Rectal tissue and vaginal tissue from intravenous VRC01 recipients show protection against ex vivo HIV-1 challenge

Rena D. Astronomo, …, Adrian B. McDermott, M. Juliana McElrath


VRC01, a potent, broadly neutralizing monoclonal antibody, inhibits simian-HIV infection in animal models. The HVTN 104 study assessed the safety and pharmacokinetics of VRC01 in humans. We extend the clinical evaluation to determine intravenously infused VRC01 distribution and protective function at mucosal sites of HIV-1 entry.

Healthy, HIV-1–uninfected men (n = 7) and women (n = 5) receiving VRC01 every 2 months provided mucosal and serum samples once, 4–13 days after infusion. Eleven male and 8 female HIV-seronegative volunteers provided untreated control samples. VRC01 levels were measured in serum, secretions, and tissue, and HIV-1 inhibition was determined in tissue explants.

Median VRC01 levels were quantifiable in serum (96.2 μg/mL or 1.3 pg/ng protein), rectal tissue (0.11 pg/ng protein), rectal secretions (0.13 pg/ng protein), vaginal tissue (0.1 pg/ng protein), and cervical secretions (0.44 pg/ng protein) from all recipients. VRC01/IgG ratios in male serum correlated with those in paired rectal tissue (r = 0.893, P = 0.012) and rectal secretions (r = 0.9643, P = 0.003). Ex vivo HIV-1_{Bal26} challenge infected 4 of 21 rectal explants from VRC01 recipients versus 20 of 22 from controls (P = 0.005); HIV-1_{Du422.1} infected 20 [...]

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Rectal tissue and vaginal tissue from intravenous VRC01 recipients show protection against ex vivo HIV-1 challenge

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BACKGROUND. VRC01, a potent, broadly neutralizing monoclonal antibody, inhibits simian-HIV infection in animal models. The HVTN 104 study assessed the safety and pharmacokinetics of VRC01 in humans. We extend the clinical evaluation to determine intravenously infused VRC01 distribution and protective function at mucosal sites of HIV-1 entry.

METHODS. Healthy, HIV-1–uninfected men (n = 7) and women (n = 5) receiving VRC01 every 2 months provided mucosal and serum samples once, 4–13 days after infusion. Eleven male and 8 female HIV-seronegative volunteers provided untreated control samples. VRC01 levels were measured in serum, secretions, and tissue, and HIV-1 inhibition was determined in tissue explants.

RESULTS. Median VRC01 levels were quantifiable in serum (96.2 μg/mL or 1.3 pg/ng protein), rectal tissue (0.11 pg/ng protein), rectal secretions (0.13 pg/ng protein), vaginal tissue (0.1 pg/ng protein), and cervical secretions (0.44 pg/ng protein) from all recipients. VRC01/IgG ratios in male serum correlated with those in paired rectal tissue (r = 0.893, P = 0.012) and rectal secretions (r = 0.9643, P = 0.003). Ex vivo HIV-1Bal26 challenge infected 4 of 21 rectal explants from VRC01 recipients versus 20 of 22 from controls (P = 0.005); HIV-1Bal26 infected 20 of 21 rectal explants from VRC01 recipients and 12 of 12 from controls (P = 0.639). HIV-1Du422.1 infected 0 of 14 vaginal explants of VRC01 recipients compared with 23 of 28 control explants (P = 0.003).

CONCLUSION. Intravenous VRC01 distributes into the female genital and male rectal mucosa and retains anti–HIV-1 functionality, inhibiting a highly neutralization-sensitive but not a highly resistant HIV-1 strain in mucosal tissue. These findings lend insight into VRC01 mucosal infiltration and provide perspective on in vivo protective efficacy.

FUNDING. National Institute of Allergy and Infectious Diseases and Bill & Melinda Gates Foundation.
Table 1. Baseline demographic and clinical characteristics of study participants separated by HVTN 104 participation and sex assigned at birth

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>VRC01-infused HVTN 104 Male</th>
<th>Control Male</th>
<th>VRC01-infused HVTN 104 Female</th>
<th>Control Female</th>
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<tbody>
<tr>
<td>Number</td>
<td>7</td>
<td>10</td>
<td>5</td>
<td>8</td>
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<tr>
<td>Sample collection time point (days after infusion)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Median</td>
<td>10</td>
<td>NA</td>
<td>6</td>
<td>NA</td>
</tr>
<tr>
<td>Range</td>
<td>6–13</td>
<td>NA</td>
<td>4–12</td>
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<tr>
<td>VRC01 dose infused (mg)</td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Median</td>
<td>1109</td>
<td>NA</td>
<td>1630</td>
<td>NA</td>
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<tr>
<td>Range</td>
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<td>610–2520</td>
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<td></td>
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<td></td>
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<tr>
<td>Median</td>
<td>31</td>
<td>32</td>
<td>28</td>
<td>30</td>
</tr>
<tr>
<td>Range</td>
<td>24–43</td>
<td>26–50</td>
<td>21–37</td>
<td>24–38</td>
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<td>Race</td>
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<tr>
<td>White</td>
<td>71% (n = 5)</td>
<td>60% (n = 6)</td>
<td>40% (n = 2)</td>
<td>62.5% (n = 5)</td>
</tr>
<tr>
<td>African American</td>
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<td>20% (n = 2)</td>
<td>40% (n = 2)</td>
<td>25% (n = 2)</td>
</tr>
<tr>
<td>Mixed race</td>
<td>14% (n = 1)</td>
<td>20% (n = 2)</td>
<td>20% (n = 1)</td>
<td>12.5% (n = 1)</td>
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<tr>
<td>American Indian, Hispanic</td>
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<td>10% (n = 1)</td>
<td>0% (n = 0)</td>
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<tr>
<td>Sex partners</td>
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<td></td>
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<td></td>
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<tr>
<td>Female</td>
<td>57% (n = 4)</td>
<td>70% (n = 7)</td>
<td>20% (n = 1)</td>
<td>12.5% (n = 1)</td>
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<tr>
<td>Male</td>
<td>28% (n = 2)</td>
<td>30% (n = 3)</td>
<td>60% (n = 3)</td>
<td>75% (n = 6)</td>
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<tr>
<td>Both</td>
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<td>20% (n = 1)</td>
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<td>100% (n = 2)</td>
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<td>HIV-1 serology negative</td>
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<td>100% (n = 10)</td>
<td>100% (n = 2)</td>
<td>100% (n = 8)</td>
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<td>HSV-2 serology positive</td>
<td>28% (n = 2)</td>
<td>30% (n = 3)</td>
<td>20% (n = 1)</td>
<td>25% (n = 2)</td>
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<tr>
<td>Female contraception</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Oral contraceptives</td>
<td>NA</td>
<td>NA</td>
<td>40% (n = 2)</td>
<td>25% (n = 2)</td>
</tr>
<tr>
<td>Condoms</td>
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<td>NA</td>
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<td>62.5% (n = 5)</td>
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<tr>
<td>Hormonal intrauterine device</td>
<td>NA</td>
<td>NA</td>
<td>20% (n = 1)</td>
<td>0% (n = 0)</td>
</tr>
<tr>
<td>None</td>
<td>NA</td>
<td>NA</td>
<td>20% (n = 1)</td>
<td>12.5% (n = 1)</td>
</tr>
</tbody>
</table>

