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microRNA-21-5p dysregulation in exosomes derived from heart failure patients impairs regenerative potential

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Exosomes, as functional paracrine units of therapeutic cells, can partially reproduce the reparative properties of their parental cells. The constitution of exosomes, as well as their biological activity, largely depends on the cells that secrete them. We isolated exosomes from explant-derived cardiac stromal cells from patients with heart failure (FEXO) or from normal donor hearts (NEXO) and compared their regenerative activities in vitro and in vivo. Patients in the FEXO group exhibited an impaired ability to promote endothelial tube formation and cardiomyocyte proliferation in vitro. Intramyocardial injection of NEXO resulted in structural and functional improvements in a murine model of acute myocardial infarction. In contrast, FEXO therapy exacerbated cardiac function and left ventricular remodeling. microRNA array and PCR analysis revealed dysregulation of miR-21-5p in FEXO. Restoring miR-21-5p expression rescued FEXO’s reparative function, whereas blunting miR-21-5p expression in NEXO diminished its therapeutic benefits. Further mechanistic studies revealed that miR-21-5p augmented Akt kinase activity through the inhibition of phosphatase and tensin homolog. Taken together, the heart failure pathological condition altered the miR cargos of cardiac-derived exosomes and impaired their regenerative activities. miR-21-5p contributes to exosome-mediated heart repair by enhancing angiogenesis and cardiomyocyte survival through the phosphatase and tensin homolog/Akt pathway.

Introduction
Mounting lines of evidence, including that from our laboratory (1–4), have demonstrated that cell-based therapy holds great promise for the regeneration of injured heart muscle (5–12). Cell-based products must be carefully preserved to maintain their viability and activity until transplantation, however, and there are also some risks involved in cell transplantation. The modes of action for cell therapy products remain elusive, making it difficult to standardize each cell lot. Recent meta-analyses indicate that cardiac cell therapies are overwhelmingly safe and only show none-to-marginal efficacy (13, 14). The development of cell-free and nonliving therapeutics (e.g., proteins, RNAs) has the potential to revolutionize cardiovascular regenerative medicine. These therapeutics have compounded the evidence showing that the benefits of stem cell therapies mainly come from paracrine mechanisms, instead of the direct differentiation of injected stem cells into cardiomyocytes (15, 16). The injected cells secrete proteins and nucleic acids to promote endogenous repair (17, 18).

Exosomes are nanosized membrane vesicles secreted by most cell types, including stem cells and cancer cells (19, 20). They are extracellular nanoshuttles that facilitate cell–cell communications and are crucial for maintaining cells’ normal physiological functions. It has been reported that exosomes, as functional paracrine units of stem cells, can partially recapitulate the regenerative activities of their parent cells, suggesting that they may provide an alternative, cell-free therapeutic option (21–24). The mechanisms that drive the exosome-mediated repair processes rely largely on the transferring of exosomal cargos, including microRNAs (miR), mRNA, and proteins, to the recipient cells (25). Accordingly, the constitution of exosomes, as well as their biological activity largely depend on the physiological state of their parent cells (26–29).

It has been reported that stem cells derived from animals or patients with physiological stresses, such as systemic inflammation, have impaired regenerative activity (30–33). Because many patients with cardiac diseases, especially heart failure, suffer from these physiological stresses, we have hypothesized that cardiac cell–secreted exosomes derived from these patients may have impaired therapeutic activity, or possibly make the condition worse. In this article, we compared the treatment effects of cardiac exosomes from healthy donor hearts (normal exosomes, or NEXO) to those of cardiac exosomes from failing hearts (failure exosomes, or FEXO).

Results

Generation and characterization of NEXO and FEXO. Information regarding patients and healthy donors is presented in Table 1. There were no substantial differences in age or body mass index
Effects of NEXO or FEXO on cardiomyocytes, endothelial cells, and cardiac fibroblasts in vitro.

