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GABA interneurons are the cellular trigger for ketamine’s rapid antidepressant actions

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A single subanesthetic dose of ketamine, an NMDA receptor (NMDAR) antagonist, produces rapid and sustained antidepressant actions in depressed patients, addressing a major unmet need for the treatment of mood disorders. Ketamine produces a rapid increase in extracellular glutamate and synaptic formation in the prefrontal cortex, but the initial cellular trigger that initiates this increase and ketamine’s behavioral actions has not been identified. To address this question, we used a combination of viral shRNA and conditional mutation to produce cell-specific knockdown or deletion of a key NMDAR subunit, GluN2B, implicated in the actions of ketamine. The results demonstrated that the antidepressant actions of ketamine were blocked by GluN2B-NMDAR knockdown on GABA (Gad1) interneurons, as well as subtypes expressing somatostatin (Sst) or parvalbumin (Pvalb), but not glutamate principle neurons in the medial prefrontal cortex (mPFC). Further analysis of GABA subtypes showed that cell-specific knockdown or deletion of GluN2B in Sst interneurons blocked or occluded the antidepressant actions of ketamine and revealed sex-specific differences that are associated with excitatory postsynaptic currents on mPFC principle neurons. These findings demonstrate that GluN2B-NMDARs on GABA interneurons are the initial cellular trigger for the rapid antidepressant actions of ketamine and show sex-specific adaptive mechanisms to GluN2B modulation.

Introduction
The NMDA receptor (NMDAR) antagonist ketamine produces rapid-acting antidepressant effects in clinical studies of depressed patients and preclinical rodent models (1, 2). Although ketamine has been traditionally used as an anesthetic, more than a decade of research has been devoted to unraveling the mechanisms responsible for its rapid antidepressant effects. These efforts have uncovered a complex signaling cascade that results in activity-dependent synapse formation and reversal of the synaptic deficits caused by chronic stress exposure (2–4). Early studies using a subanesthetic dose of ketamine found a rapid and transient increase in glutamate release in the medial prefrontal cortex (mPFC) (5). This glutamate burst is believed to initiate activity-dependent release of brain-derived neurotrophic factor and stimulation of protein synthesis–dependent synapse formation, both of which are required for the antidepressant behavioral actions of ketamine (2, 4). However, the initial cellular trigger for the rapid release of glutamate in the PFC and the antidepressant behavioral actions of ketamine remains unclear.

Two mechanistic hypotheses have been proposed for ketamine’s initial cellular trigger (6, 7). The direct hypothesis posits that ketamine’s antidepressant effects result from direct antagonism of NMDARs on principle-glutamatergic neurons, causing homeostatic synaptic plasticity and increased synaptic drive. Alternatively, the indirect hypothesis, or disinhibition hypothesis, proposes that ketamine first inhibits GABAergic interneurons, leading to the disinhibition of principle-glutamatergic neuron activity. During basal resting state conditions, pyramidal cell firing is inhibited by presynaptic tonic-firing GABA interneurons (8, 9). Because ketamine is an NMDAR open-channel blocker that requires activity to remove Mg2+ blocking entry to the channel, tonic-firing GABA interneurons should be more sensitive to low, antidepressant doses of ketamine, resulting in a rapid disinhibition of pyramidal neurons and the subsequent transient glutamate burst. Furthermore, as discussed in Homayoun and Moghaddam (10), fast-spiking GABAergic interneurons have faster excitatory postsynaptic potentials (EPSPs) than pyramidal neurons and are more effectively recruited by excitatory inputs (11, 12). These findings support earlier studies and suggest that NMDAR antagonists initially regulate the spontaneous firing of GABAergic interneurons to a greater extent than pyramidal neurons (8, 9). A recent study of hippocampal slices provides further support, demonstrating that ketamine reduces inhibitory input and disinhibits CA1 pyramidal neurons (13). Studies of GABA interneurons in depression have focused on the subtypes expressing somatostatin (Sst) or parvalbumin (Pvalb), which together make up approximately 70% of GABA interneurons (14). These 2 subtypes innervate different aspects of excitatory neurons (Sst dendrites, Pvalb cell bodies) and have different electrophysiological properties (15).
NMDARs are a heterotrimeric complex composed of 2 obligatory GluN1 subunits and 2 additional subunits that are a combination of GluN2A, GluN2B, GluN2C, or GluN2D. Previous studies have demonstrated that a selective GluN2B-negative allosteric modulator (Ro 25-6981) produces antidepressant behavioral effects and synaptic changes similar to ketamine in rodent models (2, 3, 16). In addition, clinical studies have demonstrated that a single dose of another selective GluN2B-negative allosteric modulator, CP-101,606, produces antidepressant actions in depressed patients (17). Here, we used a virus-mediated shRNA knockdown strategy and conditional deletion approach to target GluN2B subunits on different populations of cells in the mPFC, and thereby determine their role in mediating ketamine’s antidepressant behavioral effects. We combined electrophysiological, behavioral, and molecular techniques to determine whether GluN2B-containing NMDARs on excitatory neurons or GABAergic interneurons in the mPFC are necessary for ketamine’s antidepressant effects.

