MicroRNA reexpression as differentiation therapy in cancer

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Since their discovery in the early 2000s, microRNAs (miRNAs) and their penchant for RNA interference have taken the scientific community by storm, working their way into virtually every corner of biological inquiry. The very nature of their action, the ability to simultaneously extinguish the expression of a multitude of genes and negate their functions, immediately suggested therapeutic promise. In this issue of the JCI, a step toward the realization of this promise is described. Taulli et al. demonstrate that the miRNAs miR-1/miR-206, which are routinely lost in advanced, poorly differentiated rhabdomyosarcoma (RMS) but characteristically expressed in the mature skeletal muscle from which these tumors arise, restore the myogenic differentiation program and block the tumorigenic phenotype (see the related article beginning on page 2366). Their data support the notion that these small RNAs, effectively functioning as “micro-sheriffs” by restoring myogenic law and order, hold substantial clinical potential as differentiation therapy for RMS and perhaps other solid tumors. miRNA reexpression therapy constitutes a novel approach to handcuff oncogenes and arrest tumor development.

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Nonstandard abbreviations used: HDAC, histone deacetylase; miRNA, microRNA; RMS, rhabdomyosarcoma.

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Sometimes you can tell a large story with a tiny subject.
— Eliot Porter

One of the largest, most intriguing scientific stories concerns the ability of developing organisms to orchestrate the expression of a vast multitude of genes to achieve the formation of the various tissues and organs required for life, despite the fact that every cell has the same complement of genetic material. One obvious explanation that has emerged is that every tissue has its own set of transcription factors that regulate tissue-specific gene expression and development. With the discovery of epigenetics, we now know that promoters of many genes, including transcription factors, can be turned on and off during development, through covalent chromatin modifications. Most recently, the discovery of “tiny” tissue-specific microRNAs (miRNAs) has provided further complexity and coordination to the regulation of developmental gene sets, framing a broader picture in which epigenetic programming of specific transcription factors and miRNAs during development define tissue specificity.

miRNAs are small, noncoding RNAs (22–23 nucleotides long), shown to regulate gene expression at translational and posttranslational levels (1). Processed from longer transcripts by Drosha and Dicer, miRNAs mostly bind to the 3′ untranslated regions of target genes and inhibit gene expression translationally and/or by destabilizing the target mRNA (2). miRNAs, encoded in intergenic/intronic noncoding regions once thought to be “junk” DNA, contribute to differentiation when expressed in a tissue-specific manner (2) and even possess the power to shift global mRNA expression patterns with respect to differentiation signatures (3). Such miRNAs are responsible for tissue integrity and homeostasis, behaving functionally as “micro-sheriffs” (Table 1). Like a sheriff, miRNAs maintain law and order in specific tissues, by stringently regulating the expression of their target genes, especially oncogenes. Accumulating evidence now suggests that such miRNAs are assigned their working “zip codes” early during development by epigenetic modifications of miRNA promoters (4).

With respect to cancer, miRNAs are often located in genomic unstable regions and therefore are typically downregulated in tumors (reviewed in ref. 5); moreover, inhibiting miRNA biogenesis tends to enhance tumorigenesis (6). Downregulation may be achieved through mutation or by epigenetic silencing of the miRNA, resulting in loss of tissue-specific miRNA synthesis and overexpression of pro-proliferation genes (i.e., oncogenes); these miRNAs normally function as tumor suppressors (7). Of course, miRNAs can also act as oncogenes (8, 9). Many investigators have posited that reexpression of specific miRNAs may have therapeutic antitumor value (10, 11). In a report published in the current issue of the JCI, Taulli et al. demonstrate that expression of specific miRNAs regulating skeletal muscle development, miR-1/miR-206 (also known as myomiRs), is reduced in rhabdomyosarcoma (RMS) (12). RMS tumors, the most common soft tissue sarcomas in pediatric patients and young adults, are thought to arise from the skeletal muscle lineage, coexpressing markers of proliferation and myogenic differentiation (13). Reexpression of these myomiRs to physiological levels suppressed many aspects of the transformed phenotype and induced myogenic differentiation (12), raising the possibility that miRNA reexpression may represent effective differentiation therapy for RMS and perhaps other cancer types.

