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PTEN overexpression suppresses proliferation and differentiation and enhances apoptosis of the mouse mammary epithelium

Joëlle Dupont, Jean Pierre Renou, Moshe Shani, Lothar Hennighausen, and Derek LeRoith

The phosphatase PTEN regulates growth, adhesion, and apoptosis, among many other cell processes. To investigate its role during mouse mammary gland development, we generated MK-PTEN, a transgenic mouse model in which human PTEN is overexpressed in ductal and alveolar mammary epithelium during puberty, pregnancy, lactation, and involution. No obvious phenotype was observed in mammary tissue of pubescent virgin mice. However, MK-PTEN females could not lactate normally, and ~30% of pups died, with survivors exhibiting growth retardation. Transgenic offspring nursed by wild-type foster mothers, conversely, developed normally. This phenotype is consistent with a reduced number of alveolar epithelial cells due to a decrease in cell proliferation and an increase in apoptosis. Using mammary-enriched cDNA microarrays, we identified several genes that were preferentially expressed in MK-PTEN mammary tissue, including the IGF-binding protein-5 (Igfbp5) gene, and others whose expression was reduced, including the genes for c-Jun amino-terminal kinase. Secretory epithelial cell differentiation was impaired, as measured by the expression of specific milk protein genes. MK-PTEN mice also exhibited a 50% decrease in the phosphorylation state of Akt. Taken together, these results suggest that PTEN controls mammary gland development and, consequently, lactation.

Shc and FAK and inhibition of the mitogen-activated protein kinase (MAPK) pathway have also been reported (13, 14).

Stimulation of the IGF-1, insulin, EGF, and PDGF receptor tyrosine kinases activates both PI3K/Akt and MAPK pathways. These two signaling pathways can also be activated by estrogen (15), prolactin (16), and growth hormone (17), which all regulate mammary gland development. Several studies have shown that in the mammary gland epithelium, MAPK and PI3K/Akt regulate cell proliferation (18), cell survival (19), and apoptosis (20). When a constitutively active mutant of Akt was expressed specifically in the mouse mammary gland, delays in involution and in the onset of apoptosis were observed (21, 22). Thus, further characterization of proteins that regulate the PI3K/Akt and the MAPK signaling pathways is important for understanding both the regulation of normal mammary gland development and the process of mammary tumorigenesis.

In the present study, we have examined the role of PTEN in mammary gland development using transgenic mice in which PTEN is overexpressed specifically in mammary epithelium using the MMTV-LTR. We show that the overexpression of PTEN decreases mammary epithelial cell proliferation and increases apoptosis, thereby resulting in the formation of alveolar hypoplasia. In addition we demonstrate that functional differentiation of mammary epithelium is impaired. Consequently, the process of lactation is affected, and the newborn pups either die or exhibit growth retardation. At the molecular level, we show that the overexpression of PTEN in mammary gland significantly inhibits the Akt signaling pathway, but that it does not affect the MAPK pathway. Furthermore, using oncogene- or mammary-enriched cDNA microarrays, we identify new target genes for PTEN, including the c-Jun NH2-terminal kinase/stress-activated protein kinase–associated protein-1 (JNK/SAPK-1) and the IGF-binding protein-5 (IGFBP-5) genes, that could explain the reduction of the cell proliferation and the increase in the apoptosis of the MK-PTEN epithelial cells, respectively.

Methods

Generation of MK-PTEN transgenic mice. The MMTV-LTR promoter was used to generate transgenic mice expressing PTEN specifically in the mammary gland. The MMTV-KCR vector was kindly provided by Sophia Y. Tsai (Baylor College of Medicine, Houston, Texas, USA) (23). A 1,218-bp blunt-ended EcoRI fragment of the human PTEN cDNA, kindly provided by Kaz Matsumoto (National Institute of Dental and Craniofacial Research, NIH), was cloned into the blunt-ended EcoRI site of the MMTV-KCR vector. The MMTV-KCR-PTEN plasmid was then digested with Acc65I and XbaI to liberate a 4,029-bp fragment. This fragment was microinjected into fertilized eggs derived from C57BL/6J mice. These eggs were implanted into CD-1 foster mothers, and integration of the transgene in the offspring was assessed by Southern blot and PCR analyses, as described below.