Results

Ketamine reduces inhibitory postsynaptic currents in the mPFC. We first tested the influence of ketamine on spontaneous postsynaptic currents in layer V pyramidal neurons of the mPFC (Figure 1). If ketamine preferentially acts at NMDARs on GABAergic interneurons, we would expect a reduction in the inhibitory tone or frequency of spontaneous inhibitory postsynaptic currents (sIPSCs). In order to record sIPSCs and spontaneous excitatory postsynaptic currents (sEPSCs) simultaneously, whole-cell patch-clamp recordings were performed at a holding potential of −65 mV (the Cl⁻ reversal potential in the pipette solution was −72 mV). Spontaneous postsynaptic currents were determined before and during bath application of ketamine at 1 μM, the estimated concentration reached in the human brain after i.v. infusion of a dose in the therapeutic range (18) and a concentration that blocks approximately 50% of NMDA-induced currents (19); or 10 μM, a concentration that is sufficient to block approximately 80% of NMDA-induced currents (20). These concentrations have also been used in recent reports (4, 13). The results demonstrated that 1 μM ketamine significantly decreased sIPSC, but increased sEPSC in male and female mice (Figure 1, A–D); the cumulative probability curves for interevent intervals were shifted to the right for sIPSCs (i.e., longer intervals, decreased frequency), whereas for sEPSCs the curves were shifted to the left (i.e., shorter intervals, increased frequency) (Figure 1, C and D). Ketamine at 1 μM increased sEPSC amplitude in male and female mice, corresponding to increased sEPSC frequency; sIPSC amplitude was increased in males but decreased in females (the reason for this difference is unknown) (Supplemental Figure 1, E and F; supplemental material available online with this article; https://doi.org/10.1172/JCI130808DS1). In contrast, 10 μM ketamine significantly decreased basal rates of sIPSCs and sEPSCs in male and female mice (Supplemental Figure 1, A–D). These findings are consistent with the hypothesis that GABA interneuron–mediated IPSCs are more sensitive to ketamine blockade than glutamate pyramidal neuron–mediated EPSC responses.

Knockdown of NMDAR-GluN2B on GABA but not glutamate neurons produces baseline antidepressant-like effects and occludes the
actions of ketamine. First, we confirmed that glutamatergic pyramidal neurons and GABAergic interneuron subtypes in the mPFC express GluN2B using immunohistochemistry: GluN2B colabeled with CaMKII (pyramidal cell) or glutamate decarboxylase 67 (GAD67; GABA interneuron) cells (labeled using specific antibodies) and Sst- or Pvalb-tdTomato cells (Supplemental Figure 2, A–D). Furthermore, bath application of the selective GluN2B antagonist conantokin G (2 μM) reduced the NMDA-induced inward currents in pyramidal, Sst, and Pvalb cells in the mPFC (Supplemental Figure 3), demonstrating the expression of GluN2B subunits.

To target specific cell types, we used an shRNA-mediated adeno-associated virus (AAV) cell-type specific knockdown (sMACK-down) approach, combining viral expression of shRNA targeting GluN2B with cell-specific Cre recombinase (Cre') transgenic mouse lines. The GluN2BshRNA construct (Figure 2A) expressed both mCherry and EGFP, and in the presence of Cre recombinase, EGFP was deleted, resulting in expression of only mCherry (as a marker of recombination) and GluN2BshRNA. The GluN2BshRNA construct was packaged into an adeno-
regions) of Camk2aCre+ and Gad1Cre+ adult (8–12 week) male mice and WT Camk2aCre+ and Gad1Cre+ littermate controls (Figure 3A). The mPFC was targeted because prior studies have shown that this region is necessary and sufficient for the antidepressant actions of ketamine (2, 21). Three weeks after viral infusion to allow for GluN2BshRNA expression and GluN2B knockdown, the mice underwent behavioral testing, and then brains were collected for histology. Camk2aCre+ and Gad1Cre+ mice with AAV2GluN2BshRNA infusions (referred to as Camk2aCre+/AVV and Gad1Cre+/AVV, respectively) displayed different subsets of cells with mCherry only (Figure 3, B and F), demonstrating cell-specific recombination and expression of the GluN2BshRNA construct in each transgenic line. In WT-Camk2aCre+/AVV and WT-Gad1Cre+/AVV controls, the labeled neurons were yellow (i.e., both mCherry and EGFP), demonstrating that there was no recombination as expected.

Baseline testing was conducted to evaluate immobility, referred to as preswim (before ketamine) as part of the forced swim test (FST, a measure of behavioral despair); in addition, preswim is often used as an initial stressor before testing antidepressants in this paradigm. Baseline behavior was also analyzed in an open field test (OFT), a measure of ambulatory activity (distance traveled) and anxiety behavior (time spent in center); the OFT also serves as an additional mild stressor prior to ketamine treatment. One day after the OFT, mice were administered vehicle (saline, i.p.) or a single dose of ketamine (10 mg/kg, i.p.) and then tested 1 and 3 days later in the FST and novelty-suppressed feeding test (NSFT, a measure of anxiety-like behavior), respectively. This dose of ketamine produces a rapid transient burst of glutamate and antidepressant behavioral responses (2–5, 15, 19). The behavioral testing times (1 and 3 days) were used to avoid the acute locomotor and sensorimotor effects of ketamine administration and still show the rapid and sustained antidepressant effects of ketamine (Figure 3A). In Camk2aCre+/AVV male mice, there were no differences in preswim immobility, or on center time or distance in the OFT (Figure 3C). Ketamine significantly decreased immobility in the FST in WT-Camk2aCre+/AVV and Camk2aCre+/AVV male mice (Figure 3D). In the NSFT, ketamine administration significantly reduced the latency to feed in WT-Camk2aCre+/AVV and Camk2aCre+/AVV mice (Figure 3E). No significant differences were observed in home cage feeding (Figure 3E).

Using the same design, we found that Gad1Cre+/AVV mice displayed significantly decreased baseline immobility in the preswim test, an antidepressant-like effect, compared with WT-Gad1Cre+/AVV controls (Figure 3G); there were no significant effects on baseline behaviors in the OFT center time or total distance (Figure 3G). Saline-treated Gad1Cre+/AVV mice showed a significant baseline effect, similar to the pretest, and ketamine administration had no further effect beyond the baseline changes (Figure 3H). In WT-Gad1Cre+/AVV controls, ketamine administration significantly decreased FST immobility time (Figure 3I). In the NSFT, ketamine treatment significantly reduced latency to feed compared with saline in the WT-Gad1Cre+/AVV controls (Figure 3I). In Gad1Cre+/AVV mice, there was a nonsignificant decrease in baseline latency to feed, and the actions of ketamine were blocked or occluded (Figure 3I). No significant differences were observed in home cage feeding (Figure 3I). To address the possibility that GluN2B knockdown could produce a floor effect on immobility, we treated a separate cohort of Gad1Cre+/AVV male mice with 20 mg/kg desipramine (DMI), a norepinephrine-selective reuptake inhibitor (22). DMI administration significantly reduced immobility in the FST in WT-Gad1Cre+/AVV and Gad1Cre+/AVV mice (Supplemental Figure 4).

