Introduction

The immune system serves to protect the organism from external and internal danger by using an elaborate network of immune sensors. In the past, immunologists believed that most of these sensors served to detect pathogens, but there is a growing recognition, elegantly formulated by P. Matzinger (1), that the ability of immune sensors to recognize pathogens may have an evolutionary origin in the detection of molecules on distressed cells from within the organism itself. This point of view is of particular interest for our understanding of immune responses in neurodegeneration in the absence of pathogenic triggers. Importantly, immune sensors are not restricted to dedicated immune cells but specialized sensors are expressed by most cell types including neurons; what’s more, immune sensors are also present inside cells, allowing them to detect internal dangers. Once triggered, the immune system can employ cellular or soluble arbiters, which attempt to remove the cause of injury and initiate repair processes, thereby employing all or part of what is called an inflammatory response. However, in the event that the injurious trigger cannot be removed, the immune response may turn into chronic inflammation or cell death.

It is now clear that the immune response forms an integral part of neurodegeneration, and we are gaining a better understanding of how this system may promote or inhibit neurodegenerative processes. The complement system and microglial cells, for example, are invariably activated in brain regions with neurodegeneration, and genetic studies in animal models demonstrate that these immune effectors can be either protective or harmful (Table 1). A growing number of studies also show associations between neurodegeneration in the brain or spinal cord and immune changes in the periphery. These findings have been discussed extensively (2–4) and will not be reiterated here. Instead, we explore immune responses in neurodegeneration at three levels: (a) cell-autonomous immune mechanisms in neurons, (b) immune responses in the extra-neuronal brain tissue, and (c) immune responses associated with neurodegeneration at a systemic level (Figure 1). Theoretically, neurodegeneration may start at any of these levels, and a failure of the immune responses at each of them may thus contribute to or result in neurodegeneration. While the basic understanding of immune responses in neurodegeneration is growing, the clinical implications for humans are still largely unclear.

Immune responses by neurons

Neurodegeneration ultimately targets neurons and can range from damage to synapses or neurites to cell death. It seems therefore obvious to ask whether cell autonomous immune responses in neurons may have a role in neurodegeneration. However, only in the past few years have scientists started to explore how neurons sense danger using intracellular or surface immune sensors (Figure 2). Once a danger signal is detected, neurons can activate intracellular defense mechanisms and can alert neighboring cells via cell-cell interactions or the release of signaling proteins, neurotransmitters, and other messengers.

Innate immune sensors of cell-autonomous distress. The best studied innate immune sensors are TLRs, which signal through various adaptor molecules to activate NF-κB and/or Stat signaling pathways to induce the production of mostly pro-inflammatory cytokines and chemokines (reviewed in ref. 5). Mammalian TLRs not only recognize pathogen-derived molecules such as lipoproteins, peptidoglycans, single-stranded RNA (ssRNA), double-stranded RNA (dsRNA), and unmethylated CpG motifs, but also endogenous molecules produced by stressed and injured cells, including hsp, mRNA, and fibrinogen. Neurons and neuro progenitor cells express several TLRs, including TLR2, -3, -4, -7/8, and -9, and their functions range from regulating neurogenesis to triggering neurite retraction or cell death (reviewed in ref. 6). Because TLRs can be functionally expressed in the ER and in endolysosomes (7, 8), molecules associated with injury or aging could in principle trigger a pathogenic cascade from within neurons. In support of this idea, mitochondrial DNA (mtDNA), which is released from damaged mitochondria, is recognized by TLR9 in the endosome and can lead to an inflammatory and injurious response (9). While very little is known at this point about the role of innate immune sensors in neurons, studies in macrophages show that stimulation of TLR3, -4, and -7 can be beneficial by inducing autophagy (10, 11). Autophagy, which is a degradation pathway for long-lived proteins and organelles (12), has largely beneficial functions in postmitotic neurons, and it is possible that damaged mitochondria or intracellular protein aggregates in aged neurons may trigger TLR-dependent autophagy for clean up. Defects in autophagy, as they occur in neurodegenerative diseases (13–16), might thus interfere with such an innate immune process.

