Autoantibody selection and production in early human life

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Natural antibodies are autoreactive/polyreactive antibodies believed to be secreted in the absence of xenoantigens. The origin and functional role of this limited and selective autoimmune reaction are not clear, nor is the specificity and range of autoantibodies that drive the development of B cells producing natural antibodies. In this issue of the *JCI*, Merbl et al. report that in utero, humans generate natural IgM and IgA antibodies that recognize a uniform set of autoantigens (see the related article beginning on page 712), some of which are associated with autoimmune diseases. The authors postulate that this “autoimmunity” at birth favors the emergence of autoimmune diseases in later life. We present a molecular basis for the limited and common repertoire of antibodies produced by fetal B cells, which may be distinct from the abnormalities in B cell development described in patients with autoimmune diseases.

B cell tolerance checkpoints remove many developing self-reactive B cells from the adult repertoire, yet the presence of certain autoreactive B cells in normal individuals is revealed by the identification of serum autoantibodies referred to as natural antibodies (1, 2). Most natural antibodies are low-affinity IgM autoantibodies that are often polyreactive and bind to a broad range of self-antigens (2, 3). In mice, natural antibody–secreting B cells belong to a specific B cell subpopulation called B-1 cells, which preferentially reside in the peritoneal cavity (4). The fact that these peripheral autoreactive B cells are positively selected by self-antigens seems paradoxical to B cell tolerance, but raises the possibility that autoantibodies may in some cases perform useful functions (5). In humans, the origin of natural antibodies and the autoantibodies that they recognize remain to be characterized. In this issue of the *JCI*, Merbl et al. (6), using an antigen microarray, identified specific autoantigens recognized by natural IgM and, to a lesser extent, IgA antibodies produced during human fetal and neonatal life (6). In addition, they found that autoantibodies recognized by serum IgMs from newborns were remarkably similar among individuals, suggesting that in the absence of xenoantigens, specific autoantibodies may select and stimulate autoreactive B cells to produce self-reactive natural IgM antibodies in fetuses (6). In contrast, IgM and IgA antibodies from mothers showed an antigenic recognition pattern different from that of their newborns. Furthermore, IgM and IgA recognition patterns were extremely diverse among the mothers studied, reflecting the distinct immunological histories of each individual. However, encounters with some common pathogens, such as Gram-negative bacteria, stimulated the production of anti-LPS antibodies in all mothers (6).

Potential origins of common natural antibody reactivity at birth

What might account for the recognition of the same autoantigens by different individuals during early human development? Antibodies are generated by random recombination of Ig variable (V), diversity (D), and joining (J) gene segments during early B cell development. Analysis of B cells from fetuses and neonates showed that this was not the case in early life (7, 8). Rather, Ig gene segment usage in early life is biased toward specific genes, thereby lim-

Nonstandard abbreviations used: BCR, B cell receptor; CD8α, complementarity-determining region 3; D, diversity; H, heavy chain; IgH, Ig heavy chain; J, joining; NMHC-II, nonmuscle myosin heavy chain type II; TDT, terminal deoxynucleotidyl transferase; V, variable; XLA, X-linked agammaglobulinemia.

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Voting the extent of Ig diversity (Figure 1). For example, fetal liver B cells have been shown to have a recombination bias toward heavy chain (H) V_{H}6–1 and D_{7}–27 segments located at the 3′ end of the V_{H} and D loci (Figure 1) (7, 8). J_{H} gene usage is also skewed toward more upstream J_{H}1–4 genes, and J_{H}6 is rarely found in rearrangements from fetal and neonatal B cells (Figure 1) (7, 8). Ig diversity in early life is further limited by low activity of terminal deoxynucleotidyl transferase (TDT), an enzyme that adds nontemplate nucleotides between V_{H}–D and D–J_{H} joining regions (7, 8). As a consequence of the lack of insertion of nontemplate nucleotides combined with the fact that D_{7}–27, the segment preferentially used in fetal liver B cells, is the shortest D segment, Ig heavy chain (IgH) complementarity–determining region 3 (CDR3) involved in antigen recognition is particularly short in fetuses and neonates compared with that in adults (7, 8).

Taken together, these restrictions to V(D)J recombination that occur during fetal and neonatal life may explain both the generation of B cell clones that recognize common autoantigens and the similarities in antibody repertoires among different individuals. It is likely that this incomplete fetal/neonatal antibody repertoire is not detrimental to neonates, because protective maternal IgG antibodies cross the placenta and provide potent and properly selected antibodies required for host defense.