*All enrolled females were premenopausal. HSV-2, herpes simplex virus 2.

numeros monoclomonal bnAbs (bn-mAbs) of exceptional potency and breadth have been cloned from B cells of HIV-1-infected donors (19, 20). Availability of these bn-mAbs has renewed enthusiasm for passive immunization strategies to prevent HIV-1 infection.

VRC01, a human IgG1 anti–HIV-1 bn-mAb, is the first prototype used to test the concept that passively transferred bn-mAbs can provide antibody-mediated protection against HIV-1 in at-risk populations (21). VRC01 neutralizes HIV-1 by binding to the conserved CD4-binding site on HIV-1 Env (22), thus preventing viral entry into target cells (23). Against a global panel of 190 viral reference strains, VRC01 neutralizes approximately 90% of these viruses in vitro, with a geometric mean 50% inhibitory concentration (IC50) of 0.33 μg/mL and IC80 of 1.0 μg/mL (22, 24). Passively administered VRC01 protects against rectal and vaginal challenge in several animal models of HIV transmission (25–31). In nonhuman primate challenge studies, the infused bn-mAb was detected in mucosal secretions and tissues (26). Notably, tissue rather than serum or secretion levels of VRC01 were associated with protection, and these decayed more rapidly in rectal compared with vaginal tissue (26).

In phase 1 human trials (VRC 601, VRC 602, and HVTN 104), VRC01 infusions proved safe and well tolerated, and demonstrated expected pharmacokinetic (PK) properties for a human IgG1 in both adults without HIV-1 (32, 33) and adults living with HIV-1 (34, 35). Moreover, a single infusion of the bn-mAb in people living with HIV-1 demonstrated a modest clinical antiviral effect (34–36). VRC01 PK and functionality following subcutaneous and/or i.v. administrations were evaluated in a multisite, phase Ib study in healthy adults (HVTN 104; ClinicalTrials.gov NCT02165267; ref. 32). Two groups in HVTN 104 received 3 i.v. infusions of VRC01 at 10 mg/kg (T4) or 30 mg/kg (T5) administered at 2-month intervals. Over a 24-week period, functional bn-mAb was maintained in the participant sera at concentrations predicted to neutralize a majority of circulating HIV strains in vitro (IC50 > 5 μg/mL) (32). Thus, these VRC01 infusion regimens were tested in parallel phase Ib efficacy trials of antibody-mediated prevention (AMP) in the Americas (HVTN 704/ HPTN 085; NCT02716675) and in sub-Saharan Africa (HVTN 703/ HPTN 081; NCT02568215) for prevention of HIV infection in populations at high risk of sexual exposure to HIV-1 (37).

Both the level and the function of bnAbs in gastrointestinal and genital tract secretions and tissues are potentially key to the success of passive immunoprophylaxis approaches that aim to prevent HIV-1 transmission through sexual exposure (26, 38). To address these features in humans for the first time to our knowledge, we designed and implemented a nested observa-
of the bn-mAb. Enrollment criteria in HVTN 104 (32) included healthy participants of both female and male sex assigned at birth, aged 18 to 50 years, weighing between 53 and 115 kg, HIV-1 sero-negative, and amenable to HIV risk reduction counseling. Female participants were not pregnant and agreed to consistent use of contraception or abstinence during the conduct of the study.

Seven male and five female eligible and consenting HVTN 104 participants (VRC01-infused group) underwent peripheral blood, mucosal secretion, and biopsy collections in a single visit at the Seattle site (Seattle Vaccine Trials Unit, Fred Hutchinson Cancer Research Center) (Table 1 and Figure 1, left). Male participants donated serum, rectal biopsies, and secretions at a median postinfusion interval of 10 days (range 6–13 days); female participants donated serum, vaginal biopsies, and cervical secretions outside of menses at a median postinfusion interval of 6 days (range 4–12 days). Most participants (9/12) completed all three HVTN 104-
scheduled infusions (1 infusion every 2 months) prior to their nested mucosal visit. One male participant (PTID4) and 2 female participants (PTID8 and PTID12) completed only 2 infusions, either the first and second with sample collection conducted after the second infusion (PTID8), or the first and third infusions with sample collection conducted after the third infusion (PTID4 and PTID12).

All HVTN 104 placebo recipients were already outside of our study window at the time of nested study enrollment. Thus, we recruited and screened uninfused control participants in Seattle. Ten male and 8 female control participants donated similar specimens from June 2015 to June 2016 and shared demographic and clinical characteristics similar to those of participants enrolled from HVTN 104 (Table 1 and Figure 1, middle and right). No procedure-related complications were reported.