Knockdown of GluN2B in Sst interneurons produces sex-specific baseline behavioral effects and blocks the antidepressant-like effects of ketamine. To extend the results of the Gad1Cre+ study, we investigated the role of 2 major GABAergic interneuron subtypes, Sst and Pvalb, in the rapid antidepressant actions of ketamine (23). Sst and Pvalb interneurons display different molecular, electrophysiological, morphological, and circuit connectivity properties that could differentially contribute to the actions of ketamine (23). Postmortem studies of depressed subjects and rodent studies of chronic stress highlight a selective vulnerability of Sst-expressing GABAergic interneurons in PFC subregions (24, 25). We have reported that the initial cellular trigger for another rapid-acting antidepressant, scopolamine, is dependent on Sst but not Pvalb interneuron M1-type muscarinic acetylcholine receptors (26). Based on these findings, we initiated studies of Sst interneurons, including virus-mediated shRNA GluN2B knockdown and development of conditional deletion mutants (i.e., SstCreGluN2Bflox); we also compared male and female mice and conducted electrophysiological and RNA-Seq studies.

Electrophysiology studies have demonstrated that Sst interneurons are responsive to NMDA, and that this effect is blocked by a selective GluN2B antagonist, conantokin G (Supplemental Figure 3); immunohistochemistry studies have demonstrated that Sst+ interneurons in the mPFC express GluN2B (Supplemental Figure 2, C and D). Sst GluN2B knockout studies were carried out as described above. SstCre+ mice were infused with AAV2GluN2BshRNA (SstCre+/AVV) into the mPFC, resulting in cell type-specific recombination (mCherry only, red cells), compared with WT-SstCre+AVV control mice with no recombination (mCherry + EGFP double-labeled yellow cells) (Figure 4, A and B). Functional knockdown was also shown in SstCre+/AVV male and female mice by slice physiology, with decreased NMDA- but not AMPA-induced inward currents (Figure 2, H and I). In male SstCre+/AVV mice, there was a significant decrease in immobility time in the preswim test compared with WT-SstCre+AVV controls (Figure 4C); however, in female SstCre+/AVV mice, there was no significant effect on immobility time (Figure 4C). There were no effects in male or female SstCre+/AVV mice in the OFT (Figure 4, D and E). A single dose of ketamine significantly decreased FST immobility time in male and female WT-SstCre+AVV mice compared with saline controls (Figure 4, F and I). In saline-treated SstCre+/AVV mice, there was a significant decrease in baseline FST immobility in males and a trend for a decrease in females, and ketamine administration had no further effects in either sex (Figure 4, F and I). In the NSFT, ketamine significantly reduced latency to feed in male and female WT-SstCre+AVV mice when compared with saline controls, but these effects were completely absent in SstCre+/AVV mice (Figure 4, G and J). There was a small decrease in baseline latency to feed in female SstCre+/AVV mice, although this effect was not significant. No significant differences were observed in home cage feeding (Figure 4, G and J). In males, we also examined a reward-seeking behavior, the female urine sniffing test (FUST) (27). Male WT-SstCre+AVV mice administered ketamine spent significantly more time sniffing female urine.
when compared with their genotype controls, and this effect was blocked in SstCrem/AAV mice (Figure 4H). There was no effect on baseline FUST in male SstCrem/AAV mice, and no significant differences in time sniffing water across any group (Figure 4H).

To examine possible cellular determinants of the baseline behavioral differences in male and female SstCrem/AAV mice, we measured sIPSCs and sEPSCs in layer V pyramidal neurons in mPFC slices (Figure 4, K and L). In SstCrem/AAV males, there was a small but significant decrease in the frequency of sIPSCs, but a larger increase in sEPSCs (Figure 4K). In female SstCrem/AAV mice, there was a significant decrease in the frequency of sIPSCs, but no effect on sEPSCs (Figure 4L). The increase in basal sEPSCs in male mice could contribute to the baseline antidepressant-like effects observed in the preswim in male versus female SstCrem/AAV knockdown mice (Figure 4C). Despite there being no significant difference in sEPSCs in female mice, there was still an occlusion of ketamine’s effects in female Sst Cre+/AAV knockdown mice. These findings suggest that there may be varying molecular and cellular profiles that account for the observed sex differences.

To further examine cellular differences between males and females, we examined NMDAR subunit composition in Sst interneurons. Sst-tdTomato interneurons from mPFC were sorted by FACS and levels of NMDAR subunits determined by RNA-Seq. We found that expression of the GluN2B subunit, encoded by the Grin2b gene, in the male and female mPFC is higher compared with Grin2a, similar to reports of Grin2b predominance in Sst+ interneurons of the adult male mouse visual cortex (28). In addition, there were no significant sex-specific differences in the expression of Grin1, Grin2a, and Grin2b in Sst+ interneurons (Supplemental Figure 5, A and B). Because NMDAR channel opening...
requires AMPA receptor (AMPAR) activity, we also examined the relative abundance of AMPAR subunits 1 and 2, encoded by the Gria1 and Gria2 genes, respectively. Here, we observed significantly higher levels of Gria2 and a trend for higher levels of Gria1 in female mice compared with male mice (Supplemental Figure 5C). Further studies are required to determine whether differences in AMPAR expression contribute to the behavioral differences in males and females and whether GluN2B knockdown alters the expression patterns of NMDAR and AMPAR subunits.

Because ketamine is a nonselective NMDAR channel blocker, we tested whether knockdown of another subunit, GluN2A, would influence the behavioral actions of ketamine. The efficacy of AAV2 GluN2A shRNA was confirmed by Cre-dependent GluN2A knockdown in primary neuronal cultures (Supplemental Figure 6, A–D). In male SstCre mice with infusion of AAV2 GluN2A shRNA into the mPFC (SstCre/AAV2), there was no significant effect on baseline immobility time in pretreatment compared to WT-SstCre/AAV2 control mice (Supplemental Figure 6F). Ketamine administration significantly decreased immobility time in the FST in WT-SstCre/AAV2 and SstCre/AAV2 mice (Supplemental Figure 6G). In the NSFT, ketamine administration significantly reduced the latency to feed in WT-SstCre/AAV2 and SstCre/AAV2 mice (Supplemental Figure 6D). No significant differences were observed in locomotor activity or home cage feeding after ketamine administration (Supplemental Figure 6, H and I).

