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Graphical abstract

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Enhancing glycolysis attenuates Parkinson’s disease progression in models and clinical databases

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Parkinson’s disease (PD) is a common neurodegenerative disease that lacks therapies to prevent progressive neurodegeneration. Impaired energy metabolism and reduced ATP levels are common features of PD. Previous studies revealed that terazosin (TZ) enhances the activity of phosphoglycerate kinase 1 (PGK1), thereby stimulating glycolysis and increasing cellular ATP levels. Therefore, we asked whether enhancement of PGK1 activity would change the course of PD. In toxin-induced and genetic PD models in mice, rats, flies, and induced pluripotent stem cells, TZ increased brain ATP levels and slowed or prevented neuron loss. The drug increased dopamine levels and partially restored motor function. Because TZ is prescribed clinically, we also interrogated 2 distinct human databases. We found slower disease progression, decreased PD-related complications, and a reduced frequency of PD diagnoses in individuals taking TZ and related drugs. These findings suggest that enhancing PGK1 activity and increasing glycolysis may slow neurodegeneration in PD.

Introduction

Parkinson’s disease (PD) is the second most common neurodegenerative disease. It is estimated to affect approximately 6 million people worldwide, and its prevalence will increase further as populations age (1). Patients with PD suffer debilitating motor symptoms as well as nonmotor symptoms including dementia and neuropsychiatric abnormalities (2, 3). Dopamine neurons in the substantia nigra pars compacta (SNc) and their projections into the striatum are especially susceptible to disruption in PD (4). Loss and impaired function of dopamine neurons cause the motor abnormalities that are a hallmark feature of PD. Although current treatments can sometimes relieve PD symptoms, no therapies prevent the neurodegeneration (5).

PD may have a number of different causes, and several pathogenic mechanisms have been proposed to contribute to the apoptotic death of neurons (6–10). In the majority of cases, the etiologies are unknown and probably complex. Aging, environmental toxins, and genetic mutations are all risk factors. In many cases, energy deficits and decreased ATP levels are observed in PD (11).

First, aging, the major risk factor for PD, impairs cerebral glucose metabolism, reduces mitochondrial biogenesis, and decreases ATP levels (12). Second, glycolysis and mitochondrial function are decreased in individuals with PD (13, 14). Third, mitochondrial toxins (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine [MPTP], rotenone, paraquat) induce PD and PD-like phenotypes in cells and animals, including humans (15). Fourth, mutations associated with familial PD (e.g., PINK1, LRRK2, α-synuclein, parkin, DJ-1, CHCHD2) disrupt various aspects of energy metabolism (16). It is also hypothesized that SNc dopaminergic neurons may be particularly susceptible to PD neurodegeneration, because their highly branched, unmyelinated axonal arbor, their many neurotransmitter release sites, and their rhythmic firing engender a large metabolic burden (17). These considerations suggested that impaired bioenergetics and reduced ATP levels might contribute to the pathogenesis of PD, might modify the risk of developing PD in the face of PD risk factors, and/or might modify the course or severity of the disease.

We recently discovered that terazosin (TZ) binds and activates phosphoglycerate kinase 1 (PGK1) (18), the first ATP-generating enzyme in glycolysis (Figure 1A). TZ is an α1-adrenergic receptor antagonist that can relax smooth muscle and is prescribed to treat benign prostatic hyperplasia and, rarely, hypertension (19). However, biochemical and functional studies show that the effects of TZ on PGK1 are independent of α1-adrenergic

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administration of MPTP to mice, pyruvate and ATP levels fell (Figure 1, F and G, and Supplemental Figure 2, A–C), and administration of TZ prevented the fall in pyruvate and ATP levels. Mitochondrial content (assessed by the ratio of mitochondrial DNA to nuclear DNA and by VDAC and PHB1 levels) also fell (Supplemental Figure 2, D–F). TZ partially prevented the decrease. As previously suggested (23), the increased pyruvate levels may have stimulated mitochondrial biogenesis. It would be difficult to measure ATP specifically in neurons, however, we observed similar changes in human neuroblastoma cells (Supplemental Figure 3, A–E). These data indicate that TZ activates glycolysis in vivo. Together with measurements of brain TZ levels (Supplemental Figure 3F), they also indicate that TZ readily crosses the blood-brain barrier.