**VRC01 levels in systemic, rectal, and cervicovaginal compartments following i.v. infusion.** We determined total protein, total IgG, and VRC01 levels in blinded serum and mucosal samples collected from VRC01-infused and uninfused control participants. Because of the anticipated low concentrations of VRC01 in mucosal samples, we used an anti-idiotypic mAb, 5C9, in a highly sensitive, custom Erenna Single Molecule Counting (SMC) immunoassay (Singulex/EMD Millipore) to measure VRC01 levels in all samples (39). Serum VRC01 levels were uniformly higher in the infused (median 96.2 μg/mL, IQR 60.7–105.5) compared with the control group (median and IQR below the lower limit of quantitation [LLOQ]; Mann-Whitney P < 0.0001; Figure 2A and Supplemental Table 1; supplemental material available online with this article; https://doi.org/10.1172/JCI146975DS1). Serum total protein and IgG levels were comparable among all participants (Mann-Whitney P = 0.051 and P = 0.211, respectively; Figure 2A). Infusion dose–normalized VRC01 serum concentrations in female participants (median 1.23/L, IQR 1.23–1.26) tended to be higher than but not significantly different from those in male participants (median 1.12/L, IQR 1.09–1.16; P = 0.090) after accounting for differences in the collection times (Table 1). The 10 mg/kg VRC01 dose group (T4) had lower serum bn-mAb concentrations (median 61.6 μg/mL, IQR 48.3–74.3) than the 30 mg/kg group (T5) (median 104.2 μg/mL, IQR 98.6–198.3; P = 0.004; Figure 2B).

All serum VRC01 concentrations measured by the Erenna assay in infused participants were within the range of values predicted by statistical modeling of participant-specific, serum VRC01 pharma-
Table 2. Protein- and IgG-normalized VRC01 levels

<table>
<thead>
<tr>
<th>Sample type</th>
<th>Protein-normalized VRC01</th>
<th>IgG-normalized VRC01</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>N</td>
<td>Median</td>
</tr>
<tr>
<td>Serum (male)</td>
<td>7</td>
<td>1.21</td>
</tr>
<tr>
<td>Serum (female)</td>
<td>5</td>
<td>1.31</td>
</tr>
<tr>
<td>Rectal secretion</td>
<td>7</td>
<td>0.13</td>
</tr>
<tr>
<td>Rectal tissue lysate</td>
<td>7</td>
<td>0.11</td>
</tr>
<tr>
<td>Cervical secretion</td>
<td>5</td>
<td>0.44</td>
</tr>
<tr>
<td>Vaginal tissue lysate</td>
<td>5</td>
<td>0.10</td>
</tr>
</tbody>
</table>

*Concentrations reported as pg VRC01 per ng total protein. *Concentrations reported as pg VRC01 per ng IgG. "Sex assigned at birth.

in female participants, VRC01 levels were also highest in serum but were not significantly different from those in matched cervical secretions (Figure 2C and Table 2). Vaginal tissues had approximately 13-fold lower VRC01 levels than sera. These findings suggest that distinct VRC01 levels are achieved within the cervical and vaginal compartments within 2 weeks of i.v. administration. As with PTID4, serum and mucosal VRC01 levels in the 2 donors who had received only 2 infusions (PTID8 and PTID12) were not lower than those of the other participants who had not missed a scheduled infusion (Figure 2C), suggesting that the last infusion received by the participants largely determined the antibody levels at the study collection time point. This is consistent with the previously reported approximately 15-day half-life of serum VRC01 (32), and it allows us to combine participants receiving 2 and 3 infusions in our comparisons.

We also compared VRC01 levels relative to total IgG across matched samples (Table 2 and Figure 2C). In male participants, IgG-normalized VRC01 levels in serum were not significantly different from the levels in rectal secretions or rectal tissues. Similarly, no differences were identified between IgG-normalized VRC01 levels in sera, cervical secretions, and vaginal tissues from female participants. The similar ratios of VRC01 to total IgG found across participant-matched sample types indicate that VRC01 transport into tissues is proportional to other IgGs in circulation. The differences between protein-normalized and IgG-normalized comparisons reflect that IgG is a less dominant protein in mucosal samples than serum and that the proportion of IgG in total protein varies across participant-matched sample types.

Protection of rectal and vaginal biopsies from VRC01-infused participants against ex vivo HIV-1 challenge. To assess the antiviral functionality of VRC01 in rectal tissue explants from infused (n = 7) and control (n = 10) male participants, we used an ex vivo HIV challenge assay employing 2 recombinant strains, one sensitive (HIV-1_Bal26; Figure 3) and one resistant (HIV-1_Du422.1; Figure 4) to VRC01 in vitro neutralization. Sections of rectal biopsies were challenged with replication-competent, secreted nanoluciferase (sNLuc) reporter viruses expressing the Env coxsackievirus B (CIB) reporter viruses expressing the Env of the Erenna (Supplemental Table 2). For simplicity, these reporter viruses are hereafter referred to as Bal26 and Du422.1. Since sNLuc is generated during viral stock preparation, productive infection was defined as sNLuc activity in relative light units (RLU) that is above the level corresponding to residual input sNLuc that dilutes out at each change of medium (black dashed lines in Figure 3A and Figure 4A). Total replication was summarized using area under the curve (AUC) for statistical comparisons (Figure 3B and Figure 4B).

As expected, ex vivo rectal challenge with either virus resulted in productive infection of explants from the male controls, thus verifying the infectivity of both HIV-1 reporter viruses (Figure 3B and
Figure 4B). Addition of exogenous VRC01 24 hours before challenge of rectal explants from control donors demonstrated that Bal26 explant infection could be blocked by exogenous VRC01 in a concentration-dependent manner (Figure 3B), unlike Du422.1 infection (Figure 4B). Whereas 1 μg/mL of VRC01 significantly inhibited Bal26 infection (Dunn’s P = 0.046), the same or even a 10-fold higher concentration did not affect Du422.1 infection (Dunn’s P = 0.999 and P = 0.819, respectively). The addition of seminal proteins during the Bal26 viral challenge did not alter rectal tissue infectivity or VRC01-mediated protection (Supplemental Figure 4), so all challenges in the study were performed in the absence of semen.

