A genetic model of substrate deprivation therapy for a glycosphingolipid storage disorder

Yujing Liu,1 Ryuichi Wada,1 Hiromichi Kawai,1 Kazunori Sango,1 Chuxia Deng,1 Tadashi Tai,2 Michael P. McDonald,3 Kristlyn Araujo,3 Jacqueline N. Crawley,3 Uwe Bierfreund,4 Konrad Sandhoff,4 Kinuko Suzuki,5 and Richard L. Proia1

Introduction

The glycosphingolipid (GSL) storage diseases are caused by genetic disruption in the lysosomal degradation pathway of GSLs (1, 2). These disorders include Tay-Sachs disease, Sandhoff’s disease, Gaucher’s disease, Fabry’s disease, Krabbe’s disease, and several others. In most of these diseases, GSLs accumulate to massive levels in cells, particularly in neurons, causing neurodegeneration and a shortened life span.

Neurons are primary sites of GSL storage because gangliosides — sialylated GSLs — are normal and relatively abundant constituents of neuronal membranes (1). GSLs are synthesized by the sequential addition of monosaccharides to ceramide by multiple Golgi membrane–bound glycosyltransferases (Fig. 1). The relatively simple gangliosides GM3 and GD3 are produced by the monosialylation and disialylation, respectively, of lactosylceramide. Synthesis of the more complex gangliosides is dependent on the subsequent action of β1,4-N-acetylgalactosaminyltransferase (GalNAcT, Ga3/ Ga2 synthase; EC 2.4.1.94). After transport to the plasma membrane, the GSLs are ultimately delivered to lysosomes for degradation. Their degradation involves the stepwise removal of individual saccharides through the concerted action of lysosomal enzymes and activator proteins. In the GSL storage diseases, the absence of particular enzymes or activator proteins blocks the degradation pathway and leads to accumulation of GSL substrates unique to each disease.

An emerging strategy for the treatment of these diseases is substrate deprivation therapy (3, 4). In this approach, a specific inhibitor of GSL biosynthesis is used to reduce lysosomal storage by lowering the synthetic rate to more closely approximate the compromised rate of degradation. Platt et al. (4) demonstrated the potential validity of this approach with the oral administration of N-butyldeoxynojirimycin (NB-DNJ), an inhibitor of the first glycosyltransferase in GSL synthesis, to a Tay-Sachs disease mouse model that had central nervous system (CNS) storage of GSLs, but which was without clinical manifestations. This regimen substantially lowered brain levels of abnormally stored GSLs in the model mice.

Although the results suggest that the strategy may ultimately lead to an effective treatment for the GSl storage diseases, a number of important issues remain. These include the expected clinical benefit of the therapy, potential deleterious effects of GSL depletion, and the diseases most suitable for the therapy. We have addressed these issues by constructing a genetic model of substrate deprivation therapy for a GSL storage disease through the establishment of mice with simultaneous blocks in GSL synthesis and degradation pathways.
Methods

Generation of GalNAcT–/– and Hexb–/– GalNAcT–/– double knockout mice. Genomic clones from a 129/Sv strain library (catalog no. 946306; Stratagene, La Jolla, California, USA) containing the mouse GalNAcT gene were isolated by using a mouse GalNAcT cDNA probe (5). One genomic clone was used to construct the targeting vector containing a 7.8-kb genomic DNA sequence. The targeting vector (20 µg) was linearized and electroporated into 2 × 10^5 TC1 embryonic stem (ES) cells (6) in a volume of 0.8 ml at 400 V and 25 µF (Bio-Rad Laboratories Inc., Hercules, California, USA). The G418, ganciclovir-resistant ES cells were obtained and screened by Southern analysis. Chimeric mice were established from targeted ES cells as described (6).

Male Hexb–/– mice were crossed to female GalNAcT–/– mice to generate Hexb–/– GalNAcT–/– mice. These double heterozygous mice were crossed to generate Hexb–/– GalNAcT–/– double knockout mice. The knockout mice were a combination of C57BL/6 and 129/Sv strains.

