Reactivation of the p53 pathway as a treatment modality for KSHV-induced lymphomas

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Kaposi’s sarcoma herpesvirus (KSHV) is the etiologic agent for primary effusion lymphoma (PEL), a non–Hodgkin type lymphoma manifesting as an effusion malignancy in the affected individual. Although KSHV has been recognized as a tumor virus for over a decade, the pathways for its tumorigenic conversion are incompletely understood, which has greatly hampered the development of efficient therapies for KSHV-induced malignancies like PEL and Kaposi’s sarcoma. There are no current therapies effective against the aggressive, KSHV-induced PEL. Here we demonstrate that activation of the p53 pathway using murine double minute 2 (MDM2) inhibitor Nutlin-3a conveyed specific and highly potent activation of PEL cell killing. Our results demonstrated that the KSHV latency-associated nuclear antigen (LANA) bound to both p53 and MDM2 and that the MDM2 inhibitor Nutlin-3a disrupted the p53-MDM2-LANA complex and selectively induced massive apoptosis in PEL cells. Together with our results indicating that KSHV-infection activated DNA damage signaling, these findings contribute to the specificity of the cytotoxic effects of Nutlin-3a in KSHV-infected cells. Moreover, we showed that Nutlin-3a had striking antitumor activity in vivo in a mouse xenograft model. Our results therefore present new options for exploiting reactivation of p53 as what we believe to be a novel and highly selective treatment modality for this virally induced lymphoma.

Introduction
Kaposi’s sarcoma herpesvirus (KSHV) is a DNA tumor virus and causative agent in 3 different tumor types: Kaposi’s sarcoma (KS), a plasmablastic variant of multicentric Castelman’s disease (MCD), and an AIDS-related form of B cell lymphoproliferative disorder called primary effusion lymphoma (PEL) (1–3). Additionally, KSHV infection is suggested to be linked to other types of lymphoproliferations (reviewed in ref. 4). PEL is a non–Hodgkin type lymphoma latently infected with KSHV and manifests as an effusion malignancy in KS patients with advanced AIDS, but it may also occur in HIV-negative individuals (reviewed in refs. 5, 6). The KSHV genome encodes several homologs of cellular proteins, which engage cellular signaling pathways, govern cell proliferation, and modulate apoptosis (reviewed in ref. 7). Latent viral genes include a cluster of 3 genes transcribed from the same promoter, encoding latency-associated nuclear antigen (LANA), viral cyclin, and viral FLICE-inhibitory protein. Similar to oncogenic proteins of other tumor viruses, these proteins are known to regulate the major tumor suppressor pathways (cell cycle, apoptosis, and cell survival), suggesting a role for them in oncogenesis of this lymphoma.

p53 is a transcription factor that plays a central role in protecting cells from tumor development by inducing cell-cycle arrest or apoptosis via a complex signal transduction network referred to as the p53 pathway (reviewed in ref. 8, 9). The p53 gene is mutated or deleted in 50% of all malignant tumors (reviewed in ref. 10). The other half of human cancers express WT p53 protein, which upon reactivation is capable of inducing apoptosis in cancer cells, thus offering a potential therapeutic opportunity applicable to a wide range of human tumors. Because tumor cells are prone to p53-induced apoptosis as a result of oncogene activation, it is possible that p53-based anticancer strategies may not require selective targeting of tumor cells (11–13). A recently discovered strategy for p53 activation targets the interaction of p53 with its negative regulator, murine double minute 2 (MDM2), an E3 ubiquitin ligase that binds p53 and facilitates its ubiquitin-dependent degradation (14). Vassilev et al. (15) have developed potent and selective small-molecule inhibitors of p53-MDM2 interaction, the nutlins, which activate the p53 pathway in vitro in cells with WT p53 and cause cell-cycle arrest via induction of p21(CIP1) and, in some cases, apoptosis. Although mechanisms converting the Nutlin-3a–induced cytostatic pathways to cytotoxic pathways are not fully understood, the fact that Nutlin-3a has shown potent antitumor activity in certain mouse xenograft models suggests that it is a potential treatment option for cancers with WT p53 (15, 16).

