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The forkhead transcription factor Foxo1 links insulin signaling to Pdx1 regulation of pancreatic β cell growth

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Introduction

Type 2 diabetes results from combined defects of insulin action and pancreatic β cell function (1, 2). Classically, the two abnormalities have been considered as separate entities. However, the recent demonstration that insulin/IGF signaling plays a role in insulin secretion and β cell proliferation has led to a critical reassessment of this view (3). For example, inactivation of insulin receptor (Insr) (4) or IGF-1 receptor (Igf1r) (5, 6) leads to impaired insulin secretion, while inactivation of insulin receptor substrate-2 (Irs2) impairs β cell proliferation (7, 8). These observations indicate that peripheral insulin resistance and β cell failure may share a common pathogenesis. However, the mechanism by which insulin signaling regulates β cell function remains unclear. Forkhead transcription factors of the Foxo subfamily, previously known as Fkhb (9), can promote (10) or repress (11) gene expression. Insulin inhibits Foxo through Akt-mediated phosphorylation and nuclear exclusion, and Foxo1 mutations affect insulin sensitivity in mice (12). Here we report the identification of Foxo1 as an effector of insulin action in pancreatic β cells, and propose a model in which Foxo1 links insulin signaling to regulation of β cell mass.

Methods

Reagents. We maintained and transfected kidney epithelial cells and β cells according to standard protocols (13). We purchased anti-insulin antibody from Linco Research Inc. (St. Charles, Missouri, USA), anti-Hemagglutinin (12CA5), and anti–5-bromo-2-deoxyuridine (anti-BrdU) antibodies from Sigma-Aldrich (St. Louis, Missouri, USA); anti–c-Myc antibody (9E10) from Roche Molecular Biosystems (Indianapolis, Indiana, USA); anti-Foxo1 and anti–phospho-Foxo1S253 from Cell Signaling Technology Inc. (Beverly, Massachusetts, USA); and anti-Foxa2 monoclonal antibody from the Developmental Studies Hybridoma Bank at the University of Iowa (Iowa City, Iowa, USA). We described previously the anti-Foxo1 antiserum used for gel shift assays (13) and the antibody against pancreas/duodenum homeobox gene-1 (Pdx1) (12). Expression vectors and Foxo1 adenoviruses have been described previously (13). All primer sequences are available upon request.

Animal production and phenotypic analysis. We have described Insr, Irs2, and Foxo1 mutant mice (12). We measured glucose and insulin as indicated in previous publications (12). Real-time RT-PCR and Northern analyses of gene expression. We isolated mRNA using the Micro-Fast Track 2.0 kit (Invitrogen Corp., San Diego,
California, USA). We carried out Northern blots according to standard methods and semiquantitative RT-PCR using the GeneAmp RNA PCR kit (Applied Biosystems, Foster City, California, USA) with amplification primers corresponding to Foxo1, Foxo3, Foxo4, Pdx1, Irs2, insulin, glucagon, pancreatic polypeptide, and β-actin sequences. We performed realtime PCR using primers encoding insulin 2, Pdx1, Slec2a1, glucokinase (Gck), Foxo2, Bad, Bcl-2 interacting mediator (BimL), Fas ligand (Fasl), and p27Kip1. For these experiments, we isolated mRNA from handpicked islets of similar sizes derived from three 4-week-old animals for each genotype and amplified it using a LightCycler PCR instrument and LightCycler RT-PCR kit (Roche Molecular BioSystems). We carried out each reaction in triplicate using a standard curve with the relevant cDNA for each primer set.

Immunohistochemical and morphometric analysis of pancreatic islets. We isolated and fixed pancreata from 8-week-old wild-type, Irs2–/–, and Irs2–/–Foxo1+/– mice. After 4 hours, we removed and fixed pancreata in PBS (pH 7.4) containing 2% paraformaldehyde and permeabilized them in 0.2% Triton X-100 for immunofluorescence as described previously (13). We visualized endogenous Pdx1 with anti-Pdx1 polyclonal antibody and FITC-conjugated anti-rabbit IgG. We detected transiently expressed c-Myc–Foxo1 in the same sections with a monoclonal anti–c-Myc antibody followed by CY3-conjugated anti-mouse IgG.

