Islet transplantation in patients with autoimmune diabetes induces homeostatic cytokines that expand autoreactive memory T cells

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Successful transplantation requires the prevention of allograft rejection and, in the case of transplantation to treat autoimmune disease, the suppression of autoimmune responses. The standard immunosuppressive treatment regimen given to patients with autoimmune type 1 diabetes who have received an islet transplant results in the loss of T cells. In many other situations, the immune system responds to T cell loss through cytokine-dependent homeostatic proliferation of any remaining T cells. Here we show that T cell loss after islet transplantation in patients with autoimmune type 1 diabetes was associated with both increased serum concentrations of IL-7 and IL-15 and in vivo proliferation of memory CD45RO+ T cells, highly enriched in autoreactive glutamic acid decarboxylase 65–specific T cell clones. Immunosuppression with FK506 and rapamycin after transplantation resulted in a chronic homeostatic expansion of T cells, which acquired effector function after immunosuppression was removed. In contrast, the cytostatic drug mycophenolate mofetil efficiently blocked homeostatic T cell expansion. We propose that the increased production of cytokines that induce homeostatic expansion could contribute to recurrent autoimmunity in transplanted patients with autoimmune disease and that therapy that prevents the expansion of autoreactive T cells will improve the outcome of islet transplantation.

Introduction
Lymphocyte loss is a hallmark of T cell depletion therapy and certain infections. The immune system can sense T cell loss and responds with a vigorous cytokine-dependent expansion of the remaining T cells in the periphery, a process known as homeostatic proliferation (1). Homeostatic proliferation is largely controlled by cytokines of the common γ chain receptor family. IL-7 is required for expansion of CD4 cells (2), and expansion of CD8 cells is promoted by IL-7 and IL-15 (3, 4).

Homeostatic proliferation affects the T cell repertoire by increasing the size of clonal populations. Homeostatic proliferation of peripheral naive T cells requires the presence of specific peptide, whereas memory T cells can expand independently of T cell receptor engagement (5–7). Cells that undergo homeostatic proliferation develop properties that are remarkably similar to antigen-expanded memory cells (8, 9). As a consequence, homeostatic proliferation is suggested to promote T cell–mediated pathologies, including autoimmunity (10, 11), and to hinder tolerance induction in transplantation (12).

Islet transplantation in patients with type 1 diabetes mellitus (T1DM) is performed in the presence of a memory autoimmune response, and immunosuppression must control islet graft rejection caused by alloimmunity and autoimmunity. An increase in autoimmunity to islet autoantigens after islet transplantation has previously been observed (13, 14), and the presence of high-titer autoantibodies is associated with poor islet graft survival (15). Thus, mechanisms that expand autoreactivity can occur in the presence of a heavily compromised immune system. Studies in the autoimmune nonobese diabetic (NOD) mouse model showed that autoimmunity and diabetes are promoted by a chronic state of lymphopenia and consequent homeostatic expansion of autoreactive T cells (16). Conversely, common γ chain blockade in NOD mice substantially reduces a population of memory-like autoreactive T cells (17). We therefore asked whether mechanisms akin to homeostatic T cell proliferation are active after islet transplantation and could expand the islet-autoreactive T cell pool. We studied patients with T1DM who received islet allografts under immunosuppression composed of anti–IL-2 receptor (anti–IL-2R) mAb induction therapy followed by low-dose FK506 (tacrolimus) and rapamycin (sirolimus) maintenance therapy as described in the Edmonton protocol (18). The findings in this clinical model demonstrated that a reduction in peripheral lymphocyte count was associated with a chronic elevation of circulating IL-7 and IL-15 in vivo T cell proliferation that led to the expansion of autoantigen-specific T cells.

Results
Reduced blood lymphocyte counts after islet transplantation with immunosuppression. All 13 patients who received islet allografts using the Edmonton protocol experienced a significant, immediate decrease in peripheral lymphocyte counts (13). In all 13 patients, the circulating levels of IL-7 and IL-15 were significantly increased, as were the absolute numbers of CD4 cells (13). In 11 patients, the expansion of autoreactive T cells was more marked than that of memory and naive T cells. In 10 patients, the memory CD45RO+ T cell pool was expanded compared with the naive CD45RA+ T cell pool. In 9 patients, the CD8 cell pool was expanded.

