M-current preservation contributes to anticonvulsant effects of valproic acid

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Valproic acid (VPA) has been widely used for decades to treat epilepsy; however, its mechanism of action remains poorly understood. Here, we report that the anticonvulsant effects of nonacute VPA treatment involve preservation of the M-current, a low-threshold noninactivating potassium current, during seizures. In a wide variety of neurons, activation of Gq-coupled receptors, such as the m1 muscarinic acetylcholine receptor, suppresses the M-current and induces hyperexcitability. We demonstrated that VPA treatment disrupts muscarinic suppression of the M-current and prevents resultant agonist-induced neuronal hyperexcitability. We also determined that VPA treatment interferes with M-channel signaling by inhibiting palmitoylation of a signaling scaffold protein, AKAP79/150, in cultured neurons. In a kainate-induced murine seizure model, administration of a dose of an M-channel inhibitor that did not affect kainate-induced seizure transiently eliminated the anticonvulsant effects of VPA. Retigabine, an M-channel opener that does not open receptor-suppressed M-channels, provided anticonvulsant effects only when administered prior to seizure induction in control animals. In contrast, treatment of VPA-treated mice with retigabine induced anticonvulsant effects even when administered after seizure induction. Together, these results suggest that receptor-induced M-current suppression plays a role in the pathophysiology of seizures and that preservation of the M-current during seizures has potential as an effective therapeutic strategy.

Introduction
Valproic acid (VPA) is used to treat epilepsy and bipolar disorder (1). VPA is known to have multiple pharmacological actions, such as prolongation of sodium-channel inactivation (2), elevation of GABA content in the brain (3), and inhibition of HDAC activity (4). However, underlying mechanisms for the anticonvulsant effect of VPA are still unclear.

The M-current is a low-threshold noninactivating voltage-gated potassium current generated by tetrameric channels composed of various combinations of subunits encoded by the KCNQ gene family (KCNQ2, 3, 4, and 5) (5, 6). A recent study showed that the Kv7.2/KCNQ2 is the dominant subunit in pyramidal neurons in the hippocampus (7). Various neurotransmitters that activate Gq-coupled receptors, including acetylcholine, suppress the M-current and induce neuronal hyperexcitability. Accordingly, loss-of-function mutations in KCNQ genes are known to cause epilepsy and encephalopathy (5, 8), and M-channel openers are used to treat epilepsy (9, 10).

Intensive studies have been conducted to elucidate the molecular pathway mediating M-current suppression induced by neurotransmitters, especially by the muscarinic action of acetylcholine (5, 6, 11, 12). These studies revealed that multiple signaling pathways converge at the M-channel complex (12–15). One mechanism is through the depletion of phosphatidylinositol 4,5-bisphosphate (PIP2), an essential cofactor for ion conduction. In this mechanism, activation of Gq-coupled receptors activates phospholipase C (PLC), which consumes PIP2 leading to its loss from KCNQ subunits. PIP2 deficient M-channel cannot conduct potassium ions, since PIP2 is required to prevent collapse of the ion-conducting pore (16). Another mechanism is mediated by PKC phosphorylation of the KCNQ2 subunit, which reduces PIP2 efficacy. Upon activation of m1 muscarinic receptor, PKC bound to the KCNQ2 subunit via A-kinase anchoring protein (AKAP79/150) phosphorylates the KCNQ2 subunit. Phosphorylation of KCNQ2 dissociates calmodulin (CaM), thereby destabilizing KCNQ2-PIP2 interaction and leading to the shutdown of M-channel activity (15).

AKAP79/150 (human AKAP79/rodent AKAP150) is a critical signaling-scaffold protein for the M-channel, which tethers several signaling enzymes, facilitating local signaling events. AKAP79/150 is known to be important not only for the M-channel, but also for glutamate receptors and other ion channels (14, 17, 18). AKAP150 (Akap5) gene knockout mice show impaired learning and resistance to pilocarpine-induced seizures (19). In addition, recent studies have shown that palmitoylation of AKAP79/150 plays essential roles in cellular signaling (20, 21). Palmitoylation is a reversible posttranslational modification, which adds predominantly palmitic acid to selective cysteine residues (22). Palmitoylation is required for neural plasticity, as it targets relevant proteins to lipid rafts, which often creates signaling hot spots (23). Furthermore, changes in neuronal activities induce depalmitoylation (21, 24). Therefore, palmitoylation is gaining greater attention due to its dynamic regulation during signaling events.