Detection and quantitation of β cell replication. We injected BrdU intraperitoneally (0.1 g/kg) into 2-week-old wild-type, Irs2–/–, and Irs2–/–Foxo1+/– mice. After 4 hours, we removed and fixed pancreata in PBS (pH 7.4) containing 2% paraformaldehyde, embedded them in paraffin, and immunostained consecutive 5-μm sections for β and α cells using anti-insulin and anti-glucagon antibodies, respectively. We performed morphometry using NIH Image 1.60 software (NIH, Bethesda, Maryland, USA) as described (14). We expressed results as percentage of total surveyed pancreatic area occupied by β and α cells.

Immunofluorescence. We incubated frozen islet sections with anti-insulin, anti-Pdx1, and anti-Foxo1 antibodies at dilutions of 1:1,000, 1:5,000 and 1:30, respectively. We visualized immune complexes using a CY3-conjugated anti–guinea pig IgG for anti-insulin antiserum, and FITC-conjugated secondary anti-rabbit IgG for anti-Pdx1 and anti-Foxo1 antisera. For colocalization of Foxo1 and Pdx1, we co-stained two adjacent sections with insulin and Pdx1, or insulin and Foxo1 antibodies, respectively. Thereafter, we acquired images using a SPOT-RT digital camera (Morrell Instruments, Melville, New York, USA) and identified matching islets in the two sections by overlaying the insulin immunostaining. We rendered Pdx1 immunoreactivity as mauve pseudocolor and scored cells in which we could unambiguously identify the nucleus on both sections for either Pdx1 or Foxo1 immunoreactivity. We analyzed a total of ten sections and 50 islets from three mice for each genotype. We fixed transfected βTc-3 cells in 2% paraformaldehyde and permeabilized them in 0.2% Triton X-100 for immunofluorescence as described previously (15). We visualized endogenous Pdx1 with anti-Pdx1 polyclonal antibody and FITC-conjugated anti-rabbit IgG. We detected transiently expressed c-Myc–Foxo1 in the same sections with a monoclonal anti–c-Myc antibody followed by CY3-conjugated anti-mouse IgG.

Luciferase and secreted alkaline phosphatase assays. The sequence used for these experiments spanned nucleotides –2,233 to –2,097 of the Pdx1 promoter and includes a single consensus forkhead binding site (15). We carried out cotransfections in βTc-3 cells at 70–80% confluence with Pdx1/luciferase reporter gene (pGL3-PDX1-PH2) and control pCMV5 vector or pCMV5–Foxa2, pCMV5–ADA-Foxo1, and pCMV5–wild-type Foxo1. We used secreted alkaline phosphatase (SEAP) (20 ng) as a control for transfection efficiency. We measured luciferase and SEAP activity according to the manufacturer’s specifications (Great EscApe SEAP kit; Clontech Laboratories Inc., Palo Alto, California, USA). In cotransfection experiments, we added recombinant adenovirus encoding ADA-Foxo1 to cells at different moi’s 12 hours after transfection with Foxa2. Forty-eight hours later, we determined luciferase activity in triplicate samples and normalized it by SEAP activity.

Gel shift assays. We isolated nuclei of LLC or βTc-3 cells by sequential extraction in NaCl (16). In some experiments, we transduced cells with Foxo1 adenovirus 48 hours prior to nucle isolation. We incubated double-stranded 32P-labeled oligonucleotide probes (20,000 cpm; 5 fmol) corresponding to the consensus forkhead binding region in the Pdx1 PH2 domain (5′-AGTGCTAAGCATAACCATGCCTG-3′, 50 mM NaCl, 1 mM MgCl2, 0.5 mM EDTA, 4% glycerol, 0.5 mM DTT, and 100 ng of poly(dI-dC) × poly(dI-dC)). For competition experiments, we used either the wild-type oligonucleotide indicated above, or a mutant oligonucleotide: 5′-AGTGCTAcGcgggTTCCTG3′ (mutant nucleotides are in lower case). The concentrations of cold competitor used were 0, 100-, 300- and 900-fold excess for both wild-type and mutant probes. We resolved the DNA/protein complexes on 6% nondenaturing polyacrylamide gel and visualized them by autoradiography. For supershift experiments, we incubated anti-Foxa2, anti-Foxo1, or non-immune sera with nuclear extracts for 10 minutes at room temperature. We incubated competitor wild-type and mutant probes in the reaction mixture for 10 minutes prior to the addition of the radiolabeled probe.
Results