Nonstandard abbreviations used: CyA, cyclosporine A; GAD, glutamic acid decarboxylase; IL-2R, IL-2 receptor; IQR, interquartile range; MMF, mycophenolate mofetil; NOD, nonobese diabetic; T1DM, type 1 diabetes mellitus.

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in blood lymphocyte counts after transplant (pretransplant, mean 2,068 cells/μl; 1 d after transplant, mean 1,364 cells/μl; \( P < 0.0001 \); Figure 1A and Supplemental Figure 1; supplemental material available online with this article; doi:10.1172/JCI35197DS1). Reductions ranged between 15% and 63% of pretransplant values (mean, 33%). Moreover, reductions were seen after each islet infusion (mean reduction after second and third infusions, 33%). Reductions in lymphocyte counts after transplant were similar in patients who received rapamycin pretreatment or the Edmonton protocol, and lymphocyte counts were unaffected during rapamycin pretreatment (data not shown). Lymphocyte counts partially recovered, but, with the exception of a few patients, did not return to pretransplant levels (6 mo after last infusion, mean 1,610 cells/μl; \( P = 0.04 \) versus pretransplant). Reduced counts after transplant were observed for CD3\(^+\), CD4\(^+\), and CD8\(^+\) lymphocytes, whereas CD19\(^+\) lymphocyte counts were minimally affected (Figure 1B).

**In vivo proliferation of lymphocytes after islet transplantation.** In order to determine whether the reduction in lymphocyte count following transplantation promoted homeostatic-like proliferation, we examined fixed freshly isolated PBMCs for the cell proliferation–associated antigen Ki-67 (Figure 2A). Between 0.01% and 0.1% of lymphocytes from normal control subjects stained positive for Ki-67. The percentage of Ki-67\(^+\) cells in patients with T1DM prior to transplantation (median, 0.03%; interquartile range [IQR], 0.01%–0.1%) was similar to that in control subjects. Rapamycin pretreatment, given to 4 patients, did not increase the percentage of Ki-67\(^+\) cells. After islet transplantation, the percentage of Ki-67\(^+\) cells rose significantly in all patients. The percentage of Ki-67\(^+\) cells was already significantly increased within the first weeks after transplantation (mean, 1.4%; range 0.3%–2.7%; \( P = 0.001 \) versus pretransplant) and remained elevated throughout follow-up long after cessation of anti-IL-2R treatment (mean, 1.6%; range, 0.55%–2.6%; \( P = 0.0001 \) versus pretransplant). Although CD3\(^+\) lymphocyte counts returned to pretransplant values in some patients, the percentage of Ki-67\(^+\) cells remained increased (Figure 2B) unless immunosuppression was stopped or changed (see below). In vivo proliferation after islet transplantation was confirmed by propidium iodide (PI) staining to distinguish cells in S-G\(_1\)-M phase (DNA content, >1n to 2n) from cells in G\(_0\)-G\(_1\) phase (DNA content, 1n; data not shown). Almost all Ki-67\(^+\) lymphocytes had surface CD3 and CD45RO, consistent with memory or memory-like T cells (Figure 2C). Both CD4\(^+\) and CD8\(^+\) cells were identified in the Ki-67\(^+\) fraction. IL-7 receptor (CD127) was detected on the majority of the Ki-67\(^+\) cells.