Cardiomyocytes, endothelial cells, and cardiac fibroblasts (CFs) are the 3 major cell types in the heart. We first measured the uptake efficiency of NEXO and FEXO by those cells. DiI-labeled exosomes were readily internalized by neonatal rat cardiomyocytes (NRCMs) (Figure 1A). There was no significant difference in uptake efficiency between NEXO and FEXO by NRCMs (Figure 1B). NEXO significantly promoted cardiomyocyte proliferation, as evidenced by the higher percentage of Ki-67-positive nuclei, while treatment with FEXO suppressed cardiomyocyte proliferation (Figure 1, C and D). In addition, cardiomyocytes treated with NEXO exhibited fewer terminal deoxynucleotidyl transferase nick-end labeling (TUNEL)-positive nuclei, indicating less apoptosis. In contrast, FEXO did not show any antiapoptotic effects (Figure 1, E and F). To further validate the effects of NEXO and FEXO on adult cardiomyocytes, human cardiomyocytes (HCs) were cultured on HC cell culture extracellular matrix, then treated with NEXO or FEXO for 24 hours. NEXO significantly promoted cardiomyocyte proliferation, as indicated by the higher percentage of Ki-67, while treatment with FEXO suppressed cardiomyocyte proliferation (Supplemental Figure 4, A and D). This result was supported by the assessment of 2 additional proliferation markers: phosphohistone H3, a characteristic marker of mitosis, and aurora kinase B, a cytokinesis marker, (Supplemental Figure 4, E–H).

A tube formation assay using human umbilical vein endothelial cells (HUVECs) was used to evaluate the proangiogenic effects of exosomes. Treatment with FEXO inhibited tube formation of HUVECs, while NEXO promoted tube formation, compared to saline control (Figures 1, G and H). A-Smooth muscle actin (α-SMA) was employed as an indicator of transition from fibroblasts to myofibroblasts. The expression of α-SMA was upregulated in FEXO-treated CFs (Figures 1, I and J), suggesting a phenotype transition to myofibroblasts. Taken together, our data suggest that cardiac cells in the failing heart secreted exosomes with an impaired ability to promote cardiomyocyte proliferation, decrease programmed cell death, and stimulate angiogenesis in vitro. Moreover, such diseased exosomes drive the differentiation of fibroblasts to myofibroblasts.

Effects of NEXO and FEXO therapy on cardiac function and morphometry in a mouse model of acute myocardial infarction. We induced an acute myocardial infarction (MI) in CD1 mice by coronary vessel ligation, then intramyocardially injected NEXO, FEXO, or phosphate-buffered saline (PBS) into the MI border zone. Left ventricular ejection fraction (LVEF) was used to assess...
fractional shortening deterioration (Supplemental Figure 5A). These results demonstrate that FEXO has reduced reparative activity and may exacerbate LV dysfunction post-MI. At the histological level, Masson’s trichrome staining was performed to simultaneously explore infarcted (blue) and viable (red) cardiac tissues (Figure 2D). NEXO-treated hearts exhibited decreased infarct size (Figure 2E), increased infarcted wall thickness (Figure 2F), and increased viable tissue (Figure 2G) compared to FEXO and PBS controls. Similarly, injured hearts treated with NEXO displayed less chamber dilation (Supplemental Figure 5B and C) and smaller infarct circumference (Supplemental Figure 5D) when compared to control or FEXO-treated hearts. These data indicated that NEXO attenuated LV remodeling after MI, whereas FEXO exacerbates it.

the enhancement or preservation of heart function. There was no significant difference in LVEF between the 3 treatment groups at baseline (Figure 2A). LVEF progressively decreased in the control group over the next 3 weeks, whereas hearts injected with NEXO showed greater LVEFs, compared to those treated with FEXO or PBS control. FEXO-treated hearts exhibited a worsening of LVEFs when compared to PBS-treated control hearts (Figure 2B). To facilitate head-to-head comparisons, treatment effect was evaluated by calculating the changes in endpoint LVEF relative to baseline LVEF post-MI (Figure 2C). Control treatment had a negative effect, as LVEF declined over time, while NEXO treatment robustly prevented the decline of LVEF post-MI (Figure 2C). Likewise, the animals that received NEXO treatment showed a recovery in fractional shortening, whereas those injected with FEXO suffered from
Effects of NEXO and FEXO treatment on post-MI heart angiogenesis and apoptosis. It has been well established that adult stem cells exert their therapeutic benefits through indirect regenerative mechanisms. The secretome from the injected cells enhances cardiomyocyte proliferation, promotes neovascularization, and inhibits cardiomyocyte apoptosis (15, 35). Three weeks after MI and exosome injections, the percentage of cycling cardiomyocytes (Ki-67+/α-SA+, Figure 3, A and D) in the peri-infarct zone of NEXO-treated hearts was significantly higher than in those that received FEXO or PBS control injections. This finding was further confirmed by mitosis marker phospho-histone H3 and cytokinesis marker aurora kinase B (Supplemental Figure 6). A similar trend was detected in capillary density (von Willebrand [vWF]+ capillaries, Figure 3, B and E), suggesting the proangiogenic role of NEXO but not of FEXO. Hearts injected with NEXO displayed significantly fewer apoptotic cardiomyocytes (TUNEL+/α-SA+, Figure 3, C and F).
3, C and F), suggesting NEXO therapy led to tissue preservation. Overall, FEXO treatment is detrimental to post-MI healing, as it inhibits angiogenesis and cardiomyocyte proliferation.