PELs are aggressive KSHV-induced lymphomas, with median survival times reported to be shorter than 6 months after diagnosis (17). Despite some interesting new therapeutic leads such as inhibition of NF-kB signaling (18, 19) or RNA interference against viral latent proteins (20), the current clinical treatments based on high-dose chemotherapy regimens are neither potent nor selective for this cancer (21, 22), and PEL remains a fatal disease. Although p53 mutations are relatively common in hematopoietic malignancies, the majority of the PELs appear to have WT p53 (23–25), suggesting that genetic alterations in the p53 gene are not selected for during PEL tumorigenesis. However, the pathogenic mechanisms leading to lymphomas by this oncogenic
tumor virus are unknown. Two previous reports have suggested that upon exogenous expression, LANA interferes with p53 and inhibits its transcriptional activity (26, 27). More recently, LANA was also suggested to function as a component of the ECSS ubiquitin complex targeting p53 for degradation (28). However, these findings have not progressed toward a mechanistic explanation of how the virus overcomes normal cellular barriers or provided opportunities for therapy.

Here we studied p53 signaling and nongenotoxic activation of the p53 pathway in PEL cells. We demonstrate that a small-molecule inhibitor of the p53-MDM2 interaction, Nutlin-3a, efficiently activated the p53 pathway in several PEL cell lines expressing WT p53, leading to cell-cycle arrest and massive apoptosis. The cytotoxic effect of Nutlin-3a was specific for the KSHV-infected cells, as Nutlin-3a did not induce apoptosis in lymphoblastoid cell line (LCLs) transformed with EBV despite their expression of WT p53. We present data for the mechanisms underlying this specificity and demonstrated the antitumor activity of Nutlin-3a using a mouse xenograft model of PEL. Taken together, our results suggest that activation of the p53 pathway may be an effective treatment for KSHV-infected lymphomas.

Results

**Nutlin-3a stabilizes p53 and activates the p53 pathway in PEL cells.** Targeting the p53-MDM2 interaction by using small-molecule inhibitors is a promising strategy for anticancer therapy in tumor cells retaining WT p53. One of these compounds, Nutlin-3a, mimics a p53 peptide by competitively binding MDM2 at the p53 binding pocket, which activates the p53 pathway (15). To explore whether the p53 pathway is functional in PEL cells, we first examined the effect of Nutlin-3a on accumulation of p53 and induction of its target gene expression. Treatment of asynchronously growing PEL cell lines BC-1 and BC-3 with 7 μM Nutlin-3a for 12 hours resulted in a strong and specific increase (12- to 15-fold) in p53 levels (Figure 1). Interestingly, BCBL-1 cells have previously been shown to harbor a TP53 missense mutation (M246I; ref. 25) in one copy of the TP53 gene (29), providing a possible explanation for the attenuated response. Interestingly, PEL cells express relatively high levels of p53 binding protein 1 (53BP1), a component of the ataxia telangiectasia mutated–checkpoint kinase 2 (ATM-Chk2) DNA damage checkpoint pathway (30, 31) recently identified as a critical mediator of cellular cytotoxicity by Nutlin-3a (27). To address the specificity of Nutlin-3a treatment on KSHV lymphomas, we also treated EBV-transformed LCLs, which express WT p53. Exposure of EBV-transformed LCLs CZE and IHE to Nutlin-3a resulted in increased levels of p53, MDM2, and p21CIP1, Both PEL cells and EBV-transformed LCLs also showed a specific increase in the expression of activated p53, detected using an antibody specific for p53 phosphorylated on serine 15 (Figure 1). However, the expression of Bax did not change notably in the EBV-transformed LCLs upon Nutlin-3a treatment (Figure 1). Treatment of EBV-negative Burkitt lymphoma cells constitutively expressing high levels of mutant p53 (cell line DG-75) or the p53-deficient cell line HL-60 did not result in activation of the p53 pathway (Figure 1). This confirmed that Nutlin-3a activated the p53 pathway only in cells with functional WT p53.