miRNA reexpression as differentiation therapy in cancer

The discovery of miRNAs has added an entirely new dimension to antitumor therapeutic approaches; the potential of noncoding RNAs as drugs for cancer patients is both intriguing and compelling. As miRNA expression seems to be altered in many human diseases, including cancer, the miRNA revolution has already set the stage for “miRNA reexpression therapy” (Figure 1). Reexpressing lost miRNA in a cell can deliver a dramatic effect, because miRNAs regulate a vast number of genes and pathways. Among the many genes that miRNAs can regulate are oncogenes and tumor suppressors, targets of drugs currently used in the clinic. Although a few miRNAs are overexpressed in cancer and seem to function as oncogenes themselves
(miR-17-92, miR-155), a greater number of miRNAs have been shown to be downregulated in cancer and have the potential to act as tumor suppressors (5, 9) (i.e., Let-7, miR-15/miR-16, miR-1/miR-206, miR-29, miR-124, miR-143/miR-145; see Table 1 and Figure 1). miRNA reexpression and downregulation have both been shown to have antitumor effects (9, 14). Silencing an oncogenic miRNA could allow reexpression of tumor suppressor genes, while reexpressing a tumor suppressor miRNA could downregulate multiple oncogenes (4).

Reexpression, to physiological levels, of tissue-specific miRNAs that are lost in cancer can induce the dedifferentiation of cancer cells (12, 14). Strikingly, transfection of the myomiR miR-1 or the neural-specific miR-124 in HeLa cells downregulated hundreds of genes and shifted the miRNA expression profile toward that of the tissue in which they were originally enriched (3). Transfection of miR-124 induced neuronal phenotypic changes, by affecting the expression of neuronal-specific markers in stem cells derived from both mouse neural/brain tumor and human glioblastoma multiforme (11). It was also recently demonstrated that miR-29, a ubiquitous miRNA previously shown to suppress tumorigenicity by normalizing atypical patterns of methylation in non–small cell lung cancer cells (15), acts as an enhancer of myogenic differentiation and a suppressor of RMS (16) and is aberrantly expressed in RMS (17). Collectively, these data suggest that tissue-specific, tumor suppressor miRNAs have the potential to promote the redifferentiation of tumor cells to their normal counterparts and solid malignancies to their original tissue types.

**MyomiR-1/206 reexpression effectively targets RMS**

The two major histological types of RMS tumors are alveolar RMS, the more aggressive subtype defined by specific translocations involving the genes paired box 3 (PAX3) or paired box 7 (PAX7) and forkhead box O1 (FKHR also known as FOXO1a), and embryonal RMS, which is less genetically well defined (18). These tumors differ in occurrence, location, patient age, and, most importantly, prognosis. It is widely recognized that RMS development represents a failure of myoblast differentiation (19). A subset of miRNAs, the myomiRs, are specifically expressed in striated muscle and have been shown to help regulate myogenesis, muscle growth, cardiac function, and hypertrophy (20). MyomiRs consist of three bicistronic pairs of miRNAs (miR-1-1/miR-1-2/miR-133a-2, miR-1-2/miR-133a-1, and miR-206/miR-133b) (12). All myomiRs have very similar sequences, can target the same mRNAs, and function prominently in the development of both skeletal muscle and heart. miR-206 is a skeletal muscle–specific miRNA expressed during development and postnatally, although expression levels in mature muscle are low compared with those for miR-1 (reviewed in ref. 21). Notably, miR-1/miR-206 expression can promote differentiation of muscles, whereas myomiR-133 expression appears to promote myoblast proliferation (21). MyomiR-1/206 are also known to target the hepatocyte growth factor/scatter factor receptor MET (22, 23), miR-206 is upregulated during perinatal skeletal muscle development in mice and increases in myotubes over the course of differentiation. In chicken embryos, miR-206 is expressed in the somatic myotome (21). In zebrafish, miR-1 and miR-133 shape muscle gene expression and regulate sarcomeric actin organization (24, 25). Interestingly, changes in miR-1 abundance have also been implicated in cardiac muscle disease, including arrhythmia and heart failure (21).