From VRC01-infused male participants (n = 7), only 4 of 21 rectal explants were infected by Bal26 challenge compared with 20 of 21 explants infected by Du422.1 challenge (Figure 3A and Figure 4A). The significant difference in total replication of Bal26 compared with Du422.1 (Wilcoxon’s P = 0.016, n = 7) indicates that i.v.-infused VRC01 reaches and retains neutralization functionality in the rectal mucosa. Additionally, explants from the VRC01-infused group (n = 7) were significantly resistant to Bal26 infection compared with those from the controls (n = 5) (Dunn’s P = 0.022; Figure 3B). By contrast, rectal explants from both VRC01-infused (n = 7) and control groups (n = 6) were susceptible to Du422.1 infection, resulting in comparable levels of viral replication (Dunn’s P = 0.999; Figure 4B). Within the VRC01-infused group, the high rate and magnitude of productive ex vivo Du422.1 infection also demonstrated that these participants’ rectal tissue was not intrinsically resistant to HIV-1 (Figure 4, A and B). Together, these results show that i.v.-infused VRC01 provides significant but incomplete protection against ex vivo HIV-1 challenge of rectal tissues collected 6–13 days after infusion, consistent with its neutralization activities.
Furthermore, we investigated whether genetic resistance against VRC01 binding had developed ex vivo, which can occur early (34, 44), causing the breakthrough viral replication observed in 4 Bal26-challenged explants from VRC01-infused donors (Figure 3, A and B). We sequenced a 3036 bp fragment (corresponding to HXB2 5969–9015) of HIV-1 Bal26 env in the culture supernatants (pooled from day 15–18) of the 4 explants in question (Figure 3A, PTIDs 1, 5, and 6), 3 untreated control explants, and the original Bal26 viral challenge stock. For all but 1 sample (PTID5, no virus cDNA from 1 of 2 explant cultures), we obtained a minimum of 230× coverage across all the bases, using Illumina Nextera XT indexing and sequencing on the MiSeq NGS platform (Supplemental Methods). All viral sequences were identical within and between samples except for a single G to A substitution at position 2539 identified in 1 untreated control explant culture. Thus, we found no evidence that genetic resistance to VRC01 binding developed during the 20-day explant culture of VRC01-infused biopsies. Rather, these results suggest that the breakthrough infections in the VRC01-infused group were the result of neutralization-sensitive Bal26 virions that were able to establish infection in the face of subprotective levels of VRC01 in these explants.

To assess the anti–HIV-1 functionality of i.v.-infused VRC01 in the female genital tract, we likewise compared the ability of Bal26 to establish infection in vaginal biopsies collected from VRC01-infused (n = 5) and uninfused control (n = 8) female participants in an ex vivo challenge assay. Vaginal biopsies are less amenable than rectal explants to dissection for ex vivo challenge because of the lower density of HIV-1 target cells (45–51) and the uneven distribution of the target cells within the epithelium and stroma (52, 53). Thus, whole vaginal biopsies were challenged ex vivo with Bal26 and maintained in a manner comparable to the rectal assay (see Methods), but optimized for vaginal tissues (17) to...
achieve consistent infection in untreated controls. Notably, none of the 13 explants from the 5 VRC01-infused female participants were infected by Bal26 challenge (Figure 5, A and B), whereas 23 of 28 vaginal explants from the 8 control female participants were infected using the same viral challenge conditions (Mann-Whitney $P = 0.003$; Figure 5B). Protocol restrictions in the number of vaginal biopsies that could be collected at a single visit prevented us from conducting parallel ex vivo challenges with Du422.1. Nonetheless, these results suggest that i.v.-infused VRC01 also infiltrates human vaginal tissue and retains antiviral activity 4–12 days after infusion.

Collectively, the rectal and vaginal ex vivo challenge findings provide further evidence that i.v.-infused VRC01 may provide partial or full protection at these sites of HIV-1 transmission against a virus strain highly sensitive to its neutralization activity.

**Correlations between i.v.-infused VRC01 levels in sera and mucosal compartments.** To investigate the relationship of VRC01 levels in the sera and mucosa following i.v. infusion, we compared IgG-normalized VRC01 levels among matched peripheral blood, intestinal, and genital locations using Spearman’s correlations (Figure 6). A poor correlation would suggest that the relative concentration of VRC01 in the local IgG pool (secretion or tissues) is significantly affected by local IgG production and not just serum IgG transudation.

In male participants (Figure 6A), VRC01 levels in sera strongly correlated with those in rectal secretions ($r = 0.964$, $P = 0.003$) and in rectal tissues ($r = 0.893$, $P = 0.012$). Similarly, VRC01 levels in rectal tissues and secretions correlated with one another ($r = 0.857$, $P = 0.02$). Correlation analyses for VRC01 levels in female participants were not significant, and were limited by a sample above the upper limit of quantitation and small sample size (Figure 6B). Together, these results suggest a strong, positive relationship between serum VRC01 levels and those in the rectal compartment of male participants at a given time after infusion. However, the relationships in terms of the biodistribution of VRC01 between the female genital compartments and serum remain ambiguous.

Lastly, we examined whether ex vivo Bal26 infectivity in rectal tissues from VRC01-infused male participants was related to VRC01 levels in matched sera, rectal secretions, or rectal tissues. No significant correlation between total Bal26 replication and IgG-normalized (Figure 6C) or total protein–normalized (data not shown) VRC01 levels was observed for any compartment. These results suggest that in the time interval tested, the number of Bal26-infected rectal explants was too small to properly assess potential relationships between VRC01 levels and susceptibility to HIV-1 infection, as only 4 of 21 rectal explants tested showed any detectable viral replication.

