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PLCγ2 regulates osteoclastogenesis via its interaction with ITAM proteins and GAB2

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Excessive bone loss in arthritic diseases is mostly due to abnormal activation of the immune system leading to stimulation of osteoclasts. While phospholipase Cγ (PLCγ) isoforms are known modulators of T and B lymphocyte–mediated immune responses, we found that blockade of PLCγ enzymatic activity also blocks early osteoclast development and function. Importantly, targeted deletion of Pleg2 in mice led to an osteopetrosis phenotype. PLCγ2, independent of PLCγ1, was required for receptor activator of NF-κB ligand–induced (RANKL-induced) osteoclastogenesis by differentially regulating nuclear factor of activated T cells c1 (NFATc1), activator protein–1 (AP1), and NF-κB. Specifically, we show that NFATc1 upregulation is dependent on RANKL-mediated phosphorylation of PLCγ2 downstream of Dap12/Fc receptor γ (Dap12/FcRγ) receptors and is blocked by the PLCγ inhibitor U73122. In contrast, activation of JNK and NF-κB was not affected by U73122 or Dap12/FcRγ deletion. Interestingly, we found that in osteoclasts, PLCγ2 formed a complex with the regulatory adapter molecule GAB2, was required for GAB2 phosphorylation, and modulated GAB2 recruitment to RANK. Thus, PLCγ2 mediates RANKL-induced osteoclastogenesis and is a potential candidate for antiresorptive therapy.

Introduction
Bone erosion is a major hallmark of rheumatoid arthritis and is solely executed by the bone-resorbing cells, the osteoclasts (OCs) (1). These cells arise from macrophage precursors and differentiate into the mature polykaryon after stimulation with receptor activator of NF-κB ligand (RANKL) and M-CSF (2). OCs are recruited to sites of inflammation or differentiate at these sites owing to elevated levels of local RANKL secreted by cells in the inflamed tissue. Levels of TNF-α, IL-1, and M-CSF are also elevated during inflammatory responses and they collaborate with RANKL to further increase the rate of OC formation and bone resorption (3–6).

Binding of RANKL to receptor activator of NF-κB (RANK) leads to receptor trimerization, followed by the recruitment of TRAF6, which acts as a key adapter to assemble signaling proteins that direct OC-specific gene expression (2). Both NF-κB and activator protein–1 (AP1) are rapidly induced in response to RANKL, following the activation of IkB kinase 1/2 (IKK1/2) and JNK, respectively (2, 7–9). Interfering with IKK1/2 or JNK1 signaling pathways blocks in vitro and in vivo osteoclastogenesis (10–12). Targeted mutagenesis of the p50/p52 components of NF-κB (13, 14) as well as the c-Fos component of AP1 (15) results in osteopetrosis due to a block in osteoclastogenesis. Further, a recent finding indicates that activation of JNK and NF-κB pathways requires the phosphorylation and the recruitment of the Grb2-binding adapter protein, GAB2, to RANK (16). Consistent with these observations, GAB2−/− mice exhibit an osteopetrotic phenotype due to aberrant OC differentiation (16).

In addition to the known roles of AP1 and NF-κB pathways in osteoclastogenesis, recent studies have highlighted the role of the nuclear factor of activated T cells c1 (NFATc1), a calcineurin- and calcium-regulated transcription factor, as a master transcription factor in this process (17, 18). The importance of NFATc1 in OC development has been buttressed by the fact that NFATc1−/− embryonic stem cells cannot differentiate into OCs. Furthermore, overexpression of the WT protein stimulates OC development from embryonic stem cells in a RANKL-independent manner (18). Interestingly, the ability of NFATc1 to mediate osteoclastogenesis is strictly connected to the c-Jun pathway (19). Transgenic mice expressing a dominant negative form of c-Jun have osteopetrosis due to defective osteoclastogenesis that cannot be rescued by NFATc1 overexpression in OCs (19).