**Homeostatic cytokines IL-7 and IL-15 are increased after transplantation.** In antigen-driven immune responses, T cell proliferation is promoted by T cell autocrine production of IL-2. In a lymphopenic environment, the expansion of peripheral T cells is mainly driven by signaling through the common γ chain by homeostatic cytokines such as IL-7 and IL-15. IL-2, IL-7, and IL-15 were undetectable (<10 ng/l) in sera from normal control subjects and in patients with TIDM before transplant with or without rapamycin pretreatment (Figure 3). Serum IL-7 concentration was significantly increased in samples taken 2 weeks after islet transplantation in all 10 patients examined (median, 27 ng/l; IQR, 18–36 ng/l; \( P < 0.0001 \)) and remained increased throughout the post-transplant period (\( P < 0.0001 \)). Serum IL-15 concentration also increased after transplantation (\( P < 0.002 \)). Serum IL-2 concentration was not significantly increased after islet transplantation and showed sporadic elevations in individual patients. These data suggest that the in vivo proliferation was likely to be mediated through homeostatic cytokines such as IL-7 or IL-15.

Rapamycin, but not mycophenolate mofetil, is permissive for homeostatic proliferation. Two patients who received islet transplantation changed from rapamycin plus FK506 to mycophenolate mofetil (MMF) plus FK506 maintenance therapy because of rapamycin intolerance during the course of follow-up (Figure 4A and Supplemental Figure 2). In both cases, the change to MMF was associated with a marked reduction in Ki-67 positivity without change in the increased IL-7 levels, which indicates that MMF did not suppress compensatory cytokine production but blocked cell proliferation. Both patients remained insulin independent for more than 2 years after islet transplantation. To provide additional support to this hypothesis, we examined Ki-67 positivity and cytokine concentrations in 3 patients who received pancreas plus kidney transplants with antithymocyte globulin induction therapy and MMF plus cyclosporine A (CyA) or FK506 maintenance therapy. In each case, and despite lymphopenia (550, 850, and 1,079 lymphocytes/μl), the percentage of Ki-67\(^+\) cells was not increased (0.01%, 0.1%, and 0.1%), whereas serum IL-7 concentra-
Vigorous proliferation in post-transplant lymphocytes is sensitive to MMF, but not rapamycin. Both Ki-67 positivity and serum concentrations of IL-7 returned to pretransplant levels when immunosuppression was removed from patients. We therefore cultured patient PBMCs in vitro in the absence of immunosuppression (Figure 4B). Washed PBMCs from healthy control subjects or patient PBMCs in vitro in the absence of immunosuppression suppression was removed from patients. We therefore cultured sequential CD3+ T cell counts and the frequency of Ki-67+ lymphocytes after islet transplantation. (C) Phenotype of peripheral blood Ki-67+ cells in representative patient hSR-056-ITA-Ed07 following islet transplantation. Cells were stained for Ki-67 together with an isotype control, CD45RO, the IL-7 receptor CD127, CD3, CD4, or CD8. Numbers denote percentage of cells in the respective quadrants. of cytokines. After 7 d, 10%–80% of the total cells remaining in culture had undergone proliferation. Proliferating cells included CD4+, CD8+, and CD19+ cells. Cells that were left in culture for 3 d before labeling with CFSE did not show further proliferation (data not shown), which indicates that the in vitro proliferating cells were responsive to stimuli received in vivo, presumably homeostatic cytokines. Consistent with these data, no IL-7 was detected in supernatants of 3-d cultures.

The in vivo data on Ki-67 positivity indicated that immunosuppressive drugs varied in their ability to control lymphocyte proliferation in response to increased cytokines. In order to determine which drugs could block homeostatic proliferation, we cultured PBMCs obtained after islet transplantation in the absence and presence of increasing concentrations of different immunosuppressive drugs and quantified proliferation (Figure 4C). At therapeutic concentrations, 10 μg/ml MMF — but not 10 ng/ml rapamycin, 10 ng/ml FK506, or 100 ng/ml CyA — markedly inhibited the in vitro proliferation. Dose-dependent inhibition was observed for each drug, but only MMF inhibited proliferation by more than 50%. The combination of rapamycin plus FK506 at therapeutic concentrations was only partially effective in blocking proliferation.

