Everolimus improves the efficacy of dasatinib in PDGFRα-driven glioma

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Pediatric and adult high-grade gliomas (HGGs) frequently harbor *PDGFRA* alterations. We hypothesized that cotreatment with everolimus may improve the efficacy of dasatinib in PDGFRα-driven glioma through combinatorial synergism and increased tumor accumulation of dasatinib. We performed dose-response, synergism, P-glycoprotein inhibition, and pharmacokinetic studies in in vitro and in vivo human and mouse models of HGG. Six patients with recurrent PDGFRα-driven glioma were treated with dasatinib and everolimus. We found that dasatinib effectively inhibited the proliferation of mouse and human primary HGG cells with a variety of *PDGFRA* alterations. Dasatinib exhibited synergy with everolimus in the treatment of HGG cells at low nanomolar concentrations of both agents, with a reduction in mTOR signaling that persisted after dasatinib treatment alone. Prolonged exposure to everolimus significantly improved the CNS retention of dasatinib and extended the survival of PPK tumor–bearing mice (mutant *TP53*, mutant *PDGFRA*, *H3K27M*). Six pediatric patients with glioma tolerated this combination without significant adverse events, and 4 patients with recurrent disease (*n* = 4) had a median overall survival of 8.5 months. Our results show that the efficacy of dasatinib treatment of PDGFRα-driven HGG was enhanced with everolimus and suggest a promising route for improving targeted therapy for this patient population.

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Introduction

High-grade gliomas (HGGs) are common and aggressive pediatric and adult brain tumors. The median overall survival (OS) of adult patients diagnosed with glioblastoma (GBM), grade IV glioma, is 12.6 months (1), and the median OS for pediatric HGG is 14.1 months (2). Established therapies for adult and pediatric HGG are not currently targeted to the unique molecular attributes of each tumor, and patients frequently consider experimental therapies after up-front radiation and at recurrence.

One of the most frequently altered genes in HGG is platelet-derived growth factor receptor α (PDGFRα). PDGFRα is 1 of 2 PDGF receptor subunits, and it interacts with at least 4 PDGF ligands (3). It has been shown that PDGF activation induces multiple cellular activities including cell proliferation, transformation, migration, and survival (3). PDGFRα is mutated or amplified in 12% of adult GBMs and 21% of pediatric HGGs (4, 5). Within childhood HGGs, PDGFRα mutation is correlated with older age (10–30 years of age), whereas amplification is more commonly seen in younger patients (5–10 years of age) (5). PDGFRα alterations are drivers of aggressive glioma behavior and are associated with worse prognoses in pediatric non-brainstem HGG and adult anaplastic astrocytoma (WHO grade III glioma) (5, 6).

Preclinically, there are multiple effective agents targeting PDGFRα in glioma, including the tyrosine kinase inhibitor (TKI) dasatinib (5, 7). Unfortunately, monotherapy with dasatinib has failed to improve outcomes in adult HGG, even when gliomas were selected for by PDGFRα overexpression using IHC (8). The failure of dasatinib may be related to its use as a single agent. Recent data in other solid tumors have demonstrated synergism between TKIs and the mTOR inhibitor everolimus, including the clinical success of lenvatinib and everolimus for renal cell carcinoma (9). Osteosarcoma preclinical models treated with sorafenib overcame resultant upregulation of mTOR complex 1 (mTORC1) through cotreatment with everolimus (10).

Additionally, treatment with dasatinib for CNS tumors may be limited by CNS penetration. Given its intrinsic qualities (i.e., lipophilicity, size, and protein binding), dasatinib is a promising agent for CNS penetration (11). Although the ability of dasatinib to penetrate the blood-brain barrier (BBB) is promising, its retention...
is limited by the cerebrospinal fluid (CSF) efflux protein P-glycoprotein (P-gp) and breast cancer resistance protein (BCRP) (12). Recent work has demonstrated that coadministration of TKIs with agents that inhibit P-gp and BCRP can improve brain retention of the TKI (12–14). Everolimus has been shown to improve CNS retention of the TKI vandetanib (13).

To our knowledge, coadministration of dasatinib and everolimus has never been tested in CNS tumor models or administered to patients with PDGFRA-driven gliomas. In this study, we assessed whether cotreatment of dasatinib with everolimus could improve the efficacy and CNS retention of dasatinib in PDGFRA-driven gliomas in both preclinical models and patients. We found that everolimus acted synergistically with dasatinib in PDGFRA-driven glioma. We also found that sustained everolimus administration resulted in substantial increases in tumor concentrations of dasatinib in a genetically engineered HGG animal model. Our preliminary data on 6 patients who received dasatinib and everolimus combination therapy establish its promising feasibility. Furthermore, our findings represent a route for enhancing the efficacy of combination therapy establish its promising feasibility. Further additional everolimus and dasatinib demonstrates a synergistic effect at nearly all dose

Results

Generation of TP53, PDGFRA, and H3K27M mutant glioma primary cultures and in vitro treatment with dasatinib. In order to study the utility of dasatinib in PDGFRA-driven HGG, we adapted an intraterrorine electroporation (IUE) mouse model (ref. 15 and Figure 1A). We induced glioma in mice by injecting plasmids encoding: (a) PBase; (b) PB-CAG-DNp53-Ires-luciferase (TP53); (c) PB-CAG-PDgfraD824V-Ires-EGFP (PDGFRA D824V); and (d) PB-CAG-H3.3 K27M-Ires-EGFP (H3K27M) into the lateral ventricles (forebrain) of E13.5 embryos in CD1 mice. Transfection efficiency and tumor growth were monitored by in vivo imaging of luminescence (Figure 1A). With the injection of PPK (mutant TP53, mutant PDGFRA, H3K27M) plasmids, mice developed high-grade, invasive glial tumors (Supplemental Figure 1A; supplemental material available online with this article; https://doi.org/10.1172/JCI133310DS1) with a high penetrance (>90%) and a median survival of 52 days. IHC analysis confirmed upregulation of total PDGFRA, tumor-specific expression of H3.3K27M, and reduced expression of H3.3K27me3, as expected for H3.3K27M-mutant tumors (ref. 16 and Supplemental Figure 1B). Multiple TP53, PDGFRA, and H3K27M (PPK) mutant glioma primary neurosphere cultures were generated from dissociation of intracranial tumor tissue from these mice (Figure 1A), with confirmed mutations in cultured neurospheres (Supplemental Figure 1C).