Although PGK1 produces ATP, oxidative phosphorylation is probably important for increasing ATP on the basis of the following: (a) pyruvate, the product of glycolysis and major substrate for the citric acid cycle, increased (Figure 1, C and F, Supplemental Figure 1B, Supplemental Figure 2B, and Supplemental Figure 3A); (b) citrate synthase activity, a marker of mitochondrial activity, increased (Figure 1D, Supplemental Figure 1C, and Supplemental Figure 3B); (c) the extracellular acidification rate, a measure of glycolysis, and the O$_2$ consumption rate, a measure of mitochondrial respiration, both increased (Supplemental Figure 3, D and E); and (d) mitochondrial content was partially maintained after MPTP (Supplemental Figure 2, D and F), which may have also contributed to the increased ATP content.

We also asked whether TZ would increase energy production in mice that received MPTP. MPTP causes PD in humans and is used to model PD in other animals (21, 22). Seven days after administration of MPTP to mice, pyruvate and ATP levels fell (Figure 1, F and G, and Supplemental Figure 2, A–C), and administration of TZ prevented the fall in pyruvate and ATP levels. Mitochondrial content (assessed by the ratio of mitochondrial DNA to nuclear DNA and by VDAC and PHB1 levels) also fell (Supplemental Figure 2, D–F). TZ partially prevented the decrease. As previously suggested (23), the increased pyruvate levels may have stimulated mitochondrial biogenesis. It would be difficult to measure ATP specifically in neurons, however, we observed similar changes in human neuroblastoma cells (Supplemental Figure 3, A–E). These data indicate that TZ activates glycolysis in vivo. Together with measurements of brain TZ levels (Supplemental Figure 3F), they also indicate that TZ readily crosses the blood-brain barrier.

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by administration of TZ for the next 7 days, and an assay on day 7 (Figure 2A). Because individuals with PD present after the onset of neuron degeneration, we also asked whether delayed TZ administration would slow neuron loss and functional decline. Therefore, in some mice, we waited 7 days after delivering MPTP before starting the 7-day course of TZ treatment. We then performed an assay on day 14 (Figure 2A).

Over the course of 14 days, MPTP progressively decreased the levels of tyrosine hydroxylase (TH), the rate-limiting enzyme for generating dopamine. MPTP decreased TH levels in the SNc and striatum, reduced the numbers of TH-positive cells in the SNc, and decreased the intensity of TH immunostaining in the neuron projections into the striatum (Figure 2, B–G, and Supplemental Figure 4, A and B). As a result, the dopamine, 3,4-dihydroxyphenylacetic acid (DOPAC) and homovanillic acid (HVA) content of the striatum and SNc decreased (Figure 2, H and I, and Supplemental Figure 4, C–F). MPTP also increased the percentage of TH-positive cells that were TUNEL positive, indicating increased apoptosis (Figure 2J and Supplemental Figure 4, G and H). Beginning TZ treatment at the time we administered MPTP attenuated all these defects on day 7. When TZ delivery was delayed for 7 days after MPTP administration, it improved the abnormalities on day 14. Consistent with these biochemical defects, TZ prevented deficits in motor function on day 7, and it improved motor performance on day 14 after delayed administration (Figure 2K and Supplemental Figure 4, I and J).