**Discussion**

Advancement of human monoclonal antibodies to prevent and control infectious diseases has become an attractive, alternative approach to vaccines for HIV and other viral pathogens (7, 54–56). Very recent findings from the 2 large international
The efficacy of VRC01 and other bn-mAbs is likely influenced by their potency, distribution, concentration, and anti-HIV activities at mucosal sites of transmission (26). However, the relationship between serum antibody levels (routinely measured) and mucosal antibody levels has been poorly defined, owing in part to practical and technical challenges in mucosal sampling and analyses within a clinical trial (59, 60). In this mucosal substudy of i.v.-administered VRC01, we successfully piloted processes to obtain genital and rectal samples, both secretions and tissues, and developed sensitive assays to specifically measure and compare VRC01 levels in the mucosal samples and peripheral blood. By taking advantage of the high sensitivity and low background characteristics of a VRC01-specific Erenna Single Molecule Counting (SMC) immunoassay, we have shown that VRC01 can be detected in serum, rectal, and cervicovaginal secretions and tissue compartments of healthy human volunteers between 4 and 13 days after they received an i.v. infusion of either 10 mg/kg or 30 mg/kg VRC01. Based on the success of this pilot study, we initiated a follow-up study, HVTN 116, in parallel with the AMP efficacy trials, which will elucidate compartment-specific bn-mAb decay rates and provide insight into mucosal antibody levels associated with ex vivo and in vivo protection. This larger and longitudi-

Figure 6. Correlation analysis of systemic and mucosal levels of VRC01 between compartments and in relation to ex vivo HIV-1 infection of rectal explants. (A and B) Relationships between IgG-normalized concentrations in sera, tissue homogenates, and secretions from male (n = 7) (A) and female (n = 5) (B) participants. (C) Relationships between total ex vivo HIV-1_{Replic} replication and IgG-normalized VRC01 levels in male participants (n = 7). Black-bordered symbols indicate participants in whose rectal biopsies ex vivo HIV-1 replication was detected. Correlations were determined by Spearman’s rank coefficients.
nal mucosal study, with sampling from 1 day after infusion to 6 months after infusion, will address the primary limitations of the pilot study (i.e., cross-sectional, nonrandomized trial design and sample size; male-only rectal sampling; and lack of earlier and later time points) and will enable more in-depth investigations that were outside the scope of the pilot study.

Our study suggests that the i.v.-infused VRC01 circulates in the body and penetrates into rectal and cervicovaginal compartments similarly to in vivo-derived human IgG, since bn-mAb levels were largely proportional to total IgG across participant-matched sample types. Our methodology is well designed to investigate in the future whether Fc-modified monoclonals with enhanced FcRn binding (26) preferentially transport into tissues, continue to have activity against mucosal HIV exposures, and/or undergo any accumulation in mucosal compartments.

Our findings also indicate that protein-normalized VRC01 levels in rectal secretions, rectal tissues, and vaginal tissues were up to 10-fold lower than those in serum, 4–13 days after infusion. Since 70% of recently circulating strains in the Americas and South Africa have neutralization IC80 greater than 1μg/mL (37), our findings support prioritizing development of long-lasting, highly potent antibodies for immunoprophylaxis to maximize blockade of HIV-1 at the mucosa. Intravenous delivery, which has been well tolerated in all VRC01 infusion studies (32, 37), may be necessary to achieve those protective concentrations at mucosal surfaces. Importantly, our ex vivo data and those of others (26) suggest that antibodies both within tissues and in secretions may play a role in viral inhibition.

Interestingly, protein-normalized VRC01 levels in cervical secretions were not lower than those in serum; whether this reflects differences in bn-mAb transfer from serum into cervicovaginal secretions versus tissues, or into the cervix versus the vagina, warrants future investigation. It is also possible that infused bn-mAbs preferentially bind mucins in cervical mucus to achieve this increased mAb/protein ratio (61), potentially enhancing virion trapping and neutralization. Regardless of the mechanism for accumulation, our findings illustrate differences in the amount of bn-mAb that can be suffused into particular mucosal compartments through i.v. administration, which could define the degree and duration of protection afforded by a dosing regimen. The vaginal mucosal challenges do not yet address the potential role of antibodies in cervical secretions and how they may contribute to HIV inhibition, but we plan to address this contribution in future work that extends our assessments to ectocervical tissue challenges in the context of cervical secretions.

We also observed that IgG-normalized VRC01 serum levels correlated strongly with those in both rectal secretions and tissues of infused male participants. This observation could be explained by the high degree of vascularization of intestinal tissues (42) and the expected proportional transport of VRC01, as well as other IgG1 antibodies, by FcRn as it recycles IgG molecules (62). Because this cross-sectional relationship may not apply to other time points after infusion, extended longitudinal PK data in matched serum and rectal samples are needed for characterization of the biodistribution properties of VRC01 in and between serum and rectal tissues in order to determine the predictive utility of serum bn-mAb levels for estimating rectal levels over time.

The lack of significant correlations between IgG-normalized VRC01 levels in vaginal tissues, cervical secretions, and sera from infused females is difficult to interpret given the small sample size. Whereas the vaginal walls are also highly vascularized (63, 64) with IgG transport facilitated by FcRn (65), female sex hormones have been shown to regulate immunoglobulin levels (66–69) and other mediators of immunity (70) in the female genital tract. Interestingly, Nardelli-Haefliger and colleagues showed that human papilloma virus-specific, vaccine-induced IgG levels in serum and cervical secretions were strongly correlated for women on hormonal contraception but not for women who were naturally cycling (69). Thus, future studies should address the impact of female sex hormones on both antibody levels and HIV-1 susceptibility in order to better evaluate and predict the efficacy of antibody-based passive and active immunization strategies for HIV-1 prevention in women.

The ex vivo challenge assay provided strong evidence that most biopsies from VRC01-infused participants contained functional, neutralizing mAb. All vaginal explants from infusion recipients, regardless of dosage, remained uninfected after a high-dose ex vivo Bal26 challenge. All rectal explants tested from 4 male volunteers were likewise completely protected from ex vivo Bal26 challenge, whereas at least 1 of 3 rectal explants tested from each of the remaining 3 volunteers became infected (Figure 3, A and B). Postinfusion safety monitoring in the main study prevented biopsy collections within the first 3 days of infusion. As a result, we cannot ascertain how early protective VRC01 levels are achieved in vaginal and rectal tissues. Breakthrough infections occurred at days 8–12 after infusion, likely reflecting the decay of functional VRC01 over time to subprotective levels (Figure 3A). Follow-up studies under way will further address the relationship between ex vivo HIV-1 infectivity and compartment-specific VRC01 levels throughout multiple half-lives.