We demonstrate that VPA treatment disrupted muscarinic acetylcholine receptor–induced M-current suppression, preventing receptor-induced hyperexcitability in neurons. Consistently, pharmacological blockade of M-current diminished the anticonvulsant effect of VPA but not that of diazepam. We show that reti-
Rat superior cervical ganglion (SCG) neurons were used in the initial assessment of VPA effects on the M-current, since M-current regulation has been extensively characterized in this neuronal type and neurons in the CNS are known to use the same regulatory mechanism (6, 27). SCG neurons were pretreated with 500 μM VPA for 3 days, and M-current response to a muscarinic agonist, 0.1 μM oxotremorine-M (oxo-M), was measured. VPA-treated neurons showed reduced muscarinic suppression of the M-current (Figure 1A). In contrast, pretreatment with a structurally similar short-chain fatty acid, valeric acid (500 μM), did not affect muscarinic suppression of the M-current (Figure 1B). In addition, other antiepileptic compounds, 100 μM vigabatrin or 100 μM phenytoin, did not change muscarinic suppression of the M-current (Figure 1B).

We next tested whether VPA also affects M-current–mediated neuronal excitability in central neurons using hippocampal neurons. Cultured rat hippocampal neurons were pretreated with 500 μM VPA or 500 μM valeric acid for 3 days. Current clamp recording was used to measure neuronal excitability of these neurons. In control nontreated neurons, 50 pA current injections for 500 μM gabine, an M-channel opener, had anticonvulsant effect only when administered before induction of seizure in control mice. However, in VPA-treated mice, anticonvulsant effect of retigabine was maintained even when administered after seizure induction. We determined that disruption of receptor-induced M-current suppression was caused by reduced palmitoylation of AKAP79/150 by VPA treatment. These findings suggest that M-current suppression plays a role in the pathology of seizures and that disruption of M-current suppression is an integral part of the anticonvulsant mechanism of nonacute VPA treatment.

Results
VPA prevents muscarinic agonist–induced neuronal hyperexcitability by disrupting M-current suppression. VPA has been used to treat epilepsy and bipolar disorder for several decades (25). Accumulating genomic analyses indicate that these 2 neurological disorders are also linked with KCNQ gene mutations (5, 26). The similarity between the 2 motivated us to investigate whether VPA affects M-channel regulation.

Rat superior cervical ganglion (SCG) neurons were used in the initial assessment of VPA effects on the M-current, since M-current regulation has been extensively characterized in this neuronal type and neurons in the CNS are known to use the same regulatory mechanism (6, 27). SCG neurons were pretreated with 500 μM VPA but not by 500 μM valeric acid, 100 μM vigabatrin, or 100 μM phenytoin. **P < 0.01 using Kruskal–Wallis one-way ANOVA followed by Dunn’s multiple comparisons test. Numbers above bars represent n values. (C) Current clamp traces showing action potential firing in response to 50 pA current injection. Traces from VPA-nontreated (top), 500 μM VPA–treated (middle), and 500 μM valeric acid–treated (bottom) hippocampal neurons are shown. Control, presence of 1 μM oxo-M, or presence of 20 μM XE991. (D and E) Input-excitation relationships from hippocampal neurons before (open circle) and after application of oxo-M (filled circle) from nontreated neurons (D) or 500 μM VPA–treated neurons (E). (F) Summary of neural firing by 50 pA current injection shown in C–E. **P < 0.01 and ***P < 0.001 using repeated measures one-way ANOVA followed by Tukey’s multiple comparisons test. Error bars show ± SEM.
XE991, a voltage-dependent inhibitor of the M-channel (28), should diminish anticonvulsant effects of VPA. XE991 is a unique channel blocker that does not inhibit M-channel when the membrane potential is below –65 mV (28), which is close to the resting membrane potential of neurons. Hence, it should have minor effects on neurons showing sparse firing. Indeed, administration of XE991 (2 mg/kg) did not affect basic motor behaviors in mice (Supplemental Figure 1; supplemental material available online with this article; doi:10.1172/JCI79727DS1). However, this dose of XE991 transiently exacerbated kainate-induced seizure in VPA-treated mice reaching stage-5 seizures (Figure 2C). In contrast, 2 mg/kg XE991 did not affect either the anticonvulsant effect of diazepam (Figure 2D) or stage-3 seizures in VPA-nontreated mice induced by 25 mg/kg kainate (Figure 2E). These results suggest that VPA shows anticonvulsant effect through preservation of the M-current. Inversely, since control kainate-induced seizures and diazepam-treated seizures were not affected by XE991, these results also suggest that M-current is suppressed during seizures.