Genotyping and animals. Genotypic analyses by both PCR and Southern blot were performed on genomic DNA isolated from the tails of potential founder mice. Primers used for the detection of the transgenic product were GLE3-F (5′-TGGTTGGTGGCTGCTCATC-3′) and PTEN-R (5′-CAAAGGATAGGGCAGAATC-3′). Southern blots were performed using a 436-bp fragment of the PTEN cDNA as a probe for transgene detection on genomic DNA that had been digested with BamHI. Similar phenotypes of mammary gland hypoplasia were observed in two independent transgenic lines. Thus, the results reported are primarily from the transgenic line 7011, unless otherwise specified. Mice were kept under a 12-hour light/dark cycle and were fed standard diets. Procedures were conducted in compliance with the guidelines of the Animal Care and Use Committee of the National Institute of Diabetes and Digestive and Kidney Diseases (NIH). To determine the pregnancy stage, the mice were mated and inspected for the presence of vaginal plugs in the mornings. The day of the vaginal plug was counted as day 0 of pregnancy.

Isolation and primary culture of mammary epithelial cells. Primary mammary epithelial cells were prepared from wild-type (WT) and MK-PTEN mice pregnant for 16–19 days, as described previously (24). Cells were resuspended in DMEM with phenol red containing 10% FBS, 100 units/ml penicillin, 100 mg/ml streptomycin, 10 ng/ml EGF (Upstate Biotechnology Inc., Lake Placid, New York, USA), and 5 µg/ml insulin (Sigma-Aldrich, St. Louis, Missouri, USA). After 48 hours, the medium was removed and fresh medium was added with or without the IGFBP-5–blocking peptide for 24 hours. This IGFBP-5–blocking peptide (25) was kindly provided by D.R. Clemmons (Department of Medicine, University of North Carolina, Chapel Hill, North Carolina, USA).

Determination of caspase-3 activity. We determined caspase-3 activity by measuring the absorbance at 405 nm, following cleavage of the synthetic substrate, Ac-Asp-Glu-Val-Asp-chromophore p-nitroaniline (Ac-DEVDPNA). Briefly, mammary epithelial cells from WT and MK-PTEN mice incubated in the presence or absence of the IGFBP-5–blocking peptide were lysed on ice for 10 minutes in cell lysis buffer provided in the Clontech ApoAlert Caspase-3 colorimetric assay kit (Clontech Labs, Palo Alto, California, USA). Lysates were then incubated with 50 µM Ac-DEVDPNA in the reaction buffer containing 0.1 M HEPES, 20% glycerol, 10 mM DTT, and protease inhibitors. The lysates were maintained at 37°C for 1 hour and then analyzed in an ELISA plate reader. Levels of relative p-nitroaniline were normalized against the protein concentration of each extract.

RNA isolation and Northern blots. Total RNA from the left inguinal mammary gland (gland number 4) was extracted at the indicated times of development using TRIzol reagent according to the manufacturer’s instructions (Invitrogen Corp., Carlsbad, California, USA). Twenty micrograms of total RNA was separated in 1.5%
formaldehyde gels and blotted on nylon membranes. Expression of milk protein RNA and keratin 18 was analyzed as described previously (26). Expression of PTEN was analyzed using the entire 1.2-kb human PTEN cDNA as a probe. RNA levels were quantified using a Phospholmager apparatus (Fujifilm Medical Systems USA Inc., Stamford, Connecticut, USA). The integrity of different transcripts was assessed using the human RNA 18S probe from Ambion Inc. (Austin, Texas, USA).

**RT-PCR analysis of mRNAs.** Total RNA from mammary glands of WT and MK-PTEN mice (day 2 of lactation) was extracted as described above. One microgram of DNase-treated RNA of each group (WT and MK-PTEN) was reverse-transcribed using 2.5 U/ml of murine leukemia virus reverse transcriptase, 2.5 mM MgCl2, 1 mM dNTP, 2.5 mM oligo-dT, and 1 U/ml RNase inhibitor. The same reverse transcription products were subjected to PCR in standard conditions in the presence of specific primers: sense PTEN, 5′-GAGATATATCTCTC-CAATTCC-3′; antisense PTEN, 5′-CTTGTGACATAGACTTC-3′; sense WAP, 5′-CAGACCGTATCAGGCTTG-3′; antisense WAP, 5′-TAGACCGATTTAGAACGCT-3′; sense IGFBP-5, 5′-TGTTGCTGCTGATGACTTG-3′; antisense IGFBP-5, 5′-GAACCTTGGAGGAAAGCTT-3′; sense e-cad, 5′-CTGACTTTGCGTTTGGCAGG-3′; antisense e-cad, 5′-AAACCTGTGGCAGGTGCT-3′; sense carbonic anhydrase 3, 5′-ACATGTCTGCTGAGAAG-3′; antisense carbonic anhydrase 3, 5′-TGGAAGCCTCAGACATAC-3′; sense cdc2, 5′-TGCAGACATCTGACTAC-3′; antisense cdc2, 5′-AGATCGGGCCAGAAGCAGT-3′; sense β-actin, 5′-ACGGAAACGACTTATCAT-3′; antisense β-actin, 5′-CCATATCCCTCTGACCTG-3′; sense JNK/SAPK-1, 5′-TAACACGATGGCTTTTGAGG-3′; antisense JNK/SAPK-1, 5′-TTGCAAGACACCATAGAGACG-3′. The identity of all PCR products was verified by sequencing.