Behavioral analysis. Sixty-four mice (10 Hexb–/– GalNAcT–/–: 6 males, 4 females; 16 Hexb–/– GalNAcT–/–: 8 males, 8 females; 14 Hexb–/– GalNAcT–/–: 10 males, 4 females; 10 Hexb–/– GalNAcT–/–: 3 males, 7 females; and 14 Hexb–/– GalNAcT–/–: double knockouts, 7 males, 7 females) completed 8 weeks of behavioral testing. Mice that survived longer completed additional behavioral testing. All mice were 11 weeks old at the start of behavioral testing. Mice were group-housed by gender, 2 to 5 per cage. Food and water were freely available in the home cages. The colony room was maintained on an artificial 12-h light/12-h dark cycle, with lights on at 6 am.

Rotorod. The ability to maintain balance on a rotating cylinder was measured with a commercially available rotorod apparatus (Model 7650; Ugo Basile, Varese, Italy). The cylinder was 3.2 cm in diameter with a surface of scored plastic. Mice were confined to a section of the cylinder 6.0 cm long by black Plexiglas dividers. Each mouse was placed on the cylinder, which increased rotation speed from 5 to 40 rpm over a 5-min period. The primary measure of interest was the latency at which mice either fell off the rotorod or clung to the rod while it turned one full rotation. Mice that did not fall or cling during the 300-s trial were removed and given a score of 300. Rotorod performance was measured once every 2 weeks for 8 weeks.

Righting reflex. Mice were tested at 17 and 23 weeks old. Righting reflex was measured by turning the mouse on its back on a flat stainless steel surface. Latency to turn itself upright was recorded. Mice not righting themselves within 10 s were turned right-side up and assigned a score of 10.

Statistical analysis. Group comparisons were made using ANOVA. Analyses of performance over time were made using repeated measures ANOVA (RMANOVA), with genotype as a between-subjects factor and age at testing as the repeated measure. Follow-up comparisons of repeated-measures data were made using single-factor and two-group RMANOVA or Bonferroni-corrected t tests.

Biochemical analysis. The analysis of lipids followed a method described previously (7). Fluorophore-assisted carbohydrate electrophoresis (FACE) was used for urinary oligosaccharide analysis with a kit from Glyko Inc. (Novato, California, USA) (8). Urine samples were normalized by using a creatinine quantitative kit (catalog no. 555-A; Sigma Chemical Co., St. Louis, Missouri, USA). Aliquots of urine were incubated overnight at 37°C with the ANTS (8-aminonaphthalene 1,3,6-trisulfonic acid, disodium salt) reagent followed by high-resolution PAGE analysis with a kit from Glyko Inc. (Novato, California, USA). The analysis of lipids followed a method described previously (7). Fluorophore-assisted carbohydrate electrophoresis (FACE) was used for urinary oligosaccharide analysis with a kit from Glyko Inc. (Novato, California, USA) (8). Urine samples were normalized by using a creatinine quantitative kit (catalog no. 555-A; Sigma Chemical Co., St. Louis, Missouri, USA). The tissues for frozen sections were immersed in fixative overnight. After cryoprotection by immersion in 0.1 M phosphate buffer (pH 7.4) containing 20% sucrose, the tissues were snap frozen and sections were stained with PAS. For electron microscopic analysis, tissues were processed for embedding in plastic resin. Sections (1 µm thick) were cut and stained with toluidine blue. Appropriate areas for further thin sectioning were selected under light microscopy and trimmed for ultrathin sectioning. Sections were double stained with uranyl acetate and lead citrate and examined with a Zeiss 10a electron microscope.

Immunohistochemistry. The mice examined were 4.5-month-old wild-type, Hexa–/–, Hexb–/– GalNAcT–/–, and Hexb–/– GalNAcT–/– mice. The brains, spinal cords, kidneys, livers, and spleens...
were removed, embedded in O.C.T. compound (Sakura Finetek USA Inc., Torrance, California, USA) and quickly frozen in liquid nitrogen.