Homeostatic proliferation can enrich autoreactive T cell populations. Because in vivo proliferating cells had a memory T cell phenotype (CD45RO+ and transplanted patients had an autoimmune disease, we asked whether autoreactive T cells could be preferentially expanded by homeostatic mechanisms. First, we used PE-labeled HLA-A*0201 pentamers loaded with GAD65114–122 peptide to identify glutamic acid decarboxylase 65–autoreactive (GAD65-autoreactive) CD8+ cells (19) in all 5 patients with the HLA-A*0201 allele (Figure 5A). Uncultured cells from post-transplant samples were immediately stained with anti–Ki-67 and HLA-A*0201–GAD65114–122 pentamers. In all cases, there was an enrichment of HLA-A*0201–GAD65114–122+ cells in the Ki-67+ fraction. The total CD8+ lymphocytes, 1.65% ± 0.25% were Ki-67+ and 0.31% ± 0.13% were HLA-A*0201–GAD65114–122+ (1 × 106 cells were acquired for pentamer quantification). The proportion of HLA-A*0201–GAD65114–122+ cells within the Ki-67+CD8+ cell fraction ranged from 2.6% (0.04/1.53; patient hSR-056-ITA-Ed09) to 8.3% (0.15/1.79; patient hSR-063-ITA-Ed06), with a median of 5.6%. In comparison, the proportion of HLA-A*0201–GAD65114–122+ cells within the Ki-67–CD8+ cell fraction ranged from 0.16% (0.16/98.54; patient hSR-064-ITA-Rp04) to 0.26% (0.27/98.47; patient hSR-059-ITA-Ed09), with a median of 0.21% (P = 0.005). Enrichment of HLA-A*0201–GAD65114–122+ cells within the
Ki-67+CD8+ cell fraction was therefore 10- to 34-fold (data not shown). Virtually all HLA-A*0201–GAD65114-122 Ki-67+ cells expressed CD45RO (data not shown). CD45RO+HLA-A*0201–GAD65114-122+ cells were undetectable in nondiabetic control subjects (ref. 19 and data not shown). Enrichment was not specific for the GAD65-reactive T cells, because flu-reactive (i.e., HLA-A*0201–influenza A matrix protein158-66) T cells were also enriched 12- to 29-fold in this Ki-67+ fraction (data not shown). Thus it appears that homeostatic proliferation after islet transplantation preferentially expands memory T cells that include autoreactive clones in patients with T1DM.

We subsequently selected 2 HLA-A*0201–positive patients who were also GAD65 autoantibody–positive and monitored HLA-A*0201–GAD65114-122+ cells over time (Figure 5B). Patient hSR-055-ITA-Ed06 had increasing numbers of HLA-A*0201–GAD65114-122 CD8+ T cells during follow-up, and there was further expansion of these cells as well as GAD65 autoantibodies after immunosuppression was removed because of a nondepleting anti–IL-2R. We observed an acute, moderate reduction of lymphocytes to deplete them or inhibit their function. The conditioning for transplantation usually involves nonspecific targeting of the T cell repertoire, including the presence of autoreactive clones, is determined early in life by thymus-dependent pathways, but the size of clonal populations can be modified later in peripheral lymphoid compartments as a consequence of encountering antigens and by homeostatic mechanisms (1, 20). By examining in vivo and in vitro expansion of lymphocytes from autoimmune patients under immunosuppression after receiving islet allografts, we were able to identify homeostatic mechanisms that expand and enrich autoreactive memory T cells and to determine empirically how the expansion and effector function of these cells could be modulated using immunosuppression. These findings are relevant to the control of transplant rejection and autoimmunity.

Peripheral T cell expansion plays a central role in both maintenance and regeneration of T cells. Primary T cell development occurs largely via thymopoiesis, but efficient seeding of the neonatal peripheral T cell pool requires post-thymic expansion. In humans, there is marked homeostatic expansion of T cells in neonates, which declines rapidly soon after birth to negligible levels in adults (21, 22). In adults, a slow basal turnover of memory T cells is likely to continue in order to avoid attrition. Conditioning for transplantation usually involves nonspecific targeting of lymphocytes to deplete them or inhibit their function. The Edmonton islet transplantation protocol includes induction with a nondepleting anti–IL-2R. We observed an acute, moderate reduction of circulating lymphocyte numbers in all patients and at each islet infusion that partially reverted to pretransplant values soon after transplantation. Even with a relatively modest reduction in circulating lymphocyte numbers, increased lymphocyte turnover...
was seen. This finding indicates that there are sensitive mechanisms controlling lymphocyte homeostasis in adults in vivo.