One caveat of the above experiments is a possibility that observed effects of XE991 were derived from off-target inhibition of other potassium channels, such as Kv4 channels (29). To further test whether disruption of M-current suppression contributes to anticonvulsant effects of VPA, we performed a second set of experiments using retigabine as a probe, which does not affect Kv4 channels (30). Retigabine is a recently approved antiepileptic drug that opens the M-channel by shifting activation voltage close to the resting membrane potential (30). In addition, retigabine has been demonstrated to be ineffective at opening M-channels under receptor-induced suppression (30). We first tested whether VPA treatment modulates the pharmacological action of retigabine in vitro using tran-
retigabine even when applied together with 10 μM oxo-M, KCNQ2 current was decreased as reported (ref. 30 and Figure 3, A and B). In contrast, in 500 μM VPA-treated CHO hm1 cells, KCNQ2 currents showed augmentation by retigabine even in the presence of 10 μM oxo-M (Figure 3, A and B). Similar results were obtained from KCNQ2/3 heteromeric channels (Figure 3C), which are considered to be a common form of the endogenous M-channels. Retigabine application augmented KCNQ2/3 current. However, the current amplitude was reduced when applied together with oxo-M. Notably, KCNQ2/3 currents from VPA-treated cells showed augmentation by retigabine even in the presence of oxo-M. These results were consistent with our observation that VPA treatment preserves KCNQ current when m1 muscarinic receptor is activated. We then examined anticonvulsant effects of retigabine in kainate-induced seizures in mice. When retigabine was administered 5 minutes before application of kainate, it showed anticonvulsant effect on kainate-induced seizures, as expected (Figure 3D). However, it did not show any anticonvulsant effect when applied 20 minutes after kainate injection (Figure 3D), which suggests retigabine-sensitive M-channels are reduced after induction of seizures.

To test whether VPA modulates retigabine efficacy, we used an increased kainate dose (35 mg/kg), which induced stage-6 seizures in VPA-treated mice (5/6 mice) (Figure 3E). Retigabine administration 5 minutes before kainate injection reduced seizure severity similarly as observed in VPA-nontreated mice. Importantly, retigabine administration 20 minutes after kainate also reduced severity of seizures in VPA-treated mice (Figure 3E). These results further support that the anticonvulsant effect of VPA involves preservation of M-current during seizures.

Pharmacological modulation of behavioral seizure severity in the above experiments may merely reflect changes at the motor cortex without altering generalized seizure activity in other brain regions. Therefore, we evaluated accumulative neural hyperactivity in the brain during kainate-induced seizures by quantifying c-FOS expression in the hippocampus, a primary target brain region for administered kainate (32). We observed strong expression of c-FOS in the CA1 region and the dentate gyrus of the hippocampus 60 minutes after kainate injection, which would correspond to neuronal activity during status epilepticus in control mice, as reported previously (Figure 4 and refs. 33, 34). Kainate-induced c-FOS expression was reduced in VPA-treated mice. However, when XE991 was administered to VPA-treated mice, c-FOS expression showed equivalent induction, as compared with kainate alone (Figure 4). These results suggest that neuronal activity in the hippocampus correlates with behavioral seizures in our experimental conditions.

**VPA interfered with the PKC-mediated pathway of muscarinic suppression of KCNQ2 current.** Since VPA effects could be reconstituted in CHO hm1 cells expressing homomeric KCNQ2 channels (Figure 3, A and B), we used this heterologous expression system to further characterize the molecular process that is disrupted by VPA treatment. Overnight treatment with 500 μM VPA reduced muscarinic suppression of KCNQ2 current (Figure 5A) similar to the effects observed in SCG neurons. Dose-response experiments...
showed that VPA disrupted muscarinic suppression of KCNQ2 current with EC50 of 213 ± 9 μM (n = 7) (Figure 5B). Importantly, short exposure (5 minutes) to 500 μM VPA did not alter muscarinic suppression of KCNQ2 current (Figure 5C). In addition, pretreatment with lithium chloride (1 mM) or HDAC inhibitors (1 mM sodium butyrate or 2 μM SAHA) did not affect muscarinic suppression of the KCNQ2 current (relative currents with oxo-M were 26.3 ± 4.6% control [Figure 5C, n = 8], 23.5 ± 5.6% lithium [n = 6], 33.9 ± 2.7% SAHA [n = 9], 27.0 ± 5.8% butyrate [n = 9], P > 0.05).