**Genetic deletion of Grin2b from Sst interneurons.** In addition to the virus-mediated approach, we generated Sst interneuron–specific Grin2b deletion mice by crossing SstCre and Grin2bfl/fl mice (SstCre/Grin2bfl) and littermate controls (SstCre/Grin2afl), Baseline immobility in the FST and anxiety-related behaviors in the OFT were tested in adult male and female mice (Figure 5A). Similar to virus-mediated knockdown of GluN2B from Sst interneurons in the mPFC, male but not female SstCre/Grin2afl mice showed reduced immobility in the FST compared with controls (Figure 5C). There were no differences observed in the OFT (center time or distance traveled) in either sex (Figure 5, D and E). In male SstCre/Grin2afl mice, there was also a significant antidepressant-like effect (increased time sniffing) in the FUST compared with SstCre/Grin2afl controls (Figure 5F). In preliminary studies, we found that male and female SstCre/Grin2afl mice responded similarly to ketamine (Supplemental Figure 7), so we used a mixed cohort of males and females (24 hours after dosing). Ketamine significantly reduced immobility time in the FST in SstCre/Grin2afl controls, but not in SstCre/Grin2afl deletion mutants (Figure 5G). Ketamine also significantly reduced latency to feed in NSFT in the SstCre/Grin2afl controls, but not in SstCre/Grin2afl deletion mice (Figure 5H).

To examine the effects of genetic Grin2b deletion on neuronal activity, postsynaptic currents were recorded from layer V pyramidal neurons of the mPFC (Figure 5, I and J). SstCre/Grin2afl males displayed significantly decreased sIPSCs but increased sEPSCs (Figure 5J), similar to in vitro incubation of PFC slices with 1 μM ketamine (Figure 1C), whereas SstCre/Grin2afl females displayed significant decreases of both sIPSCs and sEPSCs (Figure 5K). The increase in sEPSCs in male mice could have contributed to the baseline antidepressant-like effects observed in male versus female SstCre/Grin2afl deletion mutant mice (Figure 5C).

**Knockdown of GluN2B in Pvalb interneurons blocks or occludes the antidepressant-like effects of ketamine.** To further examine GABAergic interneuron subtypes in the actions of ketamine and to compare with Sst results, we conducted GluN2B-knockdown studies in Pvalb interneurons. Preliminary studies demonstrated that Pvalb interneurons were responsive to GluN2B-containing NMDARs (blocked by conantokin G) (Supplemental Figure 3) and that Pvalb+ interneurons in the mPFC expressed GluN2B (Supplemental Figure 2, C and D). Functional knockdown was also shown in PvalbCre/+ male mice by slice physiology, with decreased NMDA- but not AMPA-induced inward currents (Figure 2J). Using the same design described above, male and female PvalbCre or WT-PvalbCre littermate controls were infused with AAV2 GluN2A shRNA (PvalbCre/+ or WT-PvalbCre/+ (Figure 6A), resulting in cell-specific recombination with a subset of mCherry only–labeled cells (Figure 6B).

In contrast, in WT-PvalbCre/+ mice, the labeled neurons were yellow (mCherry + GFP labeled interneuron), demonstrating that there was no recombination. In PvalbCre/+ male and female mice, there was no difference in baseline presim immobility or OFT behavior compared to WT-PvalbCre/+ controls (Figure 6, C–E). A single dose of ketamine significantly decreased immobility time in WT-PvalbCre/+ but not in PvalbCre/+ male (Figure 6F) and female mice (Figure 6H). Interestingly, PvalbCre/+ female mice given saline showed significantly reduced baseline immobility in the FST, and there was no further effect of ketamine (Figure 6H); there was a similar trend for decreased immobility in saline-treated PvalbCre/+ males. In the NSFT, ketamine significantly reduced the latency to feed in male and female WT-PvalbCre/+ control mice compared with saline (Figure 6, G and I). In male and female PvalbCre/+ mice, there was no baseline effect in the NSFT, and there were no significant differences after ketamine administration, although there was a trend for a decrease in males (Figure 6, G and I). No differences were observed in home cage feeding (Figure 6, G and I). Next, we measured sIPSCs and sEPSCs in layer V pyramidal neurons of mPFC slices from PvalbCre/+ mice (Supplemental Figure 8). In PvalbCre/+ males, there was a significant decrease in the frequency of sIPSCs (Supplemental Figure 8, A and B). In contrast to SstCre/+ male findings, we observed a small but significant decrease in the frequency of sEPSCs in PvalbCre/+ males (Supplemental Figure 8, A and B).

**Discussion**

The results of the present study demonstrated that GluN2B knockdown in GABAergic interneurons (Gadi, Sst, and Pvalb), but not Camk2a/glutamate neurons, occluded and/or blocked the antidepressant behavioral actions of ketamine (Table 1). The results also provided electrophysiological evidence that a low concentration of ketamine blocked NMDARs on GABA interneurons in the mPFC, resulting in decreased inhibitory input and increased sEPSCs on layer V principle neurons. Viral knockdown or genetic deletion of GluN2B on Sst interneurons produced a similar significant increase (disinhibition) of EPSCs, but only in male mice, which could account for the sex difference in baseline behaviors. Together, these studies provide strong evidence in support of the disinhibition hypothesis that the initial trigger for ketamine blockade is GluN2B-containing NMDARs on GABAergic interneurons in the mPFC. The mPFC layer V principle neurons project to cortical and subcortical brain regions implicated in depression-related behaviors, including the basolateral nucleus of the
The amygdala, which is necessary and sufficient for the rapid and long-lasting antidepressant actions of ketamine (19, 21). Because behavioral tests were conducted 24 hours after dosing to avoid the acute side effects of ketamine, we also acknowledge the possibility that there could be adaptations that require GluN2B on GABA interneurons that mediate the actions of ketamine.

