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Low-dose leptin reverses skeletal muscle, autonomic, and neuroendocrine adaptations to maintenance of reduced weight

Michael Rosenbaum, Rochelle Goldsmith, Daniel Bloomfield, Anthony Magnano, Louis Weimer, Steven Heymsfield, Dympna Gallagher, Laurel Mayer, Ellen Murphy, and Rudolph L. Leibel

Columbia University College of Physicians and Surgeons, New York, New York, USA.

Maintenance of a reduced body weight is accompanied by decreased energy expenditure that is due largely to increased skeletal muscle work efficiency. In addition, decreased sympathetic nervous system tone and circulating concentrations of leptin, thyroxine, and triiodothyronine act coordinately to favor weight regain. These “weight-reduced” phenotypes are similar to those of leptin-deficient humans and rodents. We examined metabolic, autonomic, and neuroendocrine phenotypes in 10 inpatient subjects (5 males, 5 females [3 never-obese, 7 obese]) under 3 sets of experimental conditions: (a) maintaining usual weight by ingesting a liquid formula diet; (b) maintaining a 10% reduced weight by ingesting a liquid formula diet; and (c) receiving twice-daily subcutaneous doses of leptin sufficient to restore 8 am circulating leptin concentrations to pre–weight-loss levels and remaining on the same liquid formula diet required to maintain a 10% reduced weight. During leptin administration, energy expenditure, skeletal muscle work efficiency, sympathetic nervous system tone, and circulating concentrations of thyroxine and triiodothyronine returned to pre–weight-loss levels. These responses suggest that the weight-reduced state may be regarded as a condition of relative leptin insufficiency. Prevention of weight regain might be achievable by strategies relevant to reversing this leptin-insufficient state.

Introduction

In rodents, imposed weight-loss decreases energy expenditure per unit of metabolic mass, reflecting metabolic opposition to loss of body fat (1). Maintenance of a 10% or greater reduction in body weight by either obese or never-obese humans is also accompanied by a reduction in energy expenditure beyond that predicted by loss of body mass (2–5). In such subjects, sympathetic nervous system (SNS) tone and circulating concentrations of leptin and bioactive thyroid hormones are reduced, while parasympathetic nervous system (PNS) tone is increased (2–9). These changes are of comparable magnitude in obese and lean individuals (2) and may be causally related to the reduction in energy expenditure that occurs following weight loss. This decrease in energy expenditure is of sufficient magnitude to account—in part—for the very high recidivism to obesity in otherwise successfully weight-reduced subjects. The major “effector organ” for this decline in energy expenditure is skeletal muscle (3). Maintenance of a 10% reduction in body weight is associated with a 20–30% decline in energy expended in physical activity regardless of whether calculated from physical activity logs (10–12), assessed by bicycle ergometry and indirect calorimetry (3), or measured by NMR spectroscopy as the ATP cost per muscle contraction during low-level physical activity (3, 12). The decrease in energy expenditure is not due to reduced time spent in physical activity (2, 3, 10).

The metabolic phenotypes of weight-reduced humans, leptin-deficient humans, and leptin–deficient rodents are similar but not identical (13–15). Murine leptin deficiency is characterized by hyperphagia, hypometabolism, decreased T cell–mediated immunity, decreased SNS activity, stunted skeletal growth, decreased hypothalamic-pituitary–thyroid and –gonadal axis activities, and increased circulating concentrations of corticosterone (13, 14, 16, 17). Congenitally leptin-deficient children have a similar phenotype, except that energy expenditure, statural growth, and circulating concentrations of cortisol are all within normal limits (15). Administration of leptin to leptin-deficient rodents in doses that restore circulating leptin concentrations to their physiological range increases energy expenditure, decreases energy intake, increases SNS output (18, 19), and normalizes hypothalamic-pituitary-adrenal, thyroid, and gonadal function (20–23). Leptin administration to leptin-deficient humans decreases energy intake and increases activity of the hypothalamic-pituitary-thyroid axis (15, 24). Yet in humans and rodents that are not leptin deficient, induction of weight loss requires doses of leptin that produce 10-fold elevations of plasma leptin concentrations (23, 25).