Immune receptor tyrosine activation motifs (ITAMs) are important signaling components for a number of receptors in the immune system, and they have been recently identified as costimulatory signals in RANKL-induced osteoclastogenesis (20). The 2 ITAM-containing adapters, Dap12 and the γ chain of Fc receptor (FcγR), have a profound impact on OCs by modulating RANKL-mediated calcium influx and NFATc1 expression. In humans, mutation in the Dap12 gene is responsible for Nasu-Hakola disease, characterized by multiple bone cysts associated with a unique form of neurodegeneration (21). Deletion of Dap12 and FcRγ (Dap12−/− FcRγ−/−) in mice, leads to severe osteopetrosis due to defective OC development (20–22). In vitro triggering of either receptor by crosslinking with an antibody accelerates RANKL-induced OC differentiation (23). However, in the absence of RANKL, these receptor-mediated signals alone are not sufficient to induce OC differentiation.

The phospholipase C (PLC) family regulates PKC activation, intracellular calcium levels, and NFAT expression in hematopoietic systems in response to the stimulation of immune receptors (24–27). PLC cleaves the membrane phospholipid phosphatidylinositol-4,5-biphosphate (PIP2) into the second messenger molecules inositol-1,4,5-triphosphate (IP3) and diacylglycerol (DAG). IP3 directly increases intracellular calcium levels by inducing the release of endoplasmic reticulum calcium stores, while DAG activates PKC at the plasma membrane.
The PLCγ family consists of 2 members, PLCγ1 and PLCγ2 (24–26). Both isoforms require phosphorylation on specific tyrosine residues for their catalytic activity. In addition, they contain 2 SH2 and 1 SH3 domains through which they can interact with other signaling proteins, suggesting adapter function. PLCγ1 is widely distributed, while PLCγ2 is primarily limited to cells of hematopoietic lineage (24). Mice deficient in PLCγ1 develop normally to embryonic day 8.5 but die soon after due to undefined defects in growth and development (28). The disruption of PLCγ2 in mice does not result in embryonic lethality, but animals show internal bleeding, decreased B cell number and function, defective platelet aggregation to collagen, and a block in Fc receptor–mediated responses in mast cells and NK cells (29–31).

Although NF-κB, AP1, and NFATc1 are all critical for RANKL-induced osteoclastogenesis (9, 18, 32), due to their ubiquitous expression, selective therapies targeting these signaling pathways in OCs are not yet in clinical use. In this study, we show that targeted deletion of Pleg2 in mice results in an in vivo osteopetrotic phenotype, independent of PLCγ1. In vitro, Pleg2-null cells fail to form multinucleated OCs and exhibit decreased RANKL-mediated NFATc1 expression; impaired JNK, c-Jun, and IκBα phosphorylation; and thus defective AP1 and NF-κB activation. Our data reveal a role for PLCγ2 in bone homeostasis apart from its contribution to B cell development. The combined effect of PLCγ2 on lymphocyte and OC differentiation could represent a novel target for the treatment of inflammatory osteolytic diseases.

Results

Inhibition of PLCγ enzymatic activity blocks OC differentiation and disrupts actin ring formation. PLCγ isoforms are known activators of the NFAT family of transcription factors in lymphoid cells, downstream of immune receptor signaling via ITAMs (33). Since NFAT and ITAMs are both critical regulators of OC differentiation and cells lacking the ITAM-containing adapters Dap12/FcRγ display severe osteopetrosis (20), we asked whether PLCγ family members were also required for efficient OC differentiation and function. We therefore tested the capacity of the PLCγ inhibitor U73122 to block OC formation and bone resorption. Addition of U73122 to the culture media blocked OC development, as demonstrated by the absence of tartrate-resistant acid phosphatase–positive (TRAP-positive) cells, compared with the numerous multinucleated OCs obtained in wells cultured only with RANKL and M-CSF (Figure 1A). Replacing the media with fresh osteoclastogenic media lacking the inhibitor restored the ability of the cells to differentiate, indicating that the compound was not toxic to the cells (Figure 1A). Culture of OCs on dentin in the presence of U73122 blocked bone resorption (Figure 1B). Furthermore, addition of U73122 to mature resorbing cells rapidly disrupted the organization of the actin rings, indicating that this compound can block bone resorption independent of its capacity to affect differentiation (Figure 1C).