**Dysregulation of miR-21-5p in FEXO.** Several studies imply that the mechanism of exosome-mediated repair involves exosomal miRNAs, which target specific signaling pathways in the recipient cells (e.g., injured cardiomyocytes) (36, 37). Therefore, we hypothesized that the functional impairment of FEXO might result from altered miR cargos. We compared the miRNA components of NEXO to that of FEXO using PCR microarrays for 84 well-reported miRNAs (Supplemental Figure 7, A and B). Twenty-three miRs were differentially expressed in the 2 groups of exosomes. Among those, miR-21-5p was the most dysregulated in FEXO, as compared to the expression in NEXO (Figure 4, A and B). We performed RT-PCR to verify the results. Interestingly, miR-21-5p, but not miR-21-3p, was dysregulated in FEXO (Figures 4C). We further verified that the myocardial tissue levels of miR-21-5p were decreased in the post-MI hearts injected with FEXO (Supplemental Figure 7C), as compared to those injected with NEXO.

**The role of miR-21-5p in exosome-mediated heart repair.** We designed various gain- and loss-of-function studies to verify the role of miR-21-5p in exosome-mediated, post-MI repair. The silencing of exosomal miR-21-5p by transfecting healthy cardiomyocytes with anti–miR-21-5p oligo (NEXO + anti–miR-21) was confirmed by RT-PCR on the exosome products (Supplemental Figure 8, A and C). Knocking down miR-21-5p in NEXO led to impaired antiapoptotic effects of NEXO on cultured H9C2 cells (Figure 5, A and B). In contrast to the control NEXO, NEXO + anti–miR-21 lost the ability to promote tube formation (Figure 5, C and D). In the meantime, we also engineered miR-21–rescued exosomes by transfecting heart failure cardiac cells with miR-21 oligo. These cells were engineered to produce exosomes with miR-21-5p overexpression (FEXO + miR-21) (Supplemental Figure 8, A and B). As expected, upregulation of miR-21-5p in FEXO rescued their ability to inhibit apoptosis (Figure 5, E and F) and promote tube formation (Figure 5, G and H). To further validate the functions of miR-21 in adult cardiomyocytes, we transfected adult HCs with miR-21 mimic or scrambled control. Overexpression of miR-21 led to improved survival of adult cardiomyocytes, as indicated by less apoptotic cells (TUNEL-positive nuclei) (Supplemental Figure 9).

To confirm those results in vivo, we employed the same mouse MI model to test the therapeutic effects of FEXO + miR-21 and NEXO + anti–miR-21. Mice injected with NEXO + anti-
miR-21 exhibited decreased pump function (Figure 6, A and B), increased scar mass, and decreased viable heart tissue, compared to the hearts injected with NEXO + miR-scr (Figure 6, C–G). In contrast, miR-21-5p enhancement rescued the therapeutic functions of FEXO. FEXO + miR-21 treatment led to improvement in cardiac function (Figure 6, A and B) and attenuation of LV remodeling (Figure 6, D–G). Histological analysis revealed that miR-21-5p inhibition abolished the ability of NEXO therapy to promote angiogenesis (Figure 7, A and B) and inhibit apoptosis (Figure 7, C and D). On the contrary, restoring miR-21-5p expression in FEXO rescued those paracrine effects (Figure 7, E–H). Moreover, the proangiogenic effects of miR-21-5p were confirmed by the increased number of functional vessels. The functionality of a blood vessel was validated by co-staining the endothelial cell marker, vWF, and the RBC marker. As shown in Supplemental Figure 10, RBCs were detected in the blood vessels in the MI border zone of the hearts treated with miR-21–overexpressed FEXO. Through those gain- and loss-of-function studies, we confirmed the pivotal role of miR-21-5p in exosome-mediated heart repair, possibly through proangiogenic and antiapoptotic mechanisms.

Together with miR-21, other miRs at the top of our list are let-7b-5p, miR125a-5p, and miR-146a-5p. We explored the effects of those miRs on cardiomyocytes, CFs, and endothelial cells as well. The results indicated that miR-146a inhibited apoptosis in cardiomyocytes but promoted tube formation in endothelial cells (Supplemental Figure 11). This finding was consistent with previous studies (36). In contrast, neither let-7b-5p nor miR-125a-5p showed beneficial effects on cardiomyocyte survival or endothelial tube formation (Supplemental Figure 11).

