Monocyte/macrophage expression of ABCA1 has minimal contribution to plasma HDL levels

Mehrdad Haghpassand, … , Omar L. Francone, Robert J. Aiello


Excess accumulation of cholesterol in macrophages results in foam cell production and lesion development. Recent studies have demonstrated that ATP-binding cassette protein A1 (ABCA1) is highly regulated in macrophages and mediates the efflux of cholesterol and phospholipids to apolipoproteins, a process necessary for HDL formation. The goal of this study was to determine the contribution of monocyte/macrophage ABCA1 to HDL formation in vivo. We generated mice expressing ABCA1 in macrophages and mice with selected inactivation of ABCA1 in macrophages by bone marrow transplantation in ABCA1-deficient (ABC1−/−) and wild-type (WT) mice. At all times, the level of HDL inABC1−/− recipient mice remained low relative to WT recipient mice irrespective of the genotype of the donor macrophage ABCA1 or high-fat feeding. Expression of WT macrophage ABCA1 in ABC1−/− mice resulted in a small but significant increase in apoA-I levels starting 2 weeks after transplantation. No further increase in apoA1 was observed up to 14 weeks after transplantation. The increase in apoA1 was accompanied by a small but significant increase in HDL cholesterol 6 weeks after transplantation. The HDL formed as a consequence of the expression of WT macrophage ABCA1 migrated to the alpha position in a two-dimensional gel electrophoresis. These results demonstrate that monocyte/macrophage ABCA1 contributes to HDL formation; however, the contribution to the overall plasma HDL levels is minimal.

Find the latest version:

https://jci.me/12810/pdf
Monocyte/macrophage expression of ABCA1 has minimal contribution to plasma HDL levels

Mehrdad Haghpasand, Patricia-Ann K. Bourassa, Omar L. Francone, and Robert J. Aiello

Pfizer Global Research and Development, Groton, Connecticut, USA

Address correspondence to: Robert J. Aiello, Pfizer Global Research and Development, Department of Cardiovascular and Metabolic Diseases, Eastern Point Road, Groton, Connecticut 06340, USA. Phone: (860) 441-6051; Fax: (860) 441-1128; E-mail: robert_j_aiello@groton.pfizer.com.

Received for publication March 26, 2001, and accepted in revised form September 4, 2001.

Excess accumulation of cholesterol in macrophages results in foam cell production and lesion development. Recent studies have demonstrated that ATP-binding cassette protein A1 (ABCA1) is highly regulated in macrophages and mediates the efflux of cholesterol and phospholipids to apolipoproteins, a process necessary for HDL formation. The goal of this study was to determine the contribution of monocyte/macrophage ABCA1 to HDL formation in vivo. We generated mice expressing ABCA1 in macrophages and mice with selected inactivation of ABCA1 in macrophages by bone marrow transplantation in ABCA1-deficient (ABC1−/−) and wild-type (WT) mice. At all times, the level of HDL in ABC1−/− recipient mice remained low relative to WT recipient mice irrespective of the genotype of the donor macrophage ABCA1 or high-fat feeding. Expression of WT macrophage ABCA1 in ABC1−/− mice resulted in a small but significant increase in apoA-I levels starting 2 weeks after transplantation. No further increase in apoAI was observed up to 14 weeks after transplantation. The increase in apoAI was accompanied by a small but significant increase in HDL cholesterol 6 weeks after transplantation. The HDL formed as a consequence of the expression of WT macrophage ABCA1 migrated to the alpha position in a two-dimensional gel electrophoresis. These results demonstrate that monocyte/macrophage ABCA1 contributes to HDL formation; however, the contribution to the overall plasma HDL levels is minimal.

in vivo. We generated mice expressing ABCA1 in macrophages and mice with selected inactivation of ABCA1 in macrophages by bone marrow transplantation. Our findings demonstrate that macrophage ABCA1 mediates the biogenesis of mature HDL; however, this contribution is insufficient to restore HDL plasma levels or modulate the overall HDL pool.