**Western blot analysis.** Proteins from mammary glands were extracted and processed for immunoblotting exactly as described previously (27). The resulting immunoreactive bands were subjected to densitometry and quantitated with MacBas V2.52 software (Fujifilm Medical Systems USA Inc.).

**Phosphatase assay.** Mammary gland protein extracts containing 500 μg of protein were immunoprecipitated overnight using a rabbit polyclonal anti-PTEN antibody (C20) developed against the 384DSDPENEPPD-EDQHTQITKV403 region of the human PTEN molecule (Zymed Laboratories Inc., South San Francisco, California, USA), and the phosphatase assay was performed exactly as described previously (28). The absorbance at 630 nm was recorded in an ELISA plate reader. A standard curve was performed with each assay, and the amount of free phosphate was calculated from the standard curve data.

**Whole mounts, histology, and immunohistochemistry.** Whole-mount analyses were performed as previously described (26). To prepare mammary gland sections, the left inguinal glands were fixed for 4 hours in 4% phosphate-buffered paraformaldehyde, then transferred to 70% ethanol. Tissues were embedded in paraformaldehyde and then cut into 5-μm sections. PTEN immunostaining was performed as described previously (29) using the mAb 6H2.1. This antibody was raised against the last 100 C-terminal amino acids of human PTEN. Specificity and characterization of the PTEN antibody 6H2.1 have been previously demonstrated by Western blot and immunohistochemistry on cell lines with known PTEN expression status (29). IGFBP-5 immunostaining was performed in the same conditions as the PTEN immunostaining. Incubating the sections with normal serum in the absence of antibody resulted in no signal (data not shown).

**Cell proliferation and cell death assays.** Two hours before sacrifice, mice were injected with 20 μl/g of body weight of cell labeling reagent (Amersham Biosciences, Piscataway, New Jersey, USA). Mammary glands were prepared and fixed in 4% paraformaldehyde for 4 hours. The tissues were then embedded in paraffin. For detection of incorporated BrdU, 5-μm sections were processed according to the manufacturer’s instructions (Amersham Life Science). On adjacent sections from the same samples, apoptotic cells were detected by TUNEL assay with ApopTag Plus Peroxidase, according to the manufacturer’s instructions (Intergen Co., Purchase, New York, USA). Between 2,000 and 3,000 epithelial cells were counted from three to four different sections of a sample, and the averages of percentages of labeled cells were calculated.

**Microarray analyses.** A cDNA array (mammochip) containing 3,500 genes assembled from various mammary gland cDNA libraries (NbMMG, NMLMG, CGAP Mam1 to -6) was generated and printed on glass slides (K. Miyoshi and L. Hennighausen, unpublished observations). In addition, the National Cancer Institute oncochip containing 2,600 genes was used. RNA was prepared from mammary tissue from MK-PTEN and control mice at day 2 after parturition (about 12 weeks of age). For each experiment, samples from two different mice were pooled. For fluorescence labeling, 20 μg of total RNA was reverse-transcribed in the presence of 300 U of SsII (Life Technologies Inc., Rockville, Maryland, USA) and labeled with cy3-dUTP and cy5-dUTP (NEN Life Science Products Inc., Boston, Massachusetts, USA). Samples were combined, purified, and concentrated with Microcon YM30 columns (Millipore Corp., Bedford, Massachusetts, USA). Slides were prehybridized for 1 hour and hybridized overnight at 42°C in 25% formamide (30). Slides were washed as described (31) and scanned on a GenePix scanner (Axon Instruments Inc., Foster City, California, USA). The images were analyzed by GenePix Pro 3.0 (Axon Instruments Inc.). Each
experiment was repeated with the cy3-dUTP and cy5-dUTP labeling switched. The data were analyzed with the mAdb tools of the National Cancer Institute–Center for Information Technology µArray Center (32) with some modifications. The differentially expressed genes were selected by signal intensity fivefold higher than background in at least one channel and an expression ratio of 3:0.3 or higher. These criteria were also used in the experiments with the labels reversed.