GM2 and N-linked oligosaccharide storage was detected by an indirect immunofluorescence method using anti-GM2 (9) anti–N-linked oligosaccharide (10) monoclonal antibody. The latter antibody binds N-linked oligosaccharides with a nonreducing terminal b-linked GlcNAc residue. Frozen sections (7 µm thick) were fixed in cold acetone for 10 min and air dried. After rehydration in PBS, the sections were incubated with 5% normal goat serum in PBS for 1 h. The culture supernatant from mouse hybridomas GMB28 (9) (anti-GM2) and OMB4 (10) (anti–N-linked oligosaccharide) were then applied to the sections, and the slides were incubated at room temperature for 1 h. After rinsing with PBS, the sections were incubated with FITC-labeled anti–mouse IgM antibody (Sigma Chemical Co.) for 1 h. The fluorescence was observed under a Leica fluorescence microscope.

Results

Experimental design. We have previously produced a clinically and pathologically accurate mouse model of Sandhoff’s disease through the disruption of the gene (Hexb) encoding the β subunit of β-hexosaminidase (11). These mice are deficient in both β-hexosaminidase A (βA) and B (βB), the major forms of lysosomal β-hexosaminidase. Their nearly total β-hexosaminidase deficiency — only a very small amount of β-hexosaminidase S (αA) remains — renders the mice compromised in their ability to degrade Gb3 ganglioside and the related Gb2 glycolipid. As in the human disease, these glycolipids accumulate in neurons causing apoptosis (12), progressive loss of motor behaviors, and a severely shortened life span (11).

To investigate consequences of totally eliminating Gb3 ganglioside and Gb2 glycolipid synthesis in Sandhoff’s disease mice, we first generated mice deficient in GalNAcT. GalNAcT transfers N-acetylgalactosamine onto glycolipid acceptors to produce complex glycolipids, including Gb2 and GM2 (Fig. 1). The GalNAcT gene was disrupted by a replacement-type targeting vector in which exon 6, exon 7, and part of exon 8 were deleted and replaced with the MC1NeoPolyA (13) selection cassette (Fig. 2a). GalNAcT–/– mice were obtained through heterozygous matings with a frequency expected from mendelian inheritance, suggesting the absence of embryonic lethality. The inactivation of the GalNAcT gene was proven by Northern analysis: no GalNAcT-related transcripts were detected in the brain and liver of GalNAcT–/– mice (not shown). Because of the absence of GalNAcT, these mice were unable to synthesize complex gangliosides normally present in the brain (Fig. 2c, lane 1). The complex gangliosides (Fig. 2c, lane 7) were replaced by Gb3, Gb2, and lactosylceramide in the brains of the GalNAcT–/– mice (Fig. 2c, lanes 3 and 4), structures synthesized before the
block at GalNAcT (Fig. 1).

Despite the striking absence of complex ganglioside structures, the \textit{GalNAcT\textsuperscript{–/–}} mice were grossly normal at 12 months old. This finding is similar to the previously generated \textit{GalNAcT}\textsuperscript{–/–} knockout mice, in which the \textit{neo} gene was simply inserted into exon 4 of the gene (14), even though our mice were created with a more severe, multiexon gene deletion.

A major phenotypic finding related to the absence of complex gangliosides in the \textit{GalNAcT\textsuperscript{–/–}} mice was male infertility caused by degeneration of seminiferous tubules (not shown). Female \textit{GalNAcT\textsuperscript{–/–}} mice were fertile.

\textbf{Life span and behavioral analysis.} We generated \textit{Hexb\textsuperscript{–/–} GalNAcT\textsuperscript{–/–}} double knockout mice through matings of double heterozygotes (Fig. 2b). In the same crosses, \textit{Hexb\textsuperscript{–/–} GalNAcT\textsuperscript{+/–}} and \textit{Hexb\textsuperscript{–/–} GalNAcT\textsuperscript{+/-}} mice were also obtained. The life span of each group of mice (\(n = 10\)) was evaluated (Fig. 3a). The mice in the \textit{Hexb\textsuperscript{–/–} GalNAcT\textsuperscript{+/-}} group died 120–150 days after birth, following the onset of a rapid neurodegenerative disease course (11). The group of \textit{Hexb\textsuperscript{–/–} GalNAcT\textsuperscript{–/–}} mice showed a very similar but slightly delayed disease course; they lived until 149–184 days after birth, a significantly longer period than the \textit{Hexb\textsuperscript{–/–} GalNAcT\textsuperscript{–/–}} mice (\(P = 0.0002\)). During the 300-day duration of the experiment, 9 of 10 double knockout mice remained viable; a double knockout mouse died at 170 days after birth for unknown reasons. All of the \textit{Hexb\textsuperscript{+/-} GalNAcT\textsuperscript{–/–}} mice survived the testing period.