Another group of innate immune sensors, nucleotide binding oligomerization domain–like (NOD-like) receptors, form cytosolic protein complexes called inflammasomes, which can activate caspase-1, IL-1, and NF-κB–dependent inflammatory responses (17, 18). Recent studies show that NOD-like receptors (NLK) can also trigger autophagy (19–21), which supports an intriguing link among...
recognition of abnormal molecular patterns, initiation of inflammatory pathways, and activation of autophagy. This mechanism may well be evolutionarily conserved and could allow cells to purge themselves of abnormal protein aggregates, aged mitochondria, or other cellular junk apart from intracellular pathogens. That such a scenario may, at least in part, take place in neurons in a cell-autonomous way is supported by elegant studies demonstrating recognition of Sindbis virus capsid protein by the adaptor protein p62, leading to activation of autophagy and protection against virus-induced cell death (22). Additionally, studies in medicinal leech, which is able to regenerate the structure and function of its CNS after mechanical lesions (23), show that both intracellular leech NLR and TLR1 are temporally and spatially associated with regeneration of neurons (24). It will therefore be most interesting to test whether intracellular innate immune sensors are capable of recognizing abnormal patterns or assemblies of the disease-related proteins β-amyloid (Aβ), α-synuclein, huntingtin, or TAR DNA-binding protein 43 (TDP43). It will also be important to determine in which cellular compartment such protein aggregates may be accessible to TLRs, NLRs, and related receptors.

**Alerting the neighbors.** Once neurons have detected cell intrinsic abnormalities, they will alert their neighbors, which include microglia, astrocytes, and possibly other, less abundant cells in the CNS. Injured neurons can release ATP, ions, neurotransmitters, growth factors, cytokines, and other injury response factors, or they can retract inhibitory molecules that keep microglia in a quiet, surveillant state (25). Neurons may release nucleic acids or hsp, which are ligands for TLRs and other receptors, into the extracellular space. For example, hsp60 released from injured neurons can activate microglia via TLR4, causing enhanced nitric oxide production and in turn, neuronal cell death (26). An important mediator alerting other cells to neuronal injury is the DNA-binding protein high mobility group box 1 (HMGB1), which can be released actively from injured cells or passively from dying cells. HMGB1 promoted neurodegeneration in a mouse model of ischemic stroke by activating microglia through ligands for TLRs and other receptors, into the extracellular space. For