Statistical analyses. Results are expressed as the mean ± SEM. Statistical analysis of the data was performed using a Student’s $t$ test.

Results

Generation of transgenic mice overexpressing PTEN specifically in the mammary gland. To determine the role of PTEN in the developing mammary gland, we overexpressed the human PTEN cDNA transgene (hPTEN) specifically in the mammary epithelium using the MMTV-LTR. The transgenic construct used is shown in Figure 1a. Six founders with transgene integration were obtained. A representative Southern blot and PCR derived from BamHI-digested genomic DNA are shown for two founders in Figure 1b. Of these six founders, five transmitted the transgene through the germline. These founders were then bred with WT mice to generate females to test the expression of PTEN.

Expression of PTEN throughout the development of the mammary gland in MK-PTEN and WT mice. We next analyzed each of the transgenic lines for the expression of PTEN. Total RNA was isolated from biopsies of inguinal mammary glands (number 4) from transgenic and WT littermates 10 days after the onset of lactation, when the MMTV-LTR has been shown to be highly active. In the mammary glands, we detected PTEN as a major transcript of 2.5 kb and two other mRNA species of 5 and 3 kb, the latter two being weakly expressed (data not shown). Two founders, F7011 and F7044, highly expressed PTEN at day 10 of lactation, as compared with the WT mice. The profile of PTEN mRNA expression during the development of the mammary gland was determined in the founder F7011 (Figure 1c). In MK-PTEN mice, the level of PTEN mRNA progressively increased, as compared with WT littermates, from the age of 6 weeks (virgins), through the beginning of puberty and throughout pregnancy, peaking at
day 10 of lactation and then decreasing during involution of the mammary gland (Figure 1c). These findings are in agreement with the hormonally regulated expression profile of the MMTV-LTR (23).

To determine whether the differences in mRNA were also reflected at the level of protein, we performed Western blot analyses on mammary gland extracts prepared from MK-PTEN and WT female mice at different stages of development. In 3- and 5-week-old virgins, two bands of approximately 54 and 58 kDa in size were detected in both WT and MK-PTEN mice. These two bands are also detected when using two other PTEN antibodies, suggesting that they are specific and correspond to two isoforms of the PTEN protein (data not shown). At the age of 6 weeks, the level of PTEN protein is about twofold higher in MK-PTEN than in WT mice (Figure 1d). This difference between transgenic and WT mice progressively increases throughout all stages of pregnancy and becomes maximal at day 10 of lactation, then decreases during involution (Figure 1d). These results show that PTEN mRNA and protein expression profiles closely correlate throughout development of the mammary gland (Figure 1, compare c and d). To determine whether the transgene was expressed in the expected tissue-specific manner, protein extracts from different tissues of MK-PTEN mice at day 20 of pregnancy and day 10 of lactation were immunoblotted for PTEN. PTEN expression was observed only in the mammary gland (data not shown). This protein expression profile is in agreement with that of other MMTV-driven transgenes (21).

We used immunohistochemistry to localize PTEN within the mammary gland of WT and transgenic mice (Figure 2). In WT mice, PTEN immunostaining is very low in virgin, pregnant, and lactating mice. In 7-week-old virgins, higher PTEN staining is observed in ductal epithelial cells in MK-PTEN as compared with WT littermates (Figure 2). Despite the higher expression of PTEN within the virgin MK-PTEN mammary gland after 6 weeks (Figure 1d), we did not detect any changes in ductal growth or in cellular morphology within the nulliparous mammary gland at the ages of 7 or 9 weeks, as determined by whole mounts and histologically stained paraffin-embedded sections (data not shown). During early pregnancy, late pregnancy, and lactation, about 70%, 80%, and 95% of the lobuloalveoli in MK-PTEN mice were immunoreactive for PTEN, respectively. PTEN staining is very weak in the ductal

Figure 2
Immunohistochemical localization of PTEN in the mammary gland of MK-PTEN and WT mice. Sections from mammary glands removed from virgin, 9-day-pregnant, 14-day-pregnant, and 2-day-lactating mice were analyzed by immunohistochemistry as described in Methods. PTEN-expressing lobuloalveoli and ductal epithelial cells are indicated by black arrows and white arrows, respectively.