Previous studies have demonstrated that tumor cells driven by the PDGFRA D824V mutation may be relatively PDGF α and β ligand insensitive (as compared with PDGFRA amplification) (7). However, we found a dose-dependent increase in proliferation when the cells were supplemented with PDGF α and β ligands (Figure 1B). Cell cultures showed strong expression levels of phosphorylated [p-PDGFRA (Tyr849)] and total PDGFRA, which were not significantly different with the addition of ligand (Figure 1C and Supplemental Figure 1D). UMPED58, a brainstem HGG primary cell culture with PDGFRA amplification, also showed increased proliferation when grown with PDGF α and β ligands (P ≤ 0.0001) (Supplemental Figure 1E).

We tested in vitro dasatinib treatment on mouse PPK and several other human pediatric HGGs with PDGFRA alteration in the following cell cultures: 2 pediatric HGG (pHGG) cell cultures with PDGFRA amplification (UMPED58 and UMPEDOS; ref. 5); 3 PDGFRA-mutant adult HGG (aHGG) cultures derived from patient-derived xenografts (PDXs) (17); 1 pHGG cell culture with enhancer-mediated PDGFRA upregulation (DIPG-XIII-P; ref. 18); and 1 pediatric HGG cell culture without any growth factor receptor alteration (KNS42) as control (5). PPK cells with a PDGFRA D824V mutation showed sensitivity to monotherapy with dasatinib, with an IC50 of 100 nM (Figure 1D). All the pHGG cell cultures also demonstrated different degrees of nanomolar sensitivity to dasatinib; UMPED05 (PDGFRA amplification) showed maximum sensitivity, and KNS42 with WT PDGFRA was the least sensitive (Figure 1D). We analyzed publicly available data on human glioma cell culture treated with dasatinib (n = 11) from the Genomics of Drug Sensitivity in Cancer (GDSC) database (19). Glioma cell cultures showed promising sensitivity to dasatinib in general (Supplemental Figure 2). Dasatinib demonstrated improved efficacy in glioma cell cultures with PDGFRA alterations (Figure 1E). Collectively, these data showed that dasatinib effectively inhibits proliferation in mouse and human HGGs with various PDGFRA alterations.

Addition of everolimus provides synergism with dasatinib treatment of pHGG cells. Previous work has established that TKIs may have improved efficacy in combination with the mTOR inhibitor everolimus (9, 10). To determine whether the addition of everolimus can improve the efficacy of dasatinib, we treated PPK cells with varying concentrations of everolimus and dasatinib, which showed that at concentrations as low as 1 nM, everolimus resulted in a pronounced reduction in the dasatinib concentration required to achieve a reduction of mouse PPK cell proliferation in vitro, allowing for efficacy in PDGFRA-altered HGG cells at nanomolar concentrations of both drugs (Figure 2A). To assess whether this efficacy was improved via inhibition of the mTOR pathway, we performed Western blot analysis on mouse (PPK) and human (DIPG-XIII-P) cells treated with dasatinib and/or everolimus to measure the activity of RTK/MAPK/mTOR pathways (Figure 2B). Treatment with dasatinib alone resulted in reduced expression of p-PDGFRα/β (Tyr849/Tyr857) in a dose-dependent manner and of the RTK/MAPK targets p-Src (Tyr416) and p-ERK1/2 (Thr202/Tyr204), even at the lowest dose (0.01 μM) (Figure 2B and Supplemental Figure 3). Everolimus effectively inhibited the mTOR pathway [p-S6 (Ser235–236)] at the lowest dose (0.01 μM). The combination of everolimus and dasatinib led to dual inhibition of mTOR (p-S6) and RTK/MAPK (p-Src) pathways at 0.01 nM dose (Figure 2B). Quantification of Western blot data showed a reduction in residual mTOR pathway (p-S6) pathway expression after dasatinib treatment alone (Figure 2C and Supplemental Figure 4). We performed a neurosphere limiting dilution assay on IUE PPK cells in the micromolar range for both dasatinib and everolimus, which indicated no enhancement of the effect beyond the cytotoxicity seen in proliferation assays at this dose, suggesting that these agents do not have an added impact on the stemness of cultured cells (Figure 2D and Supplemental Figure 5A). Using the Chou-Talalay median-effect method for drug combination analysis, everolimus and dasatinib demonstrated a synergistic effect at nearly all dose.
combinations, including single nanomolar doses of both agents (Figure 2E and ref. 20).