We have previously reported that restoration of circulating concentrations of leptin in weight-reduced individuals to levels that were present prior to weight reduction reverses the decline in circulating concentrations of bioactive thyroid hormones and in energy expenditure that characterize the weight-reduced state (4). Similarly, Welt et al. (26) reported that leptin administration (0.08 mg/kg/d) to women with secondary hypothalamic amenorrhea due to reduced fat mass reversed the decline in hypothalamic-pituitary–gonadal axis function and raised circulating concentrations of triiodothyronine (T3) and thyroxine (T4); no such effects were detected when leptin was administered in similar doses to eumenorrheic women without reduced fat mass.
In the present study, we tested the hypothesis that administration of leptin to weight-reduced subjects in doses that would restore circulating leptin concentrations to levels present prior to weight loss would reverse the changes in skeletal muscle metabolism and autonomic function that accompany maintenance of a reduced body weight (2–9).

Results

Protocol and leptin dosing. This protocol was designed to examine the effects of leptin administration on the weight-reduced phenotype of subjects maintaining a 10% reduced body weight (Figure 1). Subjects were inpatients at the General Clinical Research Center and on a liquid formula diet throughout the study. Briefly, subjects were admitted at their usual body weight (designated as “Wt_initial”), and caloric intake was titrated until weight was stable, at which time subjects underwent the measurements of metabolic, autonomic, and neuroendocrine function described in Methods. Achievement of weight stability and testing required an average of 8 weeks (range 5–10 weeks). Subjects were then placed on 800 kcal/d of the same liquid formula diet until they had lost 10% of their usual weight. Subjects required an average of 7 weeks to complete studies at Wt_initial (range 5–10 weeks). Following completion of studies at Wt_initial, and while remaining on the same daily caloric intake, subjects received 5 weeks of twice-daily leptin injections in doses calculated to maintain circulating leptin concentrations at levels present prior to weight loss (designated as “Wt_10%lep”) and within dosing ranges that have been reported to have little if any effect on the parameters measured in this study in subjects at usual weight (25, 26). Leptin doses were titrated until trough concentrations, measured just prior to leptin administration at 8 am, were comparable to those levels present prior to weight loss. When subjects who had lost 10% (Wt_10%) of their initial body weight (Wt_initial) received subcutaneous leptin (Wt_10%lep), plasma leptin concentrations at 8 am were significantly higher than those values observed at Wt_10% and not significantly different from those observed at Wt_initial (Figure 2). In studies of leptin administration to a cohort of 16 obese and lean males and females at usual weight who were assessed monthly for 6 months while receiving leptin, Heymsfield et al. (25) reported no significant effects of administration of 0.01–0.10 mg/kg/d on body weight or composition. Welt et al. (26) reported no significant effects of administration of 0.08 mg/kg/d r-metHuLeptin (over a 1-month period) on body composition, am cortisol levels, or thyroid hormones in 8 women with hypothalamic amenorrhea. In the present study, mean ± SEM total daily leptin doses relative to body mass at Wt_10% were 0.068 ± 0.01 mg/kg (range 0.019–0.121 mg/kg/d) divided q12 hours (8 am and 8 pm), i.e., within ranges that have been previously shown to have no significant effect on body composition or hypothalamic-pituitary-thyroid function in obese or lean subjects at usual weight. It should be noted that twice-daily administration of exogenous leptin does not mimic the normal pulsatility of leptin secretion (27–30) and that the effects on leptin pulsatility of maintenance of a reduced body weight or of exogenous leptin administration in humans on leptin pulsatility, to our knowledge, have not been reported. Therefore, the decline in 8 am circulating leptin concentrations at Wt_10% and restoration of 8 am circulating concentrations of leptin to those present prior to weight loss during leptin administration at Wt_10%lep do not necessarily reflect total daily amounts of circulating leptin or the pulsatility of leptin secretion during the different study periods. No significant effects of gender or initial somatotype were noted on any results, except that lean subjects consistently required less time (mean 6 weeks) to lose 10% of their weight than obese subjects (mean 9 weeks).