PLCγ2 regulates bone mass in vivo. To determine whether PLCγ1, PLCγ2, or both are required for OC formation, we turned to the analysis of mice lacking these proteins. Pleg1−/− mice die in utero by day 9, rendering the analysis of their bone phenotype difficult. Pleg2−/− mice, however, are viable. Thus, femurs from 4-month-old sex-matched WT and Pleg2−/− mice were subjected to histological examination (Figure 2, A–E) and μCT analysis (Figure 2, F–I). Pleg2−/− mice had a more than 3-fold increase in the percentage of trabecular bone volume versus total bone volume compared with their WT counterparts, as determined by histological examination (Figure 2B) and as shown in μCT reconstructed 3D images (Figure 2F). Histomorphometric analysis also indicated that the number of osteoblasts (OBs) per bone perimeter was similar in WT and Pleg2−/− mice (Figure 2C), while the number of OCs per bone perimeter and the OC surface were significantly decreased in the null animals (Figure 2, D and E). The number of the trabeculae (Figure 2G) and their thickness (Figure 2H) were significantly higher in the Pleg2−/− compared with WT mice, while the space between the trabeculae was smaller (Figure 2I). Thus, PLCγ2 regulates bone mass in vivo.

PLCγ2 is required for OC differentiation. To determine whether PLCγ2 modulates the capacity of OBs to form bone or to promote OC differentiation, primary OBs were isolated from newborn mice, and their capacity to mineralize extracellular matrix or induce osteo-

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**Figure 1**

PLCγ inhibition blocks osteoclastogenesis and actin ring formation. (A) WT OCs were generated with RANKL (100 ng/ml) and M-CSF (10 ng/ml) in the presence of the PLCγ inhibitor U73122 (5 μM) for 4 days on plastic. In some wells, the media with the inhibitor was replaced with fresh osteoclastogenic media, and cells were allowed to differentiate for 4 more days (Withdrawal U73122). (B) WT OCs were grown on dentin with or without U73122 for 10 days. Cells were then removed and pits stained with hematoxylin red (Magnification, ×200). CTR, control. (C) WT OCs were generated on dentin in the absence of the inhibitor, then treated with vehicle or U73122 for 1 hour. Cells were fixed and actin stained using FITC-phalloidin (Magnification, ×200).
clastogenesis in a coculture system was examined. *Plcg2*−/− OBs were indistinguishable from their WT counterparts in their ability to form bone nodules in response to β-glycerolphosphate (WT, 14 ± 2.08 nodules/well versus *Plcg2*−/−, 14.6 ± 1.65 nodules/well) (Figure 3A). This finding was also supported by the in vivo analysis of bone mineral apposition determined by calcein double-labeling (Figure 3B), which revealed no significant difference in bone formation rates (BFRs) and mineral apposition rates (MARs) between WT (BFR, 0.8938 ± 0.1042 μm/μm3/d; MAR, 0.5927 ± 0.0219 μm/d) and *Plcg2*−/− animals (BFR, 0.8450 ± 0.1629 μm/μm3/d; MAR, 0.4015 ± 0.0876 μm/d). To determine whether *Plcg2*−/− OBs are capable of supporting OC differentiation, BM macrophages (BMMs) from WT mice were cocultured with either WT or *Plcg2*−/− OBs in the presence of 1,25 vitamin D3 (100 ng/ml) and M-CSF (10 ng/ml) up to 4 days. The expression of TRAP, cathepsin K, and NFATc1 increased within 24–48 hours after RANKL stimulation in WT but not in *Plcg2*−/− cells (not shown). On day 4, the levels of these genes were maximal in WT cells but dramatically lower in the null cells (Figure 4, A–D). Similarly, calcitonin receptor was detected after 4 days of treatment with RANKL only in WT OCs, suggesting that *Plcg2*−/− cells have an early block of OC development. Confirming mRNA expression, protein levels of NFATc1 were significantly higher in total cell lysates of day 4 WT OCs than in null cells, both under basal conditions and following 60 minutes of RANKL stimulation (Figure 4E). Together, these data indicate that PLCγ2 is required at early stages of OC development.