Figure 2. TZ improves dopamine neuron and motor function in MPTP-treated mice. (A) Schematic for experiments in panels B–K. Eight-week-old C57BL/6 mice received 4 i.p. injections of MPTP (20 mg/kg at 2-hour intervals) or vehicle on day 0. Mice were then injected with TZ (10 μg/kg) or vehicle (0.9% saline) once a day for 1 week, and assays were performed on day 7. Other mice began receiving daily TZ or vehicle injections beginning on day 7, and assays were performed on day 14. n = 6. (B–D) Example of Western blots with TH and β-actin (protein loading control) in striatum and SNc on days 7 and 14 (B). Quantification of TH protein normalized to control (C and D). n = 6. (E) Example of immunostaining of TH in SNc and striatum. Scale bars: 100 μm (SNc) and 1 mm (striatum). Quantification of TH-positive neurons in SNc (F) and TH intensity in the striatum (G). n = 6. (H and I) Dopamine (DA) content in striatum and SNc. n = 6. (J) Percentage of TH-positive neurons that were positive for TUNEL staining. n = 6. (K) Behavioral response of mice in the rotarod test. Data reflect the duration that the mice remained on an accelerated rolling rod, normalized to mice on day 0. n = 8. Data represent examples and indicate the mean ± SEM. Blue indicates control and red indicates TZ treatment. *P < 0.05 and **P < 0.01, by Mann-Whitney U test for days 7 and 14. Supplemental Table 3 shows P values for all comparisons.
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of TH immunostaining in the striatum also decreased (Figure 3, D and E). TZ partially reverted these abnormalities toward control values. 6-OHDA progressively decreased dopamine, DOPAC, and HVA content, and TZ partially prevented the reduction (Figure 3F and Supplemental Figure 5, G and H). Seven weeks after injection of 6-OHDA into the right striatum, we observed that use of the left forepaw had fallen (Figure 3G), however, when the rats received TZ between weeks 5 and 7, they used both forepaws equally.

Previous studies have shown that MPTP and 6-OHDA can rapidly reduce TH expression (21), and consistent with this finding, we observed that TH levels, TH-positive neurons, TH intensity in the striatum, and dopamine content decreased rapidly after administration of MPTP and 6-OHDA to mice and rats, respectively (Figures 2 and 3 and Supplemental Figures 4 and 5). Cell death was also apparent. However, not all the damaged cells were rapidly killed, as cell death continued to progress for at least 14 days in MPTP-treated mice and for 7 weeks in 6-OHDA–treated rats (Figure 2J and Figure 3B). Accordingly, TH levels, TH-positive

These in vivo results in mice suggest that TZ slows or prevents MPTP-induced neurodegeneration, partially restores TH and dopamine levels, and improves motor function.

TZ enhancement of PGK1 activity slows neurodegeneration in 6-OHDA–treated rats. The compound 6-hydroxydopamine (6-OHDA) is delivered to rats to produce a model of dopamine neuron degeneration in PD (24). Previous studies have shown progressive cell death and injury between 2 and 12 weeks after administration of 6-OHDA (25–27). Therefore, we chose a 7-week course of observation. We injected 6-OHDA into the right striatum of rats, waited 2–5 weeks, and then initiated a 2-week course of TZ treatment (Figure 3A). In vehicle-treated rats, evidence of SNc cell apoptosis progressively increased from 2 to 7 weeks (Figure 3B and Supplemental Figure 5, A and B). However, irrespective of the delay before the start of treatment, we observed that TZ attenuated further cell loss. 6-OHDA also progressively decreased TH levels in the striatum and SNc (Figure 3C and Supplemental Figure 5, C–F). The percentage of TH-positive cells in the SNc and the intensity of TH immunostaining in the striatum also decreased (Figure 3, D and E). TZ partially reverted these abnormalities toward control values. 6-OHDA progressively decreased dopamine, DOPAC, and HVA content, and TZ partially prevented the reduction (Figure 3F and Supplemental Figure 5, G and H). Seven weeks after injection of 6-OHDA into the right striatum, we observed that use of the left forepaw had fallen (Figure 3G), however, when the rats received TZ between weeks 5 and 7, they used both forepaws equally.

