Effects of free fatty acids on glucose transport and IRS-1–associated phosphatidylinositol 3-kinase activity

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To examine the mechanism by which free fatty acids (FFA) induce insulin resistance in human skeletal muscle, glycogen, glucose-6-phosphate, and intracellular glucose concentrations were measured using carbon-13 and phosphorous-31 nuclear magnetic resonance spectroscopy in seven healthy subjects before and after a hyperinsulinemic-euglycemic clamp following a five-hour infusion of either lipid/heparin or glycerol/heparin. IRS-1–associated phosphatidylinositol 3-kinase (PI 3-kinase) activity was also measured in muscle biopsy samples obtained from seven additional subjects before and after an identical protocol. Rates of insulin stimulated whole-body glucose uptake. Glucose oxidation and muscle glycogen synthesis were 50%–60% lower following the lipid infusion compared with the glycerol infusion and were associated with a ~90% decrease in the increment in intramuscular glucose-6-phosphate concentration, implying diminished glucose transport or phosphorylation activity. To distinguish between these two possibilities, intracellular glucose concentration was measured and found to be significantly lower in the lipid infusion studies, implying that glucose transport is the rate-controlling step. Insulin stimulation, during the glycerol infusion, resulted in a fourfold increase in PI 3-kinase activity over basal that was abolished during the lipid infusion. Taken together, these data suggest that increased concentrations of plasma FFA induce insulin resistance in humans through inhibition of glucose transport activity; this may be a consequence of decreased IRS-1–associated PI 3-kinase activity.


Introduction

Increased plasma free fatty acid (FFA) concentrations are typically associated with many insulin-resistant states including obesity and type 2 diabetes mellitus (1–3). Furthermore, raising plasma FFA levels in healthy humans, by triglyceride/heparin infusions, can also acutely induce insulin resistance (4–11). Over thirty years ago, Randle et al. (12, 13) demonstrated that FFAs compete with glucose for oxidation in isolated rat heart and diaphragmatic muscle preparations, and they speculated that increased fat oxidation may cause the insulin resistance associated with diabetes and obesity. They proposed that increased FFA oxidation leads to an increase in the intramitochondrial acetyl-coenzyme A (acetyl-CoA) and reduced/oxidized nicotinamide adenine dinucleotide (NADH/NAD+) ratios, resulting in inactivation of pyruvate dehydrogenase activity. The consequent increase in intracellular citrate concentration causes inhibition of phosphofructokinase resulting in an increase in glucose-6-phosphate levels. The elevated glucose-6-phosphate levels would inhibit hexokinase II activity and then lead to decreased glucose uptake. However, recent studies by our group (14) and others (15, 16) have called this mechanism into question. Boden and coworkers have shown that a reduction in carbohydrate oxidation was responsible for only one-third of the fatty acid–dependent decrease in glucose uptake, while impaired non-oxidative glucose metabolism accounted for the remainder (16). These workers suggested that two different defects might contribute to the impairment in nonoxidative glucose metabolism. At FFA concentrations of ~0.75 mM, they found an increase in glucose-6-phosphate concentrations in muscle biopsies, suggesting an inhibitory effect of FFA on glycogen synthesis activity, whereas at lower FFA concentrations (~0.50 mM) they observed no difference in intramuscular glucose-6-phosphate concentration. In contrast, using carbon-13/phosphorous-31 nuclear magnetic resonance (NMR) spectroscopy under increased plasma FFA concentrations (~1.8 mM), we observed a decrease in intramuscular glucose-6-phosphate concentration associated with a 50% reduction in insulin-stimulated muscle glycogen synthesis (14). These data suggest that acute elevations in plasma FFA levels in humans cause insulin resistance by initial inhibition of glucose transport and/or phosphorylation activity that is concurrently followed by a reduction in the rate of both muscle glycogen synthesis and glucose oxidation. Because glucose-6-phosphate (and not intracellular glucose) concentration was measured, it was not possible to distinguish between...
an FFA-induced reduction in glucose transport activity or an FFA-induced reduction in glucose phosphorylation (hexokinase II) activity.

