific cells and antigen-nonspecific, falsely positive cells cannot be ignored, the frequency of DR1/HA, DR4/HA CD4+ positivity among specimens classified as positive here (0.01%–0.18%, after subtracting measured background levels) ranges above reported background levels. Confirmation of antigen specificity during the measured immune responses was achieved by: (a) excluding the possibility of contamination from maternal cells by detecting cord blood DR1/HA and DR4/HA tetramer staining among newborns who inherited the HLA type from their fathers; (b) use of multiple negative controls, including a tetramer against an unrelated antigen and DR1/HA– or DR4/HA– vaccinated adult and cord blood samples; and (c) demonstration of increased tetramer staining in postvaccination compared with prevaccination maternal specimens (Figure 7) and among 75% (6 of 8) of tested specimens.

In contrast to HA-specific CD4+ T cells, MP-specific CD8+ T cells following vaccination were difficult to detect in maternal samples. This finding is contrary to documentation of MP-specific CD8+ T cells following influenza vaccination among healthy, young, nonpregnant individuals (39, 40). Likewise, the development of
MP-specific CD8+ T cells following vaccination of the mother was extremely difficult to detect in cord blood samples, despite the observation that CMV-specific CD8+ cord cells staining positive for tetramer are readily detectable following CMV infection (20). The findings here suggest that the predominant fetal immune responses to vaccination are MHC class II and B cell mediated. The mechanism of this apparent CD4+-dominated response to influenza vaccine in the fetus is unclear, but the response has been observed after vaccination against influenza of children following stem cell transplants and among the elderly (40–42).

Cord blood T cells have been characterized as predominantly naive cells, in part because more than 90% of circulating CD3+ T cells are CD45RO– (43, 44). Nevertheless, CMV tetramer–positive CD8+ cells with increased CD45RO expression and cytolytic activity were detected in cord blood cells following CMV infection, indicating that neonatal cells can express a mature, antigen-specific phenotype (20). Similarly, in these experiments, influenza-specific cord blood T cells identified by specific tetramer staining (but not by the negative control tetramer) were repeatedly CD45RO+, albeit to a lesser degree than maternal PBMCs. This phenotype is not consistent with recent thymic emigrants and suggests successful generation of an effector memory T cell response.

This study also corroborates the findings and greatly supports the suggestion of previous reports that antigen-specific IgM antibodies develop in utero in response to vaccinations. The frequently higher level of influenza-specific IgM antibodies among newborns born to vaccinated mothers than those born to nonvaccinated mothers, and the absence of cord blood IgM antibodies against JE, both suggest that antigen-specific B cell immune responses to influenza vaccine occur in utero. The discordance between the newborns’ and mothers’ IgM seroconversion indicates that production of IgM in cord blood is independent of the mother’s production of IgM.

Some reports suggest that maternally derived antibodies may inhibit offspring antibody production during infancy. These include studies following vaccination against measles (45) and hepatitis A (46) but not herpes (47). Persistent B cell inhibitory effects of maternal vaccination against influenza among the offspring was demonstrated elegantly by measuring maternally derived versus offspring-generated IgG antibodies using allotype markers (48). In these experiments, production of maternally derived antibodies did not affect the cell-mediated responses. Maternally derived anti-influenza antibodies did suppress the ability of infant mice to generate their own serum antibody production but not their ability to mount a secondary antibody response to influenza infection (49). In comparison, the modest positive correlations between maternally derived and cord blood IgM antibodies reported here argue against an inhibitory effect of anti-influenza maternal antibodies generated during human pregnancy on cord blood production. However, the offspring’s ability to mount a secondary response following reexposure to influenza antigens was not examined.

Influenza-specific IgM antibody production was not necessarily associated with a greater likelihood of positive DR1/HA, DR4/HA tetramer staining in either the maternal or cord blood specimens. The detection of tetramer staining also occurred discordantly with influenza-induced T cell proliferation. One may predict that the sensitivity of tetramer staining would be greater than the antigen-specific proliferation assay, due to its ability to identify antigen-specific single cells and minimize the effects of nonspecific background proliferation. But the CD4+ response following vaccination against influenza is directed against multiple epitopes, and an assay such as tetramer
staining that measures the immune response against only one epitope will be less robust (38). The reduced proliferation rate observed among CBMCs when compared with PBMCs could also reflect differences in epitopes recognized by cord versus maternal cells.