The clinical condition in which we observed increased T cell turnover included chronic immunosuppression. Immunosuppression per se was unlikely to trigger the increased turnover, because we did not observe an increase in T cell cycling in patients who were receiving conditioning with rapamycin alone prior to transplantation. It is likely that immunosuppression impeded the return to homeostasis, thereby resulting in a chronic state of recovery that facilitated the detection of in vivo lymphocyte proliferation. Our findings are consistent with the detection of increased lymphocyte turnover in patients with immunodeficiency caused by HIV infection (22). Therefore, we propose that the human immune system, like that of the mouse, rapidly responds to lymphocyte loss in order to maintain the size of the T cell compartment.

Mechanisms of peripheral T cell expansion are supported by cytokines signaling through the common \( \gamma \) chain both in mice and in humans (23). The cytokines IL-7 and IL-15 play a prominent and essential role in increasing lymphocyte numbers to optimal ranges. Consistent with this, we observed elevated circulating levels of IL-7 and IL-15 after transplantation, and most cells undergoing proliferation expressed the IL-7 receptor CD127. We also observed increased serum IL-7 concentrations in conditions in which lymphocyte numbers were low but proliferation was prevented through cytostatic immunosuppression. Thus, a mechanism of cytokine release by stromal cells in response to decreased lymphocyte numbers operates in adults.

Control of cytokine-mediated homeostatic proliferation was effectively achieved by the cytostatic drug MMF. Rapamycin, FK506, and CyA hindered, but were permissive to, proliferation. Removal of the drugs in vitro resulted in proliferation of lymphocytes. In vitro proliferation was vigorous and, in the absence of stimuli, ceased within a few days. This was also the case in patients who were treated with MMF and had high circulating IL-7 concentr-
trations, but in whom in vivo proliferation was not increased (data not shown). Islet transplantation patients who stopped immunosuppression quickly returned to a state in which no increased basal turnover of lymphocytes was observed, which suggests that the removal of immunosuppression released IL-7–stimulated cells to rapidly expand and return lymphocyte numbers to homeostasis. Thus, under conditions of lymphocyte consumption, IL-7 and IL-15 are likely to be produced by stromal cells and leukocytes and bind to receptors on lymphocytes. However, proliferation of lymphocytes can be temporarily controlled by cytostatic immunosuppressive drugs and kept in a chronic intermediate state of increased proliferation by other immunosuppressive drugs. It should be noted that our in vitro data suggest that the level of control provided by immunosuppression is likely to be dose dependent.

Evidence from mice suggests that homeostatic mechanisms could expand autoreactive T cells. In the NOD mouse model of autoimmune diabetes, islet-specific memory T cells are sensitive to a γ chain blockade that prevents diabetes (17), and homeostatic proliferation is a major pathway for expansion of autoreactive T cells (16). Hence, the model of islet transplantation in patients with autoimmune diabetes provided a unique opportunity to assess whether homeostatic proliferation expands autoreactive T cells in humans. We studied CD8+ T cells against an epitope of GAD65, previously shown to be recognized by patients with T1DM (19, 24, 25). We have previously shown that pentamers loaded with this peptide can recognize GAD65-specific T cells (19). By direct staining without in vitro priming, we found that the percentage of pentamer-positive GAD65+CD8+ cells was enriched around 30-fold in the Ki-67+ cell population. Direct labeling was also shown using HLA-A*0201–influenza A matrix protein FLU (FLU) pentamers as a positive control and HLA-A*0201–HIV-1 gag p176–64 pentamers as a negative control. (B) Sequential quantification of autoreactive HLA-A*0201–GAD65114–122+CD8+ T cells in patient hSR-055-ITA-Ed06, who was strongly positive for GAD autoantibodies at the time of islet transplantation. The percentage of positive cells (region denoted R3) after in vitro culture increased progressively following transplantation, as shown. The patient ceased taking immunosuppressive therapy at day 56 after transplant as a result of graft failure. (C) In vitro expansion of HLA-A*0201–GAD65114–122+CD8+ cells from patient hSR-055-ITA-Ed06 taken before transplant by culture with cognate peptide GAD65114–122 plus IL-7. The percentage of pentamer-positive cells in the upper right quadrant is indicated. The same experiment was also performed using cells from a nondiabetic control subject.