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is highly conserved in flies and mammals, and supplying TZ reduced brain ATP levels (Figure 4, A and B). It also disrupted I inhibitor implicated in sporadic PD (29). Rotenone exposure rodegeneration in flies. As an additional model of PD, we treated MPTP and 6-OHDA but had not yet degenerated.

TH and ATP levels also decreased, and motor performance was impaired in humans; we therefore tested the human genetic models of PD. After MPTP and 6-OHDA administration, apoptotic cell death continued for 14 days and 7 weeks, respectively. Delayed TZ administration (beginning on day 7 in MPTP-treated mice and in week 5 in 6-OHDA–treated rats) slowed or prevented further apoptotic cell death. In MPTP-treated mice, dopamine levels, behavioral performance, and in some cases TH levels on day 7 exceeded those on day 5 in 6-OHDA–treated rats) slowed or prevented further cell death. In MPTP-treated mice, dopamine levels, behavioral performance, and in some cases TH levels continued to decrease further with time. Administration of TZ, even after the onset of neurodegeneration, slowed cell death, and it increased TH levels, dopamine content, and motor performance compared with vehicle-treated controls (Figures 2 and 3 and Supplemental Figures 4 and 5).

Figure 4. TZ enhances Pgk activity to attenuate rotenone-impaired motor performance. (A) Schematic for experiments in panels B–F. Flies received rotenone (0.25 or 250 μM in food) with TZ (1 μM) or vehicle for 7 or 14 days. (B) Relative ATP content in the brains of w1118 flies that received 250 μM rotenone with or without TZ for 14 days. n = 6, with 200 fly heads for each treatment in each trial. (C) Climbing behavior of flies after 250 μM rotenone with TZ (1 μM) or vehicle for 7 days. Data show the percentage of flies that climbed up a tube (see Methods). n = 3, with 200 flies tested for each treatment in each trial. (D) Knockdown of Pgk in offspring of actin-Gal4 crossed with UAS-Pgk RNAi flies. Offspring of actin-Gal4 crossed with y1 v1 P [CaryP] attP2 were used as a genetic background matched control. n = 3, with RNA collected from 30 fly heads for each sample. (E) Pgk was knocked down in TH neurons by crossing UAS-Pgk RNAi flies with flies carrying the TH neuron-specific promoter (TH-Gal4) to produce TH-Pgk RNAi flies. Rotenone (250 μM) and TZ were administered as indicated for 7 days. Climbing behavior was measured on day 7. n = 8, with 200 flies tested for each treatment in each trial. (F) Pgk (UAS-Pgk) overexpression was driven by a dopaminergic neuron promoter (TH-Gal4), a pan-neuronal promoter (Appl-Gal4), a pan-cell promoter (Actin-Gal4), and a muscle–specific promoter (Mhc-Gal4). Rotenone (250 μM) was administered for 7 days, and climbing behavior was measured on day 7. n = 3, with 200 flies tested for each treatment in each trial. Data points represent individual groups of flies, and bars and whiskers show the mean ± SEM. Blue indicates controls and red indicates TZ treatment. Supplemental Table 3 shows statistical tests and P values for all comparisons. *P < 0.05, **P < 0.01, and ***P < 0.001, by Kruskal-Wallis with a Dwass-Steele-Critchlow-Fligner test (B), 1-way ANOVA with Tukey’s test (C and E), paired t test (D), and unpaired t test (F).

TZ enhances PGK activity to attenuate rotenone-induced neurodegeneration in flies. As an additional model of PD, we treated Drosophila melanogaster with rotenone, a mitochondrial complex I inhibitor implicated in sporadic PD (29). Rotenone exposure reduced brain ATP levels (Figure 4, A and B). It also disrupted motor function as tested by climbing behavior (Figure 4C). PGK is highly conserved in flies and mammals, and supplying TZ together with rotenone minimized decrements in ATP content and motor performance.