There are certain limitations that may diminish the generalizability of these results. These include our inability to control completely for winter season (i.e., season for influenza infection) when making comparisons between the cohort born to vaccinated mothers and the cohort born to unvaccinated mothers. The H3 virus strains used in the influenza vaccines possess the PKYVKQNTLKLAT amino acid sequence, which differs from the tetramer epitope by 1 amino acid (i.e., PKYVKQNTLKLAT). The significance of this difference when tracking immune responses to vaccination is unclear. Despite the high accuracy of our HLA typing strategy, a small percentage could have been misclassified because sequencing data were not obtained in every case. In addition, direct detection of very infrequent events always raises concerns, especially when distinguishing small differences between antigen-specific staining and background staining. Collection of a large number of cells during flow cytometry (3 × 10⁵ to 1 × 10⁶) and use of appropriate and multiple controls help mitigate against effects of this problem. Moreover, tetramer staining can only analyze known MHC-specificities, however more complex epitopes may be targeted by T cells (50). We did not observe substantial enrichment of tetramer-positive cord blood cells following in vitro proliferation and tracking with CFSE, possibly consistent with the observation that CBMCs are characteristically more apoptotic than adult PBMCs and that the antigen-induced T cell apoptosis in CBMCs dominates the cell population under culture for 48–72 hours (11, 43). Finally, determination of the presence of antigen-specific T cells several months after birth using repeated tetramer staining may have provided further proof of the development of memory T cells, but such phlebotomy was beyond the scope of this study.

In summary, this study establishes that both T and B cell immune responses to antigens occur in utero following vaccination and these responses display features of immunological effector memory, a hallmark of adaptive immunity. This study
supports the theory that the human neonatal immune system is not deficient or incompetent but, rather, capable of responding to environmental exposures. It remains unclear whether these results can be generalized to MHC class II immune responses to other environmental exposures, such as inhaled antigens, or whether neonatal priming may shape the subsequent postnatal vaccine or clinical response. Nonetheless, these results indicate that B and T cell immune responses occur in the fetus following vaccination against influenza and have important implications for determining when immune responses to environmental exposures begin.

Methods

Subject recruitment, influenza vaccination, and sample collection. 177 pregnant women were screened from prenatal clinics affiliated with New York Presbyterian Hospital. Inclusion criteria included expected delivery at New York Presbyterian Hospital or the affiliated Allen Pavilion during the season when influenza is most prevalent and age greater than 18 years. Exclusion criteria included: (a) history of cardiovascular, neurological, and other systemic disorders; (b) major complications during pregnancy or history of major complications during previous pregnancy; (c) history of recent HIV, HBV, and other systemic infection; (d) history of allergic reaction to influenza and other vaccines, egg, and latex; and (e) recent flu-like symptoms. The other 51 cases, the fathers of the expected newborn were recruited and HLA typing conducted following receipt of permission from the pregnant mothers followed by informed consent from the fathers as described below.

Influenza vaccine was administered to subjects during the second or third trimester up to 34 weeks gestation according to CDC recommendations. Of the 177 subjects screened, 126 were fully enrolled and received the influenza vaccination (23 Fluzone, 3 FluShield) (Sanofi Pasteur). The other 51 withdrew prior to vaccination due to inability to contact, voluntary withdrawal by the participant, onset of influenza infection, or administration of influenza vaccination prior to scheduled visit. The presence of HA and MP in the influenza vaccine was confirmed by Western blot analysis using 4 μg of influenza vaccine and JE vaccine (negative control) and labeling with anti-HA (1:2,500) (Roche Applied Science) and anti-MP (1:2,500) (Accurate Chemical & Scientific Corp.) antibodies (data not shown).

At the time of delivery, 87 adequate cord blood samples were collected successfully. The remaining 39 were not collected due to failure of the participant to contact research staff during labor, emergent delivery prohibiting timely collection of specimen, technical problems with the collection procedure, or delivery at home or another hospital. In cases of uncollected cord blood, collection of the maternal blood samples was deferred. Sera were not isolated from another n = 15 of these cord blood samples due to technical problems (e.g., heparin contamination, inadequate amount collected).

