function mutations in mice and humans results in abnormal DYSF internalization (10, 16). Whether mislocalization of CAV3 and DYSF within the myofiber contributes to CNM disease progression remains to be determined. These data suggest that a common pathway links MTM1, DNM2, BIN1, DYSF, and CAV3 in the biogenesis and maintenance of muscle, specifically at the T-tubule. If DNM2 levels are found to be upregulated in these other forms of myopathy, then targeting DNM2 becomes a common therapeutic strategy for a wider range of muscle disease.

Acknowledgments

This work was supported by NIH grants NS047726, AR053646, and NS070207.

Address correspondence to: Elizabeth M. McNally, University of Chicago, 5841 S. Maryland, MC6088, Chicago, Illinois 60637, USA. Phone: 773.702.2684; Fax: 773.702.2681; E-mail: emcnally@uchicago.edu.


PPARγ in emphysema: blunts the damage and triggers repair?

Neil J. Kelly and Steven D. Shapiro

Division of Pulmonary, Allergy, and Critical Care, Department of Medicine, UPMC and University of Pittsburgh, Pittsburgh, Pennsylvania, USA.

Cigarette smoke is the most common cause of pulmonary emphysema, which results in an irreversible loss of lung structure and function. Th1 and Th17 immune responses have been implicated in emphysema pathogenesis; however, the drivers of emphysema-associated immune dysfunction are not fully understood. In this issue of the JCI, Shan and colleagues found that pereoxisome proliferator–activated receptor γ (PPARγ) is downregulated in APCs isolated from the lungs of emphysematous chronic smokers and mice exposed to cigarette smoke. Furthermore, treatment with a PPARγ agonist prevented emphysema development and appeared to reduce emphysema- associated lung volume expansion in mice exposed to cigarette smoke. Further work will need to be done to evaluate the potential of PPARγ agonists to restore lung capacity in emphysematous patients.

Pulmonary emphysema is a major component of chronic obstructive pulmonary dis- ease (COPD) and involves the loss of alveo- lar units distal to the terminal bronchioles. Even though COPD holds an unenviable position as the world’s fourth-leading cause of death, current medical interven- tions have little to offer beyond symptomatic relief. Meanwhile, the prevalence of COPD is expected to continue to rise as low- and middle-income countries join in the developed world’s tobacco addiction. If we are to avoid this grim projection, we must expand our knowledge of the basic mechanisms behind lung injury and repair before translating these findings into novel therapeutic treatments.

Pulmonary emphysema results from an imbalance between elastases and anti-elastases

It has been more than 50 years since La- rell and Eriksson first identified a deficien- cy in α-1 antitrypsin, the major inhibitor of neutrophil elastase (ELANE), as the cul- prit behind hereditary pulmonary emphy- sema (1). Since the involvement of ELANE in pulmonary emphysema was first report- ed, our understanding of the disease’s

Conflict of interest: The authors have declared that no conflict of interest exists.

Citation for this article: J Clin Invest. 2014; 124(3):978–980. doi:10.1172/JCI74417.
Figure 1
Proposed role of PPARγ in pulmonary emphysema pathogenesis. Smoking leads to the acquisition of a pathogenic phenotype in lung mDCs, characterized by ciglitazone-reversible downregulation of PPARγ and upregulation of SPP1. Pathogenic mDCs then direct the differentiation of Th1 and Th17 helper T lymphocytes, whose cytokines IFN-γ and IL-17A, respectively, trigger the release of the macrophage elastase MMP12 and accompanying alveolar destruction.

pathogenesis has grown around a core paradigm, whereby a protease-antiprotease imbalance results in degradation of the elastic fibers that impart structural stability and compliance on the lung. Beginning with a report by Gross and colleagues (2), subsequent studies revealed that intratracheal instillation of elastases, including ELANE, reproduces the morphologic characteristics of emphysema in rodents. In addition to neutrophils and their associated elastases, the scope of emphysema research has expanded to include macrophages — the predominant inflammatory cells of the lung — and their elastolytic matrix metalloproteinases (MMPs). Mice lacking macrophage elastase (MMP12) exhibit total resistance to smoke-induced emphysema (3), and MMP12 appears to act synergistically with ELANE in vivo, with each elastase degrading the other’s inhibitor. Ultimately, the cooperative actions of ELANE and MMP12 bring about the defining elastolysis and airspace enlargement of emphysema.

The implication that neutrophil- and macrophage-derived elastases influence emphysema pathogenesis has justifiably led to an intense research focus on innate immunity; however, the observed increase in T lymphocytes in the lungs of emphysema patients suggests that the adaptive immune system facilitates the innate response (4). Our understanding of the crossstalk between innate and adaptive immunity in emphysema is far from clear, partly because murine research has thus far shown an inconsistent role for lymphocytes in disease progression. For example, mice with deficient T and B cell responses developed full-blown emphysema in response to cigarette smoke (5), yet CD8-deficient mice are completely resistant to the same insult (6). However, IFN-γ (through the action of the cytokine IP10) and IL-17A, the signature cytokines produced by T helper subsets Th1 and Th17, respectively, have the potential to link the adaptive and innate responses by mediating the release of MMP12 from alveolar macrophages (7, 8). In addition to being elevated in smoke-exposed lungs, IFN-γ and IL-17A are strongly associated with autoimmune disorders (9), and their preponderance supports the idea that auto-reactivity perpetuates the sustained decline of lung function observed in a subset of former smokers (10). In fact, some studies have shown that elastin fragments elicit a recall response in the peripheral T lymphocytes of emphysema patients (8), underscoring a potential link between autoimmunity and human emphysema.