An early in vivo electrophysiology study reported that ketamine administration resulted in blockade of GABA interneuron activity in the mPFC, which preceded an increase in glutamate neuronal activity, although the association between these 2 events could only be inferred (10). A recent in vitro electrophysiology study reported that ketamine blocks NMDARs on tonic-firing GABAergic interneurons, decreasing inhibitory inputs onto excitatory pyramidal neurons. An interesting observation from these experiments was...
designed and validated a virus-mediated, Cre recombinase strategy (AAV2GluN2BshRNA) to knock down GluN2B or GluN2A on specific populations of neurons in the mPFC, a region that is necessary and sufficient for the actions of ketamine (2, 21). The GluN2B subunit was targeted based on evidence that selective GluN2B antagonists produce rapid antidepressant actions in depressed patients and rodent models (3, 16, 17). The results demonstrated that knock-down of GluN2B on Gad1/GABA interneurons, but not Camk2a/pyramidal neurons in the mPFC, blocked and/or occluded the antidepressant-like behavioral effects of ketamine (Table 1). GluN2B

ments is that female mice had more clustered high-frequency IPSCs compared with males, possibly due to a greater number and/or excitation of burst-firing interneurons in females (29); however, additional studies are needed to address these differences. In contrast, a higher concentration of ketamine (10 μM) significantly decreased sIPSCs and sEPSCs in layer V mPFC neurons of male and female mice, demonstrating the selective effect of the lower 1 μM concentration on sIPSCs.

To identify the subunit type and cellular localization of NMDARs underlying the antidepressant action of ketamine, we...
Our results of GluN2B knockdown in Camk2a/pyramidal neurons differed from a previous study by Hall and colleagues reporting that genetic deletion of Grin2b using a different excitatory neuron promoter (NEX or Neuro6d) produced baseline antidepressant effects in the FST that occluded the actions of ketamine (30). However, the NEX-Cre+ Grin2b deletion mutant mice displayed 3-fold higher levels of locomotor activity than WT controls (31), which could very likely mediate the decreased baseline immobility and occlusion of the ketamine response in the FST (30). We did not observe any changes in locomotor activity in our viral Camk2aCre+/AAV or Gad1Cre+/AAV knockdown mice, or in any other lines tested. Another study by Hall and colleagues used a viral approach to delete Grin2b from pyramidal neurons in the mPFC (Grin2bfl/fl mice plus AAV-Camk2a–driven Cre recombinase), reporting decreased baseline immobility time in the FST (32); the influence of ketamine was not examined in this study. The reason for this baseline difference is not clear, but could be due to the complete deletion of Grin2b in excitatory neurons and targeting knockdown on Gad1/GABA interneurons also produced baseline antidepressant-like effects on immobility in the preswim test, as well as the FST, a measure of behavioral despair, and occluded any further action of ketamine. To determine whether the occlusion of the ketamine response was due to a floor effect, we tested whether GluN2B knockdown in Gad1Cre mice also occluded the response to another antidepressant, DMI, which has a completely different mechanism (i.e., selective inhibition of norepinephrine reuptake). DMI administration produced a significant antidepressant effect in the FST in the Gad1Cre mice, demonstrating that there was not a floor effect that occluded the response to ketamine. These baseline effects suggest that GluN2B knockdown in Gad1Cre mice reduced NMDAR drive of GABA interneurons, similar to ketamine. The reason for the differential baseline effects in the FST and NSFT is unknown, but could result from different cellular or circuit mechanisms that contribute to each behavior. Nevertheless, these results demonstrated a requirement for GluN2B on GABA interneurons in the actions of ketamine.
of prelimbic PFC because infralimbic, but not prelimbic, PFC is required for the antidepressant actions of ketamine (21).

We next examined GABA interneuron subtypes, with a focus on Sst interneurons based on evidence from human postmortem studies reporting selective loss of Sst interneurons in the PFC of depressed subjects (24, 25) and our previous studies of scopolamine (26). Sst interneurons can be regular spiking, low threshold spiking, and burst firing (15), and blockade of these responses could underlie ketamine-mediated disinhibition. We found that either viral knockdown of GluN2B in the mPFC of SstCre/+ or constitutive knockout of GluN2B in SstCre+Grin2bfl mice blocked the antidepressant effects of ketamine, demonstrating an association with antidepressant behavior in males and females (Table 1). In contrast, knockdown of GluN2A, another prominent subtype in the cerebral cortex, on Sst interneurons did not block the antidepressant effects of ketamine, suggesting that the disinhibition resulting from the actions of ketamine on Sst interneurons contributes to its greater therapeutic efficacy relative to scopolamine (15). We found that viral knockdown of GluN2B in the mPFC of PvalbCre/+ male and female mice blocked the antidepressant behavioral effects of ketamine (Table 1). This differs from our previous study of scopolamine, in which we found that knockdown of M1-AChR on Sst but not Pvalb interneurons blocked the antidepressant effects of scopolamine (26), raising the possibility that the disinhibition resulting from the actions of ketamine on both Sst and Pvalb interneurons contributes to its greater therapeutic efficacy relative to scopolamine (41). Indeed, animals that have been subjected to inescapable footshock have reduced excitatory drive onto prefrontal Pvalb interneurons, and chemogenetic activation of these interneurons can lead to susceptibility in resilient mice, suggesting that Pvalb interneurons are also important in regulating depression-like behavioral output (42). Saline-treated PvalbCre/+ mice displayed decreased immobility, which could result from blockade of increases that are observed with repeated swim exposure that is observed in rodents, suggesting a role for GluN2B subunits on mPFC Pvalb interneurons in memory or stress resiliency processes. For example, blocking synaptic transmission in mPFC Pvalb interneurons impairs spatial working memory and reversal learning, but has no effect on anxiety as measured by the OFT (43). A previous study reported that constitutive global genetic