In their new study, Taulli et al. demonstrate that myomiR-1/206 expression/function is very low or undetectable in both primary embryonal and alveolar RMS (12) (Figure 1). Moreover, upon growth factor deprivation, which normally promotes a switch from proliferation to differentiation, miR-1/miR-206 could not be induced in RMS cell lines. Strikingly, Taulli and colleagues went on to show that forced expression of pre–miR-1 and –miR-206 reduced the ability of the RMS to grow in soft agar and induced myogenic differentiation (12) (Figure 1). An inducible lentivirus vector was employed to confirm that miR-206 could promote in vitro molecular and morphological differentiation, cell cycle exit and apoptosis, and impede invasiveness. The role of miR-206 as a tumor suppressor was confirmed by transplantation of the transduced RMS cells into immunocompromised mice, in which expression of miR-206 was shown to

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**Table 1**

miRNAs are expressed in a tissue-specific manner and play an important role in maintaining tissue-specific functions and cellular differentiation

<table>
<thead>
<tr>
<th>Organ</th>
<th>miRNA</th>
</tr>
</thead>
<tbody>
<tr>
<td>Brain, neural</td>
<td>miR-124, miR-125, miR-128, miR-9, miR-379-410 cluster (miR-124), miR-200, Let-7</td>
</tr>
<tr>
<td>Muscle</td>
<td>miR-1, miR-206, miR-133</td>
</tr>
<tr>
<td>Heart</td>
<td>miR-1-2, miR-133, miR-134, miR-208</td>
</tr>
<tr>
<td>Prostate</td>
<td>miR-125b, miR-143, miR-145, Let-7C</td>
</tr>
<tr>
<td>Breast</td>
<td>Let-7a, miR-10b, miR-125, miR-145</td>
</tr>
<tr>
<td>Liver</td>
<td>miR-122, miR-152, miR-30</td>
</tr>
<tr>
<td>Lung</td>
<td>miR-29, Let-7, miR-26a, miR-199, miR-30</td>
</tr>
<tr>
<td>Pancreas</td>
<td>miR-216, miR-217 miR-7, miR-9, miR-375, miR-376</td>
</tr>
<tr>
<td>Lymph node</td>
<td>miR-142, miR-34</td>
</tr>
<tr>
<td>Cervix, uterus</td>
<td>miR-98, miR-130, miR-99, miR-195, miR-26a, miR-320, miR-28, miR-143, miR-145, miR-218, miR-424, miR-143, miR-145, miR-17</td>
</tr>
<tr>
<td>Colon, small intestine</td>
<td>miR-152, miR-215, miR-341, miR-321, miR-143, miR-145, miR-17</td>
</tr>
</tbody>
</table>

Tissue-specific/tissue-enriched miRNAs are often downregulated and play a role in cancer (shown in bold) (5, 7, 34). For example, brain-specific neuromiR-124 is downregulated in glioblastomas, and Taulli et al. now demonstrate, in this issue of the *JCI*, that myomiR-1/206 are downregulated in RMS (12). Lung-specific pneumomiR-29 suppresses tumorigenicity in non–small cell lung cancer cells (15). Let-7, which is normally expressed at higher levels in normal lung, is downregulated in lung cancer and associated with poor survival. Moreover, miR-143 and miR-145 have been shown to be downregulated in breast, cervical, and colorectal cancers (5, 7, 10, 11).
promote differentiation and inhibit tumor growth under a variety of conditions. The authors also used microarray analysis to show that reexpression of miR-206 can differentiate RD18 RMS cells by switching the global mRNA expression profiles back to their original myogenic phenotype. Upregulated genes included many muscle-specific genes, such as those encoding titin, muscle creatine kinase, myosin light chain, troponin C, myomesin 2, and tropomyosin 2. Many of the detected downregulated genes have been associated with cancer, including those encoding the cell cycle, metabolism, and DNA repair. Thus, miR-206 was sufficient to force neoplastic cells into resuming and completing the full myogenic program.

