Integrins and vascular extracellular matrix assembly.

E Ruoslahti, E Engvall

*J Clin Invest.* 1997; 99(6):1149-1152. [https://doi.org/10.1172/JCI119269](https://doi.org/10.1172/JCI119269).

Find the latest version:

http://jci.me/119269-pdf
Introduction
Integrins, a family of heterodimeric membrane proteins, serve in one of their main roles as adhesion receptors for various extracellular matrix components (1, 2). The integrin–matrix interaction is a two-way street. Most extracellular matrix proteins are ligands for integrins, allowing integrins to mediate cell attachment, and the matrix may, at least to a degree, control the expression and activity of integrins (3). Integrins, in turn, can control the formation of the matrix; this has been most clearly demonstrated for fibronectin (4), but may be true of some other matrix components as well.

The structural and morphological variations in the vasculature throughout the body are matched by an equally large variation in the molecular composition of the vessels. While clear in broad outline, the distribution of integrins and extracellular matrix components in blood vessels is incompletely known in detail.

The various integrins and matrix types are expressed in the different compartments of blood vessels and at different times in development. In principle, the endothelial cells manufacture a basement membrane matrix, the main components of which are laminin, type IV collagen, and heparan sulfate proteoglycan. The smooth muscle fibers in the media are surrounded by another basement membrane with a different set of isoforms of the same basic components. In addition, the media contain sheets of collagenous and elastic tissue, while the adventitia consists of connective tissue rich in collagens. Fig. 1 schematically represents the distribution of some matrix components of recent clinical interest in an arterial wall.

The integrins that have been detected in resting endothelia include α6β1, α5β1, α2β1, and αvβ3 (5), which are receptors for laminins, fibronectin, collagens, and vitronectin. Smooth muscle cells possess a large complement of β1 and αv integrins, of which αvβ3 may be particularly important (6).

Pathological lesions in the vasculature display various changes in integrins and matrix composition. Owing to an improving understanding of the roles of integrins and matrix in these pathologies, drugs modulating integrin activities are beginning to reach the clinic. We review here some of the recent advances in this field.

Integrins and matrix in vascular development
Gene knockout experiments, conducted both by Nature and by experimentalists, have established an important role for the matrix and its receptors in the development of the vasculature.

Fibronectin and type I collagen are necessary for the development and integrity of the vasculature. Mice that lack fibronectin have defects in somitogenesis and vascular development; the embryos die in utero during the 9th and 10th day of gestation (7). Mice that lack type I collagen develop surprisingly far; they die of circulatory collapse just before birth (8).

The absence of the α5β1 integrin, the main fibronectin receptor, is also incompatible with vascular development; the embryos of mice lacking this integrin die around gestational days 10–11. Mice lacking the αv integrin subunit, defining receptors for vitronectin and other RGD-containing proteins, die soon after birth owing to abnormalities in the cerebral blood vessels (7). The reasons for the selective effect of αv on cerebral vasculature are not known, but they may relate to the importance of the αvβ3 and αvβ5 integrins for angiogenesis (see below) or to the preferential expression of two potential ligands of these integrins, vitronectin (9) and merosin (10, 11), in brain vessels. Perhaps brain vessels follow a different developmental path than other blood vessels.

The laminin β2 chain is normally expressed in kidney glomeruli and in smooth muscle basement membranes of blood vessels (in addition to locations such as synapses in the peripheral nervous system and in muscle). In the vascular system, its absence primarily manifests itself in the kidney (12). Similarly, mutations in certain type IV collagen chains cause a kidney disease, Alport’s syndrome (13), although some endothelial basement membranes, including those of the eyes, normally contain these collagens (14). Thus, glomerular function appears to be particularly dependent on the correct structure of vascular basement membranes and vascular extracellular matrices.

Integrins and matrix in the maintenance of vascular structure and function
The cell–matrix interactions mediated by integrins are thought to be important in maintaining the polarity and positioning of the cells that make up the vessel wall. In addition, the integrins are likely to be involved in the sensing of and subsequent responses to changes in flow conditions within the vessels. Moreover, certain integrin ligands in the matrix, such as von Willebrand factor, participate in hemostasis and other injury responses (15, 16).