We have previously demonstrated that PKC-mediated phosphorylation of serine 541 is important for suppression of the M-current (15). Therefore, we tested a mutant channel that lacks the phosphorylation acceptor residue, an alanine substitution of serine 541 in the KCNQ2 subunit: KCNQ2 S541A. As we reported previously (15), KCNQ2 S541A exhibited smaller responses to oxo-M compared with the WT KCNQ2 current (Figure 5C). Importantly, VPA treatment did not further modify oxo-M responses of KCNQ2 S541A current (Figure 5C), which suggests that the KCNQ2 S541A mutation occluded VPA effects.

Since PKC-mediated phosphorylation of KCNQ2 S541 triggers CaM dissociation from KCNQ2 channel (15), we measured KCNQ2-CaM dissociation by total internal reflection fluorescence Förster resonance energy transfer analysis (TIRF-FRET) using KCNQ2-mCitrine (KCNQ2-mCit) and CaM-mCerulean (CaM-mCer). Basal FRET efficiency showed positive FRET signals between KCNQ2-mCit and CaM-mCerulean, indicating association of these proteins, as we previously reported (15). These basal FRET efficiencies were not altered by VPA treatment (0.18 ± 0.02 [n = 28] for VPA nontreated cells, 0.16 ± 0.01 [n = 34] for VPA treated cells, P = 0.327, t test). Application of oxo-M reduced FRET signal in control cells, which corresponds to dissociation of CaM, as described (ref. 15 and Figure 5D). However, VPA treatment abolished oxo-M induced decrease in FRET efficiency (Figure 5D). These results support the hypothesis that VPA treatment disrupts PKC-dependent KCNQ2 regulation.

Next, we asked whether this effect was due to the general inhibition of PKC activation or focal inhibition within the KCNQ2 channel complex. To address this question, we measured oxo-M induced cellular PKC activity using a cytosolic version of FRET based PKC kinase activity reporter, cytCKAR (35). Cellular PKC activity induced by muscarinic agonist was not altered by VPA treatment (Figure 5E), suggesting that interference of PKC signaling occurred only within the KCNQ2 channel complex.

Another key pathway of muscarinic suppression of the M-current is through PIP2 depletion. Therefore, we examined whether VPA treatment altered PIP2 depletion after muscarinic stimulation using CFP-PH, a CFP fusion protein with the PH domain of PLCδ (36). In control CHO hm1 cells, PIP2 depletion — triggered by 3 μM oxo-M — induced translocation of CFP-PH into the cytoplasm (Figure 5F). VPA treatment did not evoke any changes in oxo-M-induced translocation of CFP-PH (Figure 5F). These results suggest that VPA selectively disrupted local PKC-mediated phosphorylation of the KCNQ2 channel without interfering with global PKC activation or phosphatidylinositol turnover.

VPA inhibited palmitoylation of AKAP150 and other neuronal proteins. AKAP79/150 anchors PKC to the KCNQ2 subunit and is critical for PKC-mediated KCNQ2 S541 phosphorylation (14, 18). Therefore, we next focused on AKAP79/150-dependent mechanisms. We first suspected that VPA decreased AKAP150 expression, since similar disruption of PKC-mediated KCNQ2 phosph...
phorylation was observed in AKAP150-deficient cells (18, 19). However, VPA treatment did not change AKAP150 protein expression or protein interaction profiles of AKAP150 binding proteins (Supplemental Figure 2).