miR-1/miR-206 are known to target MET (22, 23), which is overexpressed in many cancers including RMS. Notably, deregulated MET signaling in genetically engineered mice induces embryonal RMS (26). MET is known to be rapidly downregulated at the onset of myogenic differentiation (27). Taulli and colleagues show that MET downregulation correlates with miR-206 upregulation in normal myogenic cells and, in fact, directly suppresses MET expression in RMS cells (12). This was confirmed through reexpression of constitutively active oncogenic Tpr-Met, which lacks the MET 3′ untranslated region, in miR-206–expressing RMS cells, abrogating myomiR-mediated differentiation. The authors correctly concluded that MET is a key target for the anticancer effect of miR-1/miR-206 in RMS, and their data suggest that reexpression of miR-1/miR-206 to its physiological levels may be useful as a therapeutic differentiation approach for RMS by restoring a normal muscle phenotype (12).

miR-1/miR-206 has been reported to possess activities that strongly influence other solid tumors. miR-206 was shown to target estrogen-responsive genes, including those encoding estrogen receptor, steroid receptor coactivators (SRC1, SRC3) and GATA binding protein 3 (GATA3), and has been suggested as a biomarker for basal-like breast cancers (28). Introduction of miR-206 into estrogen-dependent MCF-7 breast cancer cells inhibited cell growth in a dose-
and time-dependent manner. miR-1 expression was also markedly reduced in primary human hepatocellular carcinomas (HCCs) compared with normal liver tissues. miR-1 reexpression in HCC cells downregulated its direct/indirect targets, including forkhead box P1 (FOXP1), histone deacetylase 4 (HDAC4), and MET, and inhibited cell growth and proliferation (23). Recently, miR-1 has been implicated in lung cancer therapy (22). miR-1 is expressed in the normal bronchial epithelium but downregulated in human primary lung cancer tissues and cell lines, along with its activator C/EBPα (CCAAT/enhancer–binding protein). Notably, HDAC inhibitor treatment of lung cancer cells induced the expression of repressed miR-1, downregulating oncogenes such as MET, PIM1, FOXP1, and HDAC4, and reversing the tumorigenicity of lung cancer cells. Interestingly, miR-1 reexpression made a lung cancer cell line more sensitive to the chemotherapeutic doxorubicin by inducing apoptosis (22).

**microRNA reexpression therapy: the pros, the cons, and the promise**

Like siRNAs, miRNAs are easy to synthesize and can potentially target any gene, including otherwise non-druggable targets. However, miRNA therapy has many advantages over the originally envisioned RNA interference–based therapeutics (siRNA therapy). The major advantage of miRNA therapy is that miRNA reexpression can influence the expression of hundreds of genes involved in many cellular pathways. While siRNA therapy is more gene-specific, miRNA therapy can target an array of different gene products, more closely resembling the action of the so-called “dirty drugs” used in the clinic today; in fact, both the sense and antisense strands of miRNAs might target different miRNAs. Moreover, dirty drugs typically target only a handful of gene products, whereas miRNAs can target hundreds of genes, casting miRNAs as “super” dirty drugs. Of course, as with any such drug, the advantage of hitting a broad spectrum of targets is also a potential disadvantage. However, miRNAs are evolutionarily conserved, and targeting the upregulation or downregulation of a tissue-specific tumor suppressor miRNA or oncogenic miRNA, respectively, to its “physiological level” may incite fewer of the nonspecific, off-target effects often associated with artificial siRNAs or currently available dirty drugs.

miRNA therapy shares many of the disadvantages of siRNA-therapy, including delivery limitations, instability, and off-target effects. A major obstacle to effective miRNA-based therapy is the requirement for successful delivery. Unlike many other drugs, miRNAs do not freely diffuse into cells; therefore, miRNAs may require special delivery approaches to achieve the desired effect. Moreover, small RNAs tend to be unstable and might be degraded upon entering a cell; new methods may be required to stabilize these small sequences. Another factor is that double-stranded RNA and unmethylated CpG sequences are potentially immunogenic; their presence might increase IFN production and induce an immune response in patients. Moreover, in the case of miRNA reexpression therapy of cancer, preventing miRNA expression from exceeding physiological levels also represents a therapeutic challenge. Ultimately, miRNA-based gene therapy (i.e., delivering miRNAs encoded in a vector) has many of the same disadvantages as gene therapy (29–31). However, developing smarter/safer ways to deliver stable miRNAs/miRNA inhibitors into cells should go a long way toward overcoming the majority of these limitations, and preclinical models will be extremely helpful in this regard.