Figure 3
PTEN phosphatase activity and phosphorylation state of Akt and MAPK in mammary glands of MK-PTEN and WT mice. (a) Mammary gland protein extracts from MK-PTEN and WT mice (500 µg protein per sample) were subjected to immunoprecipitation with the PTEN polyclonal C20 antibody, and the phosphatase activity was measured as described in the Methods. The phosphatase activity in MK-PTEN and WT is represented as mean ± SD in the graph (n = 4 MK-PTEN and WT mice for each group except in virgin and involution, where n = 2 mice in each group). *P < 0.05, ***P < 0.001. (b) Akt and Erk1/2 activation in mammary gland extracts from MK-PTEN and WT mice at 9 and 14 days of pregnancy. Extracts containing 120 µg of protein were analyzed by immunoblotting with anti-Akt and anti-Erk1/2 antibodies that either recognize the phosphorylated form (pAkt or pErk1/2) or equally recognize the phosphorylated and dephosphorylated forms, i.e., total levels (Akt and Erk1/2). The blots were also analyzed for phospho-FKHR immunoreactivity. These results are representative of at least two identical experiments.
epithelial cells at each of these stages, as shown in Figure 2, suggesting that overexpression of PTEN in the mammary gland of MK-PTEN mice is more specific to the lobuloalveoli. Thus, the immunohistochemical results are in close agreement with those obtained from PTEN Northern and Western blots.

**PTEN phosphatase activity and phosphorylation of Akt and MAPK in the mammary gland of MK-PTEN mice.** To determine the functionality of PTEN in the mammary gland of MK-PTEN mice, we measured the PTEN phosphatase activity. We also determined the phosphorylation state of Akt and Erk1/2, which are two known targets of PTEN. PTEN was immunoprecipitated from mammary gland extracts at different stages, and phosphatase activity was measured using PIP3 as substrate. As shown in Figure 3, PTEN phosphatase activity levels remain relatively constant throughout mammary gland development in WT mice. In striking contrast, PTEN phosphatase activity progressively increases throughout development, pregnancy, and lactation in MK-PTEN mice and decreases after involution (Figure 3a). We also found that during early and middle pregnancy (9 and 14 days), the phosphorylation state of Akt and FKHR was decreased by about 50% in MK-PTEN mice, as compared with WT mice (Figure 3b). However, the level of Erk1/2 phosphorylation was similar in WT and MK-PTEN mice (Figure 3b). Thus, these results suggest that PTEN functions through the PI3K/Akt pathway in the mammary gland of MK-PTEN mice.

Decreases in epithelial cell number and milk protein expression in the mammary gland of MK-PTEN mice are associated with a defect in lactation. The primary biologic function of the mammary gland is to provide nourishment to suckling young in the form of milk. To determine whether PTEN is involved in the functionality of the mammary gland, we examined the ability of MK-PTEN mice to adequately nourish pups (measured by pup weight). We observed that 2 days after the onset of lactation, 30% of the pups from MK-PTEN females died (data not shown), and the survivors exhibited significant growth retardation until 9 days old (Figure 4a). However, if, immediately after the delivery, WT female mice nursed MK-PTEN pups, the pups survived and grew normally (Figure 4a). These findings suggest that the MK-PTEN newborns are viable but die of malnutrition during nursing. To understand the origin of these physiological perturbations, we performed histological examinations on the mammary glands from MK-PTEN mice. We found that the number of lobuloalveoli in MK-PTEN mice was reduced by 30–50% in pregnant MK-PTEN mice, as compared with the WT littermates (Figure 2). We also investigated whether these lobuloalveoli were functional by measuring the expression levels of various milk mRNAs during early and middle pregnancy (Figure 4b). At 9 days of pregnancy, WDNM1 and β-casein mRNA levels were reduced by...
about 40% in MK-PTEN as compared with WT littermates, as normalized to 18S RNA levels. Similar results were observed for the mRNA expression of Stat5a and 5b, lactalbumin, and WAP after 14 days of pregnancy (Figure 4b). Keratin 18 serves as a marker of the epithelial cells. No differences in litter size were observed between MK-PTEN and WT mice (data not shown), although MK-PTEN male mice expressed higher levels of PTEN in certain reproductive organs (data not shown).