PLCγ2 is required for efficient RANKL-induced activation of AP1 and NF-κB in developing OCs. To further understand whether PLCγ2 is a downstream mediator of RANKL, M-CSF, or both during OC differentiation, we analyzed MAPK activation by BMMs in culture with M-CSF for 3 days. Activation of ERK and JNK in response to RANKL (Figure 5A). Similarly, ERK was normally phosphorylated in the null cells in response to M-CSF (Figure 5A). Together, these data indicate that PLCγ2 is required at early stages of OC development.

**Figure 2**

Osteopetrosis in mice lacking PLCγ2. (A) TRAP staining of decalcified histological sections of WT and *Plcg2*−/− proximal femurs. Magnification, ×40. (B–E) Quantitative analysis of bone parameters from histological sections of WT and *Plcg2*−/− bone femurs (n = 8) showing: percentage of bone volume versus total bone volume (BV/TV) (B); number of OBs (nOB) per bone perimeter (C); number of OCs (nOC) per bone perimeter (D); fraction of trabecular surface covered by OCs (E). (F) 3D reconstitution of μCT scans of WT and *Plcg2*−/− femurs. (G–I) 3D trabecular quantitative parameters of bone structure (n = 6). Graphs show mean ± SEM, with significant differences compared with WT indicated.
To determine whether lack of PLCγ2 was abrogating the activation of PLCγ1 in response to RANKL and M-CSF, WT and Plcg2−/− OC precursors (BMMs grown in RANKL-containing media for 2 days) were stimulated with RANKL or M-CSF. Interestingly, RANKL-mediated PLCγ1 phosphorylation was barely measurable in both WT cells and cells lacking PLCγ2 (Figure 6B) despite detectable levels of PLCγ1 protein in both cell types. In contrast, a strong PLCγ1 phosphorylation signal was observed after M-CSF stimulation in WT and Plcg2−/− cells (Figure 6C).

Since costimulatory signals are required for RANKL-mediated calcium oscillation and NFATc1 expression, we determined whether PLCγ2 phosphorylation was dependent on the RANKL costimulatory receptors Dap12 and FcRγ. We found that PLCγ2 phosphorylation and NFATc1 nuclear localization were decreased in Dap12−/− FcRγ−/− cells treated with RANKL (Figure 6E and F). Reexpression of Dap12, but not empty vector, in the double null OCs (Figure 6D) completely restored PLCγ2 phosphorylation (Figure 6E) and expression of NFATc1 in the nucleus (Figure 6F). Thus, PLCγ2 is phosphorylated downstream of Dap12 in an SFK-dependent manner, following RANKL stimulation.

PLCγ2 catalytic activity is required for NFATc1 upregulation but not for JNK and IκBα phosphorylation. Our data indicate that PLCγ2 regulates NFATc1 and activates JNK and IκBα. In contrast, Da...
indistinguishable from that of WT PLCγ2 (Figure 7D). These data indicate that PLCγ2 can modulate RANK-mediated signaling independent of its catalytic activity by acting as an adapter molecule. One potential interacting protein is GAB2, which associates with RANK and mediates RANK-induced activation of NF-κB and JNK but not NFATc1 (16). Furthermore, GAB2 has been shown to bind PLCγ2 in mast cells (36). To determine whether PLCγ2 binds to GAB2 in OCs, we reciprocally immunoprecipitated GAB2 and PLCγ2 and found that the two molecules (Figure 7E). Immunoprecipitation with IgG control antibodies revealed the absence of non-specific bands (data not shown). To further determine whether PLCγ2 modulates GAB2 activation or its recruitment to RANK, WT and Plcg2−/− BMMs, treated with RANKL, were immunoprecipitated with an anti-GAB2 Ab (Figure 7F) or with rabbit IgG control Ab (data not shown), followed by immunoblotting with phospho-tyrosine (4G10) and RANK. Results showed that PLCγ2 modulates GAB2 phosphorylation and its association with RANK, suggesting that this may be the mechanism by which PLCγ2 controls AP1 and NF-κB activation.

