Septic patients frequently develop cognitive impairment that persists beyond hospital discharge. The impact of sepsis on electrophysiological and molecular determinants of learning is underexplored. We observed that mice that survived sepsis or endotoxemia experienced loss of hippocampal long-term potentiation (LTP), a brain-derived neurotrophic factor–mediated (BDNF-mediated) process responsible for spatial memory formation. Memory impairment occurred despite preserved hippocampal BDNF content and could be reversed by stimulation of BDNF signaling, suggesting the presence of a local BDNF inhibitor. Sepsis is associated with degradation of the endothelial glycocalyx, releasing heparan sulfate fragments (of sufficient size and sulfation to bind BDNF) into the circulation. Heparan sulfate fragments penetrated the hippocampal blood-brain barrier during sepsis and inhibited BDNF-mediated LTP. Glycoarray approaches demonstrated that the avidity of heparan sulfate for BDNF increased with sulfation at the 2-O position of iduronic acid and the N position of glucosamine. Circulating heparan sulfate in endotoxemic mice and septic humans was enriched in 2-O– and N-sulfated disaccharides; furthermore, the presence of these sulfation patterns in the plasma of septic patients at intensive care unit (ICU) admission predicted persistent cognitive impairment 14 days after ICU discharge or at hospital discharge. Our findings indicate that circulating 2-O– and N-sulfated heparan sulfate fragments contribute to septic cognitive impairment.
Circulating heparan sulfate fragments mediate septic cognitive dysfunction

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Introduction

Septic patients frequently experience cognitive dysfunction that persists beyond hospital discharge, impairing survivors’ quality of life and ability to return to work (1–3). This cognitive impairment has been largely attributed to brain injury occurring early in sepsis, arising from pathogenic processes including cerebral microvascular thrombosis, metabolic derangements, and IL-1β–dependent neuroinflammation (4–8). Less is known about the long-term functional effects of sepsis on the electrophysiological and molecular neuronal pathways responsible for learning.

Classic neurosurgical studies have identified the hippocampus as the anatomic center of memory. Learning requires synaptic plasticity between key hippocampal neuronal networks, including the strengthening of cornu ammonis region 3 (CA3) to cornu ammonis region 1 (CA1) neuronal connections through a process called long-term potentiation (LTP), the key molecular mechanism driving spatial memory (9). Hippocampal LTP is dependent on the neuronal growth factor brain-derived neurotrophic factor (BDNF), as demonstrated by the induction of cognitive impairment by experimental BDNF sequestration (10). Interestingly, experimental and human sepsis has been associated with hippocampal pathology, including acute dysfunction of the hippocampal blood-brain barrier as well as later loss of hippocampal volumes in sepsis survivors (11–13).

There is increasing appreciation for the importance of the endothelial glycocalyx in sepsis pathophysiology. The glycocalyx is a ubiquitous endovascular layer enriched in heparan sulfate, a linear polysaccharide of repeating glucosamine-hexuronic (iduronic or glucuronic) acid disaccharide units. These disaccharides may be sulfated at specific sites (including the 2-O position of iduronic acid and/or the 6-O or N position of glucosamine), imparting a domain patterning of negative charge. During sepsis, the glycocalyx is fragmented, releasing heparan sulfate hexa- and octasaccharides into the bloodstream (14–16). These fragments have the capacity to interact with soluble proteins (such as growth factors) with remarkable specificity through sulfation-based electrostatic interactions, influencing a variety of homeostatic and/or pathologic signaling pathways (17). Shed fragments circulate for several days after sepsis onset and correlate with clinically significant outcomes including acute kidney and lung injury (15, 18).