Mammary epithelium overexpressing PTEN exhibits decreased proliferation and increased apoptosis. The observed decrease in the epithelial content of mammary glands harboring PTEN-overexpressing epithelium may reflect a decrease in the rate of epithelial proliferation and/or an increase in the level of epithelial apoptosis. We tested these two possibilities by quantifying BrdU incorporation and by TUNEL assays, respectively. To assess the levels of epithelial cell proliferation in mice overexpressing PTEN specifically in the mammary gland, mice were labeled with the BrdU. Based on immunohistochemical analysis, the number of BrdU-positive nuclei in mammary epithelium overexpressing PTEN appeared to be lower than that in mammary epithelium of WT mice (Figure 5). To quantify this decrease, the percentage of BrdU-positive nuclei relative to the total number of nuclei was calculated at three different stages of pregnancy (P-9, P-14, and P-20; Figure 5b, left). The percentage of BrdU-positive epithelial nuclei was decreased by approximately 50% in mammary epithelium overexpressing PTEN compared with that in WT mice. In accordance with these results, we also found that cyclin D1 immunoreactivity was significantly reduced in mammary epithelium of MK-PTEN as compared with WT mice (data not shown). These data demonstrate that the mammary epithelium of MK-PTEN mice is indeed hypoplastic at all stages of pregnancy.

To assess the level of apoptosis in mammary epithelium of MK-PTEN mice, TUNEL analysis was performed on sections prepared from MK-PTEN and WT mice at different stages of pregnancy. TUNEL-positive nuclei were detected by immunohistochemistry. Based on immunohistochemical analysis, the percentage of TUNEL-positive nuclei in mammary epithelium of MK-PTEN mice was approximately two- to threefold greater than that observed in the mammary epithelium of WT mice (Figure 5b, right). Thus, these data suggest that the lower cell density in mammary epithelium of MK-PTEN mice is due to both a decrease in cell proliferation and a concomitant increase in apoptosis.

Profiling gene expression in MK-PTEN transgenic mice. To further understand the developmental defects in the MK-PTEN transgenic mice, we profiled the gene expression pattern in mammary tissue 2 days after parturition. Since most genes expressed in mammary tissue are absent from commercial cDNA arrays, we generated an array, the so-called mammochip, that contains 3,600 cDNAs selected from different cDNAs of mammary origin. While 11 genes were expressed preferentially in mammary tissue from MK-PTEN mice, nine genes were preferentially expressed in WT tissue (Table 1). Out of the 11 genes overexpressed in MK-PTEN mammary tissue, six corresponded to known genes. Most notably, the expression of the IGFBP-5 gene was 26 times higher in MK-PTEN mammary tissue. Interestingly, a database search demonstrated that expression of IGFBP-5 is preferentially seen in mammary tumors of transgenic mice. The expression of the carbonic anhydrase 3 was also increased fivefold in MK-PTEN mammary epithelium. The higher expression of both IGFBP-5 and carbonic anhydrase 3 genes in MK-PTEN than in WT mice was confirmed by RT-PCR (Figure 6a). We used immunohistochemistry to localize IGFBP-5 within the mammary gland of WT and MK-PTEN mice (Figure 6b). IGFBP-5 immunostaining is very high in epithelial cells of MK-PTEN mice at day 2 of lactation as compared with the WT littermates (Figure 6b).

Expressed sequence tag database searches demonstrated that the genes overexpressed in MK-PTEN tissue were also highly expressed in mammary tumors of transgenic mice. Three known genes, those encoding
δ-casein, JNK/SAPK-1, and WAP, were expressed 27-, 15-, and 7-fold more highly, respectively, in WT tissue (Table 1). Again, these results were confirmed by RT-PCR (Figure 6a). The genes and gene products corresponding to the differentially expressed sequence tags have not been investigated. We further monitored the expression of 2,600 known genes, including many oncogenes, on a commercial oncochip. While six genes were expressed preferentially in mammary tissue from MK-PTEN mice, six other genes were preferentially expressed in WT tissue (Table 1). Most of the identified genes were involved in the cell proliferation or apoptosis. Indeed, the cdc2, complement component 3, thymosin β 10, and cell cycle G0/G1 genes were expressed 11-, 6-, 5-, and 3-fold more highly, respectively, in MK-PTEN mammary tissue, whereas the high-mobility group protein 1 isoform C (HMG-IC), glucocorticoid-induced leucine zipper (GILZ), and receptor her3 genes were expressed 13-, 4-, and 4-fold more highly, respectively, in WT mammary tissue (Table 1).