TNF-α cannot reverse the arrested capacity of PLCγ2-null cells to become OCs. The data presented above show that PLCγ2 is central in RANKL signaling. In OCs, TNF-α appears to be the dominant cytokine mediating inflammatory osteolysis and augments the
Due to the capacity of TNF-α to phosphorylate JNK and IkBα in BMMs and OCs (37), we tested its effect in modulating the above-mentioned signaling pathways in cells lacking PLCγ2. TNF-α-mediated phosphorylation of JNK and IkBα occurred similarly in WT and null cells (Figure 8A). Furthermore, TNF-α was not capable of inducing PLCγ2 phosphorylation (Figure 8A), suggesting that the mechanism by which TNF-α regulates JNK and IkBα is PLCγ2 independent. This result prompted us to determine whether TNF-α could rescue the osteoclastogenic defect of Pleg2−/− cells. Interestingly, while TNF-α augmented RANKL-mediated osteoclastogenesis of WT cells (WT OCs without TNF-α, 19.3 ± 1.5/well; WT OCs with TNF-α, 45.3 ± 1.99/well), the cytokine did not have any effect on the null cells, which remained blocked at the early stage of OC development (Pleg2−/− OCs, not detected) (Figure 8B). Consistent with this finding, TNF-α, compared with RANKL, was not capable of promoting NFATc1 upregulation in WT or in PLCγ2-null cells (Figure 8C).

**Discussion**

The interaction between the immune and skeletal systems is a significant cause of pathological bone loss in rheumatoid arthritis, periodontal diseases, and tumor-associated bone metastasis (38). This exaggerated bone loss is most frequently due to abnormal activation of the immune system, leading to hyperstimulation and differentiation of the OCs (2, 39). The identification of signaling pathways or molecules common to the immune and bone systems could help in designing therapies for the variety of diseases that affect both bone and immunity. Recently, an activating point mutation in the murine PLCγ2 gene has been shown to lead to severe spontaneous inflammation and autoimmunity, mediated by B cells and non-B and non-T hematopoietic cells (40). The chronic inflammation resulted in severe arthritis of the small joints, leading to phalangeal erosion (40). This observation suggested that the activating mutation in PLCγ2 gene could also be responsible for the hyperactivation and recruitment of the OCs at the inflammatory sites.

In this study we have identified what we believe to be a novel role for the immunomodulatory protein PLCγ2 as a key factor in the regulation of bone homeostasis. Targeted deletion of Pleg2 led to increased bone mass due to blockade of OC differentiation in unmanipulated mice, with decreased RANKL-mediated signaling to API and NF-κB and defective upregulation of NFATc1, independent of PLCγ1. Despite the presence of normal levels of PLCγ1 in the PLCγ2-null cells, PLCγ1 could not compensate for the lack of its cognate isoform. Furthermore, we found that PLCγ2, and not PLCγ1, was
phosphorylated by RANKL. This result suggests that the 2 PLCγ isomors exert different, nonoverlapping functions in OCs.

Differential regulation of the 2 PLCγ isoforms within the same cell type has been previously demonstrated in platelets, where activation of glycoprotein VI by collagen resulted in a substantial increase in tyrosine phosphorylation of PLCγ2 but not PLCγ1 (41, 42). Since activation of PLCγ isoforms is tightly regulated by the action of a number of receptor tyrosine kinases (RTKs) (24–26), we asked whether M-CSF, via its receptor c-Fms, could lead to phosphorylation of PLCγ1, PLCγ2, or both. We found that PLCγ1, and to a lesser extent PLCγ2, was phosphorylated by M-CSF. In support of this finding, analysis of Plcg1−/− ES cells generated few hematopoietic cells with a reduced level of monocyte/macrophage differentiation (29). Furthermore, we did not detect an M-CSF–dependent proliferative defect in BMMs and OCs lacking PLCγ2, and we observed that M-CSF–mediated activation of ERK and JNK occurred normally. These results indicate that PLCγ2 is not involved in M-CSF signaling and suggest that PLCγ1 may regulate this pathway in OC lineage cells.