B cell tolerance checkpoints in fetuses and neonates

IgH CDR3s generated by V(D)J recombination play an essential role in conferring polyreactivity to natural antibodies (3). In adults, higher TDT activity and usage of J_{H}6 (the longest of all J_{H}s) are responsible for the production of longer IgH CDR3s than those in fetuses/neonates. In addition, TDT-dependent nontemplate nucleotides often encode positively charged amino acids rarely found in D segments, which, when combined with long IgH CDR3s, often lead to the production of autoreactive antibodies (1, 7). Although this random V(D)J recombination described in healthy adults generates a large number of autoreactive B cells, such autoreactive clones are counterselected at two major checkpoints (1, 9). Central B cell tolerance is established in the bone marrow at the first checkpoint between the early immature and immature B cell stages, and this silences most polyreactive and anti-nuclear antibody–expressing B cells.

A second counterselection step mediates the peripheral B cell tolerance checkpoint and serves to remove autoreactive B cells that recently emigrated from the bone marrow before they enter the long-lived mature naive B cell pool (1, 9). The regulation of central B cell tolerance requires proper B cell receptor (BCR) signaling. Alterations in BCR signaling described in patients with X-linked agammaglobulinemia (XLA) result in a defective central B cell tolerance checkpoint, which is manifested by a failure to counterselect polyreactive developing B cells in these patients (10). Little is known about the molecules and pathways that regulate human peripheral B cell tolerance checkpoints. Our recent studies suggest that T cells mediate peripheral B cell tolerance through CD40/CD40 ligand and MHC class II/T cell receptor interactions (E. Meffre, personal communication).

What could account for the production of autoreactive B cells during fetal and neonatal life? The restricted V(D)J recombination in fetuses leads to a limited B cell repertoire, which has been clearly shown by Merbl et al. to contain autoreactive/poly-
Natural antibodies, polyreactive B cells, and autoimmunity

Natural antibodies are believed to have beneficial functions. They have been shown to be an essential first line of defense against pathogens through either activation of complement or formation of immune complexes that are delivered to follicular dendritic cells in secondary lymphoid organs (reviewed in ref. 12). Natural antibodies also participate in the clearance of apoptotic cells and thereby limit circulation of autoantigens and reduce the risk of autoimmunity. By extension, autoreactive antibodies encoded by the conserved V_{H}6–1 gene may help eliminate fetal apoptotic cells and debris containing DNA molecules because those antibodies have anti–nucleic acid reactivity (11). Later in life, autoreactive V_{H}6–1–expressing B cells are rare, but autoreactive clones expressing another intrinsically self-reactive V_{H}16, V_{H}13–4–34, develop and accumulate in the mature naive B cell compartment. B cells expressing antibodies encoded by V_{H}13–4–34 variable regions recognize I/i carbohydrate self determinants displayed on red blood cells, and such autoantibodies may participate in the clearance of dying erythrocytes (13). Germline-encoded V_{H}13–4–34 antibodies also cross-react with bacterial LPS, suggesting a role in host defense. Despite a potential protective role, the V_{H}13–4–34 gene encodes all pathogenic anti-1/I cold-agglutinin antibodies, indicating that natural antibodies can have damaging effects (13). Indeed, serum natural IgM anti–nonmuscle myosin heavy chain type II (anti–NMHC-II) antibodies have recently been shown to mediate local reperfusion injury after ischemia associated with myocardial infarction, stroke, or surgery (14). Intracytoplasmic NMHC-II molecules are expressed at the cell surface as a result of hypoxic stress, and binding of natural IgM antibodies that recognize these newly exposed self antigens induces complement activation and tissue damage (14).

Whether the limited and selective autoimmunity, termed the immunusculus, is protective or whether it represents the substrate on which pathogenic autoantibodies develop will depend upon the effectiveness of checkpoints following birth. Patients with SLE and rheumatoid arthritis display defective central and peripheral B cell tolerance checkpoints that lead to the accumulation of a large number of autoreactive/polyreactive B cells in the mature naive B cell compartment (9). The alteration of these autoreactive/polyreactive B cells through somatic hypermutation and isotype switching may yield clones expressing high-affinity pathogenic autoantibodies. In mice, nucleotide changes in the Ig sequence of an antibody against phosphorylcholine, the dominant hapten on the pneumococcal cell wall, gave rise to antibodies cross-reacting with DNA (15). In healthy adults, V_{H}4–34–expressing autoreactive B cells are excluded from germinal centers and the memory B cell compartment (16). However, in SLE patients, V_{H}4–34–expressing B cells are recruited in germinal centers, where they may produce pathogenic autoantibodies (16). Indeed, such serum V_{H}4–34 antibody levels correlate with lupus disease activity (17).