Body composition. Administration of leptin to subjects ingesting the same number of calories required to maintain their weight at Wt_10% resulted in further reduction in body weight. Loss of fat mass accounted for 67.2% ± 8.5% of the weight lost during leptin administration, similar to 71% ± 6.7% of weight lost as fat observed during weight loss on an 800-kcal/d diet from Wt_initial to Wt_10% (Figure 3).

Energy expenditure. At all study periods, the respiratory quotient (RQ) at rest was not significantly different from the diet formula quotient (by bomb calorimetry) of 0.85. At Wt_10%, 24-hour energy expenditure (TEE) and nonresting energy expenditure (NREE; energy expended in physical activity above resting) were decreased out of proportion to changes in body composition from Wt_initial. The relationships among TEE, NREE, and body composition were restored at Wt_10%lep to levels present at Wt_initial (Figure 4). No significant effects of weight loss or leptin administration on resting energy expenditure (REE) or the thermic effect of feeding (TEF) beyond those predicted by changes in body composition were noted.

Skeletal muscle. At Wt_10%, skeletal muscle work efficiency was increased and RQ was decreased at low levels of work (pedaling a bicycle ergometer to generate 10 W of power) compared with sub-
ects at Wt\textsubscript{initial}, similar to our previous observations (3). At Wt\textsubscript{−10%}, the gross mechanical efficiency (GME) of skeletal muscle while bicycling to generate 10 W of power was approximately 23% higher than at Wt\textsubscript{initial} and was fully reversed during leptin administration at Wt\textsubscript{−10%lep}. Extrapolating from RQ data (31), at Wt\textsubscript{−10%} there was an approximate 12% decrease — compared with Wt\textsubscript{initial} — in the percentage of calories derived from the oxidation of glucose during bicycling to generate 10 W of power. This decrease in the use of glucose as fuel following weight loss was only evident at low levels of work (bicycling to generate 10 W but not 25 W or more of power) (Figure 4). The effect was fully reversed at Wt\textsubscript{−10%lep} (Figure 4).

**Thyroid.** At Wt\textsubscript{−10%}, circulating concentrations of T3 and T4 were reduced compared with Wt\textsubscript{initial}. At Wt\textsubscript{−10%lep}, circulating T4 and T3 concentrations were, respectively, similar to and higher than those at Wt\textsubscript{initial}. At Wt\textsubscript{−10%} and Wt\textsubscript{−10%lep}, circulating thyroid-stimulating hormone (TSH) concentrations were significantly reduced compared with Wt\textsubscript{initial} values; no significant effects of leptin administration on circulating TSH concentrations were noted (Figure 5).

**Autonomic function.** Maintenance of a 10% reduced body weight was associated with decreased SNS activity (measured by heart rate analysis) and by urinary catecholamine excretion and increased PNS activity (measured by heart rate analysis). Administration of leptin to weight-reduced subjects reversed the decline in SNS activity and 24-hour urinary epinephrine excretion but did not significantly alter the decline in 24-hour urinary norepinephrine excretion or the increase in PNS activity (Figure 6). There was no significant effect of weight loss or leptin administration on the intrinsic heart rate.

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**Figure 3**
Subject characteristics. Administration of exogenous leptin to weight-reduced subjects ingesting an isocaloric diet was associated with a significant decline in body weight and fat mass but not fat-free mass. Data refer to 10 subjects: 5 males, 5 females; 7 obese, 3 never-obese. Mean ± SEM values at each weight plateau are presented in Supplemental Table 1 (supplemental material available online with this article; doi:10.1172/JCI25977DS1). *P < 0.05 versus Wt\textsubscript{initial}; †P < 0.05 versus Wt\textsubscript{−10%}.