### Table 1. Summary of behavior and electrophysiology findings by sex and genotype

<table>
<thead>
<tr>
<th>Sex</th>
<th>Genotype</th>
<th>Baseline Behavior</th>
<th>Electrophysiology Parameters</th>
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<th>Ketamine ADT effects: NSFT</th>
<th>Ketamine ADT effects: FUST</th>
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<tr>
<td></td>
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<td>↑ sEPSCs, ↓ sIPSCs</td>
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<td></td>
<td>SstCre+/Grin2bfl</td>
<td>↓ Immobility in preswim, ↑ Time in FUST</td>
<td>↓ sIPSCs, ↑ sEPSCs</td>
<td>Blocked</td>
<td>Blocked</td>
<td>–</td>
</tr>
<tr>
<td>Females</td>
<td>SstCre+/AAVshift</td>
<td>None</td>
<td>–</td>
<td>ADT effect</td>
<td>ADT effect</td>
<td>ADT effect</td>
</tr>
<tr>
<td></td>
<td>PvalbCre+/AAV</td>
<td>None</td>
<td>↓ sEPSCs, ↓ sIPSCs</td>
<td>Ocluded</td>
<td>Blocked</td>
<td>Blocked</td>
</tr>
</tbody>
</table>

↑, increase; ↓, decrease; ↔, no change, when compared to controls. ADT, antidepressant; –, no data.

neurons upon knockdown or deletion of Grin2b. Sex differences in NMDAR and/or AMPAR expression on excitatory neurons, which are known to be influenced by estrogen, could also contribute to the observed behavioral effects in males and females (34, 35). Female rats have higher baseline PFC GluN1 expression and increased stress-induced expression of GluN2B compared with males (36). Postmortem studies have reported that female depressed subjects had increased expression of several glutamate receptor subtypes in the dorsolateral PFC, and that female depressed suicide subjects had significantly increased expression of Grin2b (37). Only one study to date has analyzed sex differences in ketamine response and found no differences in efficacy or tolerability (38, 39). These studies highlight the need for further studies of sex differences in the behavioral, neural, molecular, and genetic phenotypes of major depressive disorder to inform the development of more effective treatment options (40).

Pvalb interneurons are noted for their fast-spiking properties at rest, and could also be involved in ketamine-mediated disinhibition (15). We found that viral knockdown of GluN2B in the mPFC of PvalbCre/+ male and female mice blocked the antidepressant behavioral effects of ketamine (Table 1). This differs from our previous study of scopolamine, in which we found that knockdown of M1-AChR on Sst but not Pvalb interneurons blocked the antidepressant effects of scopolamine (26), raising the possibility that the disinhibition resulting from the actions of ketamine on both Sst and Pvalb interneurons contributes to its greater therapeutic efficacy relative to scopolamine (41). Indeed, animals that have been subjected to inescapable footshock have reduced excitatory drive onto prefrontal Pvalb interneurons, and chemogenetic activation of these interneurons can lead to susceptibility in resilient mice, suggesting that Pvalb interneurons are also important in regulating depression-like behavioral output (42). Saline-treated PvalbCre/+ mice displayed decreased immobility, which could result from blockade of increases that are observed with repeated swim exposure that is observed in rodents, suggesting a role for GluN2B subunits on mPFC Pvalb interneurons in memory or stress resiliency processes. For example, blocking synaptic transmission in mPFC Pvalb interneurons impairs spatial working memory and reversal learning, but has no effect on anxiety as measured by the OFT (43). A previous study reported that constitutive global genetic
deletion of Grin1 from PvalbCre+ mice did not block the effects of ketamine (44). The differences in the scope and developmental targeting of Grin1 deletion in Pvalb interneurons could underlie the different behavioral outcomes in these studies.

The results of the current study provided direct evidence that the initial cellular trigger for the antidepressant actions of ketamine was blockade of NMDARs on GABAergic interneurons, not excitatory neurons, leading to disinhibition and a burst of glutamate that drives synaptic and behavioral responses. Ketamine blockade of GABA activity appeared to be in opposition to evidence of a reduction of GABA, particularly Sst interneurons, in depression (24, 25). Our results also appeared to contradict rodent studies reporting that increased Sst interneuron function produced antidepressant behavioral responses (45). However, ketamine blockade of NMDARs and the resulting glutamate burst were transient, returning to baseline levels by 80 minutes after dosing (5), but led to long-lasting synaptic and antidepressant behavioral actions (2, 22, 46, 47). In addition, recent work reported that a single dose of ketamine also led to increased GABA function in the mPFC 24 hours after dosing, evidence of further adaptations to the initial glutamate burst that would contribute to correction of GABA, as well as glutamate deficits (48). This could also explain the apparent contradictory effects reported in another rodent study, which found that repeated ketamine administration increased Pvalb interneuron activity in juvenile stressed mice (49), but this likely represents an adaptation to repeated induction of the glutamate burst. Thus, the impact of transient versus sustained effects is critical when interpreting these experiments.

In conclusion, conventional monoaminergic antidepressant treatments take weeks to months to produce a therapeutic response and are ineffective for treatment-resistant depression. The novel, rapid-acting antidepressant actions of ketamine are effective even for patients considered treatment resistant, but the abuse potential of ketamine remains a limitation. Dissection of the cellular and molecular mechanisms underlying ketamine’s antidepressant effects could lead to the development of more targeted and safer treatments for depression. The results of the current study reveal an important cellular mechanism by which ketamine produces rapid antidepressant actions, indicating that activity-related neurotransmitter receptors and channels on Sst and Pvalb interneurons in the mPFC are promising targets for the development of novel rapid-acting antidepressants.