*Everolimus provides P-gp blockade at a higher dose.* Previous work has established that dasatinib is a substrate for the efflux proteins P-gp and BCRP (21-24). As P-gp is probably the predominant substrate affecting CNS efflux (14), we focused our attention on P-gp. To assess the P-gp–inhibitory effects of everolimus, we performed an in vitro P-gp inhibition assay. Everolimus fully blocked P-gp activity at a dose of 1 μM at 4 hours (Figure 2F and Supplemental Figure 3B). Lower doses of everolimus (10 nM) did not produce an effect, which was significantly different from what was observed in nontreated cells (Figure 2F). The effect at 1 μM concentrations was similar to that seen with slightly lower-than-previously-levels of everolimus (3-5 μM) necessary for in vitro P-gp inhibition in Madin-Darby canine kidney (MDCK) cells overexpressing multidrug resistance genes (P-gp and BCRP) (25).

*Pharmacokinetic analysis of dasatinib and everolimus CNS retention.* We then proceeded to test the effect of everolimus on CNS concentrations of dasatinib in nontumor-bearing mice. Mice were treated with dasatinib (10 mg/kg via tail vein injection) with or without a single dose of everolimus (5 mg/kg via oral gavage) 2 hours prior. Mice were sacrificed at 1-, 2-, 4-, and 7-hour time points (n = 3 each) for collection of serum, brainstem, and cortex (Supplemental Figure 6A). We performed a separate analysis of brain and brainstem pharmacokinetics (PK), because previous literature and clinical experience suggest that the BBB is more restrictive in the brainstem (26). We found no statistically significant differences for dasatinib concentrations between the 2 conditions (dasatinib alone and dasatinib plus everolimus) within the plasma, cortex, and brainstem (Supplemental Figure 6, B–F). Average plasma concentrations of dasatinib were higher in the cotreated mice, although this finding was not statistically significant. PK analysis of everolimus monotherapy in nontumor-bearing mice revealed an increase in everolimus concentrations (ng/g) within mouse brain tissue over time (Supplemental Figure 7, A and B).
A. PPK dasatinib and everolimus

- Vehicle
- Dasatinib 1 nM
- Dasatinib 10 nM
- Dasatinib 100 nM
- Dasatinib 1 μM

B. PPK (mouse HGG)

- p-PDGFRαβ (Tyr579/Tyr567)
- PDGF γ total
- p-Src (Tyr416)
- Src total
- p-S6 (Ser235–236)
- S6 total
- α-Tubulin (total)

C. PPK (mouse)

- XIII-P (human)

D. Neurosphere limiting dilution assay

- DMSO
- Dasatinib 10 μM
- Dasatinib 1 μM
- Everolimus 10 μM
- Everolimus 1 μM
- Dasatinib + everolimus 5 μM

E. PPK dasatinib and everolimus synergism (Chou-Talalay)

F. P-gp inhibition assay

- Verapamil (neg. control)
- No treatment
- Everolimus 10 nM
- Everolimus 1 μM
- Na3VO4 (pos. control)
Sustained exposure of everolimus improves dasatinib tumor delivery and increases the survival of mice bearing PPK tumors. We then proceeded to study our IUE murine model of pHGG to assess whether the intratumoral concentration of dasatinib would be affected by cotreatment with everolimus. We performed treatment studies in mice with large cortical PPK tumors that met the enrollment luminescence criteria (average of 32 dpi). As seen in our nontumor-bearing mice, single-dose treatment of everolimus did not influence the tumor concentration of dasatinib (Figure 3A). However, with the addition of a second dose of everolimus 24 hours before dasatinib administration, tumor concentrations of dasatinib were increased in proportion to the dasatinib dose (Figure 3A). In additional tumor-bearing mice treated with 2 doses of everolimus before dasatinib administration, immediately before sacrifice, we detected reduced expression of P-gp by IHC (Supplemental Figure 8, A and B).

To assess the effect of sustained cotreatment on survival, we treated IUE PPK mice under 4 separate conditions: vehicle, daily treatment with dasatinib alone (10 mg/kg), daily treatment with everolimus alone (5 mg/kg), or daily cotreatment with dasatinib and everolimus (10 mg/kg and 5 mg/kg, respectively). Cotreatment with dasatinib and everolimus significantly improved median survival (86 days) compared with dasatinib alone (61 days), everolimus alone (55 days), and vehicle alone (52 days) (Figure 3B). Forty-seven days after IUE injection, the dasatinib- and everolimus-treated PPK mice demonstrated reduced luminescent tumor signals when compared with control mice (Figure 3, C and D). We further analyzed tumors from each treatment group for p-Src and p-S6 expression. IHC analysis and quantification showed reductions in p-Src and p-S6 expression after dasatinib and everolimus monotherapy, with retained p-S6 expression in tumors treated with dasatinib alone (Figure 3, E and F, and Supplemental Figure 9, A and B).

Treatment of children and young adults with relapsed PDGFRα-driven glioma with dasatinib and everolimus. Patients with recurrent HGG have no proven therapies and an expected OS of 4 to 6 months (27, 28). Given the promising preclinical data on dasatinib and everolimus, we moved forward in our pursuit of clinical treatment with daily dasatinib and everolimus for PDGFRα-driven glioma in patients. We treated a series of children and young adults with confirmed PDGFRα-driven glioma with continuous dasatinib (60 mg/m² orally, twice daily) and everolimus (3 mg/m² orally, once daily) at 4 academic institutions. We treated 4 patients with recurrent grade II–IV glioma and 2 patients with HGG after initial radiation (Table 1). All patients tolerated the combination with the exception of rash (n = 4 of 6), mucositis (n = 1), and fatigue (n = 1), none of which required a dose reduction. In patients with recurrent glioma (n = 4), the median progression-free survival (PFS) was 3 months (range, 2–5 months), and the median OS was 8.5 months (range, 3–16 months and ongoing) (Figure 4, A and B), although some patients received additional concurrent therapies (Table 1).