Mice of different genotypes were tested on two behavioral tasks, righting reflex and rotorod, that have been proven to monitor the progressive disease course of the \textit{Hexb\textsuperscript{+/-}} mice (11, 15). The \textit{Hexb\textsuperscript{+/-}} mice likely become progressively impaired in these tasks as a result of CNS storage, motor neuron damage, and muscle wasting evident in the disease (16). In the righting reflex task, the mice are placed on their backs, and the time to right themselves is recorded. Normal mice right themselves immediately. As they age, the \textit{Hexb\textsuperscript{+/-}} mice show a decline in righting reflex. In the rotorod task, a measure of motor function and coordination, the time that the mice can remain on a rotating cylinder is determined. The \textit{Hexb\textsuperscript{+/-}} mice show a progressive decline in their ability to remain on the rotorod (11).

There were significant differences across groups on the righting reflex, measured at 17 weeks old (\(F_{4,59} = 12.12, P < 0.0001\)) (Fig. 3b). Post hoc analyses showed that the \textit{Hexb\textsuperscript{+/-} GalNAcT\textsuperscript{+/-}} mice had a significantly impaired righting reflex, compared with all other groups (all \(t\text{'s} > 2.99; all P\text{'s} < 0.0068\)). None of the other group comparisons were significant at 17 weeks old, showing that the double knockout and \textit{Hexb\textsuperscript{+/-} GalNAcT\textsuperscript{–/–}} mice were not impaired at this time. However, at 23 weeks old, the \textit{Hexb\textsuperscript{+/-} GalNAcT\textsuperscript{–/–}} mice had a significantly impaired righting reflex, compared with all other groups (all \(t\text{'s} > 4.12; all P\text{'s} < 0.0013\)). None of the other group comparisons was significant at this age demonstrating that the double knockout mice were not impaired in this task. By 23 weeks old, the \textit{Hexb\textsuperscript{+/-} GalNAcT\textsuperscript{–/–}} mice had died.

As shown in Fig. 3c, rotorod performance — a measure of balance and mobility — during the period that the \textit{Hexb\textsuperscript{–/–} GalNAcT\textsuperscript{–/–}} mice remained viable (11–17 weeks) differed greatly across the five groups (\(F_{3,177} = 9.03, P < 0.0001\)) and over the six-week testing period (\(F_{3,177} = 10.17, P < 0.0001\)). In addition, there was a significant Genotype X Age interaction (\(F_{3,177} = 3.32, P = 0.0002\)), indicating that performance of some mice deteriorated significantly over the six-week testing period, while performance of other mice did not change significantly. Planned follow-up comparisons showed that rotorod performance of the \textit{Hexb\textsuperscript{–/–} GalNAcT\textsuperscript{–/–}} mice was signifi-
significantly worse than that of the control mice ($F_{1,22} = 15.57; P < 0.0001$), replicating results from previous studies (7, 11, 15). Rotorod performance of the double knockout mice was significantly worse than that of the control mice ($F_{1,22} = 9.22; P = 0.0061$). However, the performance of the double knockout mice remained stable with increasing age, compared with the rapidly deteriorating performance of the $Hexb^{-/-}GalNAcT^{+/+}$ mice ($F_{3,78} = 4.86; P = 0.0038$). The $Hexb^{-/-}GalNAcT^{+/+}$ group performed significantly better overall than both the $Hexb^{-/-}GalNAcT^{+/+}$ group ($F_{1,22} = 16.99; P = 0.0004$) and the double knockout group ($F_{1,22} = 8.49; P = 0.008$). However, the performance of the $Hexb^{-/-}GalNAcT^{+/+}$ mice deteriorated much faster than that of the double knockout mice from 11 to 17 weeks old ($F_{3,66} = 5.87; P = 0.0013$). The rate of deterioration in the $Hexb^{-/-}GalNAcT^{+/+}$ mice was not different from that of the $Hexb^{-/-}GalNAcT^{+/+}$ mice ($F_{3,66} = 0.96; P = 0.419$). There was no significant difference in rotord performance between the normal controls and $Hexb^{+/+}GalNAcT^{-/-}$ groups ($F_{1,24} = 0.34; P = 0.566$) during this time period.