### Table 1

<table>
<thead>
<tr>
<th>Molecule</th>
<th>Disease model for:</th>
<th>Genetic manipulation</th>
<th>Outcome</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>C1q</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased neurodegeneration</td>
<td>98</td>
</tr>
<tr>
<td>C1q</td>
<td>Prion disease</td>
<td>Deletion</td>
<td>Prevention of prion disease</td>
<td>99</td>
</tr>
<tr>
<td>C3</td>
<td>Prion disease</td>
<td>Deletion</td>
<td>Delayed neuroinvasion</td>
<td>99</td>
</tr>
<tr>
<td>Caspase-1</td>
<td>ALS</td>
<td>Deletion</td>
<td>Extension of life span, decreased gliosis and neurodegeneration</td>
<td>50</td>
</tr>
<tr>
<td>CCL2</td>
<td>AD</td>
<td>Overexpression in astrocytes</td>
<td>Increased plaque load</td>
<td>100</td>
</tr>
<tr>
<td>CD14</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased plaque load</td>
<td>101</td>
</tr>
<tr>
<td>CD21/CD35</td>
<td>Prion disease</td>
<td>Deletion</td>
<td>Delayed neuroinvasion</td>
<td>99</td>
</tr>
<tr>
<td>CD36</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased neurovascular dysfunction, reduced vascular oxidative stress; no changes in plaque pathology</td>
<td>102</td>
</tr>
<tr>
<td>CD40L</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased plaque load and gliosis</td>
<td>103</td>
</tr>
<tr>
<td>CD40L</td>
<td>AD</td>
<td>Overexpression under metallothionein promoter</td>
<td>Increased plaque load and neurodegeneration</td>
<td>104</td>
</tr>
<tr>
<td>CX3CR1</td>
<td>AD</td>
<td>Deletion</td>
<td>Prevention of neuron loss</td>
<td>45</td>
</tr>
<tr>
<td>CX3CR1</td>
<td>AD</td>
<td>Overexpression</td>
<td>Reduced plaque load</td>
<td>47</td>
</tr>
<tr>
<td>Cxcr2</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased Aβ production</td>
<td>105</td>
</tr>
<tr>
<td>H2-BF/C2</td>
<td>Prion disease</td>
<td>Deletion</td>
<td>Prevention of prion disease</td>
<td>99</td>
</tr>
<tr>
<td>IFNγR1</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased plaque load and gliosis</td>
<td>106</td>
</tr>
<tr>
<td>IL-1β</td>
<td>ALS</td>
<td>Deletion</td>
<td>Extended life span, decreased gliosis and neurodegeneration</td>
<td>50</td>
</tr>
<tr>
<td>PTGER2</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased plaque load and oxidative damage</td>
<td>107</td>
</tr>
<tr>
<td>RAG1</td>
<td>PD</td>
<td>Deletion</td>
<td>Attenuation of neuronal cell death</td>
<td>76</td>
</tr>
<tr>
<td>RAGE</td>
<td>AD</td>
<td>Overexpression in neurons</td>
<td>Accelerated abnormalities in learning and memory</td>
<td>29</td>
</tr>
<tr>
<td>RAGE</td>
<td>AD</td>
<td>Overexpression of dominant-negative form in neurons</td>
<td>Preservation of learning and memory, decreased neurodegeneration</td>
<td>29</td>
</tr>
<tr>
<td>S100B</td>
<td>AD</td>
<td>Overexpression under human S100B promotor</td>
<td>Increased plaque load and gliosis</td>
<td>108</td>
</tr>
<tr>
<td>TCRB</td>
<td>PD</td>
<td>Deletion</td>
<td>Attenuation of neuronal cell death</td>
<td>76</td>
</tr>
<tr>
<td>TGF-β1</td>
<td>AD</td>
<td>Overexpression in astrocytes</td>
<td>Increased amyloidosis in the cerebral vasculature</td>
<td>109</td>
</tr>
<tr>
<td>TGFβR2</td>
<td>AD</td>
<td>Overexpression of dominant-negative form in cells of the myeloid lineage</td>
<td>Decreased plaque load and vascular Aβ deposition</td>
<td>110</td>
</tr>
<tr>
<td>TNFR1</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased Aβ generation and plaque load</td>
<td>36</td>
</tr>
<tr>
<td>TNFR1/2</td>
<td>PD</td>
<td>Deletion</td>
<td>Protection against dopaminergic cell death</td>
<td>35</td>
</tr>
</tbody>
</table>

*aGenetic modification is present in all cells. C3, complement component 3; Crry, complement-receptor related protein; H2-BF, complement factor B; IFNγR1, IFN-γ receptor 1; PTGER2, prostaglandin E2 receptor; RAG1, recombination activating gene 1; S100B, S100 calcium binding protein B.*
Figure 1
Immune-mediated interactions in neurodegeneration at the cellular, tissue, and systemic level. Immune interactions in neurodegeneration can occur within neurons, in the brain tissue, or in the systemic environment through cytokines, neurotransmitters, cell-cell interactions, and peripheral nerves. Abnormalities at any of these levels of organization may modulate neurodegenerative processes in the CNS and may also serve as targets for therapeutic interventions. Some of the factors discussed in more detail in the text are listed in the figure.

CNS-wide genetic manipulations, and it is unclear to what extent the effects are mediated through neurons, glial cells, or both. A notable exception was the directed expression of RAGE, one of several known receptors for Aβ, in neurons. These studies found that RAGE activates NF-κB and MAPK signaling and results in increased synaptic transmission deficits and cognitive impairment (29). Also, expression of a dominant-negative TGF-β type II receptor in neurons impaired signaling in these cells and resulted in a two-fold increase in amyloid accumulation and increased neurodegeneration in mutant APP transgenic mice, a mouse model of familial Alzheimer disease (AD) (30). Because brains from patients with AD have significantly reduced levels of this TGF-β receptor compared with those without disease (30), it is possible that deficiency in neuronal TGF-β signaling may contribute to AD.