Preparation of blood samples. Phlebotomy was performed on the mothers at recruitment, immediately prior to administration of the vaccine, and within 1 day postpartum. Cord blood was collected at delivery, as previously described (5). Sera were isolated from clotted blood, aliquoted, and frozen for future use, as described below. Maternal PBMCs and CBMCs were separated by density centrifugation from whole blood (5). Cells were counted and plated for culture (as described below) or frozen at −80°C using freezing medium (10% DMSO and 90% human serum). Additional samples (n = 42) of cord blood sera were collected from an unrelated cohort of children born to mothers who did not receive the influenza vaccine during pregnancy, usually because they were pregnant outside of the influenza season, using identical techniques and used as a negative control for ELISAs (described below).

ELISA. Anti-MP and anti-Fluzone IgM and IgG antibodies were measured in duplicate by ELISA. Prior to use, the influenza vaccine (Fluzone) and the JE vaccine were dialyzed for 36 hours using 3 exchanges of PBS and 10,000 MW cutoff (MWCO) dialysis tube (Pierce). Plates were coated with MP (80 ng/ml) (Doris Bucher, New York Medical College, Valhalla, New York, USA), dialyzed Fluzone vaccine (100 ng/ml), dialyzed JE vaccine (100 ng/ml) (Sanofi Pasteur), or no antigen overnight. Unbound antigens were washed with PBS/0.1% Tween. To block irrelevant proteins, 200 μl of blocking solution (1% pig gelatin [Sigma-Aldrich] in PBS/0.1% Tween) was added to each well and incubated for 2 hours at 37°C. Sera in appropriate dilution (1:5,000 for IgM; 1:2,500 to 1:15,000 for IgG, usually 1:10,000) were added and incubated at 4°C for overnight. After incubation, wells were washed again with PBS/Tween buffer, and 100 μl/well HRP-labeled goat-anti-human IgG or IgM (BioSource) was added and incubated at 37°C for 2 hours. In the final substrate reaction, 100 μl of 3,3′,5,5′-tetramethylbenzidine (TMB) substrate (BD Biosciences—Pharminagen) was added to each well, samples were incubated in room temperature for 20 minutes, and the reaction was stopped by 1 M phosphoric acid. ODs were read at 450 nm with a Molecular Devices plate reader.

The Fluzone inhibition assay was performed on select samples by preincubating serum samples prior to ELISA with serially diluted Fluzone vaccine (dilutions: 1:1, 1:4, 1:16, 1:64, 1:256) as well as a diluent alone (control) for 5 hours at 37°C.

DNA extraction and HLA genotyping. After harvesting, mononuclear cells were incubated with lysis buffer (50 mM Tris, 100 mM EDTA, 0.5% SDS) in the presence of proteinase K (1:15 dilution) (Sigma-Aldrich) at 55°C overnight. DNA was isolated from CBMCs or PBMCs using phenol sevag extraction and alcohol precipitation according to conventional techniques (51).