Additionally, to measure CSF and plasma levels of dasatinib with and without concurrent everolimus treatment, we collected paired CSF/plasma samples from 2 patients after 1 week of dasatinib monotherapy and then after 2 cycles of dual therapy with dasatinib and everolimus, following an IRB-approved protocol (Supplemental Figure 10A). Patient UMPEDM1 had an increase in both plasma (11.2 ng/mL to 31.5 ng/mL) and CSF (0.44 ng/mL to 0.91 mg/mL) dasatinib levels (Figure 4C and Supplemental Figure 10, A–C). Patient UMPED52 had an increase in plasma (26.4 ng/mL to 165 ng/mL) dasatinib levels, but we were unable to collect CSF samples (Supplemental Figure 11, A–D).

P-gp expression in human HGG RNA-Seq data sets. As P-gp is expressed in both brain and tumor cells and overall expression at the protein level was reduced in our mouse HGG models after sustained treatment with everolimus, we were interested in exploring whether P-gp expression correlated with clinical or genomic features in human data sets. Previous data have shown a correlation between P-gp mRNA and protein expression in human liver samples, with both predicting organ penetration of a Pgp-effluxed compound (29). We analyzed RNA-Seq data from pediatric HGGs within the PedBioPortal (Pediatric HGG ICR London data set, n = 247 with RNA-Seq data; ref. 2) and found that P-gp expression was highest in (a) pediatric hemispheric HGG, (b) tumors with WT H3F3A (gain of function mutations are found in half of pediatric HGGs), and (c) young adult–aged patients (10–15 years old) (Figure 5A). Importantly, P-gp expression was higher in pediatric HGG with PDGFRα alterations (Figure 5A, n = 147). In an analysis of adult HGGs (cBioPortal, The Cancer Genome Atlas [TCGA] data set, n = 585) (30), none of the clinical parameters showed a correlation with P-gp expression, including PDGFRα status, methylguanine-DNA methyltransferase (MGMT) status, and age at diagnosis (Supplemental Figure 12A). Future studies involving P-gp IHC of human tissue are needed to further explore possible correlations between P-gp expression and clinical or genomic features.

Discussion

Our study illustrates that everolimus improves the efficacy of dasatinib in the treatment of PDGFRα-driven glioma. Although response attribution is difficult for our heterogeneous consecutive cases, our early clinical experience with this treatment combination demonstrated excellent tolerance of the regimen and OS following disease progression that compared favorably with
Figure 3. Sustained exposure of everolimus improves dasatinib tumor concentration and efficacy in a mouse model. (A) PK data documenting an increase in dasatinib levels detected in tumors with an additional 24-hour dasatinib pre-everolimus dose of either 5 mg/kg or 10 mg/kg (*P < 0.05, by Dunnett’s multiple comparisons test). n = 5 replicates. (B) Survival curve data for mice with PDGFRα-amplified HGG via IUE shows that the median survival for the control condition was 52 days after IUE injection (n = 10), 55 days for the everolimus condition (5 mg/kg, n = 9), 61 days for the dasatinib condition (10 mg/kg, n = 9), and 86 days for the dasatinib plus everolimus condition (10 mg/kg and 5 mg/kg, respectively, n = 9) (NS P = 0.3378. ***P ≤ 0.0005, and ****P ≤ 0.0001, by log-rank test). (C) Representative bioluminescence images of PPK mouse tumors (vehicle, n = 10); dasatinib, n = 9; everolimus, n = 9; dasatinib + everolimus, n = 10) 47 days after IUE injection, displaying lower average luminescence in the cotreatment group than in the 2 groups receiving monotherapy of dasatinib or everolimus. (D) IUE PPK bioluminescence tumor monitor growth data with statistical significance between dasatinib plus everolimus and DMSO treatment groups 47 days after IUE injection (**P ≤ 0.0001, by Tukey’s multiple comparisons test). (E) In vivo IHC images of p-Src and p-S6 expression levels, documenting a reduction with everolimus and dasatinib treatment in comparison with nontreated tumors. Images are representative of 3 biological replicates. Original magnification, ×20. (F) IHC quantification data for treated PPK mice show statistical significance of p-Src and p-S6 expression levels between tumors treated with dasatinib and everolimus and nontreated tumors (**P = 0.0065 and ***P = 0.0001, by Dunnett’s multiple comparisons test). n = 3 animals per treatment group and 4 images per animal. Data represent the mean ± SEM (A, D, and F).

that of historical controls (27, 28). To our knowledge, cotreatment with everolimus and dasatinib had not previously been tested in glioma tumor models or patients. More broadly, our results demonstrate that TKI treatment targeted to alterations in HGG may have renewed clinical relevance when combined with mTOR inhibitors.

Importantly, our murine IUE tumor model experiments demonstrate that everolimus and dasatinib had improved combinatorial activity in the treatment of PDGFR-driven glioma cells. This may be due to the presence of dual inhibition within the MAPK/mTOR cell proliferation pathway, with everolimus reducing residual mTOR activation from dasatinib treatment alone (refs. 5, 31 and Figure 5B). Attribution to tumor cell toxicity from an improved intracellular dasatinib concentration versus direct mTOR inhibition was difficult to fully distinguish. However, we found no in vitro P-gp inhibition with everolimus treatment at a low dose (10 nM), whereas our treatment studies showed a marked contribution of everolimus to reduced tumor cell viability at doses as low as 1 nM. Murine PPK cells displayed dasatinib/everolimus synergism via the Chou-Talalay median-effect method for drug combination analysis (ref. 20 and Figure 2E). Future mechanistic studies may further clarify whether the benefit is additive (as opposed to synergistic).