RNA expression studies indicate that the three Foxo isoforms differ with respect to tissue localization, with Foxo1 representing the predominant mRNA isoform in liver (17), lung, and spleen, and Foxo4 in heart, skeletal muscle, and kidney (13). Foxo3 mRNA distribution mirrored that of Foxo1, albeit at lower levels (Figure 1a). Foxo1 is also the main member of the Foxo subfamily in cultured β cells (βTC-3 line) (Figure 1b, lane 2) and murine islets. Its levels decline over the first 6 months of life, as do Irs2 and Pdx1 levels. Foxo3 is expressed at lower levels and Foxo4 is undetectable in islets (not shown). Although a decrease in Pdx1 expression with age has been reported in rat islets (18), the present observation suggests that the down-regulation of Pdx1 mRNA occurs earlier than previously thought. It remains to be seen, however, whether this is a specific difference between rats and mice or a reflection of interstrain variability in mice. Immunohistochemistry of pancreatic islets with anti-Foxo1 and anti-insulin antisera demonstrates colocalization of Foxo1 and insulin to β cells. The subcellular localization of Foxo1 in β cells is heterogeneous, with some cells showing exclusive cytoplasmic or nuclear immunoreactivity, and some cells showing diffuse immunoreactivity (Figure 1d).

To address whether Foxo1 participates in β cell function, we tested the ability of a loss-of-function Foxo1 mutation to rescue β cell failure in Irs2+/− mice by crossing Irs2+/− and Foxo1+/− mice (12). In Irs2+/− mice, insulin-immunoreactive β cell area was about 70% smaller than in the wild-type at 8 weeks of age (Figure 2, a and b), with only a slight decrease in glucagon-immunoreactive α cell area (Figure 2b). No changes were observed in Foxo1+/− mice (12). In contrast, in Irs2−/−Foxo1+/− mice, β cell area was restored to about 80% of wild-type values. Irs2−/−Foxo1+/− mice have a normal life span and do not develop diabetes (Table 1).

Akt-dependent phosphorylation inhibits Foxo1 activity. When Foxo1 was expressed in islets from Irs2−/− mice by adenoviral transduction, phosphorylation of the “gatekeeper”
Akt site at serine 253 was decreased by approximately 40% compared with wild-type islets (Figure 2b), suggesting that Foxo1 is in the Irs2 signaling pathway. Since insulin receptor signaling inhibits Foxo1, we tested the effect of a loss-of-function Insr mutation on diabetes in Irs2–/– mice. We generated recombinant congenic mice carrying Irs2 and Insr targeted null alleles on the same chromosome (the two genes are about 4 Mb apart near the centromeric end of chromosome 8; ref. 19), and intercrossed them to produce Irs2–/– Insr+/– mice. Insr haploinsufficiency increased the severity and accelerated the onset of diabetes in Irs2–/– mice. Irs2–/– Insr+/– mice developed diabetes at 6–8 weeks (Table 1) and invariably died by 10 weeks of age. Pancreatic islets were virtually undetectable by 8 weeks (Figure 2a). Withers and colleagues have reported a similar phenotype in Irs2–/– Igf1r+/– mice (20).

To distinguish between increased cell proliferation and decreased apoptosis as mechanisms of β cell restoration in Irs2–/– Foxo1+/– mice, we measured BrdU incorporation and DNA fragmentation. BrdU incorporation was reduced by about 85% in islets of 2-week-old Irs2–/– mice and was restored to approximately 67% of wild-type in Irs2–/– Foxo1+/– mice (P < 0.05 by ANOVA). We observed no changes in exocrine acinar tissue (Figure 2c). In contrast to proliferation, we failed to detect apoptotic cells in any of the genotypes examined using TUNEL staining at ages 2, 3, and 4 weeks (data not shown). These data indicate that restoration of β cell mass in Irs2–/– Foxo1+/– mice is due to increased proliferation, rather than decreased apoptosis. Nevertheless, given that apoptosis in β cells is difficult to document (21), we cannot rule out its contribution to the observed phenotype.

Unlike β cell mass, Foxo1 haploinsufficiency did not affect glucose-stimulated insulin secretion from islets of Irs2–/– Foxo1+/– mice, nor did overexpression of wild-type or mutant Foxo1 affect insulin secretion in cultured βTC-3 cells (data not shown).