**Figure 1**

Nutlin-3a activates p53 and its target genes in PEL cells. KSHV-infected PEL cells (BC-1, BC-3, and BCBL-1), EBV-transformed LCLs (CZE and IHE), and cells with mutant p53 (DG-75 and HL-60) were incubated for 12 hours in the presence (+) or absence (−) of 7 μM Nutlin-3a. Whole-cell lysates were subjected to SDS-PAGE followed by Western blotting and analyzed for p53, MDM2, phosphorylated-p53(Ser15) (p-p53(Ser15)), 53BP1, p21CIP1, and Bax expression. Actin immunoblot is shown as a loading control.

**Nutlin-3a induces cell-cycle arrest in PEL cells.** Induction of p53 activity halts the cell cycle through transcriptional upregulation of the cyclin-dependent kinase inhibitor p21CIP1, which causes G1/S and G2/M cell-cycle arrest (32). To explore whether Nutlin-3a–induced p53 activation causes cell-cycle arrest, we performed flow cytometric analyses of PEL cells, EBV-transformed LCLs, and p53 mutant cells treated with the MDM2 inhibitor. Cells were exposed to Nutlin-3a for 12, 24, or 48 hours, labeled with BrdU, and analyzed by multiparameter flow cytometry. As expected, treatment of PEL cells with Nutlin-3a markedly increased the G1/S ratio of the cells over that of untreated controls, reflecting an efficient G1 arrest. We found that the proportion of S-phase cells was considerably decreased in BC-1 and BC-3 cells as soon as 12 hours after incubation. The S-phase fraction in BCBL-1 cells also decreased, but only after 24 hours, consistent with our results of delayed p53 upregulation in these cells. Incubation for 48 hours led to a complete depletion of the S-phase cells and their profound accumulation in the G1 phase in all PEL cell lines studied (Figure 2A). Interestingly, we also observed that accumulation in the sub-G1 phase was increased in Nutlin-3a–treated cells compared with untreated cells (Figure 2A, arrows). After 48 hours of treatment, the sub-G1 population reached 41%, 36.4%, and 18.7% in BC-1, BC-3, and BCBL-1 cells, respectively, suggesting increased cell death. In the KSHV-negative EBV-transformed LCLs CZE and IHE, the treatment led to an efficient G1-phase arrest, but there was no obvious increase in the sub-G1 population (Figure 2B). Importantly, even an extended 96-hour exposure to Nutlin-3a was insufficient to markedly increase the sub-G1 population in EBV-transformed LCLs (data not shown). Cell-cycle analysis of the mutant p53 cell lines DG-75 and HL-60 showed profiles indis-
Nutlin-3a selectively kills KSHV-infected cells, but not EBV-infected cells. Nutlin-3a possesses antiproliferative activity in a variety of cancer cell lines and leukemias (16, 33–36). To investigate the cytotoxic effect of MDM2 inhibition in KSHV lymphomas, we incubated PEL cell lines, EBV-transformed LCLs, and p53 mutant cell lines with Nutlin-3a and determined cell viability by trypan blue exclusion. Nutlin-3a reduced cell viability in KSHV-infected PEL cell lines BC-1, BC-3, and BCBL-1 as well as in KSHV-transformed LCL IHH (Figure 3A). After 5 days of treatment, only 2.8% of BC-1, 6.5% of BC-3, and 35% of BCBL-1 cells were viable compared with mock-treated control cells. In contrast, Nutlin-3a had no effect on cell viability of the 2 KSHV-negative EBV-transformed LCLs (CZE, 86%; IHE, 84%; Figure 3A). As expected, there was no effect on the viability of the p53 mutant cells by Nutlin-3a (Figure 3A).