**Discussion**
In human beings, maintenance of reduced body weight is met by coordinate metabolic, neuroendocrine, and autonomic responses that favor the regain of lost weight (2–9). We have previously reported that the administration of exogenous leptin to weight-reduced subjects in doses that restore circulating leptin concentrations to those present prior to weight loss reverses the declines in energy expenditure and circulating concentrations of bioactive thyroid hormones that accompany the maintenance of a reduced body weight (4). Other studies (25, 26) have demonstrated that administration of similar doses of exogenous leptin to subjects at usual weight has no significant effect on body composition or thyroid function. The major findings of the present study are that administration of leptin to weight-reduced individuals also reverses the changes in skeletal muscle energetics and SNS activity that would otherwise act coordinately to promote weight regain (32, 33). These in vivo responses of weight-reduced humans to leptin are concordant with in vitro and in vivo studies of leptin effects on skeletal muscle, autonomic nervous system activity, and neuroendocrine axes and demonstrate that leptin administration to weight-reduced subjects reverses many of the metabolic phenotypes that characterize the weight-reduced state.

Maintenance of a reduced body weight is associated with a significant decrease in TEE of approximately 300–500 kcal/d greater than that predicted by changes in body mass and composition (2, 3). This decrease in TEE is due predominantly to reduced energy expended in physical activity (NREE), reflecting increased work efficiency of skeletal muscle (3). We find that both the decline in

**Figure 4**
Energy expenditure and skeletal muscle. Percent change (mean ± SEM) from values at Wt\textsubscript{initial} of energy expenditure, skeletal muscle work efficiency, and fuel utilization. Administration of leptin to weight-reduced subjects reversed the significant decline in TEE and NREE associated with maintenance of a 10% reduced body weight. Effects of weight loss and exogenous leptin on skeletal muscle gross mechanical work efficiency (kcal/min energy expended above resting per kcal/min work generated) and fuel utilization (derived from RQ; ref. 87) were only evident at low levels of work (bicycling to generate 10 W of power). Mean ± SEM values at each weight plateau are presented in Supplemental Table 1, and individual values are presented in Supplemental Figure 1. *P < 0.05 versus 0; †P < 0.05 versus Wt\textsubscript{−10%}. GME, gross mechanical efficiency.
energy expenditure and increased work efficiency of muscle are reversed by leptin administration to weight-reduced subjects. Leptin could affect skeletal muscle function via direct interaction with muscle cells and/or by engaging hypothalamic/brain stem pathways regulating autonomic and neuroendocrine function. Skeletal muscle expresses the short and long isoforms of the leptin receptor (34), and incubation of skeletal muscle from lean mice with leptin stimulates oxygen consumption (35). Both systemic and intracerebroventricular leptin administration increase insulin-mediated glucose uptake in rat skeletal muscle (36–38), and leptin increases both glucose and fatty acid oxidation in rodent skeletal muscle in vitro (34, 36, 39–42). Declines in circulating concentrations of bioactive thyroid hormones have been shown to promote increased expression of the more chemomechanically efficient isoforms of the myosin heavy chain (MHC) that preferentially use free fatty acids as fuel (43, 44). Resultant shifts in MHC isoform expression may contribute to the leptin-reversible increase in skeletal muscle work efficiency and utilization of fatty acids noted following weight loss in this study.