Previous studies showed that TZ increases ATP by enhancing PGK1 activity (18, 30). We knocked down Pgk in Drosophila by expressing RNAi and found that it abolished the protective effect of TZ on motor performance (Figure 4, D and E, vs. Figure 4C). Conversely, overexpression of PGK1 in Drosophila TH neurons, all neurons (Appl promoter), or all cells (actin promoter) made flies resistant to rotenone-induced behavioral defects (Figure 4F). In contrast, we found that expression in muscle was not protective. These results, together with earlier findings (18, 30), indicate that TZ protects TH neurons by activating PGK1.
Abnormal accumulation of α-synuclein, a major constituent of Lewy bodies, is a key feature of PD (36). Transgenic mice overexpressing WT human α-synuclein (mThy1-hSNCA) exhibit PD-like neurodegeneration at an advanced age (37). We began treating mThy1-hSNCA mice at 3 months of age with vehicle or TZ. When they were 15 months old, the vehicle-treated mice had substantial expression of human α-synuclein in the striatum and SNc (Figure 5, G–J) and impaired motor performance on the rotarod and pole tests (Figure 5K and Supplemental Figure 7). TZ treatment partially prevented these abnormalities.

We also tested the effect of TZ on dopamine neurons differentiated from induced pluripotent stem cells (iPSCs). LRRK2 G2019S is the most common LRRK2 mutation and is associated with approximately 4% of familial PD cases and approximately 1% of sporadic PD cases (38). Dopamine neurons derived from LRRK2G2019S iPSCs recapitulate PD features including abnormal α-synuclein accumulation (39). We studied such neurons generated from 2 patients. After 30 days of differentiation, the dopamine neurons showed no overt signs of neurodegeneration (Supplemental Figure 8). However, approximately 60% of the LRRK2G2019S dopamine neurons had accumulated α-synuclein compared with approximately 15% of dopamine neurons from healthy individuals (Figure 6, A and B). Addition of TZ for 24 hours increased the ATP content and reduced the percentage of LRRK2G2019S dopamine neurons with elevated α-synuclein accumulation (Figure 6, A–C).

In the Parkinson’s Progression Markers Initiative database, individuals with PD who were taking TZ had a reduced rate of progressive motor disability. In the past, assessment of whether an agent might affect PD has been largely limited to animal models. Three factors allowed us to assess efficacy in humans. First, TZ is a relatively commonly used drug. Second, availability of human clinical databases allowed us to test for a TZ effect. Third, tamsulosin can serve as a control for TZ. Like TZ, tamsulosin is an α1-adrenergic antagonist, and, like TZ, tamsulosin is prescribed for benign pros-
The crystal structure of TZ with PGK1 (18) suggested that a quinazoline motif for binding to PGK1. Consistent with that, tamsulosin does not have a quinazoline motif that binds to and enhances PGK1 activity. Consistent with that possibility, doxazosin (DZ) and alfuzosin (AZ) increased glycolysis in M17 cells and tyrosine hydroxylase levels in MPTP-treated mice (Supplemental Figure 9, A and B). We identified 13 men in the PPMI database who were using TZ, DZ, or AZ (TZ/DZ/AZ) (Table 1). The progression of motor disability was slowed in those patients (Figure 7, Supplemental Figure 10, and Table 1).

In contrast to TZ, DZ, and AZ, tamsulosin lacks a quinazoline motif for binding to PGK1. Consistent with that, tamsulosin did not rescue tyrosine hydroxylase levels in MPTP-treated mice (Supplemental Figure 9B). Correspondingly, tamsulosin failed to slow the motor function decline of patients enrolled in the PPMI database (Figure 7 and Table 1). These data are also consistent with the conclusion that enhanced glycolytic activity and attenuation of cell death are mediated by the effect of TZ on PGK1 and not on α1-adrenergic receptors.