Effects of miR-21-5p on phosphatase and tensin homolog/Akt signaling. It has been established that miR-21-5p promotes cancer cell proliferation, angiogenesis, migration, and invasion by targeting pathways involving phosphatase and tensin homolog (PTEN), programmed cell death 4 (PDCD4), Forkhead box O1, SMAD family member 7, tumor necrosis factor-α-induced protein-8-like 2, Sprouty2 (Spry2), etc. (38). Recent studies have
also shown that PTEN and Spry1 are targeted by miR-21-5p in cardiovascular diseases (39, 40). To identify the potential target genes of miR-21 on 3 major cell types in the heart, we incubated cardiomyocytes (using human induced-pluripotent stem cell–derived cardiomyocytes [iCMs] and H9C2 cells), CFs (using human CFs), and endothelial cells (using HUVECs) with miR-21 mimic or scrambled control. We then examined 18 phosphorylated proteins with a signaling pathway phosphorylation array. The signaling pathway array pointed at PTEN, AKT, and BAD as the miR-21–regulated genes in cardiomyocytes and endothelial cells but not in CFs (Supplemental Figure 12).

Western blot studies further confirmed that PTEN expression was significantly attenuated in HCs and HUVECs treated with miR-21 mimics (Figure 8, A and B). This finding coincides with enhanced Akt phosphorylation, as well as the alteration of other downstream targets such as caspase-3 and Bcl-2 (Figure 8, A and C–E). Upregulation of miR-21 in cardiomyocytes and HUVECs also promoted proliferating cell nuclear antigen ([PCNA] a proliferation marker) and vascular endothelial growth factor (VEGF) expressions (Figure 8, F–H), which are consistent with the preceding data in Figure 5 showing the proangiogenic effects of miR-21. Previous studies indicated that PDCD4, a proapoptotic protein, is a known miR-21 target (41), so we checked PDCD4 expressions in CMs, HCFs, and HUVECs treated with miR-21 mimics. Western blot analysis showed reduced PDCD4 expression in CMs only, but not in HCFs or HUVECs (Figure 8, F and I).

Taken together, our data indicate that miR-21 inhibits cardiomyocyte apoptosis by targeting PDCD4, promotes angiogenesis by activating PTEN/Akt signaling, and promotes the expression of VEGF in endothelial cells. Interestingly, because the flow cytometry showed a significant proportion of mesenchymal cells or fibroblasts (CD90$, CD105$) in our cardiac cells (Supplemental Figure 2), it is highly possible that those cardiac cells secreted exosomes containing miR-21 to communicate with nearby cardiomyocytes and endothelial cells under physiological and heart failure conditions.
Discussion

Our results suggest that FEXO has impaired therapeutic potency in an animal model of MI and is incapable of promoting angiomyogenesis in vitro and in vivo. miRNA and a series of gain- and loss-of-function experiments revealed that the impairment of regenerative activities of FEXO is related to miR-21-5p dysregulation (Figure 8J). Over the past decade, cell therapy has emerged as a promising therapeutic option for cardiac regeneration and protection. As a major paracrine component, exosomes have been revealed to play a vital role in tissue repair by packaging and delivering RNAs and proteins. It has been reported that injured mouse hearts treated with cardiac stem cell–secreted exosomes exhibited regenerative and functional improvements produced by the stem cells themselves (21, 36). Exosomes derived from mouse embryonic stem cells promote neovascularization and cardiomyocyte survival and reduce fibrosis after MI, consistent with the resurgence of the cardiac proliferative response produced by embryonic stem cells (42). However, most of these exosome products are derived from young and healthy donors. Because most patients with cardiac diseases, especially heart failure, suffer from systemic inflammation and are elderly, it is possible that heart failure modifies exosome contents and further compromises their reparative properties. In this study, FEXO lost proangiogenic drive of HUVEC tube formations and failed to promote cardiomyocyte proliferation in injured mouse hearts. This line of evidence suggests a dysfunction of FEXO in ischemic tissue repair. Our finding can partly explain the compromised therapeutic effects in clinical trials using autologous stem cells. The advanced cardiomyopathy condition modifies exosome contents and renders the stem cells ineffective in therapeutic repair.