The mechanisms that may underlie these FFA-induced changes in glucose transport/phosphorylation activity are unknown but may include effects on the insulin signaling cascade. The effect of elevated FFA levels on insulin binding and post-receptor insulin-mediated signaling is presently not well understood, and contradictory data have been found (17–19). The conflicting results seen in these in vitro studies may be due to differences in cell types or conditions of the incubations. In the only human study to date, Gumbiner et al. (20) examined insulin receptor autophosphorylation in muscle biopsies taken before and after a lipid/heparin infusion and found no reduction in insulin receptor autophosphorylation.

The experiments performed in this study were designed (a) to examine whether elevations in plasma FFA concentrations cause a reduction in glucose transport activity or hexokinase II activity by measuring intracellular glucose concentrations using a novel carbon-13 NMR approach, and (b) to examine whether insulin-stimulated IRS-1–associated phosphatidylinositol (PI) 3-kinase activity in muscle is affected by elevations in plasma FFA concentrations.

Methods

Subjects. Fourteen healthy volunteers (thirteen males, one female; age range: 18–33 years; body wt: 75.8 ± 2.9 kg; body mass index: 23.8 ± 0.5 kg/m²) without family history of diabetes mellitus, dyslipidemia, or bleeding disorders were put on an isocaloric diet (30 kcal per kg/day; carbohydrate/protein/fat: 60%/20%/20%) for 3 days. They were admitted to the Yale/New Haven Hospital General Clinical Research Center the evening before the study and fasted for 12 h before participating in either an NMR or muscle biopsy study. On the morning of the study, Teflon catheters were inserted in the antecubital veins of the right and left arm for blood sampling and glucose/lipid/hormone infusions, respectively. Each subject was studied twice (basal plasma FFA concentrations [glycerol/heparin infusion] or increased plasma FFA concentrations [lipid/heparin infusion]) in either an NMR protocol (n = 7) or muscle biopsy protocol (n = 7). Informed and written consent was obtained from each volunteer. All protocols were approved by the Yale University Human Investigation Committee.

Lipid/glycerol infusions. To examine the effect of increased plasma FFA concentrations on insulin-stimulated muscle glucose metabolism, plasma concentrations of FFAs were increased by intravenous infusion of a triglyceride emulsion (1.5 ml/min; Liposyn II, Abbott Laboratories, North Chicago, Illinois, USA) combined with a prime (200 IU)–continuous (0.2 IU per kg/min) infusion of heparin to activate lipoprotein lipase. During the control experiments, glycerol (0.7 mg per kg/min) was substituted for the lipid infusion. Subjects were randomized with respect to the order in which they received the two infusions. Studies were performed within 2–4 weeks of each other.

Hyperinsulinemic-euglycemic clamp studies. After 5 h of the lipid or glycerol infusion, a hyperinsulinemic-euglycemic clamp using [1-13C]glucose (20g/dl, 40%–60% carbon-13 enriched; Cambridge Isotopes, Cambridge, Massachusetts, USA) was begun (t = 0 minutes). The subjects were also infused with [1,3-13C]mannitol (10g/dl, 99% carbon-13 enriched; Cambridge Isotopes) (prime [3.8 g]–continuous [1.4 g/min]) to assess intracellular glucose concentration, as previously described (21). Insulin (Humulin-Regular; Eli Lilly and Co., Indianapolis, Indiana, USA) was administered as a prime (18 mU/kg)–continuous (1 mU per kg/min) infusion to create conditions of hyperinsulinemia (~400 pM) for 240 min. Plasma concentrations of glucose were maintained at ~5 mM by varying the infusion rate of the [1,3-13C]glucose. Both the lipid and glycerol infusions were continued throughout the clamp for both the NMR and biopsy studies.