The IBM Watson/Truven database shows that individuals with PD who used TZ/DZ/AZ had fewer PD-related diagnoses. To evaluate a larger number of individuals with PD and to use a different database and assessment methods, we interrogated the IBM Watson/Truven Health Analytics MarketScan Database for the years 2011 to 2016. The database includes longitudinal, deidentified diagnoses (ICD-9/ICD-10 codes) and pharmaceutical claims. We identified 2880 PD patients with PD taking TZ/DZ/AZ (4821 person-years) (Table 2). For a comparison group, we chose patients with PD who were taking tamsulosin, which controlled for use of an α1-adrenergic antagonist and for the presence of benign prostatic hyperplasia. We identified 15,409 individuals with PD who were taking tamsulosin (21,409 person years). To obtain a list of diagnostic codes associated with PD, we first identified the 497 most common diagnostic codes in the group of individuals with PD. Then, 2 neurologists who care for patients with PD identified 79 potentially PD-related diagnoses (Supplemental Table 1). Using a quasi-Poisson generalized linear model, we found that the relative risk (RR) of having any of the 79 PD-related diagnostic codes was 0.78 (95% CI: 0.74–0.82) for the TZ/DZ/AZ group relative to the individuals on tamsulosin (P < 0.00001). Of the 79 PD-related codes, we found a reduced risk in 69 codes among patients with PD who were taking TZ/DZ/AZ versus those taking tamsulosin (Figure 8A). Moreover, 41 diagnostic codes were statistically significantly decreased in the PD patients taking TZ/DZ/AZ.

**Figure 6.** TZ increases ATP content and decreases α-synuclein accumulation in iPSC-derived dopamine neurons from patients with PD. (A) iPSC-derived dopamine neurons from 2 patients with PD (subjects 12 and 13) carrying LRRK2G2019S mutations and a healthy control (subject 11). Thirty-day-old dopamine neurons were plated and were treated with TZ (10 μM) 1 or 3 days later. The neurons were studied 24 hours after addition of TZ. We observed no difference between the 2 start dates and therefore combined the data. Representative immunofluorescence images of α-synuclein (SNCA, green), TH (red), and DAPI (nuclei, blue). (B) Percentage of TH-positive neurons with cytoplasmic accumulation of α-synuclein. n = 12. (C) ATP content in control and LRRK2G2019S iPSC-derived dopamine neurons. n = 12. Bars and whiskers indicate the mean ± SEM. Blue indicates controls and red indicates TZ treatment. Supplemental Table 3 shows statistical tests and P values for all comparisons. *P < 0.05, **P < 0.01, and ***P < 0.001, by Mann-Whitney U test.
AZ versus those on tamsulosin, whereas only 2 diagnostic codes were significantly increased in the TZ/DZ/AZ group.

To estimate PD-related benefits and risks attributable to TZ/DZ/AZ versus tamsulosin, we calculated the RR for clinically relevant groupings of the 79 PD-related codes. Relative to patients with PD taking tamsulosin, those on TZ/DZ/AZ had reduced clinic and hospital visits for motor symptoms (RR 0.77; 95% CI: 0.71–0.82), nonmotor symptoms (RR 0.78; 95% CI: 0.73–0.83), and PD complications (RR 0.76; 95% CI: 0.71–0.82) (Figure 8B, Supplemental Table 1, and Supplemental Table 2). Of note, dopamine analogues do not treat PD symptoms such as dementia and neuropsychiatric manifestations (3). However, the RR for these diagnostic codes was also less than 1.0.

These data suggest that under real-world conditions, TZ and related drugs that enhance PGK1 activity reduce PD signs, symptoms, and complications.

Individuals who used TZ/DZ/AZ had a decreased risk of PD diagnosis. We also used the Truven database to test whether TZ/DZ/AZ might reduce the frequency of PD diagnoses. We identified 78,444 PD-free enrollees who were taking TZ/DZ/AZ. During a follow-up duration of 284 ± 382 days (mean ± SD), a total of 118 individuals (0.15%) developed PD. The HR from the Cox proportional hazards regression for the matched cohort was 0.62 (95% CI: 0.49–0.78; P < 0.0001).