Accumulating evidence suggests that palmitoylation of AKAP150 is important for AKAP150-mediated signaling (20, 21). Therefore, we measured palmitoylation of AKAP150 by acyl-biotin exchange (ABE) analyses (37) in cultured hippocampal neurons. This technique uses hydroxylamine (NH2OH) treatment to cleave palmitic acid from palmitoylated cysteine residues, followed by biotin labeling of newly exposed cysteine thiols. Therefore, biotin labeling of NH2OH-treated samples indicates specific detection of palmitoylation. We observed that 1 mM VPA treatment reduced palmitoylation of AKAP150 (Figure 6A, \( P = 0.038, n = 3 \), paired \( t \) test). Interestingly, VPA treatment did not drastically alter the overall palmitoylation patterns in hippocampal neuron lysates (Figure 6B), suggesting some degree of selectivity for VPA-sensitive palmitoylation.

We next examined the effects of in vivo administration of VPA on AKAP150 palmitoylation. After administering VPA (i.p., 250 mg/kg/day × 4 days) to mice, cerebrums were collected and used for ABE analyses combined with immunoblots. Consistent with our results from cultured neurons, VPA treatment reduced palmitoylation of AKAP150 (Figure 6C). We also found that VPA suppressed palmitoylation of PSD95, flotillin-1, Fyn, and GRIP, but not EAAT2, confirming some degree of selectivity for VPA-sensitive palmitoylation (Figure 6C).

**Palmitoylation of AKAP150 is required for muscarinic regulation of the M-current.** One remaining question was whether palmitoylation of AKAP79/150 is involved in muscarinic suppression of the M-current. We addressed this question by using AKAP150-silenced SCG neurons. Endogenous AKAP150 was silenced in rat SCG neurons using a silencing plasmid, pSAKAP150i (18). In these AKAP150-silenced neurons, various versions of the human orthologue, AKAP79 — which are resistant to rat AKAP150 RNAi — were expressed to examine whether they can restore AKAP79/150 signaling. AKAP150 silenced SCG neurons showed attenuated muscarinic responses, as we reported previously (ref. 18 and Figure 7). The muscarinic response was rescued by expression of WT AKAP79, as expected (Figure 7). Two cysteine residues in AKAP79, C36 and C129, have been identified as the palmitoylation sites (20, 21). Expression of the double serine-substituted mutant for the palmitoylation sites, AKAP79(C36S/C129S), could not restore the oxo-M response of KCNQ2 current (Figure 7).

It has been shown that myristoylation can mimic constitutive palmitoylation (38). Therefore, a myristoylation consensus site was added to the N-terminus of AKAP79(C36S/C129S), Myr-AKAP79(C36S/C129S). Expression of Myr-AKAP79(C36S/C129S)
C129S) restored muscarinic suppression of the M-current (Figure 7B). These results confirmed that palmitoylation of AKAP150 is critical for AKAP150-mediated regulation of the KCNQ2 channel.

AKAP150 mutants carrying a myristoylation site were resistant to VPA treatment. Since distinct enzymes regulate myristoylation and palmitoylation, we wondered if the myristoylated AKAP150–restored M-channel pathway becomes resistant to VPA treatment. To test this, we overexpressed rat Myr-AKAP150(C36S/C123S), which corresponds to human Myr-AKAP79(C36S/C129S), in SCG neurons. Then, we treated neurons with 2 mM VPA for 3 days and tested for muscarinic responses to 1 μM oxo-M (Figure 8). In control experiments with overexpression of WT AKAP150, VPA pretreatment attenuated the muscarinic response of the M-current, as expected (Figure 8A). In contrast, muscarinic responses in Myr-AKAP150(C36S/C123S)–expressing SCG neurons became insensitive to 2 mM VPA (Figure 8, B and C). These results further support that VPA disrupts receptor–induced M-current suppression by interfering with palmitoylation of AKAP150.

Discussion
In the present study, we report that disruption of M-current suppression during seizure contributes to the nonacute anticonvulsant action of VPA. Conversely, we report that M-current suppression is involved in the pathophysiology of seizures. VPA has been demonstrated to have many pharmacological effects. However, some known effects require high concentrations of VPA. Therefore, it is important to evaluate effects at the concentration of VPA that is within the therapeutic range in humans (28–700 μM) (1). We were able to demonstrate that VPA in this concentration range prevented muscarinic receptor–induced suppression of the M-current, as well as M-current–dependent hyperexcitability in primary neurons.