Recently an exciting approach to studying the effect of prostaglandins in the brain was described in a stroke model (31). The COX-2 product prostaglandin E2 can bind to four different prostanoid receptors making it difficult to understand how COX-2 inhibitors, which seem to strongly reduce risk of AD (32) and PD (33), work in the brain. Liang and coworkers showed that genetic deletion of prostanoid receptor 4 in neurons worsened stroke injury and decreased cerebral perfusion, pointing to a beneficial effect of this receptor in neurons (31). Clearly, this is a direction in which the field of neurodegeneration has to move in order to gain a more complete understanding of the complex action of immune factors in neuronal distress and degeneration.

Local immune responses to distressed neurons
When neurons send out distress signals, there is a strong response from CNS-resident immune cells such as microglia. What is most puzzling about this response is that rather than helping the injured neurons, more often than not microglia seem to harm or kill neurons. The logic of this action seems counterintuitive, but it may be that it is better for the brain as a whole to get rid of damaged neurons than to have them disrupt otherwise well-functioning circuits. Additionally, there may be more subtle differences in neuronal alarm signals, such that the typical immune response is actually beneficial, but the signal we have been able to study in cell culture systems or through the use of rather crude experimental tools in vivo is lethal. Future studies aiming at the earliest immune responses to neuronal injury using more sophisticated genetic tools and injury models may bring clarity to this conundrum.

Activated microglia can produce a large array of immune mediators including cytokines, chemokines, complement proteins, and nitric oxide, many of which have neurotoxic effects (25). For example, blocking TNF-α attenuates loss of dopaminergic neurons in mouse models of PD (34, 35) and reduces pre-plaque pathology and learning and memory deficits in mouse models of AD (36, 37). Attempts to divide microglia into functional subsets have been hampered by the lack of reliable cellular markers, which may be due in part to the plasticity of these cells and the possibility that they can assume a continuum of different activation states (38). In a surveillant state (previously erroneously referred to as “resting”), microglia sample their environment for danger signals (39) and they seem to maintain this state through a number of close interactions with neurons. If the neuronal ties are severed, microglia become activated and, depending on the context, may support nearby neurons or kill them. Neuronal-microglial ties include CD200 and CD200R, which are expressed on microglia and neurons respectively (40, 41). The best-studied of these cellular interactions is between neuronal fractalkine (CX3CL1) and the microglia-restricted fractalkine receptor (CX3CR1) (42). Mice that are genetically deficient in CX3CR1 display increased neurotoxicity in response to MPTP, overexpressed mutant SOD, and systemic LPS injections (43). Importantly, LPS toxicity was transferred by transplantation of CX3CR1-deficient microglia from LPS-treated mice into brains of wild-type mice, indicating that microglial toxicity is cell autonomous (43). In contrast, lack of CX3CR1 in a model of stroke reduced neuronal cell loss (44) and produced distinct effects in four independent mouse models for AD pathology (Tables 1 and 2). While CX3CR1 deficiency also reduced neuronal loss in mice that express a combination of familial AD mutations in APP, presenilin 1, and t (45), it promoted neuronal t pathology and behavioral deficits in mice that express only mutant t (46). Furthermore, CX3CR1 deficiency reduced amyloid deposition, without reported effects on neurons, in mice expressing human mutant amyloid precursor protein (APP) alone or in combination with presenilin 1 (47).

How the same genetic defect can be both good and bad for neurons is still unclear, but it is likely that microglial function in vivo is shaped by signals from a multitude of receptor pathways. For example, the presence of other triggers of innate inflammatory pathways (e.g., TLR ligands) can induce caspase signaling in microglia, turning them into neurotoxic cells in vivo (48). Interestingly, SOD1 is required for activation of caspase-1, and thus production of mature IL-1, in response to LPS, suggesting a link between innate immune sensors and neurodegeneration. (49). Moreover, expression of an intracellular mutant SOD1 that is linked to genetic forms of amyotrophic lateral sclerosis (ALS) triggered an inflammasome response in microglia, and genetic deletion of caspase-1 and IL-1β extended life span and reduced pathology in mutant SOD1 transgenic mice (Tables 1 and 2; ref. 50).
A key functional state that microglia can assume and that has received much attention recently is the phagocytic state. The clearance of cellular debris or protein aggregates is evidently a positive function of microglia, but phagocytosis—so-called synaptic pruning—or removal of entire neurites could be a double-edged sword. For example, CX3CR1 deficiency results in increased numbers of dendritic spines and more immature synapses during postnatal brain development, suggesting a role for microglia in the removal of synapses and the maintenance of neuronal plasticity (51). After entorhinal cortex lesion, microglia are involved in the removal of denervated neural processes and reorganization of the brain, and again, CX3CR1 deficiency impairs this process (52).