The complete ex vivo protection of vaginal but not rectal explants is intriguing. Follow-up studies to compare paired rectal and vaginal tissues from the same participant and visit are in progress. Several potential mechanisms could support the hypothesis that rectal tissues are more difficult to protect than vaginal tissues. There is a greater density of HIV-1 target cells (71, 72) and potentially faster antibody turnover or degradation rate (26, 73) in the intestinal mucosa. Thus, it will be important to ascertain whether a higher bn-mAb concentration will be necessary to prevent rectal transmission. Although the addition of seminal proteins did not impact VRC01 neutralization against cell-free Bal26 in the rectal explant challenge, further work will extend previous reports to address the role of seminal proteins in the neutralization of other viruses at mucosal sites (18), as well as the role of other antibodies in the prevention of infection from cell-associated viruses in semen (74, 75).

Ex vivo HIV-1 challenge assays using human tissues to model 2 compartments primarily associated with sexual transmission of HIV-1 have been used in small-molecule pre-exposure prophylaxis drug development for years, typically employing HIV-1baa (76–78). Here, we adapted this concept to evaluate infused bn-mAbs taking into account theirEnv-specific targeting and extracellular activities. Our ex vivo assays employ Env-chimeric HIV-1 reporter viruses, selected for their specific neutralization sensitivities, at high virus challenge doses, compared with the estimates reported from semen exposure (79), followed...
by phytohemagglutinin (PHA) activation. This strategy produces consistent and reproducible infection readouts after a single challenge of untreated control tissues to facilitate the interpretation of bn-mAb doses that provide protection. Additionally, the explants were challenged quickly upon arrival with minimal manipulation to avoid bn-mAb leakage from tissues. The resulting stringency of the rectal and vaginal ex vivo challenge assays may provide conservative estimates of the overall protective efficacy of bn-mAbs. The nonpolarized virus challenge bypasses the natural barriers of intact mucosal surfaces. Also, bn-mAbs may leverage Fc-mediated effector functions from immune infiltrates in vivo and mediate systemic clearance of small, distal foci of infection (14, 80). Moving forward, ex vivo mucosal challenge assays using small, customized virus panels (including both neutralization-resistant viruses as controls and genetically diverse strains of intermediate neutralization sensitivities) are being designed to evaluate candidate bn-mAb combinations that provide better coverage than VRC01.

In summary, we have established clinical and laboratory procedures to investigate specific bn-mAb levels and function in rectal and cervicovaginal mucosa, and, importantly, our results demonstrate that VRC01 reaches mucosal tissues and secretions and exhibits functional antiviral activity in these compartments as early as 4–13 days after infusion. Currently, the development of potent bn-mAb combinations or multispecific mAbs, long-lasting Fc variants, and more efficient delivery routes is prioritized to optimize the breadth and durability of HIV-1 protection achievable by bn-mAb immunoprophylaxis (6). The mucosal assays developed here can be adapted to evaluate these parameters, and select the most promising approaches to protect sites of HIV-1 exposure, which may also be applicable to other sexually transmitted infections. Thus, integrating mucosal tissue and secretion assessments into mAb clinical development will help delineate requirements for successful passive immunization strategies, which, in turn, can provide a framework to develop vaccines that elicit the most effective antibody responses.

Methods

**Study participants.** VRC01-treated participants had received 2–3 bimonthly i.v. infusions of either 10 mg/kg (group T4) or 30 mg/kg (T5), administered by clinical staff at multiple clinical sites within the HVTN 104 trial (32). They were recruited for a single mucosal sampling visit at the Seattle Vaccine Trials Unit, within 3–14 days of their second or third VRC01 infusion. We chose this schedule to enhance the likelihood of measuring functional levels of VRC01 in the mucosa and to ensure that nested study collections, which required a flight to Seattle, did not interfere with HVTN 104 protocol in-study visits. Staff at sites conducting HVTN 104 referred interested study participants directly to the nested study, or participants self-referred after reading the study flyer.

The Seattle site investigators screened interested HVTN 104 participants for eligibility criteria related to the mucosal sample collection. All eligible and interested participants agreed to share their HVTN 104 study medical records and were assessed for HIV risk, history of sexually transmitted infection and antithrombotic usage, willingness to undergo vaginal (female) or rectal (male) biopsy and secretion collections, and willingness to abstain from sexual intercourse for 2 days before and 5 (male) or 7 (female) days after the biopsy procedure. Females were also assessed for pregnancy, tolerability of past vaginal exams, and vaginal health, including infection and pap smear history. Males were evaluated for rectal health, including bleeding and hemorrhoids. Uninfused control participants from Seattle were screened in person using enrollment criteria similar to those of both HVTN 104 and the mucosal study (Figure 1). Many female participants had previously donated biopsy samples in our clinic, and 2 female participants were asked to provide a second set of biopsy samples (Figure 1).

During the study visit, all participants consented and were reevaluated for HIV risk, medical history, and antithrombotic usage. All participants were tested for herpes simplex virus 2 serology and *Neisseria gonorrhoeae* and *Chlamydia trachomatis* infection by PCR (Aptima, Hologic); HIV-1 testing was done through their reference HVTN 104 study site. Before sample collection each participant underwent either a rectal or a vaginal exam to confirm eligibility for sample collection to proceed. During a single visit, male participants had 1 rectal sponge and 5 rectal pinch biopsies collected via anoscopy, 8–10 cm from the anal margin, using the Radial Jaw 4 instrument (Boston Scientific). Female participants had 1 cervical sponge and 5 vaginal biopsies collected from the fornices per visit, using Baby Tischler Biopsy Forceps (Wallach Surgical). Participants remained in Seattle for 24-hour observation. HVTN 104 participants received compensation for their time, as well as paid hotel and travel expenses during their stay in Seattle. Control participants were only reimbursed for their time, since they resided nearby.

**Sample collection and processing.** For each participant, peripheral blood was drawn into a single 8.5 mL BD Vacutainer serum separator tube. Serum was harvested by centrifugation at 2060g for 10 minutes at room temperature with low brake, and stored in aliquots at −80°C.