Previously, we showed that AKAP79/150 is important for receptor–induced suppression of the M-current (14, 18). In the present study, we demonstrated that palmitoylation of AKAP79/150 is critical for M-current regulation and that VPA disrupts AKAP79/150–mediated M-current regulation by interfering with palmitoylation of AKAP79/150. Palmitoylation of AKAP79/150 has been shown to stabilize its membrane localization (20), which may be required during the rearrangement of the multiprotein KCNQ2-channel complex that occurs during muscarinic suppression of the M-current (15).

We also demonstrated that VPA suppressed palmitoylation of a minor fraction of neuronal proteins, including several synaptic proteins. Interestingly, kainate-induced seizures have been demonstrated to increase palmitoylation of PSD-95 and SNAP25, as well as AKAP150, which promotes synaptic potentiation (21). Therefore, in addition to VPA effects on the M-channel, inhibited palmitoylation of synaptic proteins may also contribute to the anticonvulsant action of VPA. Palmitoylation is mediated by palmitoyl acyltransferases, which consist of over 20 subtypes of DHHC (Asp-His-His-Cys) proteins (23). GRIP1 has been shown to be palmitoylated by DHHC5 and DHHC8, while Fyn is palmitoylated by other DHHC enzymes (38). VPA suppressed palmitoylation of GRIP1 and Fyn to a similar degree, which indicates that VPA affects multiple DHHC subtypes. Therefore, VPA should be considered a palmitoyl acyltransferase inhibitor with moderate selectivity.

Although KCNQ subunit mutations have been linked to epilepsy, modulation of the WT M-channel has never been implicated in the pathophysiology of seizures. Our results suggest that M-cur-
current is suppressed during seizures. XE991 is a voltage-dependent inhibitor that is effective only when the membrane potential is above ~65 mV (28). Therefore, M-channels become sensitive to XE991 only when neurons are depolarized by neuronal activities, such as high-frequency firing. Indeed, we showed that XE991 aggravated kainate-induced seizures in VPA-treated mice. In contrast, XE991 administration did not show any effects in control kainate-induced seizures. This can happen if the M-current is already suppressed by seizure activity in affected regions, leaving few M-channels to be inhibited. Further support came from retigabine - already suppressed by seizure activity in affected regions, leaving few M-channels to be inhibited. Further support came from retigabine, which, in turn, facilitates propagation of ictal activities. Indeed, we showed that XE991 aggravated kainate-induced seizures in VPA-treated mice. In contrast, XE991 administration did not show any effects in control kainate-induced seizures. This can happen if the M-current is already suppressed by seizure activity in affected regions, leaving few M-channels to be inhibited. Further support came from retigabine experiments. Retigabine has been shown to activate M-channels by shifting the activation voltage below the resting membrane potential. However, retigabine-activated M-current can undergo receptor-induced suppression (30). Our experiments confirmed that retigabine could augment KCNQ2 or KCNQ2/3 currents when applied without a muscarinic agonist, but when applied together with a muscarinic agonist, muscarinic suppression overwhelmed retigabine-induced augmentation. However, when cells were pretreated with VPA, retigabine could augment KCNQ2 and KCNQ2/3 currents even in the presence of muscarinic agonist. In addition, efficacy of retigabine in kainate-induced seizures also supports this view. In nontreated mice, retigabine could suppress kainate-induced seizures only when applied before induction of seizures. In contrast, in VPA-treated mice, retigabine did not lose efficacy even when administered after seizure induction. These results suggest that administration of retigabine to VPA-treated patients would be a powerful procedure to cease status epilepticus. However, since M-current suppression has also been implicated in cognitive function, careful evaluation of long-term effects of this combination therapy would need to be performed. Together, these results suggest that M-current is suppressed during seizures and that VPA treatment preserves M-current by disrupting its receptor-induced suppression.

One scenario for M-current suppression during seizure is that seizures provoke excess neurotransmitter release that activates Gq-coupled receptors, thereby suppressing the M-current. Such seizure-induced M-current suppression increases the excitability of relevant neurons, which, in turn, facilitates propagation of ictal activities. Indeed, microdialysis studies have shown that several neurotransmitters — including acetylcholine (39, 40), serotonin (39, 41), noradrenaline (41, 42), and glutamate (43, 44) — are elevated during seizures. In addition, corresponding Gq-coupled receptors for these transmitters have been identified in hippocampal neurons: m1 acetylcholine receptor (45, 46), serotonin 5-HT2 receptor (47), α1 adrenergic receptor (47), and group 1 metabotropic glutamate receptor (48). However, noradrenaline and serotonin are considered to have anticonvulsant actions through distinct receptors (49, 50). Further studies are required to identify responsible neurotransmitters that suppress the M-current during seizures.