SNS tone appears to be crucial to leptin-mediated increases in energy expenditure in rats, since neonatal chemical sympathectomy severely attenuates the increased energy expenditure, loss of fat mass, and increased insulin-mediated glucose uptake that occur following intracerebroventricular leptin administration to rats (45). Leptin deficiency is associated with decreased SNS tone as measured by blood pressure changes during a cold pressor test in humans (46) and a leptin-reversible decline in brown adipose tissue thermogenesis in mice (47). We examined the effects of leptin administration to weight-reduced subjects on SNS and PNS inputs to the heart (heart rate analyses during pharmacological blockade) and on 24-hour secretion of norepinephrine (predominantly from SNS nerve endings) and epinephrine (predominantly from the adrenal gland). These organ-specific measures of autonomic activity do not necessarily reflect systemic autonomic nervous system tone. We found that leptin reverses the decline in SNS activity (measured by heart rate analysis and urinary epinephrine but not norepinephrine excretion) but does not significantly affect the increase in PNS activity associated with weight loss that has been previously reported (6, 48, 49). These data suggest that changes in SNS, but not PNS, activity following weight loss are due to decreased circulating leptin concentrations. Leptin-mediated SNS effects probably reflect both the peripheral stimulatory effects of leptin on adrenal epinephrine release and centrally mediated increases in sympathetic outflow. These inferences are supported by direct effects of leptin on epinephrine release from porcine adrenal medullary cells (50) and the increases in circulating concentrations of epinephrine and norepinephrine, renal sympathetic nerve activity, heart rate, arterial pressure, and sympathoexcitation of brown adipose tissue (51–54) following systemic leptin infusion, or injection of leptin into the dorsomedial hypothalamus in rats. To our knowledge, no other studies have examined the effects of leptin administration on PNS tone in states of leptin insufficiency.

The reversal by leptin of the weight loss–associated decline in activity of the hypothalamic-pituitary-thyroid axis activity associated with maintenance of a reduced body weight is consistent with the known leptin-induced increased hypothalamic expression of proopiomelanocortin and its cleavage product α-MSH, as well as leptin’s inhibitory effects on hypothalamic expression of agouti-related peptide (AgRP). The decline in hypothalamic α-MSH and increase in AgRP act coordinately to decrease pro-thyrotropin-releasing hormone (pro-TRH) release. In addition, leptin directly stimulates pro-TRH gene expression in the paraventricular nucleus of the hypothalamus (55, 56). The decline in α-MSH and increase in AgRP act coordinately to decrease pro-TRH release in the hypothalamus (57, 58). In addition, leptin directly stimulates pro-TRH gene expression in the paraventricular nucleus of the hypothalamus (21, 59, 60).
Leptin-stimulated TRH release alone would not account for our observation that 5 weeks of leptin administration to weight-reduced subjects raised circulating concentrations of T3 and T4 but not TSH, nor the observation of Farooqi et al. (15) that 1–6 months of leptin administration to humans with congenital leptin deficiency raised circulating concentrations of bioactive thyroid hormones but not TSH. The potent effect of leptin on circulating concentrations of bioactive thyroid hormones — without effects on circulating levels of TSH — suggests that leptin directly stimulates T4 release from the thyroid gland and/or increases the bioactivity of TSH. In rats, the thyroid gland expresses the long isoform of the leptin receptor, and administration of leptin to euthyroid, leptin-sufficient animals increases circulating concentrations of T3 and T4, thyroid gland weight, and volume of the thyroid follicle epithelium without affecting circulating concentrations of TSH (61, 62). Increased TRH secretion increases TSH bioactivity in vitro and in vivo by apparent effects on sialation of TSH (63), and TSH bioactivity is decreased in low-TRH states (analogous to the low-TRH state of weight-reduced humans) (64–67).