To examine the mechanism by which Foxo1 haploinsufficiency prevents diabetes in Irs2–/– mice, we analyzed expression of Foxo1 target genes. They include the cell cycle inhibitor p27kip (22) and the proapoptotic genes FasL (23) and BimL (24). For these experiments we used islets of similar size derived from 4-week-old mice to exclude artifacts due to loss of β cells.
However, real-time RT-PCR analysis of islet RNA failed to detect differences in Bad, BimL, and FasL, while p27kip was decreased by 40% in Irs2–/– mice and by 30% in Irs2–/– Foxo+/– mice (Figure 3), which is inconsistent with the possibility that Foxo1 inhibits β cell replication by increasing p27kip. Thus, changes in the expression of these genes do not appear to play a role in the observed phenotype. These data were confirmed by experiments in βTC-3 cells, in which a dominant-negative (δ256) Foxo1 adenovirus failed to inhibit p27kip, FasL, and BimL expression (data not shown).

In contrast, real-time RT-PCR demonstrated a greater than 80% decrease in the expression of Pdx1 and its target gene Slc2a2 (glucose transporter Glut2) in Irs2–/– mice. The decrease was also observed in islets of transgenic mice bearing a phosphorylation-defective Foxo1S253A mutant in β cells (12), and was partially reversed in Irs2–/– Foxo1+/– mice to about 50% of wild-type. Lesser changes (~50%) were observed in Gck levels, consistent with two recent studies showing that Gck is less sensitive than Slc2a2 to reduction of Pdx1 expression (25, 26). In contrast, Gck expression was increased about twofold in Foxo1S253A transgenics (Figure 3). The reasons for the differential regulation of Gck and Slc2a2 in Irs2–/– and Foxo1S253A mice are unclear. Foxa2 expression was also decreased by about 50% in Irs2–/– and Foxo1S253A mice, but was not restored in Irs2–/– Foxo1+/– islets.

We next compared Pdx1 expression by immunohistochemistry. In wild-type islets, more than 95% of insulin-immunoreactive cells scored (793 of 827) were Pdx1-positive (Figure 4a, green). In Irs2–/– mice, the percentage of Pdx1-positive cells decreased to 38.7% (245 of 632); moreover, Pdx1 was frequently mislocalized to the cytoplasm. In Irs2–/– Foxo1+/– mice, the percentage of Pdx1-positive cells was restored to greater than 90% (678 of 730) (P < 0.001 by ANOVA). These data support the real-time PCR analysis and suggest a negative correlation between Foxo1 activity and Pdx1 expression levels.

To investigate this relationship further, we analyzed the subcellular distribution of Foxo1 in Pdx1-positive and Pdx1-negative β cells. Because the anti-Foxo1 and anti-Pdx1 antisera available for immunohistochemistry have been raised in rabbits, we could not perform double staining on the same tissue section. Therefore, we stained adjacent sections with anti-Foxo1 and anti-insulin, or anti-Pdx1.

Table 1
Metabolic characteristics of 8-week-old mice

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Body weight (g)</th>
<th>Glucose (fasting)</th>
<th>Glucose (fed)</th>
<th>Insulin (fasting)</th>
<th>Insulin (fed)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wild-type (n = 13)</td>
<td>33.1 ± 0.6</td>
<td>96 ± 3</td>
<td>123 ± 5</td>
<td>0.9 ± 0.1</td>
<td>2.8 ± 0.2</td>
</tr>
<tr>
<td>Irs2–/– (n = 14)</td>
<td>31.2 ± 0.7</td>
<td>98 ± 4</td>
<td>191 ± 13^A</td>
<td>2.0 ± 0.3</td>
<td>4.5 ± 0.4</td>
</tr>
<tr>
<td>Irs2–/– Foxo1+/– (n = 14)</td>
<td>31.8 ± 0.6</td>
<td>98 ± 4</td>
<td>157 ± 9</td>
<td>1.3 ± 0.1</td>
<td>4.4 ± 0.4</td>
</tr>
<tr>
<td>Irs2–/– Insr+/– (n = 10)</td>
<td>29.4 ± 0.4</td>
<td>250 ± 13^A</td>
<td>440 ± 28^A</td>
<td>0.4 ± 0.1</td>
<td>1.1 ± 0.1</td>
</tr>
</tbody>
</table>