HLA-A*0201, DRB1*0101, or DRB1*0401 genotypes were first screened in mothers’ PBMCs and newborns’ CBMCs by nested PCR, using the human HLA allele–specific primers, as described previously (52, 53), and reverse dot-blot hybridization, using allelic sequence-specific high-resolution oligonucleotides (Tepnel LifeSciences) as described previously (54). The nested PCR for HLA-A*0201, based on reports from Krausa and Browning, amplifies products that are A*0201, A*0204, A*0207, A*0209, A*0211, A*0215N, A*0216, A*0217 with high resolution (55). Combined with high-resolution oligotyping (Tepnel LifeSciences) to eliminate the presence of HLA-A*0204 and A*0211, the remaining subset of A2-A*0201 individuals (A*0207, A*0209, A*0215N, A*0216, A*0217) is estimated to account for approximately 4% of all A2 Hispanic individuals tested (27), making misclassification unlikely. The reliability of this approach was confirmed through sequencing experiments of representative (n = 28) samples. Additional HLA typing was performed by flow cytometry using anti-HLA-A2 at 1:10 dilution (One Lambda Inc.). Equivocal DRB1*0401 results were confirmed by flow cytometry using an anti-DR4 antibody (Accurate).
Tetramer staining and flow cytometry. The 4-color tetramer experiments were performed with MHC class I and II tetramer as described previously (38). In each of the experiments, 1 × 10⁶ fresh or thawed PBMCs (mother’s pre-vaccine and postpartum sample) or CBMCs were placed in 50 µl of medium (RPMI medium [Mediatech Inc.] with 15% human serum) in a U-bottom, 96-well plate and blocked at 4°C for 30 minutes. After blocking, tetramer staining was conducted as follows: (a) MHC class I tetramer staining: 5 µl of PE-conjugated A2/M1 tetramer (HLA-A*0201/influenza MP[FLu] peptide [GILGFVFTL]; iTag; Beckman Coulter) or negative control tetramer (PE-conjugated HLA-A*0201 unrelated, undisclosed peptide; Beckman Coulter) was added and incubated at room temperature for 30 minutes; (b) MHC class II tetramer staining: 1:100 dilution of PE-conjugated DR4/HA tetramer (DRB1*0401/HA306-318 [PKYVKQNTLKLAT]; NIH Tetramer Facility, Virginia Mason Research Center) or negative control tetramer (PE-DRB1*0401/GAD67) was added and incubated at 37°C for 30 minutes. (c) MHC class II tetramer staining: 1:125 dilution of PE-conjugated DR1/HA (DRB1*0101/HA306-318 [PKYVKQNTLKLAT]); NIH Tetramer Facility, Virginia Mason Research Center) or negative control tetramer (PE-DRB1*0401/HA) was added and incubated at 37°C for 30 minutes. DRB1*0401/HA T cell clone HA136 was provided as a kind gift from Jean-François Fonteneau, Rockefeller University, New York, New York, USA, and was used as a positive control. After tetramer staining, cells were washed twice with medium, blocked for 15 minutes, and then incubated for another 30 minutes with 1:125 dilution of CD14–Alexa Fluor 700 (BD Biosciences—Pharmingen) (all experiments) combined with anti-CD19–Alexa Fluor 700 (eBioscience) (grant RR00645), NIH Tetramer Facility, the National Institute of Environmental Health Sciences (grants P01 ES09660, 5 RO1 ES08977, P30 ES09089), and the US Environmental Protection Agency (grant R827027). The authors are grateful to Tony Brown, Diana Blythe, Stephen Canfield, Raphael Clynes, Manisha Ballaney, Megan Williams, Vivek Iyer, Alina Johnson, Sullafa Kadura, Ji Yong Kong, Fangjun Liu, Jinming Liu, Claudia Maria, Vijay Mukhiya, Judy Nam, Yoshiko Ogawa, James Spencer, Danila Valmori, and Veronika Vasquez for assisting in the conduct of this study. The following tetramers were obtained from the Virginia Mason Research Center through the NIH Tetramer Facility: DR1/HA, DR4/HA.

Received for publication June 21, 2006, and accepted in revised form April 9, 2007.

Address correspondence to: Rachel L. Miller, PHBC, Columbia University College of Physicians and Surgeons, 630 W. 168th Street, New York, New York 10032, USA. Phone: (212) 305-7759; Fax: (212) 305-2277; E-mail: rlm14@columbia.edu.

Acknowledgments
This work was supported by Asthma and Allergic Diseases Research Center (grant 1 P01 AI05014), General Clinical Research Center (grant RR00645), NIH Tetramer Facility, the National Institute of Environmental Health Sciences (grants P01 ES09660, 5 RO1 ES08977, P30 ES09089), and the US Environmental Protection Agency (grant R827027). The authors are grateful to Tony Brown, Diana Blythe, Stephen Canfield, Raphael Clynes, Manisha Ballaney, Megan Williams, Vivek Iyer, Alina Johnson, Sullafa Kadura, Ji Yong Kong, Fangjun Liu, Jinming Liu, Claudia Maria, Vijay Mukhiya, Judy Nam, Yoshiko Ogawa, James Spencer, Danila Valmori, and Veronika Vasquez for assisting in the conduct of this study. The following tetramers were obtained from the Virginia Mason Research Center through the NIH Tetramer Facility: DR1/HA, DR4/HA.

Statistical analyses of the antibodies and percentage tetramer staining were performed using the Mann Whitney U test across groups (e.g., maternal versus cord blood levels) and Spearman’s rho for correlations (e.g., IgM versus IgG levels) using VassarStats online statistical computation software (http://faculty.vassar.edu/lowry/VassarStats.html). Data are reported as mean ± standard error (SEM) unless otherwise noted. Statistical significance was specified as a 2-sided P value of 0.05 or less.

References
The Journal of Clinical Investigation  Volume 117  Number 6  June 2007  1645

research article