IGFBP-5 is involved in the apoptosis observed in the MK-PTEN mammary epithelial cells. IGFBP-5 is an IGF-binding protein involved in vivo in mammary apoptosis during the involution stage. To determine the role of IGFBP-5 in the apoptosis observed in MK-PTEN mammary epithelium, we isolated and cultured mammary epithelial cells from WT and MK-PTEN mice. After 48 hours of culture, cells were incubated for 24 hours in the presence or absence of an IGFBP-5–blocking peptide, and we determined the apoptosis level by tunnel assay and measured the caspase-3 activity. The percentage of apoptotic cells and the caspase-3 activity were about three and four times higher in MK-PTEN mammary epithelial cells than in WT, respectively (Figure 6c). Interestingly, the IGFBP-5–blocking peptide (5 or 10 µg/ml) reduced by about 40% (P < 0.05) the higher apoptosis level observed in the MK-PTEN mammary epithelial cells (Figure 6c). Thus, these results demonstrate that the apoptosis observed in mammary MK-PTEN epithelial cells is due in part to IGFBP-5.

**Discussion**

To investigate the role of PTEN in the development of the mammary gland, we created transgenic mice (MK-PTEN) that overexpress an active PTEN specifically in the
mammary epithelium under the control of the MMTV-LTR. MK-PTEN mice overexpress PTEN from puberty, throughout pregnancy and lactation, and during the first 6 days of involution. Ductal development in MK-PTEN mice during puberty was normal, possibly because the level of transgenic PTEN is only twofold higher than in WT littermates. Furthermore, PTEN immunostaining was stronger in the alveolar epithelial cells than in the ductal epithelium of MK-PTEN mammary glands. However, during the early stages of pregnancy, the mammary tissue of MK-PTEN mice exhibits a decrease in the number of alveolar epithelial cells, as compared with that of their WT littermates. This reduction is accompanied by a significant defect in lactation, such that 30% of newborn pups die of malnutrition during the first 2 days of nursing and the survivors exhibit growth retardation.

Specific proteins have been identified that play important roles during the different stages of mammary gland development (reviewed in ref. 33). Mice deficient in the estrogen receptor exhibit a severe impairment in ductal outgrowth and branching receptor (34), and mice deficient in the progesterone receptor have retarded alveolar development (35). These findings suggest that the estrogen and progesterone receptors play vital roles in mammary gland development during puberty and early pregnancy. Similarly, mammary epithelium does not form in the absence of the prolactin receptor (24). Reduced alveolar proliferation during pregnancy was also observed in the MK-PTEN mice. In contrast to the Prolactin receptor- and C/EBPβ-null mice, mammary epithelium in the MK-PTEN mice acquired a partially differentiated state, as indicated by the expression of some milk protein genes. However, loss of normal expression of δ-casein and WAP, as determined by Northern blot and cDNA microarrays, suggests that MK-PTEN mammary did not acquire functional differentiation. In particular, expression of the δ-casein gene reflects the differentiation status of mammary epithelium. While other casein genes are activated around midpregnancy (36), expression of the δ-casein gene occurs at parturition. Thus, the reduced levels of WAP and δ-casein mRNA in MK-PTEN tissue reflect that this tissue has not undergone full differentiation. Furthermore, the expression of the carbonic anhydrase found preferentially in mammary stroma was significantly higher in MK-PTEN mammary tissue, reflecting again a reduced alveolar development. Thus, the tumor suppressor PTEN protein appears to play a critical role in the control of mammary cell proliferation, and to some extent in cell differentiation.

Several studies show that the proliferation of glioblastoma cell lines and breast cancer cell lines is significantly inhibited by the overexpression of PTEN (reviewed in ref. 37). This antiproliferative effect is dependent on the lipid phosphatase activity of PTEN. In the mammary tissue of MK-PTEN mice, we show that the reduction of epithelial cell proliferation is paralleled by a decrease in the phosphorylation state of Akt. To date, PTEN is the only known enzyme that dephosphorylates the D3 position in inositol phospholipids, suggesting that PTEN may function to oppose PI3K and Akt signaling pathways. It has been shown that constitutively active forms of PI3K and Akt can rescue PTEN-induced suppression of cell proliferation, suggesting that an Akt-mediated signaling plays a major role in PTEN-sensitive cell proliferation (38). The MAPK cascade is another growth factor-mediated signaling pathway. MAPK-mediated growth signaling can oppose the effects of PTEN overexpression on proliferation or migration in different cell lines (39). However, it has also been shown that overexpression of PTEN markedly reduces the phosphorylation of Erk2 in U87MG cells (13). This inhibition of MAPK by PTEN could be attributable to effects of Shc