Methods

Animals. ABCA1-knockout mice were created in DBA1lac/J background as described previously (11). Wild type (WT) DBA1lac/J mice were purchased from The Jackson Laboratories (Bar Harbor, Maine, USA). Mice were weaned at 4 weeks of age, and the study began when animals were between 10 and 12 weeks old. All mice were maintained on a 12-hour light/dark cycle and fed a chow diet. During the last 3 weeks before termination of the study, half of the animals were put on a high-fat diet containing 20% of calories as protein, 50% of calories as carbohydrate, 20% by weight lard, 1% by weight corn oil, and 0.15% by weight cholesterol.

Bone marrow transplantation. At 8–10 weeks of age, the recipient WT and ABC1–/– female mice were divided into two groups. Bone marrow was harvested from tibias of male WT and ABC1–/– mice and injected into lethally irradiated (10 Gy) recipient mice as described previously (15, 16). The resulting four groups of mice will be referred to as WT→ABC1+, ABC1+/–→ABC1+/–, WT→WT, and ABC1+/–→WT denoting donor bone marrow transplanted to (→) the recipient mouse. All mice that received bone marrow transplantation survived and appeared to be in good health during the first 11 weeks of the study. During the last 3 weeks of the study, of the four animals in each group on the high-fat diet, one WT→ABC1+, two ABC1+/–→ABC1+/–, one WT→WT, and two ABC1+/–→WT died. Also, one ABC1+/–→ABC1–/– mouse on the chow diet died during the last week of the study. The cause of death was not determined.

Peritoneal macrophages isolation and cholesterol efflux. Upon the termination of the study, the mice on the chow diet (n = 3 for each group) were injected intraperitoneally with 1 ml of sterile 6% casein, and peritoneal exudate cells were harvested after 4 days by washing the peritoneal cavity with HBSS (Life Technologies Inc., Rockville, Maryland, USA) supplemented with 1% FBS. Peritoneal cells were washed once and resuspended in RPMI-1640 plus 10% FBS plus 2 mM glutamine plus 40 µg/ml gentamicin sulfate at approximately one million cells per milliliter and plated in 96-well Costar plates (0.2 ml/well; Corning-Costar Corp., Acton, Massachusetts, USA) to determine cholesterol efflux to apoAI. Also, cells were plated at approximately two million cells per well in Costar six-well plates for DNA isolation. To determine the cholesterol efflux, cells effluxed in 96-well plates were labeled for 48 hours with 15 µCi/ml [1,2,3H]-cholesterol (NEN Life Science Products Inc., Boston, Massachusetts, USA) and treated for an additional 18 hours in the presence or absence of 0.3 mM 8-(4-chlorophenylthio) adenosine 3’, 5’-cyclic mono-phosphate (cpt-cAMP). Subsequently, cells were washed, and 0.2 ml of 20 µg/ml human apoAI in RPMI-1640 was added to each well (14). At various times, media from each treatment was filtered through a 0.45-µm multiscreen plate (Millipore Corp., Bedford, Massachusetts, USA), and the corresponding cells were washed once with PBS and lysed in 0.2 N NaOH. The radioactivity in filtered media and cell lysates were determined in a Wallac 1450 Microbeta Plus liquid scintillation counter (Perkin Elmer Life Sciences Inc., Boston, Massachusetts, USA). Percentage of radiolabeled cholesterol released (percentage of efflux) was calculated as (cpm in media/cpm in media + cpm in cell lysate) × 100. Calculations were also performed using cpm at time zero as the denominator with similar results.

DNA isolation, PCR and RNase protection assay. Peritoneal macrophages were lysed in DNAzol reagent (Life Technologies Inc.). Genomic DNA was isolated by spooling, and the DNA was resuspended in 8 mM NaOH. PCR amplification was performed using a PCR kit (Roche Molecular Systems, Branchburg, New Jersey, USA). WT ABCA1 was detected using a forward primer in exon 19 at position 167,602 (Genbank reference no. AF287263), 5’TTGGGAACCTGCTAAATA3’, and a reverse primer in the targeted sequence, 5’TGGCATCCTTGTGTTAAT3’, resulting in a 420-bp fragment. ABC1–/– mice and injected into lethally irradiated female mice were divided into two groups. Bone marrow was harvested from tibias of male WT and ABC1–/– mice and injected into lethally irradiated (10 Gy) recipient mice as described previously (15, 16). The resulting four groups of mice will be referred to as WT→ABC1+, ABC1+/–→ABC1+/–, WT→WT, and ABC1+/–→WT denoting donor bone marrow transplanted to (→) the recipient mouse. All mice that received bone marrow transplantation survived and appeared to be in good health during the first 11 weeks of the study. During the last 3 weeks of the study, of the four animals in each group on the high-fat diet, one WT→ABC1+, two ABC1+/–→ABC1+/–, one WT→WT, and two ABC1+/–→WT died. Also, one ABC1+/–→ABC1–/– mouse on the chow diet died during the last week of the study. The cause of death was not determined.