The events that drive breaks in B cell tolerance are unknown, but evidence is emerging that TLRs contribute to activation of autoreactive/polyreactive B cells. TLRs recognize microbial components as well as endogenous self antigens such as DNA and RNA (18). Nuclear autoantigens containing DNA and RNA molecules found in the serum of SLE patients may concomitantly trigger autoreactive/polyreactive BCRs and TLRs and thereby induce the production of autoantibodies binding double-stranded DNA and/or histones as well as nuclear ribonucleoproteins in these patients (19). The role of TLRs during fetal/neonatal B cell ontogeny is unknown. We speculate that TLR binding by self antigens released during developmental apoptosis contributes to activation of polyreactive B cells and secretion of natural antibodies in early human life.

In conclusion, fetal/neonatal autoantibodies are different from adult autoantibodies, especially those associated with autoimmune disease and organ damage. Fetal autoantibodies are IgMs with germline-encoded CDR3s that arise from restricted V(D)J recombination, whereas pathogenic autoantibodies in adults are mostly high-affinity, somatically mutated IgGs that may result from failures of counterselection processes against specific autoantigens. For these reasons, we believe that the immunusculus is protective and that defects in the maturation of the immune system later in life lead to abnormal B cell tolerance checkpoints and predispose the individual to autoimmunity.

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Inhaled environmental oxidants, such as ozone and particulates, have been variably linked to epithelial injury, inflammation, and perturbations in lung development, growth, and function. Reactions between ozone and lung surface lipids likely account for exposure-related pathophysiologic sequelae. In this issue of the JCI, Dahl et al. document a previously unrecognized pulmonary defense against inhaled oxidants in mice: macrophage scavenger receptors (SRs) bind proinflammatory oxidized lipids, thereby decreasing pulmonary inflammation (see the related article beginning on page 757). The study adds to our knowledge of diverse lung oxidative processes and identifies a potential regulatory mechanism governing pulmonary inflammation. Further investigations to elucidate more precise mechanisms and to determine the influence of SRs on airway epithelial injury, repair, and remodeling are warranted.

Scavenger receptors

The term scavenger receptor now describes a large family of proteins that feature an unusually broad ligand-binding specificity and are composed of 8 subclasses (1). The first scavenger receptor, scavenger receptor AI/II (SR-AI/II) was identified during studies of modified lipoprotein uptake in atherosclerotic plaques (2). It was soon apparent that macrophages (as well as other cell types) bear several forms of scavenger receptors that have both similar and distinct features. The macrophage class A scavenger receptors (SRAs) bind many, but not all, polyanionic molecules including acetylated or oxidized LDL, polynsinosic acid, phosphatidylserine, dextran sulfate, and components of Gram-negative and -positive bacteria (e.g., endotoxin and lipoteichoic acid, respectively), prompting one expert to characterize them as “molecular flypaper” (3).

The broad recognition capabilities of scavenger receptors are especially significant to the function of alveolar macrophages (AMs), the primary pulmonary innate immunity sentinel cell. AMs are largely responsible for binding, ingestion, and ultimately clearance of numerous inhaled macromolecules, particles, and pathogens that reach the lower respiratory tract. Previous studies have focused on two SRAs expressed on AMs: the founding member, SR-AI/II, and the more recently described macrophage receptor with collagenous structure (MARCO). SR-AI/II and MARCO knockout mice show increased susceptibility to bacterial pneumonia and more robust inflammatory responses to inhaled environmental particles (4, 5), supporting an important role for scavenger receptors in pulmonary innate defense against exogenous challenges. One can envision an evolutionary advantage for scavenger receptor broad recognition because oxidative stress (which occurs under numerous pathologic conditions) initiates lipid oxidation, producing a plethora of reaction/decomposition products, many of which show biological activities. Thus, if scavenger receptor–lipid binding contributes to bioactive species removal, some degree of nonspecificity in ligand recognition should augment efficacy across a spectrum of lung oxidative and inflammatory processes wherein lipid oxidation occurs.

Pulmonary oxidant challenge

Ozone and many other inhaled environmental oxidants, including particulates, produce diverse biologic effects that impact both acute and long-term public health. For obvious reasons, long-standing attention has centered on pulmonary sequelae such as physiologic impairments, inflammation, cell injury, and airway remodeling. Epidemiologic and laboratory studies indicate that environmentally relevant exposures may lead to abnormalities in lung growth and development. Importantly, both acute and chronic cardiovascular effects may also occur. In the respiratory tract, the epithelial lining fluid (ELF), which overlays all respiratory tract surfaces, is the first compartment that inhaled materials contact. The physicochemical properties of ozone (high reactivity, poor water solubility) constrain its diffusion through the ELF to the underlying epithelium, so that cellular perturbations likely result from products generated during extracellular (i.e., ELF) reactions (6, 7). Because the magnitude of specific biologic responses varies across species, the exact oxidant(s) involved, the dose received, and the time over which exposure occurs, determine the extent of lung injury.