To confirm that the cytotoxic effect of Nutlin-3a in PEL cells was caused by apoptosis, we incubated PEL cells, EBV-transformed LCLs, and p53 mutant cells with Nutlin-3a or vehicle control and collected samples up to 120 hours after incubation. Apoptosis was determined by annexin V binding assay followed by flow cytometry (Figure 3, B and C). Use of annexin V staining in combination with propidium iodide (PI) allows separation of cells at early phases of apoptosis (annexin V–positive, PI-negative) from those at the later stages of cell death (annexin V– and PI-positive). Nutlin-3a induced rapid apoptosis in BC-1 cells: 30% of the cells were at early apoptosis after 24 hours of treatment, compared with 4.3% in the control cells. By 72 hours,
63% of the Nutlin-3a–treated BC-1 cell population was at early apoptosis and 35% was at the late stage; thus, 98% of cells were apoptotic at this time point. After 96 and 120 hours, Nutlin-3a treatment dramatically increased the population of late apoptotic cells in the BC-1 line to 77% and 100%, respectively (Figure 3B). A strong apoptotic response was observed in BC-3 and BCBL-1 cells at 96 hours, whereas treatment of the EBV-transformed LCLs or the p53 mutant cell lines did not lead to increased apoptosis (Figure 3C). These results confirmed that Nutlin-3a selectively kills KSHV-associated lymphomas in a p53-dependent manner.

**KSHV infection promotes cell killing by Nutlin-3a.** We next investigated whether infection of cells by KSHV specifically promotes the cell-death program induced by Nutlin-3a. To this end, we used U2OS osteosarcoma cells and EA.hy 926 endothelial cells (37), both harboring WT p53, which were de novo infected
immunofluorescence using anti-LANA antibodies (data not shown). The establishment of latent infection was confirmed by indirect immunofluorescence using anti-LANA antibodies. EA.hy 926 cells confirmed accumulation of p53 and MDM2 in response to Nutlin-3a treatment (Figure 4B). Based on these indirect immunofluorescence analysis revealed sustained results, we conclude that the MDM2 inhibitor selectively induces positive foci in 72%, 62%, and 54% of BC-1, BC-3, and BCBL-1 cells, respectively, while the EBV-transformed LCLs showed remarkably fewer cells positive for γH2AX focal staining (CZE, 19%; IHE, 21%; Figure 5, A and B). We also analyzed the level of phosphorylation on Chk2 at threonine 68, another marker for activated DNA damage response, in BC-1, BC-3, and BCBL-1 cells as well as the EBV-transformed LCLs and the p53 mutant cell lines. Elevated levels of phosphorylated Chk2(Thr68) were observed in BC-1 and BC-3 cells 12 hours after exposure to Nutlin-3a (Figure 5C).

To obtain additional evidence in support of the role of activated DNA damage response as an effector for Nutlin-3a cytotoxicity, we subjected the EBV-transformed LCLs to gamma irradiation (1 Gy) and compared cell viability after Nutlin-3a treatment with that of nonirradiated cells. Induction of DNA damage by irradiation was confirmed by an increase in γH2AX focal staining (data not shown). After 4 days of treatment, there was a 2.6-fold increase with a recombinant KSHV (rKSHV) (38). rKSHV expresses red fluorescent protein (RFP) from the KSHV lytic PAN promoter and GFP from the cellular EF-1α promoter. All of the rKSHV-infected EA.hy 926 and U2OS cells were positive for GFP, and the establishment of latent infection was confirmed by indirect immunofluorescence using anti-LANA antibodies (data not shown). Less than 1% of the cells expressed RFP, indicating the absence of lytic replication. However, we successfully induced lytic replication by treating the cells with a baculovirus expressing RTA and Na-butyrate (data not shown). These KSHV-infected cells and their parental noninfected cell lines were treated with Nutlin-3a, and its cytotoxic effect was determined by trypan blue exclusion. The percentage of dead cells increased about 4-fold following 96 hours of Nutlin-3a treatment in the KSHV-infected cells compared with uninfected cells (Figure 4A). Western blotting of cell extracts from the noninfected and rKSHV-infected EA.hy 926 cells confirmed accumulation of p53 and MDM2 in response to Nutlin-3a treatment (Figure 4B). Based on these results, we conclude that the MDM2 inhibitor selectively induces cytotoxic activity in KSHV-infected cells.