Although these studies suggest that CX3CR1 activation promotes phagocytosis, an independent study found that treatment with CX3CL1 reduces phagocytic engulfment of latex beads in cultured microglia and that mice lacking CX3CR1 show increased uptake of stereotaxically injected Aβ fibrils into microglia (47). Clearly, more work is necessary to sort out these differences.

Microglial phagocytosis is also regulated by the purinergic receptor P2Y6, possibly upon binding of UDP released from injured neurons (53) or by complement proteins C1q and C3, which have been linked to synaptic pruning during development (54). However, the role of microglia in (abnormal) synaptic pruning and removal of other neuronal structures in neurodegeneration is still unclear. Likewise, it is debated how much microglia contribute to the clearance of protein aggregates such as those that are characteristic of AD. While several studies show that activation of microglia can lead to reduced Aβ accumulation and plaque formation in APP transgenic mice (55–58), surprisingly, conditional deletion of microglia in two transgenic mouse models did not change Aβ load or development of neuritic dystrophy (58). Since microglia are clearly capable of phagocytosing Aβ assemblies in vitro or on brain slices ex vivo (59, 60), it is possible that the engulfed peptide impairs intracellular degradation (61, 62) or that Aβ or other age-related factors render microglia ineffective.

In addition to microglia, astrocytes become activated in response to neuronal injury, and they too can produce many classical immune factors with important functions in neuronal survival and death (4). Examples include the chemokine CCL2, TGF-β1, IL-1, and IL-6, and some of their effects on amyloid deposition and neurodegeneration are listed in Tables 1 and 2.

**Immune responses in the systemic environment**

Immune activation in neurons or the CNS can propagate to the organism as a whole and induce alterations in immune status. In turn, immune activation in the periphery can affect CNS cells and neurons (Figure 1). The blood-brain barrier (BBB), which is formed by endothelial cells, astrocytic end feet, and basement membrane, restricts access of larger molecules and cells from the systemic environment to the CNS. Neurons express innate immune receptors that serve as sensors of danger signals. TLRs may recognize endogenous molecules and protein aggregates such as Aβ assemblies, ssRNA, or dsRNA aside from molecules associated with pathogens. While TLR4 and its co-receptor CD14 are present at the cell surface, TLR3, -7, -8, and -9 are located in the ER and endosomal compartments. Activation of TLRs can lead to initiation of autophagy via TRIF/RIP1 and possibly induce the clearance of defective organelles or protein aggregates. Activation of NOD1 or NOD2 may result in NF-κB-mediated transcription of pro-inflammatory genes or initiation of autophagy via Atg16L. The adaptor protein p62 can detect viral proteins in neurons and initiate clearance of viral particles via autophagy involving Atg5 and Atg7. It may also assist in the clearance of abnormal protein aggregates. Atg, autophagy-related protein; NOD, nucleotide binding oligomerization domain-like; p62, nucleoporin 62; TRIF, TIR domain-containing adaptor-inducing IFN-β; RIP1, receptor interacting protein-1.
blood to the CNS (63). In general, systemic immune molecules present in blood or tissues may signal to the brain through at least four different routes: active transport across the BBB, signaling at the luminal side of the BBB and propagation of secondary factors into the CNS, diffusion via circumventricular organs, and signaling via peripheral nerves such as the vagus (64–66). Blood-based immune cells may gain access to the brain through transendothelial migration across the BBB into the perivascular space, via the choroidplexus and the cerebrospinal fluid, or through post-capillary venules at the pial surface (reviewed by ref. 67).

**Immune cells infiltrating the brain.** While infiltrating immune cells are critical in CNS diseases such as multiple sclerosis, the extent to which blood-borne cells have a role in classical neurodegeneration is less well understood. In the APP transgenic mouse model, recruitment of peripheral macrophages to plaques or perivascular spaces appears to have beneficial effects, and several reviews have discussed the clinical relevance and mechanistic underpinnings of these findings (68, 69). The chemokine CCL2 and its receptor CCR2 are strong candidates as mediators of macrophage infiltration into the CNS and have captured the attention of researchers (refs. 70, 71; Tables 1 and 2); a recently described model expressing red fluorescent protein from the CCR2 locus in the brain of mice that expressed it in the CCR2 locus and GFP from the CX3CR1 locus may provide new opportunities to study migration of these cells (72).