**Mucosal secretion collection and extraction.** Mucosal secretions were collected by placement of a pre-wetted Weck-Cel sponge (Medtronic) into a fold of the rectal mucosa for men (5 minutes), or of a dry Merocel sponge (Medtronic) into the cervical os for women (1 minute), both of which were stored immediately after collection at −80°C. To determine mAb levels present in rectal and cervical secretions, samples were thawed on ice for 10–15 minutes and processed on ice as previously described, including the addition of 600 μL of extraction buffer (PBS [Gibco] with Protease Inhibitor Cocktail I [Calbiochem]) (81). The resulting supernatants were diluted 1:3 and 1:100 with PBS and tested for free hemoglobin using a Human Hemoglobin ELISA (Immunology Consultant Laboratories) according to manufacturer instructions.

**Preparation of tissue homogenates.** Biopsies collected for tissue mAb quantitation were immediately frozen and stored at −80°C. One frozen vaginal biopsy and 2 independent rectal biopsy sections were thawed on ice and transferred into a 2 mL conical tube containing 200 μL of ice-cold extraction buffer. The tissue was homogenized with a Bio-Gen PRO200 Homogenizer and multi-Gen 7XL generator probe (Pro Scientific) using 30-second bursts and cooling on ice. Residual material was recovered from the probe by rinsing with 100 μL of ice-cold extraction buffer. Tissue homogenates were subsequently centrifuged and supernatant clarified using a Spin-X tube with 0.22 μm sterile filter (Sigma-Aldrich) at 4°C.

**Protein and total IgG quantification.** Total protein concentrations for serum, secretion, and tissue homogenate samples were quantified by bicinchoninic acid (BCA) assay (Pierce) in triplicate. Secretion samples were run undiluted; biopsy homogenates were run at 1:5; serum samples were run at 1:100 and 1:1000.
Total IgG concentrations for the same samples were also quantified by ELISA, as previously described (39), using a SpectraMax i5X ELISA plate reader (Molecular Devices). All the test samples were diluted in 1× PBS plus 10% FBS plus 0.05% Tween-20 (assay diluent) at 1:1000 for mucosal samples and 1:1,280,000 for serum samples.

**VRC01 quantification.** VRC01 levels in serum and mucosal samples were assessed by a custom immunoassay using the Singulex Erenna platform (Singulex/EMD Millipore) as previously described (39). Based on previous experience with serum and mucosal secretions collected with ophthalamic sponges, these serum and mucosal samples were diluted at 1:10,000 or 1:100, respectively, using Singulex Discovery Standard Diluent (Singulex/EMD Millipore) and filtered through 96-well Filter Plates (Pall). A VRC01 standard curve, comprising 2-fold and 3-fold serial dilutions of an internal standard ranging from 142,666 pg/mL to 61.9 pg/mL, was likewise prepared with Singulex Discovery Standard Diluent. VRC01 was captured using freshly prepared 50 μg/mL S9 IgG2a paramagnetic microparticle beads and detected with labeled mouse anti-human IgG1 Fc secondary detection antibody (500 ng/mL in Discovery Assay Buffer, Singulex/EMD Millipore). Bound immune complexes were eluted, transferred to a 384-well plate, and then read on the Singulex Erenna instrument according to manufacturer instructions. VRC01 and SC9 were gifts from John Mascola, NIH VRC Vaccine Production Program.

Curve fitting and extrapolation for Singulex data were done in the nCal R package (82) using the event photon (EP) raw data. The Erenna detects single molecules of VRC01 as photons over a read time. Each measurement of photons above a threshold is considered a detected event, and the sum of all photons from these events over the read time (i.e., signal intensity) is called the EP measurement for the sample or standard. The EP values of the VRC01 standards were fitted to a 5-parameter logistic curve and used to calibrate the specific mAb concentrations in mucosal and serum samples using the geometric mean of the standard triplicates. The percent coefficient of variation (CV) of technical replicates was ≤20% for 41 of 46 quantifiable samples. The lower and upper limits of quantitation (LLOQ and ULOQ) were defined by nCal, using the standards, as the estimated concentrations at which the percentage CV equals 20. All samples were below the ULOQ, except for 1 cervical secretion (PTID11) for which the CV of the estimated VRC01 concentration was between 26% and 25%. Samples above the LLOQ were confirmed to have geometric mean signal/noise ratio of at least 1.5. The percentage recovery for standards within the limits of quantitation was between 120% and 80% except for a single standard on 1 of the 7 assay plates. VRC01 concentrations, both absolute (serum) and normalized VRC01 levels (serum and mucosal), served as secondary outcome measures. ANCOVA was used to compare the mean of log-transformed dose-normalized concentrations between male and female participants accounting for differences in collection times.

**Viruses.** For explant HIV-1 challenges, we titered and used secreted nanoluciferase–expressing (sNLuc-expressing), replication-competent HIV-1 proviruses (designated pNL-sNLuc.T2A) in which the ectodomain of gp160/Env is encoded by different heterologous strain env sequences Bal26 and Du422.1, which were derived from a chronic clade B isolate (83) and an early/acute clade C isolate (84), respectively (17).

**Ex vivo HIV-1 challenge of rectal and vaginal explants.** Infectivity assays were conducted unblinded to group categorization (VRC01-infused vs. controls); however, details regarding infusion timing and dosing were unknown to laboratory staff. Up to 3 biopsies designated for ex vivo HIV challenge were placed immediately into a 2 mL tube containing premixed transport medium on wet ice. Whole rectal biopsies were transported in 1 mL of medium consisting of RPMI 1640 (Gibco) supplemented with l-glutamine (Gibco), antimicrobials (100 U/mL penicillin, 100 μg/mL streptomycin, 0.25 μg/mL amphotericin B [all from Gibco], and 0.5 μg/mL Zosyn [Pfizer]), and 10% human AB serum (GemCell, Gemini Bio). The biopsies were cut into 2 mm explants (about 2-3 per biopsy), which were evenly distributed along with the transport medium into infection tubes (with 3 explants per tube per challenge virus). To test the viral neutralization with known concentrations of VRC01, some rectal pieces from control volunteers were preincubated for 24 hours with 0.3-10 μg/mL VRC01 (NIH AIDS Reagent Program) before viral challenge. Within 60–90 minutes of biopsy collection, rectal pieces were challenged for 2 hours at 37°C with 1 million or 3 million infectious particles of Bal26 or Du422.1, respectively. The virus inoculum for each strain was determined empirically by the dose of virus leading to infection in 90% of healthy rectal tissue, not incubated with bn-mAbs. A portion of the viral inoculum was saved for downstream analyses before washing of the explants 10 times using 500 μL exchanges of PBS in the infection tubes. The explants were then weighed by difference from the empty tube, transferred to individual Transwell inserts (Corning) containing Surgifoam supports (Ethicon), placed in a 24-well plate (Corning) containing 1.1 mL of growth medium (rectal transport medium with 50 IU/mL IL-2 [PropeTech]), and activated for 48 hours with 0.5 μg/mL pretreated PHA (Remel, Thermo Fisher Scientific). Thereafter, medium was collected and replenished every 3 days for the duration of the 20-day culture.