It has been reported that some VPA metabolites, which are primarily generated in the liver (51), have more potent anticonvulsant action (52). In our case, antihyperexcitation effects were reproduced in cultured cells. Therefore, we believe that VPA is the primary compound for the inhibition of palmitoylation in our conditions. However, we do not rule out the possibility that active VPA metabolites could be more potent palmitoylation inhibitors.

In summary, M-current suppression contributes to the pathophysiology of seizures, and preserved M-current during seizures is involved in the anticonvulsant effect of VPA, which is mediated by interfering with palmitoylation of AKAP79/150. These findings would provide insights and biochemical targets for developing better antiepileptic treatments.

**Methods**

**Antibodies.** Antibodies used for quantitative immunoblotting were as follows: Anti–V5 epitope monoclonal antibody (catalog R960, Invitrogen); anti–flag M2 antibody, HA monoclonal antibody, and HA rabbit polyclonal antibody (catalogos P3165, H3665, H6908, Sigma-Aldrich); PSD-95 (catalog 041066, Millipore); flotillin-1, lyn, GRIP1, and EAAT2 (catalogos 041066, 610821, 611318, 61654, BD Biosciences); c-FOS (catalog PC38; Calbiochem); mouse monoclonal AKAP150.
EGTA, 20 mM HEPES [pH 7.4], 1% Triton X-100, and complete cell lysis in 500 μl HSE buffer (150 mM NaCl, 5 mM EDTA, 5 mM MgCl₂, 3 mM EGTA, 1 mM CaCl₂, 4 mM ATP, 0.1 mM GTP, and 10 mM HEPES [pH 7.4]). SCG neurons were isolated from 14- to 19-day-old rats and were grown in alpha-modified Eagle medium with 5% FBS and 500 μg/ml G418 (Gambro). CHO hm1 (14) cells were grown in DMEM with 10% FBS. CHO hm1 cells were grown in alpha-modified Eagle medium with 5% FBS and 500 μg/ml G418 sulfate. SCG neurons were isolated from 14- to 19-day-old rats and cultured as described previously (14, 15, 18). Rat neonatal hippocampal culture was prepared as described (18). VPA solution was prepared daily from powder and was added to the medium at 500 μM. SCG neurons were isolated from 14- to 19-day-old rats and cultured as described previously (14, 18). VPA treatment (2 mM) attenuated responses to 1 μM oxo-M. Black box indicates presence of oxo-M. (B) Current traces (top) and pooled results (bottom) showing oxo-M responses of the M-current from SCG neurons overexpressing the myristoylation site–attached AKAP150(C36S/C123S), Myr-AK(dC/S). VPA treatment (2 mM) did not affect muscarinic response in Myr-AK(dC/S)–overexpressed neurons. (C) Histogram summarizing results shown in A and B. **P < 0.01 using unpaired t test with Welch's correction. Error bars show ± SEM. Numbers above bars represent n.

**Figure 8. M-current suppression became resistant to VPA treatment when a myristoylated AKAP150 mutant was overexpressed in SCG neurons. (A) Current traces (top) and pooled results (bottom) showing oxo-M responses of the M-current in rat SCG neurons overexpressing WT AKAP150. VPA treatment (2 mM) attenuated responses to 1 μM oxo-M. Black box indicates presence of oxo-M. (B) Current traces (top) and pooled results (bottom) showing oxo-M responses of the M-current from SCG neurons overexpressing the myristoylation site–attached AKAP150(C36S/C123S), Myr-AK(dC/S). VPA treatment (2 mM) did not affect muscarinic response in Myr-AK(dC/S)–overexpressed neurons. (C) Histogram summarizing results shown in A and B. **P < 0.01 using unpaired t test with Welch's correction. Error bars show ± SEM. Numbers above bars represent n.
Palmitoylation assay. ABE assay was performed following the protocol described by Wan et al. (37) with minor modifications. Briefly, cell and brain lysate were prepared in the presence of 10 mM N-ethylmaleimide (NEM) followed by denaturation with chloroform/methanol precipitation. Treated samples were further incubated overnight with 10 mM NEM. After extensive wash by chloroform/methanol precipitation, samples were subjected to NH₂OH treatment. After each chloroform/methanol precipitation, protein pellets were solubilized in aqueous buffers by sonication. After wash, HPDP-biotin (Pierce Biotechnology) was used for sulphydryl biotinylation. Biotin-exchanged samples were used for IP by mouse anti-AKAP150 antibody or affinity purification by NeutrAvidin resin (Pierce Biotechnology). Purified proteins were eluted by boiling in nonreducing SDS-PAGE loading buffer for detection with HRP-conjugated NeutrAvidin, or with diithiothreitol for detection with antibodies.