Besides myeloid cells, T lymphocytes have been detected in post-mortem tissue of AD, PD, and ALS patients (73–75), and at least in corresponding mouse models, T cells may indeed have a functional role in the pathogenesis of these diseases. Accordingly, the MPTP mouse model of PD recapitulates the infiltration of T cells into the substantia nigra (76), and lack of CD4+ T cells attenuates dopaminergic cell death in a Fas ligand–dependent manner (76), suggesting a detrimental function of T cells in PD. In contrast, mutant SOD transgenic mice models for ALS that are deficient in functional T cells display accelerated disease progression (77). It has been suggested that T cells exert a neuroprotective effect in this model by steering glial cells toward a neuroprotective phenotype (78).

Whether lymphocyte subset distribution in blood may be altered in humans with neurodegeneration remains to be shown. Though intriguing, most studies so far analyzed small numbers of patient samples and have not yet been replicated (reviewed in ref. 2). In order to determine whether immune cell abnormalities could provide information about disease state or severity, large, independent studies need to be conducted.

**Systemic immune mediators associated with neurodegeneration.** Systemic inflammation after major surgery or infection often causes delirium in people, and the probability of this complication increases with age (79) and may result in increased risk of dementia or mortality (80, 81). For example, a study in patients with moderate to severe AD showed that acute systemic inflammation leads to a more rapid decline in cognitive function, possibly due to increased levels of circulating TNF-α (82). In APP transgenic animal models, systemic challenge with LPS — to mimic peripheral inflammation — can result in enhanced amyloid deposition and more severe memory impairment (83). Similar results, with exacerbated disease progression and pathology following systemic LPS administration, have been observed in animal models of ALS and Prion disease (84–86).

### Table 2

<table>
<thead>
<tr>
<th>Molecule</th>
<th>Disease model for:</th>
<th>Genetic manipulationA</th>
<th>Outcome</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Beclin 1</td>
<td>AD</td>
<td>Deletion (haplo-insufficiency)</td>
<td>Increased plaque load and neurodegeneration</td>
<td>16</td>
</tr>
<tr>
<td>C3</td>
<td>AD</td>
<td>Deletion</td>
<td>Increased plaque load and neurodegeneration</td>
<td>111</td>
</tr>
<tr>
<td>CCR1</td>
<td>Prion disease</td>
<td>Deletion</td>
<td>More rapid disease progression</td>
<td>112</td>
</tr>
<tr>
<td>CCR2</td>
<td>AD</td>
<td>Deletion</td>
<td>Increased Aβ levels, decreased survival</td>
<td>71</td>
</tr>
<tr>
<td>CCR2/CCL2</td>
<td>PD</td>
<td>Deletion</td>
<td>Increased plaque load and memory impairment</td>
<td>113</td>
</tr>
<tr>
<td>CCR5</td>
<td>AD</td>
<td>Deletion</td>
<td>No effect</td>
<td>114</td>
</tr>
<tr>
<td>CD45</td>
<td>AD</td>
<td>Deletion</td>
<td>Increased levels of endogenous mouse Aβ42</td>
<td>115</td>
</tr>
<tr>
<td>CX3CR1</td>
<td>ALS</td>
<td>Deletion</td>
<td>Increased plaque load</td>
<td>116</td>
</tr>
<tr>
<td>CX3CR1</td>
<td>PD</td>
<td>Deletion</td>
<td>Increased neuron loss and microglial activation, decreased survival</td>
<td>43</td>
</tr>
<tr>
<td>IFN-γ</td>
<td>AD</td>
<td>AAV-mediated overexpression in brain</td>
<td>Increased neuron loss and microglial activation</td>
<td>43</td>
</tr>
<tr>
<td>IL-1R1</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased plaque load, increased microglial activation</td>
<td>117</td>
</tr>
<tr>
<td>IL-18</td>
<td>ALS</td>
<td>Deletion</td>
<td>No changes in plaque load</td>
<td>118</td>
</tr>
<tr>
<td>IL-1α</td>
<td>ALS</td>
<td>Deletion</td>
<td>No changes in life span</td>
<td>50</td>
</tr>
<tr>
<td>IL-1β</td>
<td>AD</td>
<td>Overexpression of active form in astrocytes</td>
<td>No effect on life span or motor axon degeneration</td>
<td>119</td>
</tr>
<tr>
<td>IL-6</td>
<td>AD</td>
<td>AAV-mediated overexpression in the brain</td>
<td>Decreased plaque load, increased gliosis</td>
<td>120</td>
</tr>
<tr>
<td>MyD88</td>
<td>Prion disease</td>
<td>Deletion</td>
<td>Decreased plaque load, increased gliosis</td>
<td>121</td>
</tr>
<tr>
<td>TGF-β1</td>
<td>AD</td>
<td>Overexpression in astrocytes</td>
<td>No changes in infection or histopathology</td>
<td>122</td>
</tr>
<tr>
<td>TGFBR2</td>
<td>AD</td>
<td>Deletion in neurons</td>
<td>Decreased plaque load and neurodegeneration</td>
<td>123</td>
</tr>
<tr>
<td>TLR2</td>
<td>AD</td>
<td>Deletion</td>
<td>Increased plaque load and dendritic loss</td>
<td>30</td>
</tr>
<tr>
<td>TLR4</td>
<td>AD</td>
<td>Deletion</td>
<td>Increased Aβ levels and accelerated memory impairments</td>
<td>124</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Increased plaque load</td>
<td>125</td>
</tr>
</tbody>
</table>