The mechanisms that drive exosome-mediated repair rely on the transfer of specific donor cell miRNAs and proteins to recipient cells (43). In the present study, we performed miRNA array
which is consistent with previous studies, while neither let-7b-5p nor miR-125a-5p showed beneficial effects on cardiomyocyte survival or endothelial tube formation. Although miR-21 expression has been associated with various disorders (39, 46–48), its precise functional role in cardiac diseases is still controversial. Thomas et al. reported that miR-21 expression is highly upregulated by cardiac stress, leading to activation of ERK/MAP kinase signaling through inhibition of Spry1, which in turn promoted cardiac fibrosis and heart dysfunction (39). miR-21 also reportedly induced cardiac fibrosis by inhibiting PTEN expression in CFs at the infarct zone after myocardial ischemia-reperfusion (49). However, Patrick et al. reported that miR-21 was not essential for cardiac fibrosis, as miR-21–null mice still display cardiac hypertrophy, fibrosis, and the loss of cardiac contractility (48). In the present study, miR-21 did not affect the survival of CFs (Supplemental Figure 11G). Moreover, miR-21 regulated the PTEN/Akt signaling pathway in cardiomyocytes and endothelial cells.

Figure 7. Manipulation of miR21 in exosomes modulates their proangiogenic and anti-apoptotic activities in vivo. (A) Representative images of post-MI heart sections stained with vWF (green), α-SM (red), and DAPI (blue). White circles indicate capillaries in the peri-infarct zone. Scale bar: 100 μm. (B) Quantification of capillary density (vWF+). (C) Heart sections stained with TUNEL (green), α-SM (red), and DAPI (blue). White circles indicate capillaries in the peri-infarct zone. Scale bar: 50 μm. (D) Quantitation of cardiomyocyte apoptosis (TUNEL+/α-SM+). (E) Representative images of post-MI heart sections stained with vWF (green), α-SM (red), and DAPI (blue). White circles indicate capillaries in the peri-infarct zone. Scale bar: 50 μm. (F) Quantitation of capillary density (vWF+). (G) Heart sections stained with TUNEL (green), α-SM (red), and DAPI (blue). White squares indicate apoptotic cardiomyocytes in the peri-infarct zone. Scale bar: 50 μm. (H) Quantitation of cardiomyocyte apoptosis (TUNEL+/α-SM+). (B, D, F, and H) n = 6 animals for each group and 3 heart sections for each animal. **P < 0.01. Two-tailed t test. All values are mean ± SD. FEXO + miR-scr, exosomes derived from the cardiac cells of patients with heart failure transfected with scrambled miR oligo. FEXO + miR-21, exosomes derived from the cardiac cells of patients with heart failure transfected with miR-21-5p oligo. NEXO + miR-scr, exosomes derived from the cardiac cells of the normal hearts transfected with scrambled miR oligo. NEXO + anti–miR-21, exosomes derived from the cardiac cells of the normal hearts transfected with anti-miR-21-5p oligo.

analysis to compare FEXO with NEXO. We found that exosomal miR-21-5p, secreted from healthy cardiac cells, can be delivered to recipient cardiac cells to regulate apoptosis and angiogenesis and improve heart function in a mouse MI model. The silencing of miR-21-5p in NEXO eliminated these benefits, and the restoration of miR-21-5p in FEXO rescued their therapeutic potency, further supporting the essential role of miR-21-5p in exosome-mediated heart repair. Although our studies focused on miR-21-5p, there are several other miRs dysregulated in FEXO, such as miR-146a, let-7b, miR-125a, and miR-23. It has been reported that exosomal miR-146a leads to beneficial effects in post-MI hearts by targeting Irak-1 and Traf6, both associated with the toll-like receptor signaling pathway (36). It has also been shown that miR-23 can promote angiogenesis in cardiac endothelial cells by activating proangiogenic signaling through the inhibition of Spry2 and Sema6A (44, 45). Our results indicate that miR-146a inhibited apoptosis in cardiomyocytes but promoted tube formation in endothelial cells, which is consistent with previous studies, while neither let-7b-5p nor miR-125a-5p showed beneficial effects on cardiomyocyte survival or endothelial tube formation.