Figure 5
Expansion of autoreactive T cell clones during homeostatic proliferation. (A) Direct labeling of cells from 5 patients and 1 control subject with PE-labeled HLA-A*0201–GAD65114–122 pentamers (pmr) and Ki-67 (gated on CD8+ cells). Numbers denote percentage of cells in the respective quadrants. Pentamer-positive cells were enriched in the Ki-67+ cell population. Direct labeling was also shown using HLA-A*0201–influenza A matrix protein FLU (FLU) pentamers as a positive control and HLA-A*0201–HIV-1 gag p176–64 pentamers as a negative control. (B) Sequential quantification of autoreactive HLA-A*0201–GAD65114–122+CD8+ T cells in patient hSR-055-ITA-Ed06, who was strongly positive for GAD autoantibodies at the time of islet transplantation. The percentage of positive cells (region denoted R3) after in vitro culture increased progressively following transplantation, as shown. The patient ceased taking immunosuppressive therapy at day 56 after transplant as a result of graft failure. (C) In vitro expansion of HLA-A*0201–GAD65114–122+CD8+ cells from patient hSR-055-ITA-Ed06 taken before transplant by culture with cognate peptide GAD65114–122 plus IL-7. The percentage of pentamer-positive cells in the upper right quadrant is indicated. The same experiment was also performed using cells from a nondiabetic control subject.
after islet transplantation, markedly expanded pentamer-positive GAD65<sub>114-122</sub>-autoreactive memory CD8<sup>+</sup> T cells. Importantly, the in vitro experiments were performed using cells that were depleted of the naive CD8<sup>+</sup> T cell population, thus confirming that bona fide memory autoreactive T cells could be expanded by increased IL-7. This evidence indicates that the homeostatic proliferation occurring after islet transplantation under the Edmonton protocol can lead to increased numbers of autoreactive T cells. It also implies that conditions that lead to homeostatic proliferation could exacerbate autoimmunity and precipitate disease conditions. That promote homeostatic proliferation may include certain infections previously reported to be associated with T1DM (26–28).

Expansion of autoreactive T cells does not necessarily imply that they have effector function. Indeed, after islet transplantation, proliferating T cells did not produce IFN-γ, which suggests that they were not activated. However, release from immunosuppression in vitro led to both expansion and IFN-γ production, implying that the cells were fully capable of effector function but that rapamycin and/or FK506 were able to suppress IFN-γ production. This observation is consistent with studies suggesting that rapamycin inhibits Th1 cell function, in particular IFN-γ production (29). In vitro, we found that only rapamycin inhibited IFN-γ production of a memory GAD65-autoreactive CD4<sup>+</sup> T cell clone isolated from a patient with T1DM, which is in line with an ability of rapamycin to modulate Th1 immunity (30). Autoimmunity is a potential cause of pancreas and islet transplant failure in patients with T1DM (13–15, 31, 32). Therefore, strategies to prevent both expansion and effector function of memory autoreactive T cells are likely to improve the outcome of islet transplantation. Combining rapamycin with the cytostatic properties of MMF was extremely effective in controlling both expansion and IFN-γ production of IL-7 plus antigen-stimulated autoreactive T cells in vitro. This drug combination, previously used successfully for single donor islet transplants (33), could therefore be particularly suited for transplantation in patients with autoimmunity.