Peritoneal macrophages isolation and cholesterol efflux. Upon the termination of the study, the mice on the chow diet (n = 3 for each group) were injected intraperitoneally with 1 ml of sterile 6% casein, and peritoneal exudate cells were harvested after 4 days by washing the peritoneal cavity with HBSS (Life Technologies Inc., Rockville, Maryland, USA) supplemented with 1% FBS. Peritoneal cells were washed once and resuspended in RPMI-1640 plus 10% FBS plus 2 mM glutamine plus 40 µg/ml gentamicin sulfate at approximately one million cells per milliliter and plated in 96-well Costar plates (0.2 ml/well; Corning-Costar Corp., Acton, Massachusetts, USA) to determine cholesterol efflux to apoAI. Also, cells were plated at approximately two million cells per well in Costar six-well plates for DNA isolation. To determine the cholesterol efflux, cells effluxed in 96-well plates were labeled for 48 hours with 15 µCi/ml [1,2-3H]-cholesterol (NEN Life Science Products Inc., Boston, Massachusetts, USA) and treated for an additional 18 hours in the presence or absence of 0.3 mM 8-(4-chlorophenylthio) adenosine 3’, 5’-cyclic mono-phosphate (cpt-cAMP). Subsequently, cells were washed, and 0.2 ml of 20 µg/ml human apoAI in RPMI-1640 was added to each well (14). At various times, media from each treatment was filtered through a 0.45-µm multiscreen plate (Millipore Corp., Bedford, Massachusetts, USA), and the corresponding cells were washed once with PBS and lysed in 0.2 N NaOH. The radioactivity in filtered media and cell lysates were determined in a Wallac 1450 Microbeta Plus liquid scintillation counter (Perkin Elmer Life Sciences Inc., Boston, Massachusetts, USA). Percentage of radiolabeled cholesterol released (percentage of efflux) was calculated as (cpm in media/cpm in media + cpm in cell lysate) × 100. Calculations were also performed using cpm at time zero as the denominator with similar results.

DNA isolation, PCR and RNase protection assay. Peritoneal macrophages were lysed in DNAzol reagent (Life Technologies Inc.). Genomic DNA was isolated by spooling, and the DNA was resuspended in 8 mM NaOH. PCR amplification was performed using a PCR kit (Roche Molecular Systems, Branchburg, New Jersey, USA). WT ABCA1 was detected using a forward primer in exon 19 at position 167,602 (Genbank reference no. AF287263), 5’TTGGGAACCTGCTAAATA3’, and a reverse primer in the targeted sequence, 5’TGGCATCCTTGTGTTAAT3’, resulting in a 420-bp fragment. RNase protection assay was performed using a 227-bp ABCA1 probe (between nucleotides 3,399 and 3,627 using Genbank sequence X75926 for mouse ABCA1) and a 103-bp cyclophilin probe from Ambion Inc. (Austin, Texas, USA). The RNase protection assay was performed using a Pharmingen (San Diego, California, USA) RNase protection kit according to the manufacturer’s protocol. The protected fragment was run on a Novex Quick-Point nucleic acid separation system (Invitrogen Corp., Carlsbad, California, USA), and bands were visualized by exposing the plates to autoradiography or quantified using a Strom 860 phosphorimagerr.

Plasma lipid and lipoprotein analysis. Mouse plasma was isolated from blood collected from the retro-orbital plexus. Cholesterol levels were determined using enzymatic colorimetric assays (Roche Molecular Biochemicals, Indianapolis, Indiana, USA). HDL was separated from non-HDL lipoproteins by dextran sulfate precipitation (17). An ELISA determined apoAI as described previously (17). The distribution of apoAI among HDL species were determined by two-dimensional gel electrophoresis as described previously (18) using a biotinylated rabbit polyclonal Ab (BIODESIGN International, Kennebunk, Maine, USA) and 125I-labeled streptavidin (Amersham Pharmacia Biotech, Piscataway, New Jersey, USA) to detect mouse apoAI.