Although miR-21 expression has been associated with various disorders (39, 46–48), its precise functional role in cardiac diseases is still controversial. Thomas et al. reported that miR-21 expression is highly upregulated by cardiac stress, leading to activation of ERK/MAP kinase signaling through inhibition of Spry1, which in turn promoted cardiac fibrosis and heart dysfunction (39). miR-21 also reportedly induced cardiac fibrosis by inhibiting PTEN expression in CFs at the infarct zone after myocardial ischemia-reperfusion (49). However, Patrick et al. reported that miR21 was not essential for cardiac fibrosis, as miR-21–null mice still display cardiac hypertrophy, fibrosis, and the loss of cardiac contractility (48). In the present study, miR-21 did not affect the survival of CFs (Supplemental Figure 11G). Moreover, miR-21 regulated the PTEN/Akt signaling pathway in cardiomyocytes and endothelial cells, supporting the essential role of miR-21-5p in exosome-mediated heart repair.
mesenchymal stem cell (MSC) exosomes and that it contributes to myocardial salvage via synergistic antiapoptotic activity (51). miR-21-5p plays a crucial role in the MSC exosome–mediated effects on cardiac contractility through PI3K signaling (52). One unique aspect of our study is the cell source. Although therapeutic MSCs can be readily derived from bone marrow and adipose tissues, these lial cells instead of fibroblasts (Figure 8), indicating that miR-21 does not promote cardiac fibrosis, as reported in previous studies.

In line with our findings, it was reported that exosomal miR-21 secreted by induced pluripotent stem cells promoted angiogenesis in transfected bronchial epithelial cells (50). Kristin et al. recently reported that miR-21a-5p is the most abundant miR in murine mesenchymal stem cell (MSC) exosomes and that it contributes to myocardial salvage via synergistic antiapoptotic activity (51). miR-21-5p plays a crucial role in the MSC exosome–mediated effects on cardiac contractility through PI3K signaling (52). One unique aspect of our study is the cell source. Although therapeutic MSCs can be readily derived from bone marrow and adipose tissues, these

**Figure 8.** miR-21-5p targets the PTEN pathway in post-MI pathology. (A) Representative Western blot images showing the expression of various PTEN/Akt pathway components. (B–E) Quantitation of the levels of PTEN, p-Akt, t-Akt, Bcl-2, and caspase-3 (n = 3). (F) Representative Western blot images showing the expression of PCNA (proliferation marker), VEGF, and PDCD4 (miR-21 target). (G and H) Quantitation of the levels of PCNA, VEGF, and PDCD4 (n = 3). (B–I) All miR-21 groups were normalized to the relevant scrambled controls. *P < 0.05 compared to scrambled control. **P < 0.01 compared to scrambled control. ***P < 0.001 compared to scrambled control. NS, no significance compared to scrambled control. Two-tailed t test. All values are mean ± SD. (J) Schematic showing the working model of our study. CM/H9C2/EC/CF + miR-scr, HC/H9C2 cells/HUVECs/human cardiac fibroblasts transfected with scrambled miR. CM/H9C2/EC/CF + miR-21, HC/H9C2 cells/HUVECs/human cardiac fibroblasts transfected with miR-21-5p mimic.
cells do not naturally reside in the heart. Our study is more physiologically relevant. We focused on intrinsic cardiac cells in failing or healthy hearts and demonstrated that heart failure pathology leads to a severe dysregulation of miR-21-5p in intrinsic cardiac cells, which further leads to the impaired regenerative function of the intrinsic heart exosomes. Therefore, our findings demonstrate that miR-21 not only plays a key role in therapeutic interventions (e.g., in the case of MSCs) but also represents a crucial component in the naturally occurring repair process after heart injury.

The regulation of PTEN by miR-21 was reported in carcinoma (53) and cardiovascular cells (49). Consistent with this, we found that upregulation of miR-21-5p in exosomes augmented phosphorylation of Akt via the inhibition of PTEN in the recipient cells. These effects were eliminated by the knockdown of miR-21-5p, indicating a possible molecular mechanism for the positive role of miR-21-5p in post-MI repair. Based on the flow cytometry data, cardiac stromal cells/fibroblasts might be the main source of exosomes with altered miR-21. Thomas et al. (39) revealed strong miR-21 expression primarily in CFs but was low in cardiomyocytes. This is consistent with our findings. It is quite possible that CFs released exosomal miR-21 to support survival of neighboring cardiomyocytes and endothelial cells under physiological conditions.