At 21 weeks old, the last testing time that included the $Hexb^{-/-}GalNAcT^{+/+}$ mice, there were significant group differences in rotord performance ($F_{3,44} = 18.57; P < 0.0001$). Follow-up comparisons show that the double knockout mice were now performing significantly better than the $Hexb^{-/-}GalNAcT^{+/+}$ mice ($t_{21} = 2.80; P = 0.0108$).

Rotord performance of the control mice was significantly better than both the double knockout mice ($t_{22} = 5.32; P < 0.0001$) and the $Hexb^{-/-}GalNAcT^{+/+}$ mice ($t_{17} = 8.16; P < 0.0001$) at this age. There was no difference in rotord performance between the control and $Hexb^{-/-}GalNAcT^{+/+}$ groups at 21 weeks old ($t_{21} = 0.03; P = 0.979$).

Beginning at about seven months old, the double knockout mice began to display an ataxic gait that progressively worsened over time. At nine months old, these mice exhibited severe phenotypic abnormalities, including muscle wasting and hunched posture, while the $Hexb^{-/-}GalNAcT^{+/+}$ mice appeared normal (Fig. 4).

Glycolipid and oligosaccharide accumulation. Brain gangliosides were isolated from 4.5-month-old mice and analyzed by thin-layer chromatography (TLC) (Fig. 5a). Gangliosides of wild-type mice ($Hexb^{+/+}GalNAcT^{+/+}$) consisted primarily of GM1, GD1a, GD1b, and GT1b (Fig. 5a, lane 1). In $Hexb^{+/+}GalNAcT^{+/+}$ mice, extensive GM2 ganglioside accumulation was superimposed upon the normal pattern of complex gangliosides (Fig. 5a, lane 2). $Hexb^{-/-}GalNAcT^{+/+}$ mice, unable to synthesize most complex gangliosides, exhibited primarily GM3 and GD3 in the acidic lipid fraction (Fig. 5a, lane 3). The acidic lipid pattern of the double knockout mice was virtually indistinguishable from the $Hexb^{-/-}GalNAcT^{+/+}$ mice, with primarily GM3 and GD3 gangliosides and no other abnormally accumulating lipid species (Fig. 5a, lane 4). GM2 storage was

**Figure 5**
Ganglioside accumulation and urinary oligosaccharide excretion. (a) The acidic fraction of sphingolipids from brain and liver tissues was separated by TLC. The ganglioside standards are indicated. (b) Urine was collected from 4-month-old mice of different genotypes. FACE was performed to analyze urinary oligosaccharides. The size standards consist of glucose polymers. The bracket indicates the high-molecular-weight oligosaccharides present in the $Hexb^{-/-}$ single knockout and $Hexb^{-/-}GalNAcT^{+/+}$ double knockout urine but not the wild-type and $GalNAcT^{-/-}$ urine. FACE, fluorophore-assisted carbohydrate electrophoresis.
also completely eliminated in the liver in the double knockout mice (Fig. 5, compare lanes 6 and 8). Likewise, storage of related Ga2 glycolipid was absent in the liver and brain of the double knockout mice (not shown). Because of a block in the degradation of N-linked oligosaccharides caused by the absence of both \( \beta \)-hexosaminidase A and B, Sandhoff’s disease patients and model mice excrete high-molecular-weight oligosaccharides in their urine (15, 17) (Fig. 5b, lane 3). The double knockout mice also excreted elevated levels of high-molecular-weight oligosaccharides (Fig. 5b, lane 5), although the amount was about half that in urine of the Hexa\(^{-/-}\)GalNAcT\(^{+/+}\) mice. The urinary oligosaccharides were sensitive to digestion with \( \beta \)-hexosaminidase (not shown). Abnormal high-molecular-weight oligosaccharides were not detected in the urine of the Hexb\(^{-/-}\) mice (Fig. 5b, lane 4).