^A P < 0.05 by one-factor ANOVA compared with wild-type.
and anti-insulin antisera and carefully matched the position of β cells using insulin immunostaining to outline the nuclear margins (Figure 4b). On average, approximately 30% of cells showed matching nuclear “ghosts” in two adjacent 5-µm sections, consistent with the fact that the average β cell diameter is 10 µm and the nucleus occupies about 70% of the cell surface (6). In the vast majority of this subset of cells, Pdx1 and Foxo1 showed mutually exclusive nuclear localization. Thus, in approximately 80% of Pdx1-positive cells examined, Foxo1 localized to the cytoplasm (Figure 4b, cells 1, 2, 7, and 8). By contrast, in about 80% of Pdx1-negative cells, Foxo1 localized to the nucleus (Figure 4b, cells 3, 4, 6, and 9). In addition, the occasional double-positive cells (such as cell 10 in Figure 4b) showed very weak Pdx1 immunoreactivity. These correlative findings were investigated further by expressing wild-type or a phosphorylation-defective Foxo1 mutant (ADA-Foxo1) that is constitutively nuclear (13) in βTC-3 cells. Wild-type Foxo1 was basally phosphorylated (data not shown) and was localized to the cytoplasm of Pdx1-expressing cells. In contrast, cells expressing the ADA-Foxo1 mutant in the nucleus were Pdx1-negative (Figure 5a). Moreover, we have previously shown that expression of a similar constitutively nuclear Foxo1 mutant in islets of transgenic mice results in greatly decreased Pdx1 levels (12).

In summary, Pdx1 levels decreased in three models of increased Foxo1 activity: Ir22−/− mice, in which Foxo1 phosphorylation is decreased (Figure 2b); transgenic mice expressing the phosphorylation-defective, constitutively nuclear Foxo1S253A in β cells (12); and βTC-3 cells transduced with ADA-Foxo1 adenovirus. Conversely, Foxo1 haploinsufficiency partially reverses the decrease in Pdx1 levels in Ir22−/− mice. These data support the hypothesis that Foxo1 inhibits Pdx1 and are consistent with the observation that overexpression of Pdx1 rescues diabetes in Ir22−/− mice (27).

To determine potential sites of interaction between Foxo1 and Pdx1, we surveyed pancreatic Foxo1 expression. The targeting vector used to ablate Foxo1 function contains a β-galactosidase (β-gal) cassette fused in-frame with Foxo1 exon 1 to enable detection of
Foxo1 expression using X-gal reactivity (12). Using this technique, we detected occasional Foxo1-positive cells in pancreatic ducts and exocrine acini, in addition to islets (Figure 5b, top row). In view of the potential role of duct-associated cells in β cell neogenesis, we investigated the colocalization of Foxo1 with insulin and Pdx1 in duct-associated cells. The majority of duct-associated Foxo1-positive cells did not display insulin or Pdx1 immunoreactivity (Figure 5b, red arrows). However, all the insulin/Pdx1-positive duct-associated cells were Foxo1-positive (Figure 5b, black arrows). These data are consistent with the possibility that Foxo1 regulates Pdx1 expression in duct-associated cells.

Next, we used gel shift and reporter gene assays to test the hypothesis that Foxo1 binds directly to the Pdx1 promoter and inhibits its transcription. A 6.5-kbp Pdx1 promoter contains elements required for β cell–specific expression, including three potential forkhead binding sites (15, 28, 29).

Among the forkhead proteins, Foxa2 is a known activator of Pdx1, and Pdx1 expression is blunted in mice lacking Foxa2 in β cells (30). We carried out gel shift assays using nuclear extracts from LLC (Figure 6a) and βTC-3 cells (Figure 6b). When we used a probe carrying the proximal consensus forkhead binding site of the Pdx1 promoter (PH2 domain) (15), we detected a gel retardation complex that could be competed for by excess cold probe, but not by a mutant probe (Figure 6a). In insulin-producing βTC-3 cells, we detected at least two gel retardation complexes, indicated as I and II. Addition of anti-Foxo1 antiserum caused a supershift to yield a distinct complex III. Addition of anti-Foxa2 monoclonal antibody caused a supershift of complex A to yield complex IV. Addition of nonimmune serum did not cause any supershift (Figure 6b).