In our in vivo model, administration of everolimus increased tumor concentrations of dasatinib above therapeutic (IC50) levels, and this increase required sustained (>24 hours) everolimus exposure. This requirement may be related to the time required for protein-level changes (i.e., reduced P-gp expression and/or activity) to occur. Interestingly, dasatinib concentrations increased more prominently in the CNS tumor samples than in the brain itself. This may be due to the disruption of the BBB in high-grade tumors, allowing for greater dasatinib influx (32). Indeed, our murine tumor had high-grade features, including necrosis and vascular proliferation, that may have influenced BBB integrity (Supplemental Figure 1A).

A concern with P-gp inhibition strategies that remains in our study is whether the inhibitor itself can penetrate the CNS at high enough concentrations to achieve P-gp inhibition (33). Our data showed a concentration of everolimus required for P-gp inhibition that was achievable in the plasma (>10 μM) and brain (<1 μM) of our mouse models, but may be more difficult to achieve in human brains with the current clinical trough goals (5-15 ng/mL, <5-15 nM). Interestingly, early adult phase I studies of everolimus involved weekly higher dosing of everolimus (up to 70 mg), which resulted in mean peak levels of everolimus of 175 ng/mL (190 nM) and a tolerability that was similar to or better than that seen with daily 10 mg dosage (34). Future studies may clarify whether everolimus inhibits P-gp at this dosage, or whether higher dosing of everolimus should be reconsidered. P-gp also promotes efflux on the luminal side of blood vessels (Figure 5B), and so future clinical efforts to achieve P-gp inhibition may be based on plasma (rather than brain parenchyma) concentrations of everolimus. At a minimum, based on our in vitro models, everolimus should contribute to synergistic cytotoxicity of PDGFRα-driven HGG at clinically relevant (<low nM) concentrations of everolimus and dasatinib.

Although everolimus may help prevent efflux of dasatinib from brain parenchyma and tumor cells, the efficacy of dasatinib will ultimately rely on the cells’ sensitivity to it, which may be variable based on the type of PDGFRα alteration expressed by the cells. The PDGFRα alteration in each glioma can strongly contribute to the sensitivity to dasatinib (7). Our PPK cells carried the recurrent PDGFRα D842V mutation (as did patient UMPED52) and displayed nanomolar reductions in proliferation in response to dasatinib. Previous studies have shown mixed sensitivity of the D842V mutant to dasatinib, including reduced sensitivity to dasatinib in PDGFRα D842V-mutant normal human astrocytes (NHAs) (7). However, nanomolar reductions in D842V-mutant kinase activity have been demonstrated in gastrointestinal stromal tumors (GISTs) (35). Of note, our IUE-based PPK cells retained endogenous, WT PDGFRα, which may have contributed to tumor proliferation and dasatinib sensitivity. Future studies will continue to clarify the responsiveness of various PDGFRα alterations to dasatinib sensitivity, but we believe our data demonstrate the promise of this dual therapy in both PDGFRα-mutant and -amplified gliomas.

Our clinical case series (n = 6) confirms the feasibility of this dual therapy in this very high-risk patient population. The patients tolerated this combination without significant adverse events. Efficacy is difficult to fully assess, given the heterogeneity and concurrent therapies in this group, but the median OS (8.5 months) for the 4 patients with recurrent pediatric glioma was slightly higher than that for historical controls for relapsed pediatric glioma (4-6 months) (27, 28). While we did find an increase in CSF dasatinib levels after everolimus coadministration in UMPED44, the overall CSF dasatinib concentration (<2 nM) still remained near or below the IC50 of most human PDGFRα-driven glioma cell cultures. We used CSF as the best proxy for dasatinib concentrations within the tumor, and it may not be an accurate reflection of the tumor dasatinib concentration. Indeed, in our murine model, cortex concen-
IUE was performed using sterile technique on isoflurane/oxygen-
Murine model of HGG using IUE for PDGFRα
HGG treatment, and the coadministration of these drugs for improved CNS penetration. The combination of dasatinib allowed for additional combinatorial efficacy and the potential for sustained coadministration of dasatinib and everolimus confirms that dasatinib effectively inhibited, and medicine approaches for these patient populations. Our study This shortcoming has exposed the need for new precision
failed to improve outcomes in both adult and pediatric patients.
escalation of this drug.
dosages. Future patient data will help clarify the CSF retention of
dasatinib, and future clinical studies may consider a further dose
treatment of dasatinib were consistently lower than tumor concen-
trations. Future patient data will help clarify the CSF retention of
dasatinib, and future clinical studies may consider a further dose
escalation of this drug.
Thus far, targeted treatments for PDGFRα-driven HGG have failed to improve outcomes in both adult and pediatric patients. This shortcoming has exposed the need for new precision medicine approaches for these patient populations. Our study confirms that dasatinib effectively inhibited PDGFRα, and the sustained coadministration of dasatinib and everolimus allowed for additional combinatorial efficacy and the potential for improved CNS penetration. The combination of dasatinib and everolimus represents a new avenue for adult and pediatric HGG treatment, and the coadministration of these drugs reopens the possibility of dasatinib being an effective therapy for PDGFRα-driven glioma.

Methods

Murine model of HGG using IUE
IUE was performed using sterile technique on isoflurane/oxygen-
anesthetized pregnant CD1 female mice at E13.5 (cortex), according to established methodology (36). All tumors for this study were generated by injecting plasmids into either the lateral ventricle (forebrain) or the fourth ventricle (hindbrain). In this study, we injected the following 4 plasmids together: (a) PBase, (b) PB-CAG-DNp53-Ires-luciferase (dominant-negative TP53, referred to herein as just TP53), (c) PB-CAG-PdgfraD842V-Ires-EGFP (PDGFRα D842V), and (d) PB-CAG-H3.3 K27M-Ires-EGFP (H3K27M), referred to herein as the PPK model.