A link between smoking and autoimmunity
But how does smoking lead to autoimmunity? In a series of reports, Shan and colleagues turned to APCs to address this question. In an initial human study (8), Shan et al. found that APCs isolated from emphysematous lungs induce naive CD4+ T cells to differentiate into Th1 and Th17 cells. In mouse models, which do not recapitulate the anti-elastin autoimmunity observed in human patients, Shan and colleagues showed that tobacco smoke exposure causes a phenotypic switch in lung APCs that is characterized by increased expression of the secreted phosphoprotein osteopontin (SPP1). These pathogenic APCs then direct the differentiation of Th1 and Th17 lymphocytes, mimicking the actions of APCs from human smokers (11). Furthermore, mice lacking IL-17A or SPP1 failed to develop airspace enlargement in response to long-term smoke exposure, highlighting the importance of APC-directed T cell differentiation in smoke-induced immune dysfunction.

Increasing evidence indicates that the nuclear hormone receptor PPARγ, best known as a regulator of tissue metabolism and adipogenesis, plays a role in lung development and inflammation. In this issue of the JCI, Shan and colleagues (12) provide further support that PPARγ regulates immune responses in the lung, with an initial observation that PPARγ is down-regulated in APCs isolated from human smokers with emphysema and from mice exposed to tobacco smoke. Additionally, deletion of Pparg in CD11c+ APCs, which include macrophages and myeloid dendritic cells (mDCs), caused spontaneous SPP1-dependent airspace enlargement in mice. Coupled with their previous work, the current study by Shan and colleagues suggests that by dampening PPARγ, tobacco smoke unleashes SPP1 to induce differentiation of Th1 and Th17 lymphocytes. When activated, Th1 and Th17 lymphocytes secrete IFN-γ and IL-17A, respectively, triggering the synthesis and release of destructive elastases from alveolar macrophages (Figure 1).

PPARγ activation: a treatment for emphysema?
Naturally, if functional antagonism of PPARγ by tobacco smoke worsens emphysema, then pharmacologic PPARγ activa-
tion with the thiazolidinediones (TZDs) should abrogate disease progression. As expected, Shan and colleagues (12) demonstrated that the PPARγ agonist ciglitazone prevented murine emphysema despite active smoke exposure. Less definitively, mice exposed to cigarette smoke for 3 months followed by ciglitazone treatment exhibited an apparent restoration of lung volume, suggesting that TZDs are able to reverse the course of emphysema. Should these results be substantiated, TZDs would join a growing list of agents with the potential to reverse rodent emphysema. In the first report of emphysema reversal, Massaro and Massaro (13) demonstrated the initiation of new alveolar growth by retinoic acid (RA) following elastase-induced emphysema in rats. Using a more pertinent smoke exposure model, albeit with a less rigorous physiological assessment, Seimetz and colleagues (14) reported that inhibition of inducible nitric oxide synthase (NOS2) reverses emphysema in mice. In an interesting twist, NO has also been reported to increase the expression of SPP1 (15), suggesting that TZDs and NOS2 inhibitors may have an overlapping therapeutic mechanism.

In light of the findings by Shan and colleagues, as well as the results of a recent study (16) that describes airway inflammation and emphysema onset following expression of a dominant-negative PPARγ in alveolar epithelium, PPARγ appears to play a broad role in checking destructive inflammatory processes in the lung. Is it possible, then, that suppression of inflammation is enough to initiate murine lung regeneration? Perhaps, but it may be that TZDs themselves trigger lung regeneration. In any discussion of tissue regeneration, it is helpful to look to development for insight. Based on studies of children born to mothers who smoked during pregnancy, we know that smoking impairs alveogenesis (17), a process that begins at the end of gestation in humans and occurs entirely postnatally in rodents. While our knowledge is far from complete, it is already clear that PPARγ is a major regulator of this process. In the nascent lung, the loss of alveolar epithelial Pparg impairs development with consequent enlargement of the airspaces (18). Further, in utero nicotine exposure decreases the number of lipid-laden pulmonary lipofibroblasts, which are thought to be important for lung development, by promoting their transdifferentiation into myofibroblasts. However, these transdifferentiated myofibroblasts revert to lipofibroblasts in vitro upon treatment with TZDs (19), while in vivo TZD administration accelerates alveogenesis in rats during the peak period of lung development (20).

It is interesting to speculate that treatment of emphysematous adults with TZDs could initiate a similar program of elastic fiber assembly, which historically has been considered to be a limiting factor in repair of emphysema-associated lung damage. However, repair in rodent models does not necessarily translate to humans. For example, RA treatment, which is efficacious in rats, thus far has shown no potential to reverse emphysema in human trials (21). The greatest concern for repairing damaged lung tissue has less to do with the regenerative capability of stem cells and more to do with the restoration of the extracellular matrix and its elastic fiber cable network. Recapitulating the spatial and temporal regulation of the multiple steps required to regenerate the extracellular matrix in the adult lung is a daunting task; however, a recent case report (22) provides a glimmer of hope that alveogenesis may be possible in the fully developed human lung. As we move forward in the search for a medical cure for pulmonary emphysema, understanding and exploiting the mechanisms behind this phenomenon offers our best chance of success.

Acknowledgments

Neil J. Kelly is supported by NIH/National Heart, Lung, and Blood Institute (NHLBI) grant T32HL094295-04, and Steven D. Shapiro is supported by NIH/NHLBI grant 5P01HL103455-03.

Address correspondence to: Steven D. Shapiro, Office of the CMSO, UPMC, U.S. Steel Tower, 600 Grant Street, Suite 6250, Pittsburgh, Pennsylvania 15219, USA. Phone: 412-605-3995; Fax: 412-647.4801; E-mail: shapirosd@upmc.edu.