These findings have important clinical implications. Unlike cell-based therapy products, exosomes can offer an off-the-shelf and universal therapeutic option. The cryopreservation of exosomes normally does not require the sophisticated steps associated with cellular products. In addition, injection of allogeneic or even xenogeneic exosomes is well tolerated by the host immune system (54, 55). Our findings suggest that FEXO carries marked altered miR signatures compared with those from healthy heart cells and are functionally deficient in their ischemic myocardial repair capabilities. This may explain the reason why patients' autologous cell therapies only show modest therapeutic effects in clinical trials. More importantly, our data showed that the dysfunction of FEXO can be rescued by modulation of a specific miR in the exosomes. Our study provides mechanistic insights into the therapeutic potential of exosomes and strategies to rescue patients’ defective exosomes through modulation of specific miR cargos. This discovery paves the ground for future translational efforts in fabricating “designer” and “super” exosomes for enhanced therapeutic applications.

There are some limitations in our study. First, HUVECs are not a good representation of cardiac endothelial cells. Heart primary endothelial cells are a more preferable source for in vitro studies. Second, the iPS-derived cardiomyocytes and the adult HC cell line we employed in our study do not completely resemble primary adult HCs. A third limitation of our study is that we only showed the effects of differential miR-21 expressions at the protein level but did not design an experiment (such as luciferase reporter assays or RT-PCR) to prove that miR-21-5p directly targets PTEN and PDCD4. However, such target validation results have been confirmed in the literature (53, 56). In addition, the CD1 mouse has been a tangible resource for biomedical research, including cardiovascular diseases. Nevertheless, we admit that the mouse MI model we employed has some limitations. The CD1 mice we used were relatively young. Therefore, their endogenous reparative ability does not resemble that of aged humans whose innate repair systems may be compromised, especially those suffering long-term heart failure.

Methods

Cell culture and exosome isolation. Diseased heart tissues were received from patients who underwent heart transplantation at UNC Hospital after heart failure. Healthy heart tissues were received from normal, nondamaged, human donor hearts. Cardiac cells were separated and cultured as previously described (6). In brief, myocardial tissues harvested from donors’ hearts were separated and washed with PBS, cut into smaller 2-mm³ fragments, and partially enzymatically digested at 37°C with collagenase type IV (C1889, Sigma-Aldrich). The tissue samples were plated onto fibronectin-coated Petri dishes in Iscove’s modified Dulbecco’s medium (IMDM, Gibco) infused with 20% fetal bovine serum (FBS; Corning). After 3 to 5 days, cardiac cells began to grow from the explants. Exosomes were isolated from the conditioned medium of these cardiac cells. Cardiac cells from passages 1 through 3 were cultured to 80% confluency. The medium was then switched to serum-free IMDM and conditioned for 14 days. Then, exosomes were isolated from the conditioned medium by ultracentrifugation, as previously described (54, 55). In brief, conditioned medium was filtered through 0.22-μm Steriflip filters to remove cellular debris and large vesicles. The filtrate was then added to Amicon Ultra-15 100 kDa filters (SCG00525, Millipore) to centrifuge at 5,000 × g for 5 minutes. The flow-through was discarded. The concentrated exosomes were collected and washed with PBS 3 times and stored at ~80°C. Labeling was performed using 10-μM DiI (V22889, Thermo Fisher Scientific).

Flow cytometry. Flow cytometry was performed to examine the antigenic phenotypes of cardiac cells. Cells were incubated with antibodies against CD90 (555595, BD), CD105 (ab11414, Abcam), CD31 (555445, BD), CD 34 (ab81289, Abcam), CD 45 (555482, BD), and c-kit (550412, BD) for 60 minutes at 4°C. Both unstained and isotype controls (55754, 559320, BD) were used as negative controls. Flow cytometry was conducted with a Cytoflex Flow Cytometer (Beckman Coulter) and data were analyzed with FCS Express software (De Novo).

Exosome characterization. Nanoparticle tracking analysis (NanoSight NS300, Malvern) was used to measure the concentration and size of exosomes. Each sample was imaged 5 times for 60 seconds and analyzed. Transmission electron microscopy was used to assess the morphology of exosomes. In preparation for transmission electron microscopy, exosomes were fixed with 4% paraformaldehyde and 1% glutaraldehyde and were stored at room temperature. Microscopy was conducted with a transmission electron microscope (JEM-2000FX, Jeol).