Following anesthesia induction, carprofen was administered subcutaneously for additional analgesia. Uterine horns were exposed through a 1 cm incision, and individual embryos were digitally manipulated into the correct orientation for intraventricular injection. A pulled capillary needle was used to inject either the lateral or fourth ventricles with the DNA-dye mixture. Three to 4 plasmids were injected simultaneously, each at a final concentration of 1 μL total solution was injected a final concentration of 1 μg/μL, and 1–2 μL total solution was injected per embryo. DNA was then electroporated into cortical neural progenitors using 5 mm tweezer electrodes (BTX), applying 5 square pulses at 35 V, 50 ms each at 950 ms intervals. The embryos were then returned to the juveniles were weaned and separated by sex. The mice with a positive signal were monitored for tumors every day by observation and biweekly by bioluminescence imaging on an IVIS Spectrum imaging system until the signs of intracranial tumor burden ensued.

Generation of IUE primary HGG cell cultures
Mouse HGG primary cell cultures were generated by harvesting IUE tumors at the time of euthanasia. Tumors were located by GFP expression under an epifluorescence microscope (Olympus CKX41) at the time of resection. The tumor mass was gently homogenized and dissociated with nonenzymatic cell dissociation buffer (Gibco, Thermo

<table>
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<tr>
<th>Patient ID</th>
<th>Age (yr)</th>
<th>Diagnosis</th>
<th>Molecular</th>
<th>Cycles on therapy</th>
<th>PFS on therapy</th>
<th>Best response</th>
<th>OS after recurrence</th>
<th>Toxicity/other notes</th>
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<td>PDGFRα amplification, TP53 (R65*), CDKN2A loss</td>
<td>3 cycles</td>
<td>3 mo</td>
<td>SD (cycle 2)</td>
<td>8 mo</td>
<td>Skin rash (did not require dose reduction)</td>
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<td>Recurrent DIPG, H3 WT</td>
<td>PDGFRα mutation (D842V), ACVR2B (E347*)</td>
<td>4 cycles</td>
<td>5 mo</td>
<td>SD (cycles 2 + 4)</td>
<td>9 mo</td>
<td>No toxicity noted</td>
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<td>Recurrent diffuse astrocytoma, H3 WT (frontal-temporal)</td>
<td>PDGFRα indel (S1045fs), EGFR (A289D, P596L), PTPN11(R498W)</td>
<td>5 cycles</td>
<td>3 mo</td>
<td>SD (cycle 2)</td>
<td>16 mo (ongoing)</td>
<td>Skin rash (did not require dose reduction); received focal radiation between cycles 3 and 4</td>
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<tr>
<td>SCH01</td>
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<td>Recurrent HGG, H3 WT, secondary toependymoma radiation (posterior fossa)</td>
<td>PDGFRα amplification, KIT amplification, CDKN2A deletion, MTAP deletion, GAB2 amplification</td>
<td>2 cycles</td>
<td>2 mo (presumed, no further imaging)</td>
<td>Not assessed (no further imaging)</td>
<td>3 mo</td>
<td>Mild rash and fatigue (did not require dose reduction); oral etoposide added after the first cycle</td>
</tr>
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| NYU01     | 15      | Metastatic HGG, H3 WT, secondary to leukemia (multiple hemispheres) | PDGFRα mutation (D842V) and amplification, KIT mutation (D579N) and amplification | 5 cycles (ongoing) | N/A (no progression) | SD/minor response (cycle 3) | 8 mo (ongoing) | No toxicity noted; therapy started adjuvantly after initial radiation |
| UCSF01    | 8       | DIPG, H3K27M-mutant | PDGFRα upregulation (RNA); H3F3A (R27M), ATRX (R2111L), MTDOR (E1799K); PHK (E545K); PPM1D (E487*) | 3 cycles | 3 mo | SD (cycle 2) | 11.8 mo | Mucositis, oral pain, rash (did not require dose reduction); oral metformin and etoposide treatment given concurrently |

Table 1. Characteristics of patients with glioma

Patient characteristics are shown for patients who underwent treatment with dasatinib and everolimus across 4 academic institutions (n = 6).
In vitro treatment and synergism calculations

IUE-generated PDGFRα-driven HGG primary (PPK) cells were cultured in the conditions described above. Cell viability in vitro was monitored either with an XTT Cell Proliferation Assay Kit (Cayman Chemical) using the included protocol, or by bioluminescence readings collected using the Synergy HTX Multi-Mode Microplate Reader (BioTek Instruments). Additionally, cell proliferation was monitored via IncuCyte ZOOM (Sartorius) live-cell analysis. Through use of the IncuCyte ZOOM live imager, the growth of 3000 PPK primary cells, plated in 96-well plates, was monitored according to units of average green fluorescence intensity (green calibrated unit [GCU] × μm²). PPK primary cell culture was grown with 200 μL media and 1 μL PDGF-AA and PDGF-BB at a working concentration of 10 ng/mL in each 96-well plate (Shenandoah Biotechnology). All in vitro treatments were completed with research-grade dasatinib and everolimus (Selleck Chemicals). UMPED58 is a primary human cell line generated from a 7-year-old male with diffuse intrinsic pontine glioma (DIPG) at autopsy. At diagnosis, the tumor was sequenced and found to carry H3F3A K27M, ATRX pQ199*, PDGFRα, KIT, and KDR amplifications. UMPED05 is a primary cell line generated from a 2-year-old female with thalamic HGG with PDGFRα amplification at autopsy, as previously described by Koschmann et al. (5).