Methods

Animals. For electrophysiology experiments looking at the effects of ketamine, WT C57BL/6 male and female mice were used (Jackson Laboratories). For electrophysiology experiments looking at the effects of ketamine on conantokin G on NMDA-induced inward currents, WT C57BL/6, Sst-tdTomato, or Pvalb-tdTomato male mice were used. To obtain Sst- and Pvalb-tdTomato mice, Sst-Cre and Pvalb-Cre mice were bred with Ai9(RCL-tDT) mice (catalog 007909), all from Jackson Laboratories. For FACS and RNA-Seq, Sst-tdTomato male and female mice were used. For viral studies, male and female transgenic mice and WT C57BL/6 littermates were obtained from in-house breeders. Gad1-Cre mice were originally obtained from Marina Picciotto (Yale School of Medicine) (50), and Camk2a-Cre mice were obtained from Günter Schütz (German Cancer Research Center, Heidelberg, Germany) (51). Pvalb-Cre (catalog 008069) and Sst-Cre (catalog 013044) mice were obtained from Jackson Laboratories. Grin2b+/−/Sst-Cre+ mice were produced by crossing Grin2b floxed mice (Eric Delpere, Vanderbilt University, Nashville, Tennessee, USA) (52) with Sst-Cre heterozygous mice to produce Grin2b+/−/Sst-Cre+ and Grin2b+/−/Sst-Cre− offspring. For lines that required additional breeding, at least 6 backcrosses were performed. All studies were performed with adult mice. Pregnant female rats were used as a source of embryonic tissue for primary neuronal cultures (Jackson Laboratories). Animals were housed in standard ventilated rack under a 12-hour light/12-hour dark cycle with ad libitum access to water and rodent chow. All experiments were performed during the 12-hour light cycle. Animal use and procedures were in accordance with the NIH guidelines and approved by the Yale University Animal Care and Use Committees.

Brain slice electrophysiology. Brain slices containing the mPFC were prepared from male and female mice (8 to 16 weeks old). Brains were placed in artificial cerebrospinal fluid (ACSF) (pH 7.35–7.38) equilibrated with 95% O2/5% CO2. Coronal slices of 300-μm thickness containing the mPFC were transferred to the fixed stage of an Olympus BX50WI scope for whole-cell recording. The chamber was continuously perfused with normal ACSF at a rate of 2 to 3 mL/minute and its temperature maintained at 33°C ± 0.5°C. Patch pipettes (3–5 MΩ) were pulled from glass tubing with a Flaming/Brown horizontal puller. The pipette solution contained the following: 115 mM K-gluconate, 5 mM KCl, 2 mM MgCl2, 2 mM Mg-ATP, 2 mM Na-ATP, 10 mM Na+, phosphocreatine, 0.4 mM NaGTP, and 10 mM Hepes, pH 7.3. Neurons were visualized by videomicroscopy using a microscope (40× IR lens) with infrared differential interference contrast (IR/DIC). Whole-cell recordings were made with an Axoclamp-2B amplifier. Postsynaptic currents were studied in the continuous single-electrode voltage-clamp mode (3000 Hz low-pass filter). sIPSCs and sEPSCs were simultaneously recorded (53–55) at a holding potential of −65 mV. Because the reversal potential of Cl− was ~72 mV (calculated from pipette solution), at −65 mV membrane potential, sIPSCs and sEPSCs were distinguished by their outward and inward polarity, respectively. Synaptic events were analyzed using Mini Analysis Software (Synaptosoft). We tested 10 μM NMDA- and 5 μM AMPA-induced inward currents with no added magnesium in the ACSF.

Immunohistology. After 3 weeks to allow for viral expression, brains were collected from mice infected with AAV2GluN2BshRNA after transcardiac perfusion with sterile PBS and 4% paraformaldehyde (PFA). Brains were postfixed in 4% PFA for 24 hours and incubated in 30% sucrose for an additional 24 hours. Fixed brains were frozen and sectioned at 30 μm using a Microm HM550 cryostat. To examine the colocalization of GluN2B with different cell markers, brain sections were washed and incubated with blocking buffer (PBS, 0.3% Triton X-100, 4% normal goat serum; 1 hour at room temperature), followed by overnight incubation with primary antibodies at 4°C: anti-Camk2a (Enzo Life Sciences, catalog KAM-CA002-D, 1:1000, rat), anti-GAD67 (Sigma-Aldrich, catalog G5419, 1:1000, mouse), and anti-GluN2B (EMD Millipore, catalog AB1557, 1:500, rabbit). Brain sections from Sst-tdTomato and Pvalb-tdTomato reporter mice were used for immunolabeling of GluN2B on Sst+ and Pvalb+ interneurons (anti-GluN2B, Alomone Labs, catalog AGC-003, 1:200, rabbit). To match the style of Camk2a and GAD67 images, Sst+ and
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*Pna*<sup>II</sup> stains were flipped to green, and GluN2B stains were flipped to red. Sections were then washed and incubated overnight at 4°C or 2 hours at room temperature with fluorange-conjugated secondary antibodies (Alexa Fluor, Thermo Fisher Scientific, 1:1000 dilution). Immunofluorescence was visualized using a confocal laser scanning microscope (Olympus FV1000).

**shRNA and virus preparation.** For knockdown of GluN2B or GluN2A in specific neuronal cell types, an shRNA sequence was designed targeting the GluN2B or GluN2A subunit (56). GluN2B shRNA (5'-TgacaaccaggtctcatttaaTTACAGAGAGTGTTCTGTTCTTTTC-3') or GluN2A shRNA (5'-TgacaaccaggtctcatttaaTTACAGAGAGTGTTCTGTTCTTTTC-3'); Integrated DNA Technologies) was ligated into a plasmid (pGluN2BshRNA or pGluN2AshRNA) designed to restrict shRNA expression to cells that express Cre recombinase (57). The TATA-loxP-flanked CMV-EGFP cassette was subcloned from pSico (from Tyler Jacks, Massachusetts Institute of Technology, Boston, Massachusetts, USA; Addgene plasmid 11578) into pAav-mCherry (Virovek) to generate the sMACKdown backbone. This plasmid allows ubiquitous mCherry expression and conditional EGFP expression driven by floxed CMV cassette that prevents shRNA expression with U6 promoter disruption.

**Primary cortical cultures.** Pregnant female rats were euthanized and cortices from E18 embryos were dissected. After incubation in trypsin-EDTA (0.25%, Gibco) for 10 minutes, cortices were dissociated and neurons were plated on 1 million cells per well in 6-well poly-D-lysine-coated plates in DMEM (Gibco)/10% FBS. Medium was changed the following day to a serum-free medium containing neurobasal and B27 (Gibco), which was changed every 4 days. Cells were maintained at 37°C, 5% CO<sub>2</sub> and 95% humidity. After 21 days (DIV26) of infection and shRNA expression, cells were collected for Western blot analysis.