The phosphorylation of PLCγ2 in response to RANKL was mediated in an SFK-dependent manner. PP2, a well-known Src kinase inhibitor, completely blocked RANKL–mediated PLCγ2 activation. Moreover, Dop12 was required for phosphorylation of PLCγ2 in response to RANKL stimulation. It is possible that SFKs phosphorylate PLCγ2 directly. However, it is more likely that SFKs phosphorylate the tyrosine residues in the ITAM motif of Dop12, leading to recruitment and activation of Syk, which in turn can activate PLCγ1, as occurs in other cell types (B cells and mast cells) (43, 44). We did not determine which Src family member is involved in PLCγ2 activation, since the inhibitor used blocked various SFKs. These results indicate that PLCγ is isoforms are differentially phosphorylated in OCs, PLCγ2 primarily by RANKL and PLCγ1 by M-CSF. AP1, NF-κB, and NFATc1 are members of well-described osteoclastogenic pathways activated by RANKL. We found that PLCγ2

Figure 7
PLCγ2 forms a complex with GAB2 and modulates its activation. (A) WT preOCs cultured with or without the PLC inhibitor U73122 (5 μM) for 3 days were stimulated with RANKL and subjected to Western blot analysis for phospho-JNK, phospho-IκBα, and NFATc1. β-Actin served as control. (B) WT and Plcg2−/− BMMs retrovirally transduced with empty vector (pMX), WT PLCγ2, or catalytically inactive PLCγ2 (PLCγ2 H/F) were cultured with RANKL (100 ng/ml) and M-CSF (10 ng/ml) for 7 days, and multinucleated OCs were detected by TRAP staining. Objective, ×10. (C) The same cells as shown in B were subjected to RANKL stimulation and Western blot analysis to detect NFATc1 expression. (D) Plcg2−/− BMMs retrovirally transduced with pMX, WT PLCγ2, or PLCγ2 H/F were treated with RANKL, and phosphorylation of IκBα and JNK was determined by Western blot analysis. PLCγ2 expression levels are shown. β-Actin served as loading control in C and D. (E) PLCγ2 and GAB2 were reciprocally immunoprecipitated in WT and Plcg2−/− BMMs treated with RANKL and subjected to Western blot analysis using anti-PLCγ2 and anti-GAB2 Abs, respectively. TCL, total cell lysate. (F) GAB2 was immunoprecipitated in WT and Plcg2−/− BMMs and subjected to Western blot analysis using anti-phosphotyrosine Ab (clone 4G10), anti-RANK, and anti-PLCγ2.
Based on the observation that PLCγ2 for activation of NF-κB and JNK, but not NFATc1, pathways (16). Based on the observation that PLCγ2 forms a complex with Gab2 in OCS and in other cell types (36), we hypothesized that PLCγ2 could regulate some RANKL signaling pathways by acting as an adapter molecule. One possibility is that PLCγ2 modulates the Gab2/RANK complex. In support of this hypothesis, we found that PLCγ2 is required for Gab2 phosphorylation and mediates Gab2 recruitment to RANK, suggesting that API and NF-κB activation might be dependent on the capacity of PLCγ2 to form a complex with Gab2. In addition, the finding that the PLC inhibitor U73122, which blocks IP3-mediated calcium influx (45), and a catalytically inactive form of PLCγ2 completely abrogate osteoclastogenesis by affecting NFATc1 upregulation suggest that both the catalytic activity and the adapter function of PLCγ2 are required for osteoclastogenesis. Thus, overall our data support the model (Figure 9) in which the catalytic activity of PLCγ2 is required for NFATc1 upregulation downstream of the ITAM motif of Dap12, while PLCγ2, acting as an adapter molecule, mediates Gab2 phosphorylation and its recruitment to RANK, leading to API and NF-κB activation.