Figure 4. Patient survival curves and treatment of UMPED44 with relapsed PDGFRα-amplified HGG. (A) PFS curve for patients with recurrent PDGFRα-driven HGG treated with dasatinib and everolimus (n = 4). Dashed lines represent a 95% confidence interval. (B) OS curve for patients with recurrent PDGFRα-driven HGG treated with dasatinib and everolimus (n = 4). Dashed lines represent a 95% confidence interval. (C) UMPED44 axial FLAIR MRI at baseline, before cycle 3, and before cycle 4 (MR spectroscopy in cycle 4 confirmed tumor progression).
Figure 5. P-gp expression in human pediatric HGG and proposed schema. (A) PedcBioPortal (pediatric HGG) P-gp RNA-Seq data by location (upper left), histone H3 mutation status (upper right), PDGFRA status (lower left), and age (lower right) (****P ≤ 0.0001, by Tukey’s multiple comparisons test). Data presented in the upper 2 graphs and lower left graph reflect the mean ± SEM. (B) Proposed schema of dasatinib and everolimus targeting of PDGF-driven HGG. Red line depicting inhibition of P-gp/BCRP by everolimus is dashed to reflect the higher concentration (μM) required to achieve P-gp inhibition than to achieve MAPK/mTOR inhibition (nM). yo, years old.
PK) analysis of tumors. Some mice (n = 3) were given an initial treat-
ment dose for their assigned arm 4 hours before euthanization. For
intratumoral concentration studies, all doses of everolimus were
administered to mice to 24 hours before dasatinib administration, con-
sistent with previous methods used for in vitro analysis of mTOR/
MAPK regulation after treatment with everolimus (10). For immuno-
histochemical analysis, mice were perfused with Tyrode’s Solution
followed by 4% paraformaldehyde fixative solution to preserve the
structures of the brain.

PK analysis
Mouse PK sample procurement. Drug administration to nontumor-
bearing C57/Bl6 mice for PK studies was performed by oral gavage of
everolimus 2 hours before a tail vein injection of dasatinib. One, 2, 4,
and 7 hours after the dasatinib injection, the mice were anesthetized
with isoflurane/oxygen, and 500 μL to 1 mL blood was drawn from the
apex of the heart within the mouse’s enclosed cavity. Immediately, the
withdrawn blood was centrifuged within a microvette EDTA-coated conical
tube for 10 minutes at 2000g, and the plasma was separated and stored
at −80°C until PK analysis was performed. Following the blood draw, the mouse was sacrificed, and the brain, brainstem, and/or
or tumor were extracted separately and stored at −80°C until PK anal-
ysis was performed.

Chemicals and reagents. For PK studies, dasatinib powder was
procured from MilliporeSigma. Liquid chromatography-mass spec-
trometry-grade (LC-MS-grade) acetonitrile was purchased from
MilliporeSigma. Formic acid (98%, LC-MS grade) was obtained from
Fluka. A Milli-Q water system from MilliporeSigma was used to obtain
ultrapure deionized water.

Stock solutions, working solutions, and quality control samples
Dasatinib and the internal standard were individually weighed and
dissolved in acetonitrile to stock solutions and then stored at −20°C. The
dasatinib stock solution was then diluted with acetonitrile to a series of working solutions from 2.5 to 5000 ng/mL. The quality control
working solutions at low, medium, and high concentrations were
prepared using a separately prepared stock solution. For sample prepa-
ration, the dasatinib stock solution was diluted to 1000 ng/mL with
acetonitrile. Quality control samples were evenly distributed among
samples from each batch.

Sample preparation. Plasma (40 μL) was dispensed into a Fisher
Scientific 96-well plate, to which 40 μL ice-cold acetonitrile (100%) and
120 μL internal standard solution (1000 ng/mL) were added. Next, the plate was vortexed for 10 minutes. The plate was then cen-
trifuged at 3500 rpm for 10 minutes at 4°C to precipitate the protein.
LC–tandem mass spectrometry (LC-MS/MS) was used to analyze 5 μL of the supernatant. The plasma samples were sonicated before being
transferred into the 96-well plates. Tissue samples were weighed and
suspended in 20% acetonitrile (80% water; 1:5 wt/vol). The samples
were then homogenized 4 times for 20 seconds each time at 6500 rpm
in a Precellys Evolution system (Bertin Instruments). For LC-MS/MS
analysis, the dasatinib in brain tissue homogenates was extracted from
the samples in the same manner as for the dasatinib in plasma. Prior
to extraction, samples that were above the upper limit of qualifica-
tion were diluted with the same matrix. Calibrator-standard samples
and quality control samples were prepared by mixing 40 μL blank bio-
matrix, 40 μL working solution, and 120 μL internal standard solution.

Calibration curve. Analytical curves were made with 12 nonzero
standards by plotting the peak area ratio of dasatinib to the internal
standard versus the concentration. The curve was created with linear
regression and weighted (1/x²). The correlation coefficient demon-
strated the linearity of the relationship between peak area ratio and
concentration.

LC-MS/MS. The concentrations of dasatinib were determined with
a SCIEX AB-5500 QTRAP mass spectrometer with an electrospray ionization source, interfaced with a Shimadzu high-performance LC system. The LC-MS/MS system was controlled with Ana-
lyst Software, version 1.6 (Applied Biosystems), which was also used for data acquisition and processing. Separation was performed on a Waters Xbridge C18 column (50 × 2.1 mm ID, 3.5 μm); the flow rate
was 0.4 mL/min. A (100% H₂O with 0.1% formic acid) and B (100% acetonitrile with 0.1% formic acid) comprised the mobile phase. The gradient began with 5% B for 30 seconds and then linearly increased to 99% B at 2 minutes, and was then reduced to 5% B at 4.1 min-
tues to 5.5 minutes with a runtime of 6 minutes in total. The mass spectrometer was operated in positive mode, and multiple reaction
monitoring was used for analysis. The Q1 m/z and Q3 m/z were 487.9 and 401.1, respectively.