Related Commentary: https://doi.org/10.1172/JCI127679

Authorship note: PSH and EPS contributed equally to this manuscript.
Conflict of interest: The authors have declared that no conflict of interest exists.
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Submitted: August 27, 2018; Accepted: January 29, 2019.
Reference information: J Clin Invest. https://doi.org/10.1172/JCI124485.
through i.p. injection of LPS (10 μg/g BW). Animals injected with LPS had a mortality rate of 18%, with 70% of deaths occurring within 3 days of LPS injection (Supplemental Figure 1A; supplemental material available online with this article; https://doi.org/10.1172/JCI124485DS1). Endotoxemic mice displayed sickness behavior for 3 days, followed by resolution of normal behavior and weight gain by day 7 (Supplemental Figure 1B). To determine whether post-septic mice had memory deficits that persisted despite resumption of normal activity, we performed hippocampus-dependent neurobehavioral testing (contextual fear conditioning) on WT C57BL/6 mice 7 days after i.p. LPS or saline (control) injection. Contextual memory was apparent in saline-treated mice but was impaired in post-LPS–treated mice (Figure 1A). This post-LPS deficit in memory coincided with loss of hippocampal

**Results and Discussion**

To establish a model of septic cognitive dysfunction, we induced endotoxemia in 8- to 12-week-old male and female C57BL/6 mice through i.p. injection of LPS (10 μg/g BW). Animals injected with LPS had a mortality rate of 18%, with 70% of deaths occurring within 3 days of LPS injection (Supplemental Figure 1A; supplemental material available online with this article; https://doi.org/10.1172/JCI124485DS1). Endotoxemic mice displayed sickness behavior for 3 days, followed by resolution of normal behavior and weight gain by day 7 (Supplemental Figure 1B). To determine whether post-septic mice had memory deficits that persisted despite resumption of normal activity, we performed hippocampus-dependent neurobehavioral testing (contextual fear conditioning) on WT C57BL/6 mice 7 days after i.p. LPS or saline (control) injection. Contextual memory was apparent in saline-treated mice but was impaired in post-LPS–treated mice (Figure 1A). This post-LPS deficit in memory coincided with loss of hippocampal
LTP, as measured using 300-μm-thick living hippocampal brain slices isolated from 8- to 12-week-old C57BL/6 mice 7 days after LPS (or saline) treatment (Figure 1B). We observed a similar loss of LTP in mice 7 days after cecal ligation and puncture (CLP) (Figure 1C, CLP or sham), a widely used model of polymicrobial peritonitis–induced sepsis. Intriguingly, we observed that post-LPS hippocampi maintained BDNF content (Figure 1D and Supplemental Figure 2. Sepsis-associated circulating heparan sulfate fragments penetrate the hippocampus and impede LTP. Liquid chromatography–tandem mass spectrometry MRM (LC-MS/MS MRM) analyses demonstrated shedding of heparan sulfate (HS) into the plasma of (A) mice 24 hours after i.p. LPS administration (10 μg/g BW vs. saline control) and (B) human patients with sepsis (enrolled in the MESSI cohort and followed longitudinally; control samples represent normal blood donors). (C) Accordingly, an increase in heparan sulfate content was detected in the hippocampus of LPS-injected mice that persisted for 7 days after injection. (D) Fluorescein-labeled, highly sulfated heparan sulfate (heparin) octasaccharides (250 μg) administered i.v. to mice 24 hours after i.p. LPS (10 μg/g) or saline treatment penetrated the hippocampal blood-brain barrier, as observed by confocal microscopy of freshly isolated hippocampal slices. Scale bars: 100 μm. (E) Highly sulfated heparan sulfate (heparin) octasaccharides (degree of polymerization 8 [dp8]) induced loss of LTP when perfused (2.5 μg/ml) over hippocampal slices isolated from healthy (non-septic mice). LTP was rescued by simultaneous perfusion with the TrkB agonist 7,8-DHF (250 nM). *P < 0.05, **P < 0.01, and ***P < 0.001, by ANOVA with Tukey’s correction for multiple comparisons (A, C, and E) or Kruskal-Wallis with Dunn’s test for multiple comparisons (B and D).
findings indicate that post-endotoxemic/post-septic mice have impaired memory and loss of LTP, despite the presence of BDNF within the hippocampus and persistence of hippocampal responsiveness to TrkB activation, suggesting induction of a competitive inhibitor of BDNF within the septic hippocampus.