Leptin administration in doses that raise circulating concentrations of leptin to levels within the physiological range for nonobese individuals reverses the hyperphagia and decreased circulating concentrations of gonadotropins that characterize congenital leptin deficiency (14, 24, 68, 69). Similar doses of leptin decrease hunger and increase satiety in individuals who are hypoleptinemic by virtue of lipodystrophy (70). We find that 5 weeks of leptin administration in doses similar to those that are effective in individuals who cannot synthesize leptin (24) reverses many metabolic/neuroendocrine aspects of the weight-reduced phenotype. Why should leptin have such striking physiological effects in weight-reduced subjects when, in individuals at usual body weight, doses that elevate circulating concentrations of the hormone nearly 10-fold have little effect? The primary functional role of leptin is apparently to defend — not reduce — body fat by increasing food seeking and decreasing energy expenditure when fat stores are insufficient (20, 32, 71). The threshold for determination of leptin sufficiency in this context is determined by anatomical and functional characteristics of the hypothalamus and other parts of the brain that are, in turn, determined by genetic and developmental factors. Recent indications of the structural plasticity of the hypothalamus, and of the role of leptin in mediating some of that plasticity, suggest that the threshold for leptin action may be influenced by developmental process and chronic changes in somatic fat stores (72, 73). Physiological responses to concentrations of leptin below and above this threshold are very asymmetrical: decreased concentrations of leptin trigger full-strength counterregulation to what is “perceived” as a threat to survival; concentrations of leptin above the threshold — signaling “sufficient” or excess fat stores — are not responded to vigorously, or at all (74). This asymmetry permits the deposition of additional fat stores in environmental circumstances when this is possible, conveying survival advantage in most instances. Variations in this threshold determine the minimum amounts of body fat defended by individuals. Hence the comparable responses to reductions in body fat in both lean and obese individuals (2). In this model, the experiments described here are examples of restoration of circulating leptin concentrations to points above each subject’s threshold with resultant reversal of leptin-insufficient/weight-reduced phenotypes. An alternative hypothesis is that the reversal of weight-reduced metabolic phenotypes in subjects receiving leptin might be secondary to temporal resolution of “carry-over” effects of the metabolic changes occurring during weight loss (2), rather than to effects of leptin per se. This hypothesis is not supported by the observation that reduction in TEE persists in subjects who have sustained weight loss for extended periods of time (6–24 months) in circumstances of enforced caloric restriction in the Biosphere 2 project (75), lifestyle modification (76), or bariatric surgery (77).

Studies of longer duration are needed to determine whether chronic reactivation of the leptin axis following weight loss might assist in the long-term maintenance of a reduced body weight. Physiologically, the induction of weight loss and the maintenance of reduced body weight are very different (2, 78). Pharmacologic and other strategies designed to maintain lost body weight may, for these reasons, be more effective than those designed to induce weight loss (74).

Methods

Subjects. Ten subjects (5 male, 5 female [3 never-obese, 7 obese]); mean ± SEM age 32.2 ± 2.9 years) were admitted to the General Clinical Research Center at Columbia University Medical Center and remained as in-patients throughout the study (Table 1). All subjects had been stable at their maximal lifetime weights for at least 6 months prior to admission, were in good health, and were taking no medications. Subjects with BMI (weight [kg]/[height (m)]²) greater than 30 kg/m² were classified as “obese” while subjects with BMI less than 30 kg/m² were classified as “never-obese” (79). Studies were approved by the Institutional Review Board of The New York Presbyterian Medical Center and are consistent with guiding principles for research involving humans (80). Written informed consent was obtained from all subjects.

Protocol. The protocol is schematized in Figure 1. Subjects were inpatients at the Clinical Research Center at Columbia University Medical Center throughout this study. As described previously (81), subjects were