Tissue storage of GM2 ganglioside and N-linked oligosaccharides were assessed using anti-GM2 ganglioside (9) and anti-N-linked oligosaccharide monoclonal antibody (10) (Fig. 6). The Tay-Sachs mouse model (18) was examined as a comparison, because although they accumulate GM2 ganglioside in tissues like the Sandhoff’s disease mice, they should not accumulate oligosaccharides because of the retention of \( \beta \)-hexosaminidase B, which is able to degrade N-linked oligosaccharides but not GM2 ganglioside.

In both Hexb\(^{-/-}\) (Sandhoff’s disease) and Hexa\(^{-/-}\) (Tay-Sachs disease) mice, the distribution of GM2 storage detected with antibody was consistent with previous descriptions (11, 19, 20). GM2 immunostaining was extensive and widely distributed in the CNS of Hexb\(^{-/-}\) mice (Fig. 6a, and data not shown). Positively stained neurons were also found in the Hexa\(^{-/-}\) mice, although the GM2 distribution was restricted to particular regions in the CNS (Fig. 6d, and data not shown). GM2 storage was not apparent in the double knockout mice (Fig. 6g) or wild-type mice (data not shown).

In Hexa\(^{-/-}\) mice (Fig. 6, e and f) and wild-type mice (not shown), anti-oligosaccharide immunostaining revealed no significant storage of oligosaccharides in CNS or visceral organs. In Hexb\(^{-/-}\) mice, oligosaccharide storage was detected in cerebellar Purkinje cells (described below), neuronal cells of the cerebral cortex, thalamic nuclei, brainstem, spinal cord (Fig. 6c), and, weakly, in the hippocampus. Double staining with anti-macrophage and anti-oligosaccharide antibodies revealed that significant storage of oligosaccharides was in microglial cells throughout the CNS (not shown).

In the double knockout mice, the neuronal staining with anti-oligosaccharide was detectable but reduced when compared with the Hexb\(^{-/-}\) GalNAcT\(^{+/+}\) mice. Microglial cells storing oligosaccharides were easily detected and found scattered throughout regions of the brain and spinal cord (Fig. 6i).

In the visceral organs, oligosaccharide storage was detectable in macrophages in the liver (Fig. 6, b and h) and spleen as well as in renal tubular epithelium in both the Hexb\(^{-/-}\) GalNAcT\(^{+/+}\) and double knockout mice.
Pathologic analysis. Pathologic differences in 4.5-month-old mice of different genotypes were evaluated. At this age, the Hexb−/−GalNAcT+/− mice showed severe axonal degeneration in the anterior column of the spinal cord (Fig. 7a, arrowheads). By contrast, a similar section from the double knockout mice appeared relatively normal (Fig. 7b). Using combined Luxol fast blue (LFB) and periodic acid-Schiff (PAS) staining, storage neurons containing large amounts of blue-staining inclusions were readily observed throughout the cerebral cortex of the Hexb−/−GalNAcT+/− mice (Fig. 7c, arrowheads). Again, the double knockout mouse did not show this severe and conspicuous pathology (Fig. 7d). Perivascular macrophages stained reddish-pink — indicative of storage — were present in the brains of both double knockout and Hexb−/−GalNAcT+/− mice (Fig. 7, c and d; arrowheads). Although massive neuronal inclusions were not present in the double knockout (Hexb−/−GalNAcT+/−) mice, some neurons were found to contain fine to coarse vacuoles in their perikarya. These neuronal vacuoles were examined by electron microscopy and compared with the neuronal inclusions found in Hexb−/−GalNAcT+/− mice. As described previously, the inclusions of the Hexb−/−GalNAcT+/− mice were electron-dense, lamellar structures consistent with GSL storage (11, 20) (Fig. 7e). Vacuoles in the double knockout mice were very different in appearance. They were relatively empty with some electron-dense debris (Fig. 7f). This appearance is consistent with storage of water-soluble compounds such as oligosaccharides. These pathologic changes described in the double knockout mouse were not observed in the Hexb−/−GalNAcT−/− mice (not shown).

A striking pathologic finding in the double knockout mice was an extensive loss of cerebellar Purkinje cells by 8.5 months old (Fig. 8a), a time when these mice were exhibiting severe ataxia. No evidence of Purkinje cell degeneration was found in 8.5-month-old Hexb−/−GalNAcT−/− mice (Fig. 8b).