Statistics. Statistical significance was determined by unpaired student’s t test using the StatView statistical
program (Abacus Concepts Inc., Berkeley, California, USA). Results are reported as mean plus or minus SD.

Results

Characterization of macrophages in recipient mice. To assess the contribution of macrophages to HDL synthesis, ABC1−/− mice were transplanted with bone marrow from WT mice. This resulted in mice that expressed ABCA1 in macrophages. Furthermore, mice with macrophage-specific inactivation of ABCA1 were generated by transplanting bone marrow from ABC1−/− mice in WT recipients. In addition, ABC1−/−→ABC1−/− and WT→WT transplantations were performed to generate the respective controls.

The repopulation of donor macrophages in the recipient mice was assessed by PCR analysis of elicited peritoneal macrophages. Elicited peritoneal macrophages were obtained from irradiated mice transplanted with donor bone marrow (donor bone marrow→recipient mice). Total DNA was isolated and subjected to PCR amplification using specific primers for WT ABCA1 (a) or the disrupted ABCA1 (b) gene. RNase protection assay for ABCA1 and cyclophilin was performed on 10 µg total RNA obtained from the liver and spleen of recipient mice (c). Protected ABCA1 fragments were visualized by autoradiography and quantitated by a phosphorimager. Similar levels of cyclophilin were seen in all samples.

The Journal of Clinical Investigation | November 2001 | Volume 108 | Number 9

Figure 1
PCR amplification of ABCA1 gene from WT and ABC1−/− peritoneal macrophages. Elicited peritoneal macrophages were obtained from irradiated mice transplanted with donor bone marrow (donor bone marrow→recipient mice). Total DNA was isolated and subjected to PCR amplification using specific primers for WT ABCA1 (a) or the disrupted ABCA1 (b) gene. RNase protection assay for ABCA1 and cyclophilin was performed on 10 µg total RNA obtained from the liver and spleen of recipient mice (c). Protected ABCA1 fragments were visualized by autoradiography and quantitated by a phosphorimager. Similar levels of cyclophilin were seen in all samples.

Figure 2
Cholesterol efflux from peritoneal macrophages in response to cpt-cAMP. Pooled peritoneal macrophages were obtained 14 weeks after transplantation. Cells were labeled with 3H-cholesterol for 48 hours and incubated for 18 hours in the presence (filled circles) or absence (open circles) of 0.3 mM cpt-cAMP. Cholesterol efflux to 20 µg/ml of human apoAI was measured by determining the radioactivity in the media after various times as a percentage of total radioactivity in the cells and media (n = 3 ± SE).
efflux assay demonstrate complete repopulation of donor macrophages in the recipient mice.

Plasma lipid and lipoprotein analysis. Changes in total plasma cholesterol (TPC), HDL cholesterol (HDL-c), and apoAI levels were monitored for 6 weeks after the bone marrow transplantation. HDL-c and apoAI levels remained low in \( ABC1^{-/-} \) recipient mice compared with the WT recipient mice throughout the study. Also, compared with the baseline, apoAI and HDL-c in \( ABC1^{-/-} \) recipients gradually decreased after transplantation (Table 1). By 2 weeks after transplantation, an approximately 50% greater level of mouse apoAI was observed in WT \( \rightarrow ABC1^{-/-} \) mice compared with \( ABC1^{-/-} \rightarrow ABC1^{-/-} \) control mice. The greater level of apoAI in WT \( \rightarrow ABC1^{-/-} \) mice was maintained throughout the study. There was also a higher level of HDL-c in WT \( \rightarrow ABC1^{-/-} \) compared with \( ABC1^{-/-} \rightarrow ABC1^{-/-} \) mice at 6 weeks after the transplantation (3.95 ± 1.7 vs. 1.6 ± 1.2 mg/dl, respectively; \( P < 0.001 \)). However, no differences in apoAI or HDL-c were observed in WT mice that received \( ABC1^{-/-} \) bone marrow compared with WT \( \rightarrow \) WT controls at any time point. Also, no significant differences were observed in non-HDL cholesterol levels.