Table 1

Subject characteristics

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<td>10.0 ± 0.6</td>
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<td>10.0 ± 0.6</td>
<td>55.4 ± 6.5</td>
<td>66.6 ± 6.0</td>
<td>55.4 ± 6.5</td>
<td>16.0 ± 0.5</td>
<td>10.0 ± 0.6</td>
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<td>Leptin (ng/ml)</td>
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*P < 0.05 versus Wt_initial; **P < 0.05 versus Wt-15%.
fed a liquid formula diet (40% of calories as fat [corn oil], 45% as carbohydrate [glucose polymer], and 15% as protein [casein hydrolysate]) plus vitamin and mineral supplements, in quantities sufficient to maintain a stable weight (defined as an average daily weight variation of less than 10 g/d for at least 2 weeks). This weight plateau is designated as Wt_maint. To prevent changes in physical fitness as a result of the 6–7 months in-patient stay, and to insure that subjects did not substantially vary their in-patient physical activity levels during the study, either of which might affect skeletal muscle metabolism/performance, each subject’s aerobic fitness was measured by bicycle ergometry upon admission and was then maintained at that level throughout the study. Aerobic fitness was defined as the anaerobic threshold, the point at which pulmonary ventilation increases out of linear proportion to O2 consumption due to the buffering of lactate by endogenous bicarbonate. Supervised exercise (treadmill walking or stationary bicycling) was performed 3 times per week at specified intensities and durations that were adjusted to maintain each subject’s anaerobic threshold at their initial level throughout the study.

Following completion of studies (described below) at Wt_maint, subjects were provided 800 kcal/d of the same liquid formula diet until they had lost 10% of Wt_maint. Once 10% weight loss had been achieved, caloric intake was adjusted upward until subjects were again weight stable as described above. This weight plateau is designated as Wt_–10%.

Following completion of studies at Wt_10s, subjects received 5 weeks of bid (8 am and 8 pm) s.c. injections of recombinant human leptin (A-100; provided by Amgen Inc.) in doses that were calculated to achieve circulating concentrations of leptin at 8 am (preinjection) equal to those measured at Wt_maint (4, 25). Initial total daily leptin doses were: 0.08 mg/kg fat mass at Wt_10s/dose in males and 0.14 mg/kg fat mass at Wt_10s/dose in females (4). Circulating leptin concentrations at 8 am were measured weekly in subjects receiving leptin, and dosages were adjusted until circulating leptin concentrations were similar to those measured at 8 am at Wt_maint. This weight plateau is designated as Wt_10lep.

At the end of each of the 3 weight plateaus, the following measurements were made starting at 9–10 am with subjects in a postabsorptive state:

**Body composition.** Body composition by dual energy x-ray absorptiometry (DXA) (7).

**Energy expenditure.** TEE was measured by empirical determination of weight-maintenance caloric intake (2). The constancy of body composition, as well as weight stability, was confirmed by demonstrating that the RQ (VCO2/VO2) for subjects at rest in the postabsorptive state was not significantly different from the liquid formula diet formula quotient (FQ) of 0.85 (see Results) (2). Since weight and body composition are constant, the energy ingested as liquid formula calories must match the total daily energy expenditure. We have previously shown that TEE measured by such caloric titration is highly correlated with TEE directly measured by the doubly-labeled water method (r2 = 0.88) (81). Since at Wt_10lep each subject remained on a diet that is isocaloric to that at Wt_maint, any change in TEE as a result of leptin administration must be reflected by change in stored energy. At Wt_10lep, TEE was calculated as: calories fed plus minus any change in whole body energy content between Wt_10s and Wt_10lep, divided by the number of days over which leptin was administered. The energy content of intercurrent weight change was calculated using DXA measurements of body composition obtained at the end of the Wt_10lep weight plateau compared with that obtained at the end of the Wt_10s weight plateau. Chemical energy content was assigned as 9.4 kcal/g wet weight of fat mass and 0.91 kcal/g wet weight of fat-free mass excluding bone (82). REE was measured by indirect (hood) calorimetry. TEF was calculated as calories expended above REE following ingestion of liquid formula calories equivalent to 60% of REE measured from 8 to 9 am on the day of testing, as previously described (81). Daily energy expenditure above REE and TEF was designated as NREE (energy expended above resting in physical activity). NREE was calculated as NREE = TEE - (REE + TEF). For the non-zero intercept of regressions relating energy expenditure to body composition (83), residual values of TEE, REE, and NREE were calculated for subjects at Wt_10s and Wt_10lep as the difference between measured energy expenditures and values predicted from the regression equations relating energy expenditure to fat mass and fat-free mass at Wt_maint (83). Data regarding the effects of weight loss and leptin administration on energy expenditure in 4 subjects (3 never-obese, 1 obese; 2 males, 2 females) have been reported previously (4).