Activated DNA damage response in KSHV lymphomas enhances cytotoxicity by Nutlin-3a. One plausible mechanism for the conversion of the cytostatic Nutlin-3a response to a cytotoxic response is suggested by a recent report indicating that intrinsic DNA damage signaling in cancer cells is critical in regulating Nutlin-3a-induced apoptosis (27). We therefore investigated whether PEL and KSHV-negative EBV-transformed LCLs show signs of activated DNA damage signaling. To this end, the KSHV-positive BC-1, BC-3, and BCBL-1 cell lines and the EBV-transformed LCLs CZE and IHE were immunostained with an antibody against γH2AX, the phosphorylated form of histone H2AX. Phosphorylation of histone H2AX (detected by γH2AX focal staining) is a commonly used marker of the DNA damage response activated by replication stress and double-stranded DNA breaks (39, 40). Quantitative indirect immunofluorescence analysis revealed sustained γH2AX-positive foci in 72%, 62%, and 54% of BC-1, BC-3, and BCBL-1 cells, respectively, while the EBV-transformed LCLs showed remarkably fewer cells positive for γH2AX focal staining (CZE, 19%; IHE, 21%; Figure 5, A and B). We also analyzed the level of phosphorylation on Chk2 at threonine 68, another marker for activated DNA damage response, in BC-1, BC-3, and BCBL-1 cells as well as the EBV-transformed LCLs and the p53 mutant cell lines. Elevated levels of phosphorylated Chk2(Thr68) were observed in BC-1 and BC-3 cells 12 hours after exposure to Nutlin-3a (Figure 5C).

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**Figure 4**

Nutlin-3a selectively kills KSHV-infected cells. (A) U2OS and EA.hy 926 cells in the absence or presence of latent rKSHV infection were treated with 7 μM Nutlin-3a, and cell death was assessed by trypan blue exclusion at 24, 48, and 96 hours of treatment. Values represent the percentage of dead cells induced by Nutlin-3a treatment. The percentage of dead cells in the DMSO control was subtracted as a background. Each value represents the mean of 3 independent experiments. (B) Noninfected and rKSHV-infected EA.hy 926 cells were incubated for 12 hours in the presence or absence of 7 μM Nutlin-3a. Whole-cell lysates were subjected to SDS-PAGE followed by Western blotting and analyzed for p53 and MDM2 expression. Actin immunoblot is shown as a loading control.

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hours resulted in a dramatic redistribution of p53, MDM2, and LANA in the eluted fractions. Importantly, this abrogated the co-elution of MDM2 and LANA with p53, suggesting disruption of interactions among these proteins (Figure 6A).

To analyze the formation of the p53-MDM2-LANA complex in BC-3 cells and to explore the effect of Nutlin-3a on this complex, we performed reciprocal immunoprecipitations for p53 and MDM2. High–molecular weight gel filtration fractions (600–400 kDa) from mock- and Nutlin-3a–treated BC-3 cells were subjected to immunoprecipitation with anti-p53 or anti-MDM2 antibodies. Western blot analysis of the resulting coprecipitates from mock-treated fractions revealed that LANA and MDM2 coprecipitated with p53 antibodies and, conversely, that LANA and p53 coprecipitated with MDM2 (Figure 6B). This demonstrates that p53, MDM2, and LANA associate in vivo in PEL cells. Treatment of BC-3 cells with Nutlin-3a destroyed the interaction of LANA with p53 and markedly decreased the amount of LANA coprecipitating with MDM2 (Figure 6B). As expected, the interaction between p53 and MDM2 was also abolished upon 12 hours’ treatment with Nutlin-3a (Figure 6B). Taken together, these results identify LANA as a component of the p53-MDM2 complex and demonstrate that Nutlin-3a disrupted the complex in KSHV lymphoma cells. This may contribute to the specificity and efficiency of Nutlin-3a–mediated cell death in KSHV lymphomas.