Immunocytochemistry. NRCMs and CFs were isolated and cultured as previously described (57). The H9C2 cell line was purchased from Sigma-Aldrich (88092904, Sigma-Aldrich). Adult HCs were purchased from Celprogen and cultured on HC cell culture extracellular matrix (36044-15, Celprogen). iCMs were kindly provided by Dr. Freytes (UNC/NC State Joint Department of Biomedical Engineering, Chapel Hill/Raleigh). After 3 days of cultivation, the culture medium was supplemented with 7 × 10⁸ exosomes or PBS for an additional 24 hours. They were fixed with 4% paraformaldehyde and blocked with Protein Block Solution (DAKO) with 1% saponin (Sigma-Aldrich). Subsequently, the cells were stained with anti-α-sarcomeric actin (α-SA, a7811, Sigma-Aldrich), anti-Ki-67 (ab15580, Abcam), anti-α-smooth muscle actin (ab5694, Abcam), or anti-vWF (ab6994, Abcam) antibodies. Flow 488 or Texas-Red conjugated secondary antibodies (ab150117,
Heart morphometry. Animals were sacrificed 3 weeks after treatments. Six cryosections from each heart, collected at 400-μm intervals, were stained with Masson’s trichrome. Infarct size, infarct circumference, infarct wall thickness, and viable tissue in the risk area were measured with NIH Image J software, as previously described (1–4).

Antibody array and Western blot analysis. Adult HC (CM, 36044-15, Celprogen), iCMs, H9C2 cells (88092904, Sigma-Aldrich), human CFS (HCF, 306-05A, Sigma-Aldrich), and HUVEC cells (PCS-100-010, ATCC) were transfected with miR-21-5p mimic or scrambled control (miScript miRNA Mimic, 219600, Qiagen), according to the manufacturer’s protocol with slight modifications. Briefly, cells were cultured to 70% confluency. miR-21-5p mimic or scrambled control was mixed with HiPerFect Transfection Reagent and culture medium without serum, at a final concentration of 5 nM. Cells were returned to the incubator for 48 hours. Cells were collected, lysed by RIPA Lysis Buffer (89900, Thermo Fisher Scientific), and centrifuged at 14,000 x g for 15 minutes to pellet the cell debris. The protein concentration was quantified with a BCA Protein Assay (23227, Thermo Fisher Scientific).

Protein from transfected CM, H9C2, HCF, and HUVEC cells was used on the signaling pathway phosphorylation array (AAH-AKT-1-2, RayBiotech). The protein array assay was performed according to the manufacturer’s protocol with slight modification. Briefly, the antibody membranes were blocked for 30 minutes at room temperature. The samples were then added and incubated overnight at 4°C. The detection antibody cocktail was prepared, pipetted onto the membranes, and incubated overnight at 4°C. Then, the samples were incubated with HRP anti-rabbit IgG and visualized with the ChemiDoc Touch Imaging System (Chemidoc, Bio-Rad). Quantitative analysis was performed with ImageJ or Image Lab software.

Protein from transfected CM, H9C2, HCF, and HUVEC cells was used on Western blot analysis. The equivalent of 25 mg of total protein per lane was loaded onto Mini-PROTEAN TGX Stain-Free Protein Gels (4568083, Bio-Rad) and transferred to Immuno-Blot PVDF Membranes (1620177, Bio-Rad). Membranes were blocked with 5% blocking buffer for 1 hour and incubated with primary antibodies against Pten (ab13192, Abcam), p-akt (phospho T308) (ab88449, Abcam), t-akt (ab8805, Abcam), BCL-2 (ab59348, Abcam), caspase-3 (ab13847, Abcam), PDCD4 (ab51495, Abcam), PCNA (ab29, Abcam), or VEGF (PA5-16754, Thermo Fisher Scientific) overnight, at 4°C on a rocker. Subsequently, the samples were incubated with appropriate horseradish peroxidase-conjugated secondary antibodies at room temperature for 1 hour on a shaker. The blots were exposed with Clarity Western Blotting Substrates (1705061, Bio-Rad) and visualized with the ChemiDoc Touch Imaging System (Chemidoc, Bio-Rad). Quantitative analysis was performed with ImageJ or Image Lab software, and expressions were normalized to GAPDH.

Statistics. GraphPad Prism (GraphPad Software) was used for statistical analysis. Results were presented as mean ± SD. All comparisons between 2 groups were performed with 2-tailed unpaired Student’s t-test. One-way ANOVA analysis, with post hoc Bonferroni correction, was used to compare means among more than 2 groups. Differences were considered statistically significant when P < 0.05.

Study approval. All animal procedures were approved by the Institutional Animal Care and Usage Committee (IACUC, North Carolina State University).

Author contributions
YL and KC designed the overall experiments. LQ, SH, SL, HZ, HM, KH, AV, TS, ZL, JT, JC, TA, PUD, and QY performed the

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experiments and analyzed the data. LQ, JC, and KC wrote the article. All authors have approved the final article. All authors have provided the corresponding author with written permission to be named in the article.

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