The specific requirement of PLCγ2 for activation of NF-kB, API, and NFATc1 signaling pathways is manifested in a basal in vivo osteopetrotic phenotype characterized by defective OC recruitment. Although the number of OCs in vivo was significantly reduced in Plcg2-null compared with WT animals, the presence of a few TRAP-positive cells could reflect the presence of a milieu of cytokines circulating in vivo that partially rescue the differentiation of PLCγ2-null OCs. TNF-α potently stimulates osteoclastogenesis by augmenting RANKL-mediated signaling pathways (46). TNF-α can also induce JNK and IkBα phosphorylation (37). Thus, we sought to determine whether TNF-α could reverse the ability of the PLCγ2-null cells to become OCs. Interestingly, while TNF-α could activate JNK and IkBα in Plcg2-null BMMs, it could not reverse their differentiation defect, most likely due to the inability of TNF-α to induce NFATc1 upregulation. In conclusion, our data indicate that PLCγ2 is required for the maintenance of bone homeostasis. The inability of Plcg2-null mice to respond to both basal and inflammation-mediated cytokines circulating in vivo that partially rescue the differentiation of OCs (29). The inability of Plcg2-null cells to respond to both basal and inflammation-mediated cytokines circulating in vivo that partially rescue the differentiation of OCs (29).
osteoclastogenesis in vitro clearly supports the important role of PLCγ2 in osteoclast differentiation. Thus, the intrinsic defect of Pleg2−/− OCs together with the deficient B cell–mediated immune response suggest that targeting PLCγ2 could be a novel approach for antiresorptive therapies. Antirheumatic therapies directed to selectively deplete human CD20-positive B cells (e.g., Rituximab) or to directly target OC activation and function (RANKL inhibitor) have been very useful approaches in recent clinical trials (47). However, in most cases, only a combination of both treatments will resolve both the inflammatory and the ongoing osteolytic responses. Thus, PLCγ2 becomes an interesting target for the cure of rheumatoid diseases due to its dual role in the activation of a particular subset of immune cells, B lymphocytes, and for its capacity to control OC development in vivo. Therefore, a combined therapy affecting both the inflammatory and the osteolytic components of the rheumatoid disease might increase the hope that disease modification and remission in rheumatoid arthritis is a realistic goal.

Methods

Mice. Pleg2−/−, Dap12−/−/FcRγ−, and WT mice were on a C57BL/6 background and have been previously described (20, 29). All mice used in these experiments were 6–16 weeks old and bred and maintained with sterilized food, water, and bedding at the Animal Facility of the Washington University School of Medicine. All experiments were approved by the Animal Studies Committee of Washington University.

Histology and μCT. Five-micron sections of fixed, decalcified, paraffin-embedded long bones were stained with either H&E or histochemically for TRAP to detect OCs. Calvariae from mice injected with calcein (20 mg/kg dissolved in 2% NaHCO3 Sigma-Aldrich) were embedded in plastic. For μCT, 3D images from intact mouse femurs were obtained on a μCT40 scanner (Scanco Medical).

Mouse OCs. BMMs were isolated from long bones of 6- to 8-week-old mice as described previously (48). We obtained OCs by culturing BMMs with glutathione-S-transferase–RANKL (GST-RANKL) (100 ng/ml) and M-CSF (10 ng/ml) for 5 days. For coculture experiments, OBs and BMMs were mixed and cultured for 10–14 days in the presence of 1,25 Vit D3 (10−8 M). For rescue experiments, macrophages were obtained as described above; transduced with Flag-Dap12, Flag-PLCγ2, or Flag-PLCγ2 H/F; selected in puromycin– (for Dap12 constructs) or blastocystin-containing media (for PLCγ2 constructs) for 3 days; and cultured with M-CSF and RANKL to obtain OCs, as previously described (49). The PLCγ2 H/F mutation was made using a QuikChange XL Site-Directed Mutagenesis Kit (catalog 200517; Stratagene) following the manufacturer’s instructions.

Mouse OBI. We isolated OB precursors from calvariae of mice that were 4 days old, expanded them in media alone until confluent, and then cultured them with β-glycerophosphate and ascorbic acid to induce OB differentiation until bone nodules formed.