A: Genetic modification is present in all cells. C3, complement component 3; IL-1R1, IL-1 receptor 1; MyD88, myeloid differentiation primary response gene 88; TGFBR2, TGF-β receptor 2.
A growing number of studies recognize TNF-α as a main perpetra
tor in cognitive dysfunction associated with systemic inflamma	tion or injury. Thus, peripheral injection of LPS into wild-type mice
can lead to prolonged increases in cerebral TNF-α levels and loss of
dopaminergic neurons in the substantia nigra, effects which are ameliorated in mice lacking both TNF-α receptors (87). TNF-α was also shown to be a main mediator of postoperative cognitive dys	function following orthopedic surgery in mice (88). Interestingly, TNF-α may also have a role in familial forms of frontotemporal dementia (FTD) with loss-of-function mutations in progranulin. This anti-inflammatory protein was shown recently to bind to TNF-α receptors and inhibit TNF-α-mediated inflammation in a mouse model of arthritis (89), and it is conceivable that it has simi	lar functions in FTD.

Systemic immunity can be coordinated by neural circuits that operate reflexively. The central components of this immunologi
cal reflex are the afferent and the efferent arc of the vagus nerve,
which sense and inhibit the production of pro-inflammatory cytokines, respectively. Vagus nerve stimulation leads to release of noradrenaline and subsequently acetylcholine in the spleen, which suppresses the generation of pro-inflammatory cytokines such as TNF-α (65). Not much work has been done on the effects of the vagus nerve in neurodegenerative diseases, but clearly this exciting field warrants a close look. Interestingly, vagus nerve stimulation seems to reduce or slow cognitive decline in some patients with AD (90).

Additional evidence linking systemic inflammation with neu	rodegeneration comes from epidemiological studies, which show that prolonged use of nonsteroidal anti-inflammatory drugs (NSAIDs) reduces the risk of developing AD (32) or PD (33) by as much as 50%. These beneficial effects do not extend into the clinical phase of disease, and clinical trials using several different NSAIDs have not resulted in cognitive improvement (91). While it may simply be too late to target inflammation with these drugs once clinical symptoms are apparent, it is equally possible that more specific modulators of inflammatory processes targeting, for instance, prostanooid receptors, would show benefits.