NMR studies. In vivo carbon-13/phosphorous-31 NMR spectroscopy was performed on the right calf muscle to assess glycogen, glucose-6-phosphate, and intracellular glucose concentrations. Subjects remained in the supine position inside a 2.1 T Biospec NMR spectrometer system (Bruker Instruments Inc., Billerica, Massachusetts, USA), and phosphorous-31 NMR spectra were acquired (22) to measure intracellular glucose-6-phosphate concentration before the clamp (t = –40 to 0 min) and from 20 to 60 min during the clamp. Carbon-13 NMR spectra for glycogen concentrations were acquired before the clamp (t =

**Figure 1**

Plasma concentrations of glucose (a), insulin (b), and FFAs (c) during the hyperinsulinemic-euglycemic clamp following 5 h of glycerol (open circles) or lipid (solid circles) infusion. FFA, free fatty acid.
–60 to –40 min) and from 120 to 180 min during the clamp to measure rates of muscle glycogen synthesis as previously described (23, 24). Intracellular carbon-13 concentrations were measured by carbon-13 NMR spectroscopy from 180 to 240 min as described previously (21). The radio frequency (RF) coil assembly consisted of two circular hydrogen-1 coil loops (13-cm diameter each) arranged spatially to generate a quadrature field and an 8.5-cm diameter circular surface coil for carbon-13 detection (25). Unlocalized shimming was performed using FASTERMAP (26). The B2 field was calibrated using a stimulated echo y column profile that generated a 180° null at the gradient isocenter. Heteronuclear nuclear Overhauser enhancement was achieved by a train of inversion pulses to the hydrogen-1 nuclei during the relaxation delay. A numerically optimized adiabatic half-passage pulse was used for carbon-13 excitation (27) followed by WALTZ-16 hydrogen-1 decoupling. The repetition rate was 2 sec per scan. The creatine (Cr) signal observed in the carbon-13 NMR spectrum is from both creatine and phosphocreatine (PCr). Assuming the total creatine (Cr + PCr) signal is compartmentalized to the same intracellular muscle space and is natural abundance carbon-13, we calculate the extracellular to intracellular volumes from the in vivo NMR signal intensity of Crtotal to mannitol as described previously (21).

**Muscle biopsy studies.** After the subjects were supine and resting quietly for 60 min, the right vastus lateralis muscle was sterilely prepared with betadine, and 1% lidocaine was given subcutaneously. A 2-cm incision was made using a scalpel, and a 5-mm Bergstrom biopsy cannula (Warsaw, Indiana, USA) was used to perform a baseline punch muscle biopsy. The muscle tissue was blotted and snap frozen in liquid nitrogen. After this, a 5-h infusion of lipid/heparin or glycerol/heparin was given. At the fifth hour, while the infusions continued, a hyperinsulinemic-euglycemic clamp was begun as described above. After 30 min of hyperinsulinemia, a repeat punch muscle biopsy was performed. All samples were stored at –80°C until assay.

**Materials.** Phosphatidylinositol was purchased from Avanti Polar Lipids (Arlington, Alabama, USA) and phosphatidylcholine from Sigma Chemical Co. (St. Louis, Missouri, USA). Reagents for the detection of Western blots by enhanced chemiluminescence, rainbow-colored molecular weight markers for SDS-PAGE, and [y-32P]ATP (6000 Ci/mmol) were purchased from Amersham Life Sciences Inc. (Arlington Heights, Illinois, USA). Protein GPLUS/Protein A-Agarose immunoprecipitation reagent was purchased from Calbiochem (Cambridge, Massachusetts, USA). Antibodies against IRS-1CT (rabbit polyclonal) and PI 3-kinase (p85 subunit, rabbit polyclonal) were a gift from M.F. White (Joslin Diabetes Center, Boston, Massachusetts, USA). Horseradish peroxidase–labeled anti–mouse and anti–rabbit secondary antibodies were obtained from Rockland Inc. (Gilbertsville, Pennsylvania, USA).