Ex vivo vaginal challenges were performed similarly to the rectal challenges with the following modifications: Bal26 challenge virus was added directly to the original transport tubes containing whole vaginal biopsies (2-3 per tube) and transport medium, which also included 500 U/mL of IL-2, 25 ng/mL of IL-7, and 5 ng/mL of IL-15 (all from PropeTech), bringing the final volume up to 500 μL. After 2 hours at 37°C, each biopsy was weighed by difference and transferred to individual Transwell inserts (EMD Millipore) for washing and subsequent culture in a 24-well plate. After a portion of the viral inoculum was collected, the migrated cells were collected and distributed evenly among all Transwell inserts that contained individual biopsies. For 2 participants, a small vaginal biopsy disintegrated, leaving only 2 intact biopsies for explant culture. To wash the explants, PBS was added to the top of the inserts and to the bottom chambers, the Transwell plates were centrifuged, and the inserts were transferred to new wells. After 10 washes, the explants were activated for 24 hours with 0.2 or 0.6 μg/mL pretreated PHA in transport medium (660 μL). Medium was collected and replenished again on day 2 after challenge, then every 3 days thereafter, to coincide with the collection schedule for rectal explant cultures.

To monitor HIV-1 infection, the sNLuc activity in 20 μL of conditioned explant culture medium was measured using the Nano-Glo Luciferase Assay System (Promega). The relative light unit (RLU) values were scaled by culture volume (RLU × culture volume [mL]/Nano-Glo assay volume [mL]) and plotted for each collection time point to visualize HIV-1 production over time. To quantify overall virus replication, log area under the curve (AUC) from days 2 to 20 after infection was calculated in Prism software version 8 (GraphPad), which served as the primary outcome measure for this mucosal study. The volume-scaled RLUs were multiplied by 1000 before log transformation such that all values were greater than 1, and these values were used to calculate AUC using a baseline equal to 0. Adjustment for tissue mass...
differences was not performed, as we found no correlation between explant mass and AUC (data not shown).

Statistics. The study was designed based on preliminary experiments of rectal infectivity assays indicating that n = 4 provided 80% power to detect differences between tissues cultured with 1 μg/mL VRC01 and no mAb when challenged with Bal26.

Statistical comparisons were done with Prism software version 8 (GraphPad), and all tests were 2-sided. A P value less than 0.05 was considered significant. For evivo challenges, Dunn’s test was used to account for multiple comparisons of infectivity between controls and VRC01-infused samples, or between controls and exogenous VRC01-treated samples. Total HIV-1 replication levels are not directly comparable between rectal and vaginal assays because of differences in culture conditions (see above) that reflect the unique anatomical and immunological characteristics of each tissue type. Normalization of measured VRC01 concentrations by total IgG and total protein concentrations served as 2 methods to account for the different dilution factors between samples before the performing of statistical comparisons. Normalized VRC01 levels were compared within paired serum, secretions, and tissues using Friedman’s test, and comparisons between 2 specific locations were then analyzed using Dunn’s tests. Correlations were evaluated using Spearman’s rank correlation coefficients. For consistency, IgG-normalized VRC01 measurements were used for all correlation analyses, especially since total IgG levels are known to fluctuate in the cervicovaginal tract during the menstrual cycle [85].

Study approval. All study protocols and study materials were approved by the appropriate institutional review board(s), including participating HVTN 104 sites (New York Blood Center Clinical Research Site, New York, New York; Brigham and Women’s Hospital Infectious Disease Clinical Trials Center, Boston, Massachusetts; Cleveland AIDS Clinical Trials Unit at Case Western Reserve University, Cleveland, Ohio; Columbia University HIV Vaccine Unit, New York, New York; University of Pennsylvania HIV Vaccine Trials Unit, Philadelphia, Pennsylvania) and the Seattle Vaccine Trials Unit at Fred Hutchinson Cancer Research Center. All participants provided written informed consent prior to participation, which included information about the risks of mucosal sample collections. All participant samples were deidentified at collection and handled in the laboratory with no access to any participant’s identifier information. All data presented were recoded with new participant IDs for publication purposes.

Author contributions
RDA and MPL contributed equally to this work and are co-first authors; they are listed alphabetically. RDA and MPL designed the study, performed experiments, provided data, analyzed the data, and wrote the initial draft manuscript and further edited the manuscript. SN, MP, KW, and LBF helped design and perform experiments, provided data, and edited the manuscript. KES, VH, LZ, and GT provided data and analyzed the data. YL analyzed data. JC designed the study, enrolled subjects, provided data, and wrote portions of the manuscript. MG, JL, and JH helped with the conduct of the substudy and edited the manuscript. CO and ABM contributed new reagents/assays and edited the manuscript. HVT, LB, SH, IF, NG, and KM were responsible for conduct of HVTN 104 and contributed to the nested study recruitment. MJM conceived, designed, and led the study, obtained funding for the study, contributed to data analysis and interpretation, and edited the manuscript.

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75. Ducloy C, et al. HIV transmission from infected CD4+ T cells to allogenic T and dendritic cells is inhibited by broadly neutralizing antibodies. AIDS. 2018;32(10):1239-1245.