**Nutlin-3a is a potent agent for treating KSHV lymphomas in vivo.** Because in vitro experiments cannot fully mimic all aspects of tumorigenesis and predict potential therapeutic value, we assessed the effect of Nutlin-3a on KSHV lymphomas in vivo. The antitumor activity of Nutlin-3a was evaluated in a BC-3 cell–based xenograft model (41). The most crucial preclinical evaluation of an antitumor agent is to determine its ability to induce responses in established tumors. BC-3 cells were implanted subcutaneously into Balb/c nude female mice and allowed to grow until the tumors were palpable. Administration of 20 mg/kg Nutlin-3a every other day for 2 weeks (7 doses) caused marked regression of all tumors in the treated animals, whereas none of the animals receiving vehicle control showed tumor regression during this time course (Figure 7). Nutlin-3a was well tolerated without any weight loss or other obvious signs of toxicity. All animals (n = 8) treated with Nutlin-3a every other day for 2 weeks (7 doses) caused marked regression of all tumors in the treated animals, whereas none of the animals receiving vehicle control showed tumor regression during this time course (Figure 7). Nutlin-3a was well tolerated without any weight loss or other obvious signs of toxicity. All animals (n = 8) treated with Nutlin-3a every other day for 2 weeks (7 doses) caused marked regression of all tumors in the treated animals, whereas none of the animals receiving vehicle control showed tumor regression during this time course (Figure 7). Three mice showed complete tumor regression, and follow-up of these mice for over 90 days did not indicate any recurrence of the tumor growth. After cessation of the treatment, tumors with partial responses started to grow again, but the size of these tumors remained radically smaller than in the control mice (223 mm³ versus 436 mm³), and they remained susceptible to Nutlin-3a treatment. As expected,
tumors originating from the p53 mutant cell line HL-60 did not respond to the Nutlin-3a treatment described above (data not shown). These results demonstrate that p53 reactivation via Nutlin-3a is an efficient treatment for KSHV-lymphomas in mice and suggest a potential therapeutic strategy for treatment of these fatal virus-induced malignancies in humans.

Discussion
This study provides what we believe to be a novel principle for the efficient treatment of KSHV-induced lymphomas through reactivation of the p53 pathway by a small-molecule inhibitor of the p53-MDM2 interaction, Nutlin-3a. Although p53 mutations occur rarely in KS or PELs (25), our results demonstrate that inactivation of p53-mediated processes occurred through binding of the viral protein LANA to the p53-MDM2 complex in these lymphomas. Nutlin-3a treatment resulted in disruption of interactions among all 3 proteins in this complex and induced cytotoxicity at concentrations that were nonapoptotic in EBV-transformed LCLs, which express WT p53. As a control, a duplicate sample from the same fraction was immunoprecipitated with mouse IgG. Immunocomplexes were resolved by SDS-PAGE and analyzed by Western blotting with antibodies against p53, MDM2, and LANA.

Antitumor activity of Nutlin-3a in vivo. Growth curves of Nutlin-3a– and vehicle control–treated BC-3 tumors. Balb/c nude mice were injected subcutaneously with 6 × 10^6 BC-3 cells. When the tumors had grown to palpable size, the mice were treated intraperitoneally with the vehicle control (filled squares) or 20 mg/kg of Nutlin-3a (open circles). Nutlin-3a treatment resulted in regression of tumors; at the end of treatment, these tumors were significantly smaller than those treated with vehicle (P = 0.02). Dashed line indicates the volume of Matrigel in the tumor. Data (mean and SEM) are representative of 2 independent experiments. Inset shows a photograph of BC-3 tumor–bearing Balb/c nude mice treated with either vehicle control or 20 mg/kg Nutlin-3a 7 times over the course of 2 weeks.