Although successful use of small RNAs as therapeutic agents represents a substantial challenge, miRNA therapeutics also carry great expectations based on the ability of a single miRNA to specifically target multiple oncogenes, many of which are being individually targeted by drugs already in the clinic or under preclinical development. But cancer remains a complex disease and patients with the same diagnosis will harbor gene mutations and polyvarientic polymorphisms in a myriad of permutations; therefore, incorporation of personalized medicine approaches will continue to be advisable (reviewed in ref. 32).

Cancer differentiation therapy has seen success in hematological cancers (e.g., acute promyelocytic leukemia) (5, 33) but not yet for solid tumors. The recent work of Taulli et al. and others (10, 11) on solid tumors demonstrates that coaxing the micro-sheriff out of retirement and back into town represents a promising and novel approach for arresting tumor growth and reestablishing cellular law and order.

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Genomic amplicons target vesicle
recycling in breast cancer
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Aberrant endocytosis, vesicle targeting, and receptor recycling represent emerging hallmarks of cancer. In this issue of the *JCI*, Zhang and colleagues demonstrate that RAB-coupling protein (RCP; also known as RAB11FIP1) is a “driver” of the 8p11–12 amplicon in human breast cancer and mouse xenograft models of mammary carcinogenesis (see the related article beginning on page 2171). Their finding that RAB GTPase function enables genomic amplification to confer aggressiveness to mammary tumors adds significantly to the body of evidence supporting pivotal roles for receptor trafficking in the proliferation and metastasis of cancer.

**RCP is a potential driver of the 8p11–12 amplicon in breast cancer**

DNA copy number aberrations often increase the number of copies of key genes that can drive cancer progression. However, despite improved approaches for mapping regions of copy number increases (known as amplicons), the complex structure and large size of most amplification events make it challenging to identify true “driver” genes.

In this issue of the *JCI*, Zhang and colleagues apply a novel algorithm, termed TRIAGE (triangulating oncogenes through clinico-genomic intersects), to a collection of microarray expression profiles of primary human breast cancers in an effort to identify candidate genes in amplicons that could contribute to patient outcome (1). The TRIAGE algorithm is based on the concept that transcript levels of genes located in amplicons are frequently coordinately elevated. Thus, by mapping RNA levels onto the chromosome, genomic regions deranged by amplicons can be identified. The authors’ application of TRIAGE identified a 1-Mb region contiguous with the well-characterized 17q12 amplicon, which is known to harbor multiple genes, including the receptor tyrosine kinase (RTK) ERBB2 (also known as HER2 and NEU), that are associated with breast cancer development and progression (2). This region was the strongest “hit” returned, supporting the contention that the TRIAGE algorithm provides a novel approach to identifying candidate genes within amplicons that contribute to tumorigenesis.

Using this same approach, Zhang et al. (1) also identified the gene RAB11 family–interacting protein 1 (RAB11FIP1; also known and herein referred to as RAB-coupling protein [RCP]) as a candidate driver of the 8p11–12 amplicon, which is present in 10%–25% of breast cancers and is strongly associated with breast cancer subtype and outcome (3, 4). RCP is an effector and binding partner of the RAB11 family (including RAB11A, RAB11B, and RAB25) of RAB small GTPases that control vesicle recycling. The studies by Zhang et al. (1) demonstrating that RCP (8p11–12) is overexpressed as a consequence of genomic amplification, combined with previous studies of genomic amplicons including RAB25 (1q22) (5) and RAB23 (6p11; ref. 6), suggest that genomic amplicons frequently target vesicle function in cancer.

**RCP exhibits the characteristics of an oncogene**

Zhang et al. performed detailed functional studies to determine whether RCP has oncogene-like characteristics (1). Based on transfection and knockdown studies, they found that RCP is not sufficient to transform naive cells. However, in breast cancer cell lines, RCP decreased growth factor dependence; increased survival under anoikis conditions and induced motility, invasion, and epithelial-mesenchymal transition (EMT) in vitro; and increased tumor growth and progression in vivo, compatible with RCP being a key regulator of tumor aggressiveness.

The authors further show that RCP could be coprecipitated with the H-RAS proto-

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Nonstandard abbreviations used: RCP, RAB-cou-
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