Additional evidence that neurodegeneration is associated with systemic changes in the immune response comes also from screens of peripheral blood cell transcriptomes (92) and of cel	lular signaling proteins (93) in blood from patients with AD and controls. While such studies are still underpowered and pro	teomic studies suffer from technical challenges, our group was recently successful in identifying several overlapping proteins in two studies, which used different detection methods and samples (94). Among these proteins, G-CSF and macrophage-CSF were reduced in the plasma of AD patients in studies conducted by our group and led to reduced amyloidosis and neuroinflamma
tion in APP transgenic mice following systemic administration by two independent groups (95, 96). These results suggest that changes in peripheral immune factors can influence neurodegen	erative processes. Interestingly, cognitive decline with aging may be linked, in part, to increased levels of inflammatory protein in the systemic environment. These factors in the blood of old mice were capable of inhibiting hippocampus-dependent spatial learning and memory, long-term potentiation, and adult hippocampal neurogenesis in young mice, and systemic injection of one of these factors, CCL11/eotaxin, was sufficient to elicit some of these effects (97). Together, these observations provide the intriguing possibility that the peripheral immune environment in neuro
degenerative diseases could be targeted toward the development of novel therapies that regulate CNS function to slow cognitive decline and neurodegeneration.

Conclusion

While it took 36 years to generate the first 1,200 publications con	aining the key words “immunology” and “neurodegeneration,” it took only 9 more years to double that number. The small frac	tion of these studies discussed here make it clear that immune responses are integral to neurodegeneration and that we are just at the beginning of understanding this complex relationship. Exciting new studies linking innate immune receptors to cellular protein homeostasis through phagocytosis and autophagy may provide new insight into immune dysfunction and neurodegen	eration. Rapid progress is also being made in our understanding of microglia, both as protectors of neurons and as their possibly worst enemy. Fortunately, we can make active progress toward the development of useful therapies even as research into the basic mechanisms continues.

Acknowledgments

The authors thank Kira Mosher and Kurt Lucin for insightful comments on the review. Additionally, the authors thank Gregor Bieri for drawing Figure 1. This work was supported by funds from the Department of Veterans Affairs (to T. Wyss-Coray), the NIH Institute on Aging (grant R01 AG030144, to T. Wyss-Coray), a California Institute for Regenerative Medicine Award (to T. Wyss-Coray), and funds from the Alexander von Humboldt Foundation (to E. Czirr).

Address correspondence to: Tony Wyss-Coray, Stanford University School of Medicine, 1201 Welch Road, Rm. P208, MSLS Bldg., Stanford, California 94305-5489, USA. Phone: 650.852.3220; Fax: 650.849.1983; E-mail: twc@stanford.edu.

6. Okun E, Griffioen KJ, Mattson MP. Toll-like recep
11. Jaeger PA, Wyss-Coray T. All-you-can-eat: autophag


88. Murray CL, Skelly DT, Cunningham C. Exacerbation of CNS inflammation and neurodegeneration by systemic LPS treatment is independent of circulating IL-1beta and IL-6. *J Neuroinflammation*. 2010;7:3.

89. Cunningham C, Wilcockson DC, Campion S, Lunnon K, Perry VH. Central and systemic endo


94. Imbimbo BP. An update on the efficacy of non-stero

95. Boyd TD, et al. GM-CSF upregulated in rheuma

96. Sanchez-Ramos J, et al. Granulocyte colony stimu

97. Villeda SA, et al. The ageing systemic milieu nega


100. Reed-Geaghan EJ, Reed QW, Cramer PE, Landreth GE. Deletion of CD14 attenuates Alzheimer’s dis
ease pathology by influencing the brain’s inflammatio

101. Park L, et al. Inhibiting CXCR2 in mice reduces Erk1/2 activity for the cerebrovascular oxidative stress and neuro


105. Bakshi P, Margenthaler E, Reed J, Crawford F, Mullan M. Depletion of CCR2 receptors in mice decreases Erk1/2 activity for the cerebrovascular oxidative stress and neuro


107. Lee YK, et al. CCR5 deficiency induces astrocyte acti

108. Das P, et al. Interferon-1 receptor 1 knockout has no effect on amyloid deposition in Tg2576 mice and does not alter efficiency following Abeta immu

109. Nguyen MD, Julien JP, Rivest S. Induction of proinflammatory molecules in mice with amylo

110. Shaffel SS, Kyrkanides S, Olschawka J, Miller JN, Johnson RE, O’Banion MK. Sustained hippocampal IL-1 beta overexpression mediates chronic neuro


114. Richland RL, Filali M, Prefontaine P, Rivest S. Toll


review series