Structural components of the matrix are particularly im-

Address correspondence to Erkki Ruoslahti, La Jolla Cancer Research Center, The Burnham Institute, La Jolla, California 92037.

Received for publication 7 February 1997.
important in maintaining the integrity of blood vessels. Fibrillin, which is also an integrin ligand, appears to be important for the maintenance of vascular integrity. Mutations in fibrillin that cause its abnormal assembly result in Marfan’s syndrome, one hallmark of which is weakness, enlargement, and rupture of large blood vessels (17, 18). Similarly, Ehlers-Danlos type IV patients, who have mutations in type III collagen, often suffer an aortic rupture (19). Interestingly, the absence of elastin, which like fibrillin is concentrated in the elastic laminae of the media, causes aortic constriction rather than widening (20).

**Integrins and matrix in vascular pathologies**

Disturbances in integrin–matrix interactions underlie or contribute to many vascular pathologies.

Abnormal extracellular matrix accumulation is an important component in the pathophysiology of lesions involving intimal hyperplasia and in the somewhat analogous situation of glomerulosclerosis. Intimal thickening after a lesion, such as that inflicted in angioplasty, occurs in two stages. The first one is characterized by smooth muscle cell proliferation and migration into what will become the neointima (Fig. 2). During the second stage, smooth muscle cells fill the neointima with matrix. The role of integrins in these processes has been probed with antibodies and peptides capable of blocking the binding of integrins to their ligands.

<table>
<thead>
<tr>
<th>ECM components</th>
<th>Clinical importance</th>
</tr>
</thead>
<tbody>
<tr>
<td>collagen XVIII</td>
<td>angiogenesis</td>
</tr>
<tr>
<td>thrombospondin</td>
<td>angiogenesis</td>
</tr>
<tr>
<td>osteopontin</td>
<td>response to injury</td>
</tr>
<tr>
<td>fibrillin</td>
<td>Marfan syndrome</td>
</tr>
<tr>
<td>elastin</td>
<td>supravalvular aortic stenosis</td>
</tr>
<tr>
<td>collagen III</td>
<td>Williams syndrome</td>
</tr>
<tr>
<td></td>
<td>Ehlers-Danlos type IV</td>
</tr>
</tbody>
</table>

Many integrins recognize the peptide motif RGD (arginine-glycine-aspartic acid) as the key feature in their matrix ligands, and peptides containing this motif, or chemicals that mimic it, inhibit ligand binding (2). These RGD peptides and their mimetics can be designed to be specific for individual integrins. In vitro studies show that RGD peptides and antibodies capable of blocking the ligand binding of the αvβ3 integrin can inhibit the migration of smooth muscle cells. A possible ligand for the αvβ3-mediated migration is osteopontin, an RGD protein the levels of which are greatly increased in intimal hyperplasia (6). Indeed, an antibody that blocks αvβ3, as well as the platelet integrin αIIbβ3, is on the market for the restenosis indication, and drugs based on the RGD motif are also being developed for this purpose (21, 22).

The effectiveness of integrin-blocking agents in preventing restenosis may not be limited to their ability to inhibit cell migration; integrins also control cell proliferation (23) and help assemble matrix, at least fibronectin matrix (4). The αvβ3 integrin is one of the integrins that promotes the assembly of fibronectin matrix (24).

Increased integrin activity, if it plays a role, is not likely to be a major factor in the accumulation of neointimal matrix, because this matrix includes components that are not integrin ligands, such as the proteoglycan versican (25, 26). Instead, increased production of the growth factor TGFβ is the likely
cause of the pathological matrix accumulation in intimal hyperplasia. TGFβ stimulates the synthesis of several matrix components, including proteoglycans. TGFβ activity is increased in intimal hyperplasia and inhibiting this activity with antibodies has been found to reduce the matrix accumulation in both in vitro and in vivo experimental models (26). This facet of intimal hyperplasia seems to resemble many other diseases in which overproduction of TGF is an important underlying factor (27). The pathogenetic mechanism is characterized by TGFβ-induced synthesis and accumulation of excessive matrix, accompanied by degeneration of the epithelial component of the tissue; these diseases include glomerulonephritis, liver cirrhosis, pulmonary fibrosis, and others.