**Muscle preparation for insulin signaling studies.** Muscle extracts were made from the frozen specimens. Muscles were first powdered under liquid nitrogen with a mortar and pestle and then homogenized in ice-cold buffer (20 mM HEPES, pH 7.4, 50 mM β-glycerophosphate, 2 mM DTT, 1 mM Na3VO4, 2 mM EDTA, 1 mM PMSF, 1% Triton X-100, 10% glycerol, 10 μM Leupeptin, 3 mM Benazamide, 5 μM Pepstatin A, 10 μg/ml Aprotinin, 200 μg/ml soybean trypsin inhibitor). The homogenate was solubilized on a rotating mixer at 4°C for 30–60 min and centrifuged at 15,000 rpm for 60 min in a 70.1 Ti rotor in a Beckman Ultracentrifuge (Fullerton, California, USA). The supernatant was collected and assayed for total protein content using the Bio-Rad Laboratories Inc. (Hercules, California, USA) protein assay kit.

**PI 3-kinase activity measurements.** IRS-1–associated PI 3-kinase activity was measured in immunoprecipitates obtained with antibodies to IRS-1 as described previously, with some modifications (28, 29). A 1-mg aliquot of muscle extract (total protein) was added to the immune complex composed of protein A/G agarose and anti–IRS-1 antibody and allowed to incubate overnight. The immunocomplexes were collected by centrifugation, washed twice with PBS containing 1% NP-40/100 μM Na3VO4, twice with 100 mM Tris (pH 7.5) containing 500 mM LiCl/100 μM Na3VO4, and twice with Tris (pH 7.5) containing 100 mM NaCl, 1 mM EDTA/100 μM Na3VO4. The pellets were then resuspended in 50 μl of the final wash buffer and 12 mM MgCl2, and 20 μg phosphatidylinositol was added. To start the PI 3-kinase reaction, 10 μl of 440 μM ATP containing 30 μCi [32P]ATP was added to the pellets at room temperature. After 10 min, 20 μl of 8 M HCL was added to stop the reaction, followed by 160 μl of CHCl3/CH3OH (1:1). The phases were separated by centrifugation...

**Figure 2**

Mean rates of glucose infusion during the hyperinsulinemic-euglycemic clamp following 5 h of glycerol (open bar) or lipid (solid bar) infusion.

**Figure 3**

(a) Mean rates of glucose oxidation assessed by indirect calorimetry (measured from t = 95–115 min). (b) Mean rates of muscle glycogen synthesis assessed by carbon-13 NMR (measured from t = 120 to 180 min) during the hyperinsulinemic-euglycemic clamp following 5 h of glycerol or lipid infusion.
Figure 4
Increment in intracellular [G-6-P] obtained from 20 to 60 min after beginning the hyperinsulinemic-euglycemic clamp following 5 h of glycerol or lipid infusion. [G-6-P], glucose-6-phosphate concentration.

Figure 5
Mean intracellular glucose concentration (assessed by carbon-13 NMR) and mean extracellular (plasma) glucose concentration obtained from 120 to 240 min during the hyperinsulinemic-euglycemic clamp following 5 h of glycerol or lipid infusion. *P = 0.04 versus intracellular glucose concentration during glycerol infusion.

Calculations and data analyses

Increments in muscle glycogen concentration were determined from the change in [1-13C]glycogen concentration and the plasma [1-13C]glucose atom percent excess as described previously (24). Rates of glycogen synthesis were then calculated from the slope of the least-square linear fit to the glycogen concentration curve during the given time periods as described previously (24). The intracellular glucose concentration was calculated as described previously (21). All data are presented as mean ± SEM. Statistical comparisons between control and lipid infusion experiments were performed by using the paired Student’s t test, except for the PI 3-kinase data, which were analyzed using analysis of variance followed by the Student-Newman Keuls post-hoc test.