Chronic rejection resulting from alloimmune and autoimmune reaction to islet allografts is a problem for the long-term success of islet transplantation (34). Our findings in patients with T1DM indicate that T cells can expand under conditions of homeostatic proliferation. Although we have no direct evidence that alloreactive T cells may be expanded by homeostatic proliferation, a population of expanding allospecific CD4<sup>+</sup>CD25<sup>+</sup>CD45RO<sup>+</sup>CD127<sup>-</sup> T lymphocytes has previously been described in kidney and liver transplant recipients (35). We therefore suggest that homeostatic proliferation can expand pathogenetic T cells and that such expansion could contribute to chronic or acute β cell loss, particularly if immunosuppressive drug levels fall below those necessary to control effector function of expanded memory T cells. We propose that transplantation of cells that are targeted by autoimmunity in patients with autoimmune disease such as T1DM requires specific immunosuppression to avoid expansion of autoreactive T cells. Induction therapies that reduce memory T cell numbers, such as anti-CD45RB (36) or CAMPATH 1H (37), may be more appropriate than blocking IL-2-mediated proliferation. In addition, the combination of rapamycin plus MMF may prevent the expansion and activation of remaining autoreactive Th1 effector cells. Similar strategies may also be effective in controlling autoimmunity per se.

**Methods**

**Patients.** Thirteen patients with T1DM who had fasting C-peptide concentrations less than 0.5 ng/ml received islet allografts between 2001 and 2003. Ten patients underwent transplantation of islets alone with induction therapy of anti–IL-2R antibody (Zenapax; Roche) and maintenance therapy of rapamycin (Rapamune, 10–15 ng/ml circulating target levels; Fujizawa-Astellas Pharma) per the previously described Edmonton protocol (18). The remaining 3 patients underwent rapamycin monotherapy (10–12 ng/ml circulating target levels) for 6–27 weeks before their first islet infusion, according to the rapamycin pretreatment Edmonton protocol. Patients received between 1 and 3 islet infusions (median total islets, 777,450 islet equivalents; IQR, 456,000–839,500) and were followed for a total of 94–1,000 d. The median age of patients was 36 yr (IQR, 29.5–38.5 yr), and 6 were male. In addition, we investigated 5 control subjects, 5 T1DM patients at diabetes onset, and 3 patients with T1DM who received pancreas and kidney transplant with rabbit anti-thymocyte globulin induction therapy (Thymoglobuline, 75 mg over 7 d; Sangstat) — of these, 2 received FK506 (10–15 ng/ml circulating target trough levels) plus MMF (CellCept, 2 mg/d; Roche) maintenance therapy and the third received CsA (Sandimmune, 15 mg/kg/d; Novartis).
In order to study expansion of memory CD8+ T cells, PBMCs were depleted of CD45RO CD8+ naive cells using magnetic beads (Miltenyi Biotech) according to the manufacturer’s instructions. CD8+ cells were separated by negative selection and further divided into CD45RO+ and CD45RO– fractions. The CD8+ fraction containing monocytes and B cells for optimal antigen presentation was also collected. CD45RO CD8+ cells were mixed at a 1:1 ratio with the CD8+ fraction and cultured for 7 d in the presence of 2 μg/ml GAD65114–122 peptide with or without 10 ng/ml IL-7. Cells were harvested and stained with pentamers at 4°C for 1 h. To test the expansion of autoreactive T cells in the presence of immunosuppressive drugs we used BRI-4.13 CD4 T cell clone specific for GAD65535–547 peptide (provided by G.T. Nepom, Benaroya Research Institute at Virginia Mason, Seattle, Washington, USA). T cell clone labeled with CFSE was cultured for 7 d in the presence of 2 μg/ml peptide and 10 ng/ml IL-7 in the presence of rapamycin, FK506, and/or CyA. Cells were then harvested, stained for IFN-γ, and analyzed by FACS.

Statistics. The paired t test was used to compare lymphocyte counts at different time points. The Mann-Whitney U test was used to compare the proportion of Ki-67+ cells and cytokine levels between groups. For all analyses, a 2-tailed P value of 0.05 was considered significant. Statistical analyses were performed using the Statistical Package for Social Science (SPSS version 11.0).

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