Blood vessels in different tissues are specialized and subject to dynamic changes. Lymphocytes recognize these specialized features in tissue-selective homing (28). Specialized tissue recognition may underlie the tendency of certain types of tumors to show preferential patterns of metastasis. Recently, in vivo tissue targeting with phage expressing libraries of random peptides has been used to demonstrate tissue-specific biochemical differences in vascular beds (29). Whether these tissue-specific features include differences in integrins or matrix components in normal vasculature is not known but, as detailed below, highly selective changes in the expression of these proteins do occur in abnormal blood vessels, such as those providing blood supply to tumors.

**Integrins and matrix in angiogenesis**

Angiogenesis, the formation of new blood vessels, also has important connections to integrins and the matrix. Endothelial cells migrate on extracellular matrix components in vitro and very likely also in vivo. Moreover, the αvβ3 and αvβ5 integrins are upregulated in angiogenic endothelial cells. Thus, the new blood vessels in tumors (30) and in certain inflammatory lesions, such as oxygen-induced retinal neovascularization (31), express these integrins at higher levels than normal resting blood vessels. Furthermore, cultured endothelial cells, which resemble angiogenic cells in that they also express substantial quantities of αvβ3, are dependent on αvβ3 for survival. If αvβ3 is blocked with RGD-containing peptides or antibodies selective for this integrin, endothelial cells grown in vitro and angiogenic endothelial cells in vivo will undergo apoptosis (30,32). As discussed above, αv knockout mice have problems with their cerebral vasculature (8) and this may also reflect the role of αvβ3 in angiogenesis (and vasculogenesis).

The apparent dependence of angiogenic endothelium on αvβ3 has opened up promising therapeutic possibilities. Thus, αvβ3-directed RGD peptides have been shown to suppress tumor growth and retinal angiogenesis in ocular neovascular diseases (33). Various types of cells, including cultured endothelial cells, require another integrin, the α5β1 fibronectin receptor, for survival under serum-free culture conditions (34; Zhang, Z., and E. Ruoslahti, unpublished observations). However, under other culture conditions, and when engaged in angiogenesis in vivo, endothelial cells seem to be protected against apoptosis by the αvβ3 integrin, because under these conditions apoptosis can be induced with αvβ3 blockers (30,32). It is unclear how the endothelial cell dependence on αvβ3 is mediated at the molecular level. The ligand that the αvβ3 integrin binds to in mediating angiogenesis is not known but seems unlikely to be vitronectin, the best known ligand for this integrin. Osteopontin is a possible candidate as this RGD protein is expressed at high levels in injured blood vessels (6).

Despite their apparent involvement in adhesive interactions important for the functioning and survival of the angiogenic endothelial cells, a portion of the αv integrins is present on the luminal surface of these cells (5). αv integrins also appear to be active on the luminal surface, because an αv integrin-peptide carried on a phage particle causes the phage to bind to tumor capillaries (Pasqualini, R., E. Koivunen, and E. Ruoslahti, unpublished observations). Clarifying the exact role of the αvβ3 and αvβ5 integrins and their ligands in angiogenesis and tumor targeting is clearly an important task for the future.

A matrix-related marker of angiogenic vasculature is an alternatively spliced variant of fibronectin. Fibronectin is alternatively spliced at three different regions of the molecule. One of these regions contains a type III fibronectin domain known as ED-B. The ED-B domain is preferentially expressed in angiogenic vasculature and, although it has not been proven to be exposed on the luminal side of the endothelium, it has been suggested as a marker suitable for tumor targeting (35). Another link between angiogenesis and vascular matrix is the ability of components of this matrix to inhibit angiogenesis. Such a role has been shown for thrombospondin (36). Most recently, a degradation product of a vascular matrix component, type XVIII collagen, was shown to act as an antiangiogenic agent (37). The physiological significance of this protein fragment, endostatin, may be that its production can serve as a feedback mechanism to suppress angiogenesis in tissue repair.

Clearly the integrins and extracellular matrix play important roles in many facets of vascular biology, and much needs to be learned about those roles. One of the likely payoffs will be new therapies for vascular diseases.