Kainate-induced seizure. For in vivo administration of VPA, adult C57BL/6 mice of both sexes (3–5 months old, The Jackson Laboratory) were administered (i.p.) either freshly prepared sodium VPA (250 mg/kg) or physiological saline solution twice daily for 4 days unless stated otherwise. For kainate-induced seizure experiments, 30 mg/kg kainate was injected (s.c.) 6 hours after the last VPA administration. One hour after kainate administration, mice were injected (i.p.) either with 2 mg/kg XE991 in Dulbecco’s PBS (DPBS) or DPBS alone. Mice were video-taped for the subsequent 4 hours after kainate injection, and behavior was scored by treatment-blind observers every 10 minutes according to a modified Racine scale with the following criteria: (i) immobility; (ii) rigidity; (iii) automatics with scratching, head bobbing, and circling; (iv) intermittent rearing and falling; (v) continuous rearing and falling; and (vi) tonic-clonic whole-body convulsions and rapid jumping. Animals that died during the experiments were assigned stage 6 thereafter.

c-FOS IHC. 1 hour after injection of kainate (30 mg/kg), mice were deeply anesthetized with isoflurane and perfused with 4% paraformaldehyde in PBS (pH 7.4). Brains were then removed and postfixed with 4% paraformaldehyde in PBS (pH 7.4) overnight at 4°C. Brains were sectioned into coronal slices (30 μm) using a vibratome (VT-1200, Leica Microsystems). Free-floating sections were washed with PBS in 24-well dishes and then incubated with PBS containing 0.3% hydrogen peroxide and 10% methanol for 10 minutes. After 3 washes with PBS, sections were incubated with a blocking buffer containing 0.5% Triton X-100 and 1% normal goat serum in PBS. Sections were then incubated with primary antibody mixture containing rabbit anti-c-FOS antibody (1:1000 dilution) in the blocking solution for 48 hours at 4°C. After 2 washes with PBS followed by 2 washes with Tris-buffered saline (TBS), sections were incubated with Cy3-conjugated anti-rabbit secondary antibody in TBS containing 0.5% Triton X-100 and 1% normal goat serum for 2 hours at room temperature. After wash, sections were mounted onto glass slides using VECTASHIELD with DAPI (Vector Laboratories). Immunofluorescent images were acquired using a fluorescent light microscope (DM4000B, Leica Microsystems) equipped with a CCD camera (Optronics MicroFire, OPTMIF). Areas around the dentate gyrus, CA1 and CA3, were selected, and the average fluorescence intensity within each corresponding region of interest from DAPI and c-FOS staining images was quantified by observers blind to treatments using MetaMorph (Molecular Devices). Nucleus-free regions in the stratum oriens were used as a background for DAPI and c-FOS staining.

Statistics. Data are presented as mean ± SEM. Statistical analyses were performed using GraphPad Prism. For comparing 2 groups, either unpaired or paired t tests were used with Welch’s correction. For experiments with more than 3 groups, one-way ANOVA or Kruskal-Wallis tests were used as follows. If variances were the same among groups — as evaluated by Bartlett’s test and Brown-Forsythe test — one-way ANOVA, followed by Dunnett’s multiple comparisons test, was used. If variances were different among samples, then Kruskal-Wallis test, followed by Dunn’s multiple comparisons test, was used. When same cells received multiple treatments, then repeated-measures one-way ANOVA, followed by Tukey’s multiple comparisons test, was used. P values of less than 0.05 were considered significant.

Study approval. All animal experiments are approved by the Institutional Animal Care and Use Committee at the University of California, Irvine.

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