**Discussion**

Our results indicate that in both toxin-induced and genetic models of PD in multiple animal species, enhancement of PGK1 activity slows or prevents neurodegeneration in vivo, thereby increasing dopamine levels and improving motor performance. Enhancement of PGK1 activity showed beneficial effects, even when begun after the onset of neurodegeneration. Moreover, interrogation of 2 independent databases suggested that TZ and related quinazoline agents slowed disease progression, reduced PD-related complications in individuals with PD, and reduced the risk of receiving a PD diagnosis.

Evidence from our present and earlier experiments indicates that TZ elicits its beneficial effects in PD by enhancing the activity of PGK1 and not by inhibiting the α1-adrenergic receptor. Our earlier experiments and crystal structure showed that the quinazoline motif of TZ binds PGK1 near the nucleotide binding site (18). Studies with recombinant PGK1, studies using cultured cells, and measurements in brain following in vivo delivery all revealed a biphasic relationship between the concentration of TZ and ATP levels (18). In the present study, tamsulosin inhibited α1-adrenergic receptors, but its structure lacks a quinazoline group that binds PGK1, and it did not enhance glycolysis or prevent the reduction of tyrosine hydroxylase levels in MPTP-treated mice. In contrast, 2 drugs that have a structure similar to that of TZ (DZ and AZ) enhanced glycolysis in vitro and protected MPTP-treated mice.

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**Figure 7. TZ and related drugs slow the progression of motor defects for patients with PD enrolled in the PPMI database.** Movement Disorder Society–Unified Parkinson’s Disease Rating Scale (MDS-UPDRS) Part 3 (motor) scores for patients with PD in the PPMI database. Patients were taking TZ/DZ/AZ (blue, n = 13), tamsulosin (green, n = 24), or none of these drugs (red, n = 269). Data represent scores upon entry into the PPMI database through approximately 1 year and include all measures between those times. All patients taking these drugs were men prescribed TZ/DZ/AZ or tamsulosin, without breaks for benign prostatic hyperplasia or undefined urological problems. Lines are plotted from linear mixed-effect regression analyses. By maximum likelihood estimation, TZ/DZ/AZ differed from controls (P = 0.012).
Knockdown of *Pgk1* in *Drosophila* TH neurons abolished the protective effect of TZ. Overexpression of PGK1 in flies, mice, and fish phenocopied the effects of TZ (18, 30). TZ was active in *Drosophila melanogaster*, which do not have α1-adrenergic receptors. Allosteric and covalent regulatory mechanisms have been identified for most glycolytic enzymes. For example, insulin-stimulated deacetylation increases PGK1 activity, and disrupting that regulation results in glycolytic insufficiency (42).

Previous work has identified numerous genetic mutations and several environmental factors that cause or predispose individuals to PD (11–16, 43). As indicated above, reduced energy metabolism and decreased ATP levels are a feature of many of these environmental and genetic factors, as is aging, the major PD risk factor. Therefore, enhancing glycolysis might slow progression in PD of several etiologies.

This study does not reveal how enhanced glycolysis slows neurodegeneration and progression in PD. However, the increased ATP levels produced by TZ may be key. ATP has properties of a hydrotopie; it can prevent aggregate formation and dissolve previously formed protein aggregates (44, 45). Moreover, the transition between aggregate stability and dissolution occurs in a narrow range at physiological ATP concentrations. We speculate that by elevating ATP levels, TZ facilitates the solubilization of aggregates, including α-synuclein, and prevents the neurodegeneration of PD. However, other mechanisms are also possible including ATP-dependent disaggregases and chaperones (such as hsp90) that reduce apoptosis (18, 44, 45).