To determine the distribution of HDL subspecies in mice after transplantation, pooled plasma samples from each group were subjected to a two-dimensional gradient gel electrophoresis, and the distribution of apoAI among HDL species was determined by immunoblotting (Figure 3). The presence of WT macrophages in \( ABC1^{-/-} \) mice resulted in the appearance of a small population of \( \alpha \)-migrating HDL, accounting for the greater apoAI and HDL levels in plasma samples from these mice. No apparent difference was observed in WT recipient mice transplanted with WT or \( ABC1^{-/-} \) bone marrow.

Response to high-fat diet. Several studies have demonstrated that ABCA1 expression is increased in macrophages loaded with cholesterol, resulting in greater efflux to free apoAI (6, 12). To determine if a high-fat and high-cholesterol diet increases the contribution of macrophage-derived cholesterol to HDL, four mice from each group were put on a Western-type, high-fat diet for 3 weeks before the termination of the study (between week 11 and week 14 after bone marrow transplantation). Subsequently, HDL-c and apoAI levels were determined at the end of the feeding period (Figure 4). High-fat feeding increased the HDL cholesterol and apoAI levels compared with chow diet in all animals; however, the HDL and apoAI levels in the \( ABC1^{-/-} \) recipients remained low irrespective of the genotype of the donor bone marrow.

**Table 1** Plasma cholesterol and apoAI levels in mice after bone marrow transplantation.

<table>
<thead>
<tr>
<th>Donor→recipient</th>
<th>Baseline</th>
<th>Week 2</th>
<th>Week 4</th>
<th>Week 6</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>n</td>
<td>TPC</td>
<td>HDL-c</td>
<td>apoAI</td>
</tr>
<tr>
<td>WT→( ABC1^{-/-} )</td>
<td>8</td>
<td>15.2</td>
<td>7.3</td>
<td>38.7</td>
</tr>
<tr>
<td>(2.6)</td>
<td>(1.8)</td>
<td>(9.0)</td>
<td>(7.2)</td>
<td>(4.8)</td>
</tr>
<tr>
<td>( ABC1^{-/-} \rightarrow ABC1^{-/-} )</td>
<td>8</td>
<td>17.4</td>
<td>7.5</td>
<td>42.7</td>
</tr>
<tr>
<td>(3.6)</td>
<td>(1.3)</td>
<td>(9.1)</td>
<td>(7.4)</td>
<td>(3.6)</td>
</tr>
<tr>
<td>WT→WT</td>
<td>7</td>
<td>56.2</td>
<td>40.8</td>
<td>676.2</td>
</tr>
<tr>
<td>(12.2)</td>
<td>(11.2)</td>
<td>(176.1)</td>
<td>(4.0)</td>
<td>(2.0)</td>
</tr>
<tr>
<td>( ABC1^{-/-} \rightarrow WT )</td>
<td>8</td>
<td>58.3</td>
<td>36.7</td>
<td>706.8</td>
</tr>
<tr>
<td>(7.2)</td>
<td>(2.5)</td>
<td>(118.8)</td>
<td>(11.9)</td>
<td>(3.8)</td>
</tr>
</tbody>
</table>

Plasma samples were obtained from each mouse on chow diet 1 day before (baseline) and 2, 4, and 6 weeks after the transplantation by retro-orbital bleeding. Total and HDL cholesterol were determined enzymatically, and mouse apoAI was determined by an ELISA. SDs are shown in parentheses. \( ^{a} \) mg/dl. \( ^{b} \) µg/ml. \( ^{c} P < 0.001 \) compared with \( ABC^{-/-} \rightarrow ABC1^{-/-} \). \( ^{d} P < 0.05 \) compared with \( ABC^{-/-} \rightarrow ABC1^{-/-} \).