We used cotransfection assays with a minimal Pdx1 promoter containing the proximal Foxa2 binding site to study the effect of Foxo1 in Pdx1 transcription. Expression of Foxa2 increased Pdx1/luciferase activity approximately tenfold, whereas Foxo1 failed to do so (Figure 6c). Coexpression of constitutively active Foxo1 inhibited Foxa2-dependent Pdx1 transcription in a dose-dependent manner up to about 60%, consistent with the possibility that Foxo1 acts as a transrepressor of Foxa2-dependent Pdx1 transcription (Figure 6d). The ability of Foxo1 to inhibit Foxa2-dependent Pdx1 transcription was not due to the presence of an excess of Foxo1 protein, since we detected the inhibitory effect when Foxa2 levels exceeded those of Foxo1 (Figure 6e). We obtained similar data when we used the full-length Pdx1 promoter construct (data not shown). These data provide proof of principle that Foxo1 can act as a repressor of Pdx1 transcription in a transfection system. However, it is likely that the in vivo interaction between Foxo1 and Pdx1 is complex, and

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**Figure 5**

(a) Pdx1 expression in βTC-3 cells transfected with c-Myc–tagged wild-type (left column) or ADA-Foxo1 (right column). We performed immunocytochemistry with anti–c-Myc (top row) and anti-Pdx1 (middle row). We visualized anti–c-Myc immunoreactivity with CY3-conjugated anti-mouse IgG (red) and anti-Pdx1 immunoreactivity with FITC-conjugated anti-rabbit IgG (green). We show double staining in the bottom row. (b) Foxo1 localization in pancreatic duct–associated cells. We treated frozen sections for immunohistochemistry with anti-insulin and anti-Pdx1 antisera, followed by incubation in X-gal to detect β-gal expression from the Foxo1 locus. In the top row, we show a representative endocrine islet, demonstrating that X-gal reactivity colocalizes with islets. Occasional X-gal–positive cells can also be visualized in ducts and acini. In the middle and bottom rows, we show that X-gal reactivity in ducts can be detected in cells that do not express insulin and/or Pdx1 (red arrows), as well as cells with insulin and Pdx1 immunoreactivity (black arrows).
depends on the relative abundance of Foxo1, the contributions of coactivators and corepressors, and signals regulating Foxo1 localization.

**Discussion**

Our genetic analysis indicates that Foxo1 is an effector of Irs2 signaling in pancreatic β cells. Foxo1 inactivation leads to increased Pdx1 expression and β cell proliferation. Since Foxo1 is expressed in a subset of cells embedded within pancreatic ducts, we propose that, in quiescent duct-associated cells that are not committed to a β cell fate, Foxo1 acts as a transcriptional brake on Pdx1. We propose the following mechanism of Foxo1 regulation: small quantities of insulin are released in the pancreatic duct (31), where they activate signaling (32) in the Foxo1-positive duct cell subset, leading to Foxo1 nuclear exclusion and Pdx1 expression. In this hypothesis, pancreatic islets of Langerhans are functionally associated with ducts. While controversial (33), this assumption is supported by a recent reassessment of the functional anatomy of the pancreas, showing that most islets are associated with pancreatic ducts and providing a morphological correlate for insulin release into pancreatic ducts (34). The implication of this model is that insulin is the main promoter of islet proliferation in hyperinsulinemic,
insulin-resistant patients, and that insulin resistance predisposes to β cell failure via impaired Foxo1 phosphorylation. This model provides a unifying hypothesis for the commonest abnormalities of type 2 diabetes: peripheral insulin resistance and impaired β cell compensation. Furthermore, it suggests that acute islet destruction in type 1 diabetes prevents significant islet regrowth, because there is no insulin to stimulate duc tal neogenesis. This hypothesis is consistent with the observation that insulin administration following streptozotocin-induced β cell ablation is associated with robust islet regrowth (35, 36). Whether this mechanism is physiologically relevant in the context of ongoing autoimmunity is an open question. Finally, the notion that β cell neogenesis from a subset of duct-associated cells is a compensatory response to insulin resistance, rather than a physiologic event, reconciles the proposed role of duc tal neogenesis (37) with the recent demonstration that duct epithelial cells do not physiologically give rise to endocrine cells (38). We propose that Foxo1 is an important target for the development of drugs that improve β cell proliferation.

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