**Acknowledgments**

We thank Drs. Robert Burgeson, Lynn Sakai, Alexander Ljubimov, and Ulla Wewer for discussions.

The authors are supported by grants CA28896 and Cancer Center Support Grant CA 30199 (E. Ruoslahti) and HD 31636 (E. Engvall).

**References**


---

**“Cell Adhesion In Vascular Biology”**

Series Editors, Mark H. Ginsberg, Zaverio M. Ruggeri, and Aijt P. Varki

<table>
<thead>
<tr>
<th>Date</th>
<th>Title</th>
<th>Authors</th>
</tr>
</thead>
<tbody>
<tr>
<td>October 15, 1995</td>
<td>Adhesion and signaling in vascular cell–cell interactions</td>
<td>Guy Zimmerman, Tom McIntyre, and Stephen Prescott</td>
</tr>
<tr>
<td>November 15, 1995</td>
<td>Endothelial adhesions: junctions in the control of vascular permeability and angiogenesis</td>
<td>Elisabetta Dejana</td>
</tr>
<tr>
<td>November 15, 1995</td>
<td>Genetic manipulation of vascular adhesion molecules in mice</td>
<td>Richard O. Hynes and Denis D. Wagner</td>
</tr>
<tr>
<td>December 15, 1995</td>
<td>The extracellular matrix as a cell cycle control element in atherosclerosis and restenosis</td>
<td>Richard K. Assouan and Eugene M. Marcantonio</td>
</tr>
<tr>
<td>December 15, 1995</td>
<td>Effects of fluid dynamic forces on vascular cell adhesion</td>
<td>Konstantinos Konstantopoulos and Larry V. McIntyre</td>
</tr>
<tr>
<td>January 15, 1996</td>
<td>Selectin ligands: Will the real ones please stand up?</td>
<td>Aijt Varki</td>
</tr>
<tr>
<td>February 15, 1996</td>
<td>Cell adhesion and angiogenesis</td>
<td>Joyce Bischoff</td>
</tr>
<tr>
<td>February 15, 1996</td>
<td>von Willebrand Factor</td>
<td>Zaverio Ruggeri</td>
</tr>
<tr>
<td>March 15, 1996</td>
<td>Therapeutic inhibition of carbohydrate–protein interactions in vivo</td>
<td>John B. Lowe and Peter A. Ward</td>
</tr>
<tr>
<td>March 15, 1996</td>
<td>Integrins and vascular extracellular matrix assembly</td>
<td>Erkki Ruoslahti and Eva Engvall</td>
</tr>
<tr>
<td>April 1, 1997</td>
<td>Platelet GPIb/IIIa antagonists: The first anti-integrin receptor therapeutics</td>
<td>Barry Collier</td>
</tr>
<tr>
<td>April 15, 1997</td>
<td>Importance of shear stress in endothelial adhesion molecule expression</td>
<td>Michael Gimbrone</td>
</tr>
<tr>
<td>May 1, 1997</td>
<td>Proteoglycans and proteoglycan-binding proteins in vascular biology</td>
<td>Robert Rosenberg</td>
</tr>
<tr>
<td>May 15, 1997</td>
<td>New insights into integrin-ligand interaction</td>
<td>Robert Liddington and Joseph Lofts</td>
</tr>
<tr>
<td>June 1, 1997</td>
<td>Adhesive interactions of Sickle erythrocytes with endothelium</td>
<td>Robert Hebbel</td>
</tr>
<tr>
<td>June 15, 1997</td>
<td>Cell migration in vascular biology</td>
<td>Stephen Schwartz</td>
</tr>
<tr>
<td>July 1, 1997</td>
<td>Integrin signaling in vascular biology</td>
<td>Sanford Shattil and Mark Ginsberg</td>
</tr>
<tr>
<td>July 15, 1997</td>
<td>Multi-step mechanisms of leukocyte homing</td>
<td>Eugene Butcher</td>
</tr>
<tr>
<td>August 1, 1997</td>
<td>Role of PSGL-1 binding to selectins in leukocyte recruitment</td>
<td>Rodger McEver and Richard Cummings</td>
</tr>
</tbody>
</table>