Our group and others have shown that highly sulfated heparan sulfate hexa- to octasaccharide fragments are shed into the plasma in animal (19) and human (14) sepsis as a consequence of endothelial glycocalyx degradation. We confirmed these findings using high-sensitivity mass spectrometry multiple reaction monitoring (MRM) (19) analyses of plasma collected after i.p. LPS treatment in mice (Figure 2A) and in a separate cohort of human patients with sepsis, who were enrolled in the Molecular Epidemiology of Sepsis in the Intensive Care Unit (MESSI) study (Figure 2B). Consistent with the known ability of ultra-low-molecular-weight heparins to cross the blood-brain barrier in healthy mice (20), we observed that circulating heparan sulfate fragments also penetrated the blood-brain barrier, as i.v. administered fluorescein-labeled heparan sulfate octasaccharides were observed in the hippocampus (Figure 2D) and cortex (Supplemental Figure 3) in both septic and non-septic mice. Likewise, we observed an increase in hippocampal heparan sulfate content at the point of peak circulating heparan sulfate (1 day after LPS treatment) (Figure 2C). Interestingly, the accumulation of hippocampal heparan sulfate persisted for 7 days after LPS treatment, a time point characterized by impaired cognition (Figure 1A). The persistence of brain heparan sulfate has been recognized in other neurodegenerative disease states, such as Alzheimer’s dementia (21–23). Hippocampal heparan sulfate content eventually normalized in mice 14 days after LPS treatment (compared with mice 14 days after saline treatment), which was coincident with improvement in cognition (Supplemental Figure 4).

We have previously demonstrated that circulating heparan sulfate oligosaccharides can influence growth factor signaling by electrostatically binding positively charged residues of growth factor ligands (19). To determine whether hippocampus-penetrating heparan sulfate fragments interfere with BDNF, the neurotrophic responsible for hippocampal LTP, we isolated hippocampal slices from healthy (non-septic) 8- to 12-week-old C57BL/6 male and female mice and measured LTP in the presence or absence of highly sulfated heparan sulfate (heparin) octasaccharides, approximating the known size and sulfation pattern of circulating heparan sulfate fragments after sepsis (14). Exposure of hippocampal slices from healthy mice to highly sulfated heparan sulfate octasaccharides resulted in a significant decrease in LTP compared with saline controls, suggesting that these fragments can competitively inhibit BDNF binding to its receptor, TrkB. In contrast, exposure to heparin, a non-reducing heparan sulfate oligosaccharide, had no effect on LTP, indicating that the sulfation pattern of heparan sulfate fragments is crucial for competitive inhibition of BDNF.

To further investigate the role of heparan sulfate sulfation in BDNF binding and septic cognitive impairment, we performed glycoarray analyses of plasma collected 24 hours after i.p. LPS administration in mice and on day 7 of ICU admission in patients with sepsis. Consistent with the known ability of ultra-low-molecular-weight heparins to cross the blood-brain barrier in healthy mice (20), we observed that circulating heparan sulfate fragments also penetrated the blood-brain barrier, as i.v. administered fluorescein-labeled heparan sulfate octasaccharides were observed in the hippocampus (Figure 2D) and cortex (Supplemental Figure 3) in both septic and non-septic mice. Likewise, we observed an increase in hippocampal heparan sulfate content at the point of peak circulating heparan sulfate (1 day after LPS treatment) (Figure 2C). Interestingly, the accumulation of hippocampal heparan sulfate persisted for 7 days after LPS treatment, a time point characterized by impaired cognition (Figure 1A). The persistence of brain heparan sulfate has been recognized in other neurodegenerative disease states, such as Alzheimer’s dementia (21–23). Hippocampal heparan sulfate content eventually normalized in mice 14 days after LPS treatment (compared with mice 14 days after saline treatment), which was coincident with improvement in cognition (Supplemental Figure 4).
could be reversed by the downstream activation of TrkB, as octamer mice (Figure 2E). Heparan sulfate–mediated inhibition of LTP as compared with matched control slices isolated from the same hippocampal slices to highly sulfated octasaccharides inhibited LTP that participate in heparan sulfate–BDNF binding. We found that heparan sulfate fragments bound BDNF in a length- and sulfation-dependent manner (Supplemental Figure 5). Since SPR does not inform the precise sequence or pattern of sulfation necessary for BDNF binding, we used a glycosaminoglycan microarray (glycoarray) composed of 52 heparan oligosaccharides of varying sulfation patterns (24). This approach revealed that BDNF-avid oligosaccharides commonly contained disaccharides with both N- and O-sulfation sequences (Figure 3B and Supplemental Figure 6). Intriguingly, circulating heparan sulfate fragments collected from post-LPS–treated mice (Figure 3C) and septic humans (Figure 3D, Supplemental Figure 7, and Supplemental Table 1) were similarly enriched in N and O sulfation sequences on mass spectrometry MRM analysis. This enrichment in N and O sulfation was observed in the hippocampi of post-septic mice and persisted for 7 days (Figure 3E and Supplemental Table 2), indicating that mice maintained BDNF-avid heparan sulfate fragments within their hippocampi at time points coincident with post-septic cognitive impairment. We performed additional glycoarray experiments to define the avidity of heparan sulfate oligosaccharides for pro-BDNF, a proenzyme incapable of promoting LTP (25). We observed no pro-BDNF binding to any heparan sulfate oligosaccharide sequence (Supplemental Figure 8A), suggesting that the cleavage of pro-BDNF to active BDNF exposes a heparan sulfate binding site. Other known heparan sulfate binding proteins (e.g., antithrombin III, platelet factor 4) bound to heparan sulfate sequences distinct from those that bound BDNF. Similarly, the heparan sulfate 10e4 antibody (26) failed to bind BDNF-avid sequences (Supplemental Figure 8, B–D). We leveraged this inability of the 10e4 antibody to bind BDNF-avid sequences to demonstrate that sequestration of non–BDNF-avid heparan sulfate sequences failed to prevent loss of LTP (Supplemental Figure 9), further confirming the sulfation sequence specificity of the observed BDNF-inhibitory effect of heparan sulfate.