Western blot analysis and antibodies. BMMs or OCs starved for 6 hours in serum-free medium were stimulated with RANKL (100 ng/ml) or M-CSF (100 ng/ml) and lysed at the times indicated in the Results and Figures 4-8 in RIPA lysis buffer supplemented with protease inhibitors and sodium orthovanadate. In some experiments, OCs were cultured in the presence of the Src inhibitor PP2 (5 μM; Calbiochem) or PLC inhibitor U73122 (5 μM; BIOMOL), starved, and then stimulated with RANKL. For Western blot analysis of phospho-PLCγ1 and -PLCγ2, phospho-ERK, phospho-JNK, phospho-c-Jun, phospho-ικBα, and phospho-Src (Y416), we used polyclonal antibodies from Cell Signaling Technology. Polyclonal antiserum against p65, NFATc1, and PLCγ2 were purchased from Santa Cruz Biotechnology Inc. The monoclonal antibody against β-actin was obtained from Sigma-Aldrich. Polyclonal GAB2 Ab was purchased from Upstate USA Inc. To obtain nuclear extract from RANKL-treated cells, plates were washed with H2O and cells lysed with hypotonic buffer (10 mM HEPES, 1.5 mM MgCl2, 1 mM KCl, 1 mM DT, and protease and phosphatase inhibitors), followed by addition of 0.1% NP40. After centrifugation, the supernatants were collected (cytosolic fraction), while the pellets (nuclear fraction) were suspended in high-salt buffer (hypotonic buffer plus 400 mM NaCl). Extracts were quantified by a modified Coomassie method (Pierce). Nonradioactive EMSA was performed following the manufacturer’s instructions (Pierce) using AP1 and NF-κB biotinylated oligonucleotides (for AP1: 5′-Biotin-AAGTTGAGGGGACTTTCCCAGGCT-3′ and 5′-Biotin-ATTCCCCGGTGAATGTCATGTTCT-3′; for NF-κB: 5′-Biotin-AAGTTGAGGGGACTTTCCCAGGCT-3′ and 5′-Biotin-AAGTTGAGGGGACTTTCCCAGGCT-3′). An excess (200X) of nonbiotinylated oligonucleotides was used for competition control.

Immunofluorescence. WT or Dap12−/− FcRγ− OCs retrovirally transduced with either the empty vector or full-length Dap12 were cultured on glass, fixed, and stained with anti-NFATc1 Ab (Santa Cruz Biotechnology Inc.), DAPI to detect the nuclei, and FITC-phalloidin to detect actin (48).

Real-time PCR. Total RNA was isolated from cell cultures at various times using the RNasy Mini Kit (QIAGEN) and was reverse transcribed to cDNAs using SuperScript II according to the manufacturer’s instructions (Invitrogen). Primers specific for murine NFATc1, TRAP, GAPDH, cathepsin K, and calcitonin receptor were used. For quantitative real-time PCR, NFATc1 was amplified using 5′-CCCTGACATTCTGTGCTCCAT-3′ and 5′-CAATTAACCTCGTTAGCACA-3′.

TRAP was amplified using 5′-CACGCTCCCTAGAGATGAGATCAT-3′ and 5′-GATGTGAGTTGGGACATGTG-3′; β-actin using 5′-CTTCTACAATGAGCTGCTG-3′ and 5′-TCATGAGGTATGCTGCACTG-3′; cathepsin K using 5′-ATGTGGGTGTTCAAGTTTCGC-3′ and 5′-CAATCAACCTGTTAGCACA-3′.

TRAP was amplified using 5′-GATGTGAGTTGGGACATGT-3′ and 5′-GATGTGAGTTGGGACATGTG-3′; β-actin using 5′-CTTCTACAATGAGCTGCTG-3′ and 5′-TCATGAGGTATGCTGCACTG-3′; cathepsin K using 5′-ATGTGGGTGTTCAAGTTTCGC-3′ and 5′-CAATCAACCTGTTAGCACA-3′.

SYBR green dye was used for detection of the product using the SYBR Green PCR Master Mix assay (Applied Biosystems). The standard
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