![Figure 6](image-url)

(a) mRNA expression determined by RT-PCR of some genes up- or downregulated in mammary tissue of MK-PTEN mice as compared with that of WT mice. These results are representative of two WT and two MK-PTEN mice different from those used to perform the microarray experiment. (b) Immunohistochemical localization of IGFBP-5 in the mammary gland of MK-PTEN and WT mice. Sections from mammary glands removed from mice lactating for 2 days were analyzed by immunohistoechemistry as described in Methods. (c) An IGFBP-5-blocking peptide reduces the level of the apoptotic mammary epithelial cells of MK-PTEN mice. Following incubation of mammary epithelial cells from WT and MK-PTEN mice in DMEM with serum for 48 hours, the cells were untreated or treated with 5 or 10 µg/ml of an IGFBP-5-blocking peptide. Apoptotic cells were detected by TUNEL assay and caspase-3 activity was determined by the ApoAlert colorimetric assay as described in Methods. The data are representative of three independent experiments performed in triplicates.
eral T and B cells is also decreased in PTEN+/− mice resist apoptosis (46). Fas-induced apoptosis of peripheral T cells (PTEN−/−) hyperproliferate and show that the T cells (PTEN−/−) hyperproliferate and cell-specific deletion of the PTEN gene, Suzuki et al. (41). Using the Cre-loxP system to generate a T cell-specific deletion of the PTEN gene in MK-PTEN mice, fibroblasts are highly resistant to apoptotic stimuli agrees with findings that PTEN-null mouse embryo fibroblasts are resistant to apoptosis (46). Fas-induced apoptosis of peripheral T and B cells is also decreased in PTEN−/− mice (47).

In mammary epithelium of MK-PTEN mice, the phosphorylation state of Akt is decreased and the level of lipid phosphatase activity associated with PTEN is increased. This may explain the higher level of apoptosis observed in these animals as compared to their WT littermates. From these results, it would be predicted that changes in PTEN activity, through overexpression or deletion, could alter the sensitivity of cells to apoptosis.

Using mammary-enriched cDNA microarrays to examine changes in gene expression profile in MK-PTEN as compared with WT mice, we show that MK-PTEN mice expressed IGFBP-5 26 times more highly than did the WT littermates 2 days after parturition. Furthermore, using primary culture of mammary epithelial cells, we showed that an IGFBP-5–blocking peptide reduces significantly the level of apoptosis as well as the caspase-3 activity observed in the mammary epithelial cells from the MK-PTEN mice. These results suggest strongly that the higher level of apoptosis in the mammary gland of MK-PTEN is partially due to the higher expression of IGFBP-5. IGFBP-5 is one of the major factors involved in mammary apoptosis in vivo (48). In vitro, in mammary epithelial cells in primary culture, lumen formation and DNA laddering indicative of apoptosis are also associated with an increase in the IGFBP-5 levels (49).

Using an oncogene-enriched cDNA microarray, we identified other genes involved in the cell survival or apoptosis. Amongst these genes, some were highly expressed in MK-PTEN mammary tissue. These genes include the thymosin β 10 (50) and the complement component 3 (51). GILZ is reported to inhibit the expression of Fas ligand (52), which is a mediator of apoptosis of mammary epithelial cells during the first stage of mammary involution (53). Our data show a lower expression of the GILZ gene in MK-PTEN mammary tissue, suggesting a lower inhibition of Fas and consequently a higher sensitivity to the apoptosis. Thus, our results suggest that the higher apoptosis observed in the MK-PTEN mammary tissue could be due to a decrease in Akt activation but also an induction or repression of some genes directly involved in the apoptosis process.

In conclusion, overexpression of active PTEN in mammary alveolar epithelium causes a marked reduction in cell proliferation, a marked increase in apoptosis, and an incomplete functional differentiation, which results in lactation failure. This suggests that the tumor suppressor protein PTEN can control mammary gland development; furthermore, an increase in proteins such as IGFBP-5 or a reduction in JNK/SAPK-1 may be directly responsible for this effect. The specific inactivation of the PTEN gene in mouse mammary epithelium will be needed to confirm our transgenic dominant gain-of-function experiments. Future studies with MK-PTEN mice will allow us to establish whether the high level of PTEN expression in the mammary epithelium is sufficient to reduce mammary tumor formation and/or progression.

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