To determine the translational relevance of our findings, we measured heparan sulfate in plasma samples collected on day 0 (i.e., at presentation to the intensive care unit [ICU]) from septic patients enrolled in the Neurocognitive Impairment in Respiratory Failure and Shock (NIRFS) study of the MESSI cohort (Table 1) to determine whether the presence of circulating N- and O-sulfated heparan sulfate predicted cognitive impairment in sepsis. NIRFS patients with persistent cognitive impairment after sepsis (as quantified by a Montreal Cognitive Assessment [MoCA] score <21 or a cognitive inability to perform the test either at hospital discharge or 14 days after ICU discharge) were those who had increased circulating N- and O-sulfated heparan sulfate at the time of ICU admission (Figure 3F and Supplemental Table 3). Taken together, our findings demonstrate that the presence of circulating NS- and/or 2S-enriched, BDNF-avid heparan sulfate fragments at sepsis onset predicts cognitive impairment up to 2 weeks after ICU discharge.

In summary, our findings demonstrate that septic neurocognitive dysfunction is not only a consequence of inflammatory or ischemic brain injury but may also arise from septic interference with biologic processes (i.e., hippocampal BDNF/TrkB signaling) necessary for memory and cognition. Detection of circulating N- and O-sulfated heparan sulfates may therefore allow the identification of septic patients at high risk of prolonged cognitive impairment, enabling the future development (and personalized implementation) of sulfation sequence–targeted therapeutics to improve memory and other patient-centered outcomes in sepsis survivors.