**Skeletal muscle.** Skeletal muscle work efficiency was measured by indirect calorimetry during graded bicycle ergometry (3). Following a 10-minute period of accommodation, subjects pedaled at 60 rpm to generate 10 W of power for 4 minutes. Resistance was then increased so that power output was 25 W for 4 minutes. Oxygen uptake (VO2), carbon dioxide production (VCO2), and the RQ (VCO2/VO2) were measured continuously using a SensorMedics Vmax 29 metabolic cart attached to the subject via a mouthpiece (3). Skeletal muscle work efficiency was expressed as gross mechanical efficiency (GME): the ratio of power generated in kcal/min to change in energy expenditure above REE in kcal/min at power outputs of 10 W and 25 W (3). The metabolic fuel mix being oxidized was determined from RQ values (3).

**Autonomic nervous system activity.** In 9/10 subjects, PNS and SNS activity while resting supine were measured by analysis of changes in mean ECG R-R interval during sequential pharmacological blockade with the muscarinic PNS blocker atropine, followed by the cardioselective SNS blocker esmolol. The mean R-R interval from two 5-minute pre-blockade samples was used as the baseline against which the mean R-R intervals of the post-atropine measurement and post-atropine plus esmolol measurement were compared. Parasympathetic activity was calculated as the mean R-R interval change after atropine compared with baseline. Sympathetic activity was calculated as mean R-R interval change after atropine plus esmolol compared with the atropine alone. Intrinsic heart rate was defined as heart rate following complete SNS and PNS blockade (6). SNS activity measured by pharmacological blockade reflects the action of catecholamines (predominantly norepinephrine) released at nerve endings and from the adrenal gland (predominantly epinephrine), as well as the sensitivity, number, and relative proportion of cardiac α- and β-adrenoceptors (84). In order to examine the effects of weight loss and leptin administration on catecholamine production, 24-hour urine catecholamine excretion (dopamine, epinephrine, norepinephrine) was assayed by HPLC (85) at each study period in all subjects. The intraassay coefficients of variation for these assays are less than 5%. The interassay coefficients of variation for assays of dopamine, epinephrine, and norepinephrine are, respectively, 6.2%, 5.7%, and 7.1%.

**Neuroendocrine.** 8 am circulating plasma concentrations of leptin were measured by ELISA (Diagnostic Systems Laboratories Inc.), and serum concentrations of thyroid stimulating hormone (TSH), T4, and T3 by RIA (86). The intra- and interassay coefficients of variation for each of these assays are less than 5%. Data regarding the effects of weight loss and leptin administration on thyroid hormones in 4 subjects (3 never-obese, 1 obese; 2 males, 2 females) have been reported previously (4).

All women were in the follicular phase of their menstrual cycle at the start of testing. The order of testing was similar, but not identical, in all subjects. In larger study groups, we have not found a significant effect of the order of testing or of duration of time spent at reduced weight on any of these parameters (2).

**Statistics.** Data were analyzed using the STATISTICA 6.0 software package (StatSoft Inc.). Data are presented as mean ± SEM. Intragroup comparisons (same subjects at different study periods) were made by ANOVA with
repeated measures. Between-group comparisons (males versus females, obese versus never-obese) were made by ANOVA. Statistical significance was prospectively defined as $p < 0.05$.

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Address correspondence to: Michael Rosenbaum, Russ Berrie Medical Science Pavilion, Columbia University Medical College, Room 620, 1150 St. Nicholas Avenue, New York, New York 10032, USA. Phone: (212) 305-9949; Fax: (212) 8 51-5306; E-mail: mr475@columbia.edu.