Hypothetical intracellular glucose concentration resulting from reduced hexokinase flux with normal glucose transporter kinetics. We can simplify the metabolic steps regulating glycogen synthesis flux in muscle during steady state as: $\frac{\Delta \text{Glc}_\text{in}}{\Delta t} = V_1 - V_\text{HK}$, where $\Delta \text{Glc}_\text{in}/\Delta t$ is the change in the intracellular glucose concentration per change in time, $V_1$ is the rate of glucose transport influx into the cell, $V_\text{HK}$ is the rate of glucose efflux from the cell, and $V_\text{HK}$ is the rate of hexokinase flux. Assuming Michaelis-Menten kinetics yields: $V_1 = V_\text{max} \frac{\text{[Glc}_\text{in}] / (K_m + \text{[Glc}_\text{in}])}{(K_m + \text{[Glc}_\text{in}] + \text{[Glc}_\text{ex}]})$. If we assume (a) symmetrical transport of glucose by Glut4 ($V_\text{max} = V_\text{max} = K_m = K_m = 0.01$), (b) a $K_m$ for Glu4 of 5 mM (37), and (c) that $V_\text{HK}$ is approximately equal to the glycogen synthetic rate as measured in the glycerol infusion studies (~0.10 mmol per liter muscle/min), then given an extracellular glucose concentration of 5 mM and an intracellular glucose concentration of 0.1 mM, substituting into equation 1 yields: $V_1 = V_\text{max} \frac{\text{[Glc}_\text{in}] / (K_m + \text{[Glc}_\text{in}])}{(K_m + \text{[Glc}_\text{in}] + \text{[Glc}_\text{ex}]})$ – $V_\text{max} \frac{\text{[Glc}_\text{ex}] / (K_m + \text{[Glc}_\text{ex}] + \text{[Glc}_\text{in}]})$. Assuming $V_\text{HK}$ is decreased exclusively by FFA and that it is approximately equal to the glycogen synthetic rate in the lipid-infused subjects (~0.05 mmol per liter muscle/min) then repeating this calculation and substituting $V_1 = V_\text{max} = 0.21$ mmol per liter muscle/min yields: 0.21 [5/(5 + 5)] – 0.21 [Glc] / ([Glc] + 5) = 0.05, [Glc] = 1.8 mM. This value is more than 40 times higher than that measured in the lipid-infused subjects.

Results

During the hyperinsulinemic-euglycemic clamps, plasma glucose concentrations remained constant (~5 mM) compared with the basal period in both the glycerol and lipid infusion studies (Fig. 1a). In both studies, plasma insulin concentrations increased to ~400 PM almost immediately after the clamp was begun and remained at that level throughout the clamp (Fig. 1b). In the glycerol infusion study, the plasma FFAs remained at the same levels after the five-hour glycerol/heparin infusions (~0.5 mM) and decreased to ~0.1 mM once the hyperinsulinemic-euglycemic clamp was started (Fig. 1c). During the lipid infusion, the plasma FFA concentrations increased to ~1.8 mM and remained at this level throughout the clamp (Fig. 1c). These plasma concentrations of FFA are similar to what we have reported previously (14), but...
higher than those reported by Boden et al. using a similar protocol (15). It is possible that these differences are due to some post-collection lipolysis in the present studies. However, to the extent that this occurred, plasma FFA concentrations would be overestimated and any observed effects of FFA on insulin-mediated glucose metabolism would be underestimated.

During the hyperinsulinemic-euglycemic clamp, whole-body glucose metabolism, as reflected by the mean glucose infusion rate, was ~50% lower in the lipid infusion study as compared with the glycerol infusion study from the very beginning of the clamp (Fig. 2). It was also associated with ~60% reduction in the rate of whole-body glucose oxidation (Fig. 3).

**Discussion**

In contrast to previous studies (14–16), in which the lipid infusion was started five hours prior to the start of the hyperinsulinemic-euglycemic clamp, and there was no detectable increase in activity compared with the glycerol infusion studies (400 ± 120% of basal), during the lipid infusion studies, there was a >90% reduction in the insulin-stimulated increment in intracellular glucose-6-phosphate concentration compared with the glycerol-infused study during that same interval (Fig. 4). It was also associated with ~60% reduction in the rate of whole-body glucose oxidation (Fig. 3).