**Figure 8.** TZ and related drugs reduce symptoms as assessed by diagnostic codes for patients with PD in the Truven/IBM Watson clinical database. Data are from the Truven Health Marketscan Commercial Claims and Encounters and Medicare Supplemental Databases for the years 2011–2016. Patients had a diagnosis of PD and were prescribed TZ/DZ/AZ or tamsulosin for at least 1 year. We assessed RRs for 79 previously identified PD-related diagnostic codes. (A) RR for 79 PD-related diagnostic codes for patients taking TZ/DZ/AZ versus tamsulosin. Yellow indicates a statistically significant difference in risk between TZ/DZ/AZ and tamsulosin (P < 0.05) determined by a generalized linear model with a quasi-Poisson distribution. (B) RR for the categories of PD-related diagnostic codes for patients taking TZ/DZ/AZ versus tamsulosin. Data represent the mean and 95% CIs.
tamsulosin. In PD, neurons that have not yet degenerated almost certainly have compromised cellular function (28), and we speculate that TZ/DZ/AZ improved their functional integrity.

Results from this study, together with earlier data, led us to 3 additional speculations. First, TZ is already used clinically, and in this regard, it is interesting that several studies reported that TZ improved glucose metabolism in patients with diabetes (48, 49). That observation has gone unexplained. We speculate that stimulation of PGK1 activity might have been responsible. Consistent with that conjecture and with the conclusion that an α₁-adrenergic receptor antagonistic effect was not responsible, we found that α₁-adrenergic receptor antagonists structurally unrelated to TZ lacked that effect. In addition, disruption of the α₁B-adrenergic receptor in mice had an effect opposite of that induced by TZ (50). Second, loss-of-function PGK1 mutations cause recessive hemolytic anemia, myopathy, seizures, and intellectual disability. However, other studies have reported Parkinsonism (51, 52), and the authors speculated that reduced ATP generation in the SNc may have been responsible. Third, PD occurs approximately 1.5 times more frequently in men than in women (53). Why males are more predisposed to PD is unknown. However, it may be worth noting that the PGK1 gene is located on the X chromosome. Thus, the consequences of DNA sequence variations that could subtly reduce PGK1 levels or activity might more often manifest in men than in women.

Our findings that TZ increased glycolysis and prevented progressive neurodegeneration suggest that energy deficits might either be a pathogenic factor in the pathogenesis of PD or predispose individuals to PD in the presence of environmental or genetic etiologies (11, 16). These findings identify a protein and a pathway that might be targeted to slow or prevent neurodegeneration in PD and potentially other neurodegenerative diseases with altered energy balance (54).

Methods

The Supplemental Methods contain information on the materials, reagents, experimental procedures, and analysis methods used in this study.

Statistics. For experiments to quantify animal behavior and for sample collections, the experimenters were blinded to the genotype and intervention, and the studies were conducted by 2 different experimenters. The number of animals studied was based on our past experience and preliminary data. In all figures, data points are from individual mice and rats or groups of flies. We did not exclude any data points from this study. Data in the figures indicate the mean ± SEM. Blue indicates controls, and red indicates TZ treatment. Statistical significance for comparisons between data sets was primarily done with nonparametric tests. For studies of fly motor performance, our previous studies showed that within a group of flies (15–50 flies for 1 data point), the data fit a Gaussian distribution. Moreover, data for multiple groups of flies also fit a Gaussian distribution. Therefore, parametric tests were used to evaluate statistical significance in studies using flies, and ANOVA evaluations were 1 way. All statistical tests were 2 tailed. A P value of less than 0.05 was considered statistically significant. Supplemental Table 3 shows the statistical tests used for all data and the resulting P values for comparisons.

Study approval. All experiments using mice and rats were approved by the IACUC of Peking University in Beijing (approval nos. LSC-Liul-1 and LSC-Liul-2).

Author contributions

RC designed and performed many of the experiments and analyzed the data. YZ and YL performed fly and mouse studies. IFC and AC performed LRRK2 iPSC studies. AR designed the LRRK2-mutant iPSC studies. JES and PMP performed the analysis of the Truven database. YY, ZC, and WS performed parts of the animal behavior tests. JLS and NSN performed the PPMI database analysis. YH, CZ, and LG provided technical support and data analysis. XJ designed experiments. MJW designed experiments and analyzed data. LL designed and supervised experiments and analyzed data. RC, AR, MJW, and LL wrote the manuscript.

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