**Figure 3** Distribution of mouse apoAI by two-dimensional gel electrophoresis. Fourteen weeks after transplantation, 15 µl of pooled plasma from each transplant group was applied to electrophoreses in 0.75% agarose gel, which was then applied to a 3–16% polyacrylamide gradient gel. Plasma proteins were transferred to nitrocellulose membranes, and mouse apoAI was detected using a rabbit polyclonal Ab.
ABCA1, resulting in the removal of excess cholesterol to apolipoproteins. The critical role of ABCA1 in HDL formation has prompted speculation that pharmacological regulation of ABCA1 in macrophages can directly modulate HDL levels. Our results demonstrate that the repopulation of ABC1−/− mice with WT macrophages results in a minor increase in the steady state HDL levels up to 14 weeks after the transplantation. This study also suggests that steady state HDL levels represent the contribution from many tissues that express ABCA1 as opposed to only those with the greatest need for cholesterol efflux such as macrophages.

A conservative estimate for the total number of macrophages in a mouse is approximately 10^8 cells (19). These cells comprise a very small portion of the total peripheral cells in the body. Although the exact level of cholesterol flux from macrophages has not been determined in vivo, the relative proportion of total cholesterol flux contributed by the macrophages must be small. This assumption is substantiated by the observation that tissues that have high level of resident macrophages, such as the lung and the spleen, have little contribution to the total cholesterol flux through the liver (20). It is therefore unlikely that a major part of cholesterol in HDL originates from the macrophages. However, macrophages may act as the initial donors of phospholipids and cholesterol to initiate the formation of mature HDL especially because the liver, where nascent HDL is formed from secreted apolipoproteins, contains the largest macrophage population in the body (approximately 20% of total body macrophages) (19). Once the mature HDL is formed, it can obtain cholesterol by other receptor-mediated pathways such as SR-BI (5) or from the lipolysis of remnant lipoproteins. Our results demonstrate that the contribution of macrophages to the formation of mature HDL is minimal. The expression of WT macrophages with functional ABCA1 in the ABC1−/− mice resulted in only a small increase in HDL. Nonetheless, the HDL formed as a consequence of normal ABCA1 expression by macrophages migrated to the alpha position on a two-dimensional gel, suggesting that macrophages do contribute to the formation of mature HDL but the level of contribution is very small. The formation of mature HDL is also dependent on plasma enzymes such as lecithin:cholesterol acyltransferase (21, 22) and phospholipid transfer protein (23). It is possible that these factors are markedly reduced in ABC1−/− mice, thus preventing the formation of large HDL. In our study, introducing ABCA1-deficient bone marrow to WT mice (ABC1−/−→WT) did not inhibit HDL formation, suggesting that in the presence of all of the factors necessary for mature HDL production, macrophage ABCA1 ablation has no major effect on the HDL level. (See results for ABC1−/−→WT in Table 1). Also, high-fat feeding, which induces foam cell formation and upregulates ABCA1, does not substantially increase HDL levels in ABC1−/− mice transplanted with WT bone marrow up to 14 weeks after transplantation. Nevertheless, a study by Kennedy et al. has demonstrated slow repopulation of tissue macrophages after transplantation in mice (24). Based on the RNase protection study, we can estimate that approximately 7% of WT level of ABCA1 RNA is expressed in the liver from ABCA1-deficient mice. Considering that Kupffer cells constitute, at the most, 20% of an adult liver, these results suggest a 35% repopulation of liver macrophages in ABCA1-deficient mice. These results were also confirmed by Y-chromosome quantitation in the recipient liver (data not shown).

Patients with Tangier disease or ABC1−/− mice do not have gross accumulation of lipid in their peripheral tissues. However, an abnormality in macrophage accumulation has been observed in patients with Tangier disease. Increased susceptibility to macrophage accumulation may also be responsible for moderately increased risk of coronary artery disease in these patients (10). Expression of ABCA1 in macrophages may have a beneficial role in atherosclerosis by facilitating cholesterol efflux and preventing foam cell production. Studies in apoE-knockout mice have demonstrated that small changes in plasma apoE as a consequence of apoE expression by macrophages have a profound effect on atherosclerosis (15, 25). Although the role of macrophage ABCA1 on atherosclerosis remains to be determined, the expression of ABCA1 in macrophages does not seem to be a major contributing factor to the heterogeneity observed in the plasma HDL levels.
Acknowledgments

We thank Jeff McNamara and Kathy Murray at Charles River for their excellent technical and organizational help with bone marrow transplantation and animal husbandry. We also thank Lori Royer and Kenneth Hoppe for their assistance and Barrett Rollins for graciously allowing us to use the facilities at Dana Farber Cancer Institute for irradiation of mice.