**Figure 6**

IRS-1–associated PI 3-kinase activity in muscle biopsies obtained before the glycerol/lipid infusions (basal) and after 30 min of the hyperinsulinemic-euglycemic clamp following 5 h of glycerol or lipid infusion. PI3-kinase activity was measured in muscle biopsies obtained before the insulin-stimulated increment in intracellular glucose-6-phosphate concentration should be

**Figure 6**

IRS-1–associated PI 3-kinase activity in muscle biopsies obtained before the glycerol/lipid infusions (basal) and after 30 min of the hyperinsulinemic-euglycemic clamp following 5 h of glycerol or lipid infusion. PI3-kinase activity was measured in muscle biopsies obtained before the hyperinsulinemic-euglycemic clamp following 5 h of glycerol or lipid infusion.


≥1.8 mM (see Methods). However, if the impairment was at the level of glucose transport there should be either no difference or a decrease in the intracellular glucose concentration. We found that elevated plasma FFA concentrations caused a significant reduction in the intracellular glucose concentration in the lipid infusion studies compared with the glycerol infusion studies. These data imply that the rate-controlling step for FFA-induced insulin resistance in humans is glucose transport and offers further evidence against the Randle mechanism, which predicts an increase in both intracellular glucose-6-phosphate and glucose concentrations.

This reduced glucose transport activity could be the result of FFA effects on the Glut4 transporter directly (i.e., alteration in Glut4 trafficking, Glut4 budding, Glut4 fusion, Glut4 activity, etc.) (31), or it could result from FFA-induced alterations in upstream insulin signaling events resulting in decreased Glut4 translocation to the plasma membrane. To further explore the latter possibility, we examined IRS-1-associated PI 3-kinase activity in muscle biopsy samples using the identical lipid infusion protocol. We found that similar elevations in plasma FFA levels abolished insulin-stimulated IRS-1-associated PI 3-kinase activity compared with a fourfold stimulation observed in the glycerol infusion studies. This work demonstrates for the first time in vivo that FFA-induced insulin resistance, as reflected by changes in intramuscular glucose-6-phosphate concentration, occurred in temporal relationship with alterations in insulin-stimulated PI 3-kinase activity — an important component of the insulin signaling pathway leading to activation of glucose transport activity in skeletal muscle (32, 33).

The reduced PI 3-kinase activity may be due to a direct effect of intracellular FFA (or some FFA metabolite) on PI 3-kinase and/or secondary to alterations in upstream insulin signaling events. An attractive hypothesis is that elevated plasma FFA levels leads to accumulation of intracellular fatty acyl CoAs; these then activate a serine kinase (possibly protein kinase C⁶), leading to phosphorylation of serine sites on IRS-1 and/or the insulin receptor, which reduces IRS-1-associated PI 3-kinase activity (34). In support of the latter possibility, muscle strips from insulin-resistant obese subjects showed blunted insulin-stimulated PI 3-kinase activity associated with reduced insulin-stimulated insulin receptor and IRS-1 tyrosine phosphorylation (29). Similarly, Björnholm et al. (35) found reduced insulin-stimulated IRS-1 phosphorylation associated with reduced PI 3-kinase activity in muscle strips taken from subjects with type 2 diabetes suggesting a defect in the signaling pathway upstream of PI 3-kinase. More recently, Anai et al. (36) found lower insulin-stimulated PI 3-kinase activity in the Zucker fatty rat associated with relatively mild reductions in IRS-1 and IRS-2 phosphorylation. However, in contrast to the present study, none of these studies were able to determine whether or not these alterations in insulin signaling were primary or secondary to the insulin resistance. In the only human study of the effect of increased plasma FFA concentrations on the insulin signaling pathway, Gumbiner et al. (20) found no change in insulin binding or insulin receptor tyrosine kinase in skeletal muscle biopsies taken after a lipid/heparin infusion when compared with a glucose infusion; however, IRS-1, IRS-2, and PI 3-kinase proteins were not examined.

In conclusion, we found that elevations in plasma free fatty acid concentration cause insulin resistance in human skeletal muscle by reducing insulin-stimulated glucose transport activity, and this resistance appears to be a consequence of altered insulin signaling through IRS-1–associated PI 3-kinase. This mechanism may play an important role in causing the insulin resistance associated with obesity and type 2 diabetes.

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