**Western blot analysis.** Levels of GluN2B or GluN2A from cell lysates were analyzed by Western blot analysis. Cells were collected into RIPA lysis buffer containing 50 mM Tris·HCl (pH 7.5), 150 mM NaCl, 1% Triton X-100, 0.1% SDS, 1 mM NaVO<sub>3</sub>, 5 mM NaF, and 1× protease inhibitor cocktail. Homogenates were then centrifuged at 16,200 x g for 10 minutes to remove cell debris. The supernatant was collected and total protein concentrations measured using a protein assay kit (Pierce BCA Protein Assay Kit, Thermo Fisher Scientific). Twenty micrograms of protein was electrophoretically separated in a 20% FBS/HBSS/0.2% BSA solution, centrifuged (12000 x g), and resuspended and filtered using HBSS/0.2% BSA solution. Fluorescent dyes were sorted and collected into 100 μL of lysis buffer (Norgen kit) using FACS (FACSCalibur, Becton Dickinson). RNA was extracted using Arcturus PicoPure RNA isolation kit (Applied Biosystems). Using the Pico RiboGone-Mammalian (TakaraBio), 100 ng of total mRNA was processed before cDNA synthesis. RNA-Seq was performed on an Illumina HiSeq 2500 using 75-bp, paired-end sequencing. FASTQ files were trimmed and assessed for quality control before being annotated with the mouse genome (mm9) using TopHat2. The assembled transcripts were used to estimate transcript abundance and fragments per kilobase of transcript per million mapped reads (FPKM) values using cuffdiff42 and R (The Comprehensive R Archive Network. https://www.cran.r-project.org). RNA-Seq data were deposited to NCBI Gene Expression Omnibus data set repository (GEO accession GSE138670).

**Drug administration.** For ketamine experiments, mice received injection of vehicle (0.9% saline) or ketamine (10 mg/kg, i.p.) 24 hours before behavioral testing. For DMI experiments, mice received injection of vehicle (0.9% saline) or DMI (20 mg/kg, i.p.) 30 minutes before behavioral testing.

**OFST.** OFST, referred to as preswim if administered prior to drug treatment, was conducted as previously described (2). Mice were placed for 6 minutes in a clear cylinder filled with water (24°C ± 1°C, 18 cm depth). Sessions were video recorded and scored for total immobility time by a blinded experimenter. Time immobile during the 2- to 6-minute block was reported.

**RNA preparation and RNA-Seq.** In brief, the mPFC was dissected from Sst-tdTomato reporter mice and incubated in DMEM (GIBCO)/0.25% trypsin (GIBCO) for 30 minutes at 37°C. The tissue pieces were then homogenized using a 20% FBS/HBSS/0.2% BSA solution, centrifuged (12000 g 6 minutes) to collect the cells and resuspended and filtered using HBSS/0.2% BSA solution. Fluorescent dyes were sorted and collected into 100 μL of lysis buffer (Norgen kit) using FACS (FACSCalibur, Becton Dickinson). RNA was extracted using Arcturus PicoPure RNA isolation kit (Applied Bio-systems). Using the Pico RiboGone-Mammalian (TakaraBio), 100 ng of total mRNA was processed before cDNA synthesis. RNA-Seq was performed on an Illumina HiSeq 2500 using 75-bp, paired-end sequencing. FASTQ files were trimmed and assessed for quality control before being annotated with the mouse genome (mm9) using TopHat2. The assembled transcripts were used to estimate transcript abundance and fragments per kilobase of transcript per million mapped reads (FPKM) values using cuffdiff42 and R (The Comprehensive R Archive Network. https://www.cran.r-project.org). RNA-Seq data were deposited to NCBI Gene Expression Omnibus data set repository (GEO accession GSE138670).

**FUST.** This test was conducted according to previously published procedures (27). Male mice were habituated for 60 minutes to a sterile cotton-tipped applicator placed in their home cage. For the test, mice were first exposed to a new cotton tip dipped in sterile water; 45 minutes later, mice were then exposed to another cotton tip infused with 45 μL fresh urine collected from adult females of the same strain. Estrous cycle was not measured because prior studies have reported that females in all states of estrus elicit higher investigation from...
males than ovariectomized females (60). Behavior was video recorded for 5 minutes per test and total time spent sniffing the cotton tip was determined. Time spent biting the cotton tip was excluded from the recording time.

**Locomotor activity.** Animals were placed in a clean testing cage (30 × 19 × 13 cm) for 30 minutes, during which the number of beam breaks was measured using the Med-PC software (Med Associates).

**Statistics.** Data were subjected to statistical analyses with GraphPad Prism 8. The results are expressed as the mean ± SEM. Statistical significance was determined by a 1-way analysis of variance (ANOVA) or a 2-way ANOVA, followed by the Tukey’s or Sidak’s (FUST) multiple-comparisons test for comparing the treated group with the control group and multigroup comparisons, as indicated in the figure legends. Statistical differences between any 2 groups were determined using either the Student’s 2-tailed t test or Kolmogorov-Smirnov 2-sample test. Across all analyses, differences were considered significant for P values less than 0.05. Any value greater than 2 times the standard deviation of the mean was considered an outlier and removed from further analysis of the test at hand. The specific test used for each figure is mentioned in the figure legend. Each experiment was conducted a minimum of 2 times.

**Study approval.** Animal use and procedures were in accordance with NIH guidelines and approved by the Yale University Animal Care and Use Committees.

**Author contributions**

DMG, SP, ESW, and RSD designed the study. DMG, SP, RJL, MW, MJG, and CHD conducted experiments, analyzed data, and interpreted the results. XYL helped with breeding, genotyping, and the development of the SstCre/GluN2b<sup>−/−</sup> mouse line. SRT and MP contributed essential reagents and experimental advice for cell type–selective knockout experiments. ED provided the Glun2b<sup>−/−</sup> line and helped with the design of the Glun2b<sup>−/−</sup> lines used in the study. DMG and RSD wrote the manuscript. DMG was designated as the first co–first author based on primary contributions. All authors reviewed and approved the final manuscript.

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25. Wallack O, et al. The female urine sniffing