Using the anti-oligosaccharide antibody, cerebellar Purkinje cells from double knockout (Fig. 8c) and Hexb−/−GalNAcT−/− mice (Fig. 8d) were found to display a strong granular staining pattern indicative of abnormal substrate accumulation. Little immunoreactivity was seen in the molecular and granular layers. Purkinje cells from Hexa−/− mice and wild-type mice did not show reactivity with the antibody (not shown).

Vascular pathology in the Hexb−/−GalNAcT−/− mice, which has been documented previously (11), included vacuolated cells (storage macrophages) in the liver, spleen, and kidney. Renal tubular epithelial cells were also found to be vacuolated. Similar findings were also present in the double knockout but not in the Hexb−/−GalNAcT−/− mice. The vascular pathology in the double knockout and the Hexb−/−GalNAcT−/− mice corresponded well to the distribution of oligosaccharide storage determined by immunostaining. In both the Hexb−/−GalNAcT−/− and double knockout mice, degeneration of seminiferous tubules in the testis was pronounced.

Discussion

Treatments that can reverse or alter progression of the clinical manifestations for GSL storage diseases are limited and not effective for the majority of the disorders. Enzyme replacement is an effective therapy for Type 1 Gaucher’s disease, a GSL storage disorder that is unique by the absence of CNS manifestations (21, 22). For other disorders that exhibit CNS involvement, enzyme replacement has been difficult to implement thus far because of the inability of enzyme to cross the blood–brain barrier. Recently, bone marrow transplantation has been reported to reverse CNS manifestations in the juvenile form of Krabbe’s disease (23) but has been of little value in other disorders (24).

Substrate deprivation therapy has some features that make it an attractive treatment approach for the GSL storage diseases. First, a single inhibitor may be used to treat several of the disorders. NB-DNJ, the most intensively studied drug, blocks the first glycosylation step in the synthesis of glucose-based GSLs (25) and, thus,
could be used to treat all storage diseases with defects in the degradation of glucose-based GSLs. Second, NB-DNJ crosses the blood–brain barrier to alter GSL storage levels (4) and would potentially influence the CNS manifestations dominant in the majority of the diseases. And finally, substrate deprivation therapy is noninvasive in contrast to the other available treatments: bone marrow transplantation and enzyme replacement.

We have developed a genetic model of substrate deprivation therapy for a GSL storage disease — Sandhoff’s disease — that has allowed investigation into questions of potential efficacy and limitations. To achieve specific and complete elimination of GSL storage, a GalNAcT (Galgt1) knockout mouse was generated that was unable to synthesize the majority of complex gangliosides. Introduction of the disrupted GalNAcT gene into the Sandhoff’s disease background yielded mice with simultaneous blocks in GSL synthesis and degradation.

**Extent of symptom elimination.** The double knockout mice were totally devoid of the major GSL storage substrates normally found in Sandhoff’s disease mice. This resulted in the absence of the massive neuronal storage and the very severe pathology that is a defining feature of the disease (11). The life span of the mice was extended from about 4–5 months to over 12 months, and the rapid neurodegeneration characteristic of the disease was not present.

The small reduction of GSL storage in the Hexb+/– GalNAcT+/– mice increased their life span by only 18% and illustrated the importance of balancing the rates of substrate synthesis and degradation for the prevention of a lysosomal storage disease (26). The Sandhoff’s disease model is similar to infantile disease forms, which have very little GSL degradative activity. Only a nearly complete reduction in GSL synthesis, as in the double knockout mice, would be expected to rescue the disease phenotype. The partial reduction of GSL synthesis afforded by NB-DNJ would be anticipated to be more beneficial in later onset forms of the diseases, which have significant residual degradative activities, in contrast to the infantile forms. Increased effectiveness of substrate deprivation therapy in infantile diseases may be achieved by raising degradative capacity through enzyme replacement, gene therapy, or cell transplantation therapy.