Another possible explanation for the efficiency of Nutlin-3a killing of KSHV-infected cells could be a recently described property intrinsic to cancer cells: the activation of DNA damage signaling (40, 46). Accordingly, 53BP1, a component of the ATM-Chk2 DNA damage checkpoint pathway (30), was identified as a critical mediator of cellular cytotoxicity by Nutlin-3a (27). In addition, cytotoxic drugs have been shown to synergize with Nutlin-3a in inducing apoptosis of different leukemia and multiple myeloma cell lines (33–36). We detected pronounced activation of DNA damage signaling in all PEL cell lines studied compared with EBV-transformed LCLs by analyzing the levels of phosphorylated Chk2(Thr68) or focal staining of the DNA damage marker γH2AX. Furthermore, the EBV-transformed LCLs could be sensitized to Nutlin-3a–mediated cell killing by subjecting them to a low dose of gamma irradiation. Moreover, by inhibiting DNA damage checkpoint signaling, we protected KSHV lymphoma cells as well as irradiated EBV-transformed LCLs from Nutlin-3a–induced cell killing. The activated DNA damage response may cause increased levels of p53 (and MDM2); however, these were inactivated by the association to LANA. This implies that intrinsic DNA damage signaling, together with the complex formation among p53, MDM2, and LANA in KSHV lymphomas, may contribute to the selectivity and efficiency of Nutlin-3a–induced cell death.

The AIDS epidemic has had a major impact on the prevalence of KSHV, and the vast majority of PELs occur in HIV-seropositive individuals. Currently there is no efficient treatment for PEL, and the treatment modalities in use consist mostly of cytostatic drugs with DNA-damaging activities, which are neither potent nor selective for this malignancy (reviewed in ref. 21). The highly efficient antitumor effect of Nutlin-3a on KSHV-infected cells implies that KSHV infection is a factor converting p53 pathway activation from cytostatic (i.e., p21-induced) to pro-apoptotic. Intriguingly, EBV also encodes proteins suggested to bind p53 or to interfere with its function (43–45), but this did not render the EBV-transformed LCLs sensitive to Nutlin-3a–induced apoptosis.

This study provides what we believe to be a novel principle for the treatment of KS and other virus-induced malignancies. The highly specific and efficient antitumor activity of Nutlin-3a has potential therapeutic implications for the treatment of virus-induced malignancies, particularly KSHV-related KS and PEL.
active antiretroviral therapy (HAART) to treat AIDS has also proven effective for treatment of KS and PEL, but its use is restricted to patients with asymptomatic and non-life-threatening PELs (47). Development of specific and efficient therapy for KSHV-related malignancies requires better understanding of the pathways it uses for tumorigenic conversion. Here we demonstrated that the p53 pathway was inactivated in PELs and that selective disruption of the p53-MDM2-LANA complex by Nutlin-3a showed remarkable therapeutic activity in our PEL xenograft model in vivo. Importantly, the MDM2 inhibitor showed no toxic effects in previous studies with oral administration of doses 10 times higher than were used in our study (15, 16). Taken together, our data demonstrate that Nutlin-3a selectively and efficiently kills B cells of healthy donors. DG-75 Burkitt lymphoma and HL-60 human promyelocytic leukemia cell lines were purchased from the ATCC. PEL cell lines and KSHV-negative control cells were cultured in a humidified 5% CO₂ atmosphere at 37°C in RPMI 1640 medium supplemented with 15% FCS (Invitrogen), 100 μ/ml penicillin, and 100 μ/ml streptomycin. C2A10, anti-actin (C-2), and anti-Bax (B-9; Santa Cruz Biotechnology Inc.); anti-p21 (SX118; BD Biosciences—Pharminingen); and anti-3SBP1 (Novus Biologicals); anti–phosphorylated p53 (Ser15), anti–phosphorylated Chk2 (Thr68), and anti-Chk2 (DCS-270; Cell Signaling Technology). A mixture of the 3 indicated monoclonal antibodies against MDM2 was used for its detection. HRP-conjugated antibodies specific for rabbit, mouse, or rat immunoglobulins were purchased from Chemicon International. Cyto centrifugation and indirect immunofluorescence were performed as previously described (51). PEL cell lines and EBV-transformed LCLs labeled with a mouse monoclonal antibody against H2AX (Upstate USA Inc.) and Alexa Fluor 594–conjugated antibody to rabbit immunoglobulin (Invitrogen). The fluorochromes were visualized with a Zeiss Axio plan 2 fluorescent microscope equipped with Zeiss PLAN-NEOFLUOR x40/0.50 numeric aperture objective lens. Images were acquired with a Zeiss Axiocam MRC, using Zeiss AxiosVision (version 4.5 SP1) and Adobe Photoshop software (version 7.0, Adobe).