Human treatment studies
All patients received dasatinib 60 mg/m² orally twice daily and evero-
limus 3 mg/m² orally once daily continuously, with titration of dosing to maintain an everolimus trough level of 5–15 ng/mL. Four patients were treated off-therapy (patients UMPED68, SCH01, NYU01, and
UCSF01) and 2 patients (UMPED44 and UMPED52) were treated in an IRB-approved clinical trial. Enrollment required Clinical Labora-
tory Improvement Amendments-certified (CLIA-certified) confirma-
tion of a DNA (mutation, amplification) or RNA (fusion) alteration in a PDGF-related gene (PDGFRα, PDGFRβ, PDGFRB, PDGFB). Paired
CSF and plasma samples (before and after addition of everolimus)
were collected from enrolled patients when clinically feasible. Treat-
ment cycles were repeated every 28 days for up to 24 cycles. CSF and
serum dasatinib PK analyses were performed by the University of
Michigan Pharmacokinetic Core Facility (see Supplemental Methods
to determine whether blood-CSF permeability of dasatinib was
affected by dual therapy with everolimus. Prior to the first cycle, dasatinib was taken as monotherapy for 1 week followed by CSF col-
lection in order to establish a monotherapy dasatinib CSF level. After
the second cycle, CSF was collected to establish a dual-therapy dasat-
inib level (while on everolimus). Additionally, all CSF samples were
assessed for cell-free tumor DNA (cf-tDNA) using previous methods
(39), when sample remained.

Human HGG clinical sequencing and PDGFRα alteration confirmation
For confirmation of PDGFRα status in HGG patients, tumor (formalin-
fixed, paraffin-embedded [FFPE] or frozen) and normal (cheek swab)
samples were submitted for whole-exome (paired tumor and germ-
line DNA) and transcriptome (tumor RNA) sequencing. Clinically
integrated sequencing was performed after enrollment in the Univer-
sity of Michigan’s IRB-approved Pediatric MiOncoSeq study using
standard protocols, in adherence with CLIA guidelines. All somatic
mutation information generated from this study has been uploaded to
the Database of Genotypes and Phenotypes (dbGaP) (https://www.
Human P-gp expression
Pediatric and adult human P-gp expression data set analysis was performed using RNA-Seq data accessed from the PedcBioPortal (https://d3b.center/our-research/pedcbiportal/). For pediatric HGG, data were accessed from the Pediatric HGG ICR London (2). We performed comparisons of P-gp expression by location of tumor, H3 (H3F3A or HIST1H3B) mutation status, PDGFRA mutation status, and age. For adult HGG, data were accessed from TCGA data set (30). For TCGA (adult HGG) P-gp RNA, we compared P-gp expression levels by PDGFRA mutation status, age, and MGMT status.

Statistics
For comparison of dose-response curves using DNA-damaging agents, a nonlinear regression curve [log (agent) versus normalized response with variable slope] was generated. Statistical significance in all experiments was defined as a 2-sided P value of 0.05 or less. All analyses were conducted with GraphPad Prism, version 7.0 (GraphPad Software). Expression levels of p-Src and p-S6 were quantified using IHC performed on mouse brain sections treated with everolimus and/or dasatinib. Four randomly selected JPEG images were captured from each IHC slide at ×12.3 magnification on an Aperio ImageScope (Leica Biosystems) and quantified using ImageJ software (NIH). ImageJ software was also used to quantify IHC and Western blot expression levels as well as the PPK neurosphere area in squared pixel units. Comparisons between treatment groups in experiments with only 2 groups were completed using a 2-tailed Welch’s t test. Comparisons between treatment groups in experiments with more than 2 groups were done using Tukey’s or Dunnett’s multiple comparisons test. Survival curve comparisons between treatment groups were performed using the log-rank test.

Study approval
Animal studies. All animal studies were conducted according to guidelines approved by the University Committee on the Use and Care of Animals (UCUCA) of the University of Michigan.

Human studies. Four patients (UMPED68, SCH01, NYU01, and UCSF01) were treated off-therapy according to IRB-approved protocols for molecular tumor assessment. Two patients (UMPED44 and UMPED52) were treated in an IRB-approved clinical trial with dasatinib and everolimus for children and adults with newly diagnosed (after radiation) high-grade (grade III–IV) glioma or recurrent grade II–IV glioma with PDGF alterations (NCT03352427).

Author contributions
ZM, VNY, BM, RTC, CCT, and C. Koschmann designed the in vitro experiments and performed experiments and data analysis. ZM, VNY, BM, TG, TNP, TY, RTC, CCT, JRC, DRW, and JNS designed the in vivo experiments and conducted data analysis. ZM, VNY, RS, BLM, KW, AKB, SS, HJLG, AP, IW, ML, PLR, HP, SESL, SV, TN, CG, C. Kline, SM, CKS, AMC, RM, and C. Koschmann performed human clinical studies. ZM, VNY, MPP, BLM, and C. Koschmann performed PK analyses. ZM, VNY, BLM, JRC, C. Koschmann, RS, RTC, and SV prepared the manuscript. All authors discussed and reviewed the manuscript and approved the manuscript for publication.

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