Table 1. Characteristics of the NIRFS study participants

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>58 (41–64.5)</td>
</tr>
<tr>
<td>Male sex</td>
<td>12 (60%)</td>
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<tr>
<td>White race</td>
<td>13 (65%)</td>
</tr>
<tr>
<td>Years of education</td>
<td>14 (12–16)</td>
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<tr>
<td>Comorbid conditions</td>
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</tr>
<tr>
<td>Hypertension</td>
<td>6 (30%)</td>
</tr>
<tr>
<td>Diabetes</td>
<td>6 (30%)</td>
</tr>
<tr>
<td>Chronic kidney disease</td>
<td>5 (25%)</td>
</tr>
<tr>
<td>Congestive heart failure</td>
<td>4 (20%)</td>
</tr>
<tr>
<td>Cerebrovascular disease</td>
<td>1 (5%)</td>
</tr>
<tr>
<td>Cirrhosis</td>
<td>1 (5%)</td>
</tr>
<tr>
<td>Active malignancy</td>
<td>9 (45%)</td>
</tr>
<tr>
<td>Organ transplant</td>
<td>7 (35%)</td>
</tr>
<tr>
<td>Anxiety or depression</td>
<td>5 (25%)</td>
</tr>
<tr>
<td>APACHE III score</td>
<td>100 (93.5–124)</td>
</tr>
<tr>
<td>Septic shock at presentation</td>
<td>13 (65%)</td>
</tr>
<tr>
<td>Vasopressor shock at presentation</td>
<td>12 (60%)</td>
</tr>
<tr>
<td>Any septic shock</td>
<td>15 (75%)</td>
</tr>
<tr>
<td>Acute respiratory failure at presentation</td>
<td>12 (60%)</td>
</tr>
<tr>
<td>Invasive ventilation at presentation</td>
<td>6 (30%)</td>
</tr>
<tr>
<td>Any invasive ventilation</td>
<td>15 (75%)</td>
</tr>
<tr>
<td>Acute respiratory distress syndrome</td>
<td>10 (50%)</td>
</tr>
<tr>
<td>Duration of mechanical ventilation (days)</td>
<td>3 (1–7)</td>
</tr>
<tr>
<td>ICU length of stay (days)</td>
<td>6.5 (4.5–14)</td>
</tr>
<tr>
<td>MoCA</td>
<td>24 (22–25)</td>
</tr>
<tr>
<td>Moderate/severe cognitive impairment</td>
<td>6 (30%)</td>
</tr>
<tr>
<td>Trail-making test part B (seconds)</td>
<td>95.9 (66.1–139.8)</td>
</tr>
</tbody>
</table>

n = 17; 3 patients were unable to complete the MoCA because of severe cognitive impairment. APACHE, acute physiologic assessment and chronic health evaluation.
Methods
Detailed methods are provided in the Supplemental Methods.

Statistics
Statistical analyses of data on animal experiments, SPR, glycoarrays, and human subjects heparan sulfate fragment quantification were performed using GraphPad Prism (GraphPad Software). Analysis of all other human subjects data was performed using Stata Software. All replicates refer to studies using biological replicates, which were performed on different days to avoid temporal batch effects. No statistical outliers were excluded. Data represent the mean ± SEM or, for whiskers, were evaluated according to Tukey’s criteria. A two-tailed Student’s t test was performed for single comparisons. A P value of less than 0.05 was considered statistically significant for all analyses.

Study approval
Animal experiments. All animal experiments were performed with approval of the institutional animal care and use committee of the University of Colorado Office of Laboratory Animal Resources. Experiments were performed in accordance with the Institutional Animal Care and Use Committee Guidebook, published by the NIH Office of Laboratory Animal Welfare (OLAW), as well as the Animal Research Reporting of In Vivo Experiments (ARRIVE) guidelines (27). Male and female 8- to 12-week-old C57BL/6 mice, purchased from Charles River Laboratories and The Jackson Laboratory, were used in all studies. At least 3 biologic replicates were used for microscopy and glycosaminoglycan quantification results, and at least 6 biologic replicates were used for all other experiments.

Human subjects. Human samples and accompanying clinical data were collected from ICU patients enrolled in the NIRFS study, a sub-study of the MESSI cohort, which is a single-center, prospective cohort of patients admitted to the ICU at the Hospital of the University of Pennsylvania. This study was approved by the IRB of the University of Pennsylvania. Subjects or their available surrogates provided written informed consent.

Author contributions
JAH, BJA, SAM, JL, NJM, RJL, PSH, and EPS designed the study. JAH, JEO, RMD, and JAF performed animal neuroelectrophysiology and behavioral experiments. JAH and Y. Yang performed microscopy studies. SAM and KO performed Western blot analyses. JAH, GS, Y. Yu, FZ, and KH performed analytical glycochemistry experiments. BJA and NJM handled patient enrollment and data acquisition. JAH, BJA, JEO, RMD, GS, FZ, JL, RJL, NJM, PSH, and EPS performed data analysis and prepared the manuscript. EPS was responsible for overall coordination and oversight of the project.

Acknowledgments
This work was funded by NIH grants R01 HL125371 (to EPS and RJL); R01 GM125095 (to EPS and PSH); R01 HL137915 (to NJM); K23 HL140482 (to BJA); R01 HL094463 and U01 GM102137 (to JL); and T32 HL007085 (to JAH).

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