The complete elimination of abnormal GSL storage in the double knockout mice did not completely restore neurologic function to control levels. Eventually the double knockout mice began to exhibit ataxia that progressively worsened over time. The accumulation of a second β-hexosaminidase substrate, N-linked oligosaccharide, appears a likely contributor to the disease phenotype of double knockout mice. Although N-linked oligosaccharide storage in Sandhoff’s disease patients has been well documented (17), its contribution to nervous system pathology and dysfunction has been difficult to discern because of the dominant GSL storage. Our studies with anti-oligosaccharide antibody indicate this storage is widespread in the CNS and may contribute to a striking loss of cerebellar Purkinje cells in the double knockout mice. Oligosaccharide storage was also found to be widely distributed in other tissues of the double knockout mice and may be responsible for much of the visceral pathology in Sandhoff’s disease. Storage of oligosaccharides in tissues and their excretion in urine appeared to be more pronounced in the single knockout Sandhoff’s disease mice than in the double knockout mice. The difference may be due to a nonspecific impairment of lysosomal function by the additional storage of GSLs in the Sandhoff’s disease mice, as has been described in Gaucher’s disease (27).

**Proteoglycans** are also substrates for β-hexosaminidase. However, previous studies on mice with a total hexosaminidase deficiency (Hexa−/− Hexb−/−) indicated that β-hexosaminidase S, present in Sandhoff’s disease mice, is sufficient to prevent significant pathology associated with proteoglycan storage (7, 28). While the Sandhoff’s and double knockout mice (Hexb+/– GalNAcT−/–) are without the characteristic pathology of mucopolysaccharidosis, indicating that proteoglycans are likely not a major pathogenic factor, a minor contribution to pathogenesis by this substrate is difficult to totally rule out.

**Potential for secondary storage disease after substrate deprivation therapy.** In most of the GSL storage diseases, GSLs are the exclusive substrate stored in tissues. With Tay-Sachs disease, the presence of β-hexosaminidase B is sufficient to preclude the storage of N-linked oligosaccharides that occurs in Sandhoff’s disease. Other disorders with exclu-
sive storage of GSL substrates include Gaucher’s disease, Fabry’s disease, and the activator deficiencies. In all these cases, secondary storage diseases after GSL depletion would not appear to be a concern.

GMI gangliosidosis (β-galactosidase deficiency) (29), like Sandhoff’s disease, would have the potential for a new treatment paradigm for the GSL storage diseases. All the results advance substrate deprivation therapy as a treated some potential limitations in the approach, over-

tered CNS more sensitive to damaging effects of oligosac- 
chides may stabilize or protect the CNS from insult or 
injury. Caution should be taken, however, in directly 
prevent GSL storage, may be capable of preventing the 
storage of water-soluble substrates.

Deleterious effects of GSL depletion. Slowing the rate of GSL synthesis may lower their levels in the plasma membrane of cells. Although consequences of this GSL depletion are unclear, the GalNACT+/− mice provide some insight into the effects of total elimination of most complex gangliosides. The GalNACT+/− mice have a normal life span and develop a near normal nervous system with only subtle conduction abnormalities (14). Recently, some degener-

ative changes in their axons and Schwann cells have been noted (30). Male infertility also results from impaired GSL synthesis. However, in the GalNACT+/− mice, the absence of certain gangliosides is absolute and occurs throughout embryonic development. This would not be the case in substrate deprivation therapy where reduction of GSL levels are unlikely to be reduced completely and where GSL reduction would be implemented after development of the nervous system.

The effects of GSL depletion in a compromised CNS may also be a concern. The 11-week-old double knockout mice with no GSL accumulation were unexpectedly less proficient on rotorod than the Hexb+/−/−/− mice, which may also be a concern. The 11-week-old double knockout mice with no GSL accumulation were unexpectedly less proficient on rotorod than the Hexb+/−/−/−/− mice but exhibited no significant defects in their nervous system. (Proc. Natl. Acad. Sci. USA. 93:10662-10667.


Suzuki, K., Proia, R.L., and Suzuki, K. 1998. Mice lacking both subunits of lysosomal β-hex- 


Starr, C.M., Masada, R.I., Hague, C., Skop, E., and Kleck, J.C. 1996. Flu- 

Suzuki, K., Johnson, O.N., Kozak, C.A., and Proia, R.L. 1995. β-1,4-N- 

Suzuki, K., Johnson, O.N., Kozak, C.A., and Proia, R.L. 1995. β-1,4-N- 