Drug treatment and viability determination. PEL cells, EBV-transformed LCLs, KSHV-infected LCLs, and p53 mutant cells suspended at 2 x 10⁴ cells/ml were incubated with 7 μ/ml Nutlin-3a (Alexis Biochemicals) or relative amount of the solvent (vehicle; 0.1% DMSO) for the indicated time. KSHV-infected and noninfected parental U2OS and EA.hy 926 cells were plated at a density of 0.5 x 10⁵ cells per well in 24-well plates, and after 48 hours the cells were treated with 7 μ/ml Nutlin-3a. Cell viability was determined by trypan blue exclusion (Sigma-Aldrich). The control treatment with DMSO was always included, and the relative survival in each assay was calculated as the percentage of live cells relative to the live cell population in the control. Results are from 2-3 independent experiments. To inhibit the ATM-Chk2 pathway in KSHV-infected PEL cell line BC-1, KSHV-infected LCL IHH, or EBV-transformed LCL IHE, the cells were pretreated with 2 μ/ml caffeine (Sigma-Aldrich) for 24 hours before the Nutlin-3a treatment. Caffeine was kept constant during the exposure to Nutlin-3a.

Measurement of cell proliferation and apoptosis. The proportion of cells at the S phase was determined by measuring incorporation of BrdU and PI into the DNA. Cells were grown at a density of 2 x 10⁶ cells/ml 24 hours prior to the treatment with Nutlin-3a. The cells were pulse-labeled with 25 μ/ml BrdU (Sigma-Aldrich) for 30 minutes and fixed in ice-cold 70% ethanol. After fixation, the cells were washed in PBS and treated with 3.5 N HCl for 30 minutes at room temperature. After washing in a neutralizing washing buffer (0.1% BSA/PBS), the cells were incubated with an anti-BrdU antibody (Dako) for 45 minutes. Alexa Fluor 488–conjugated (Invitrogen) secondary antibody was used for detection. Finally, the cells were stained with 30 μ/ml PI (Invitrogen) in PBS supplemented with 50 μ/ml RNase (Sigma-Aldrich) for 30 minutes at 37°C. Apoptosis was measured by dual-labeling with the Annexin V–FITC Apoptosis Detection Kit I (BD Biosciences—
Pharmingen) according to the manufacturer’s instructions and analyzed by flow cytometry. Labeled cells were acquired using a BD-LSR Flow Cytometer (BD Biosciences), and the cell populations were analyzed by CellQuest software (version 3.3; BD).

**Human tumor xenografts.** Female Balb/c nude mice (4–6 weeks old) were obtained from Taconic Europe and maintained under specific pathogen-free conditions in a temperature- and humidity-controlled environment. Mice under anesthesia were injected subcutaneously with 6 × 10^6 BC-3 or HL-60 cells in 50% Matrigel (BD Biosciences). Treatment was started intraperitoneally after the tumors were established (i.e., palpable). Nutlin-3a (20 mg/kg) or the vehicle control was administered every second day for a total of 2 weeks (7 doses). Tumor volumes were measured with a caliper and calculated according to the formula V = width × height × depth/2, derived from the formula for the volume of an ellipsoid (52). In order to monitor the health of the animals, the mice were weighed once per week. All animal studies were conducted in accordance with the guidelines of the Provincial Government of Southern Finland, and the protocol was approved by the Experimental Animal Committee of the University of Helsinki.

**Statistics.** Statistical analyses were performed using ANOVA. P values less than 0.05 were considered significant.

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**References**