The acute respiratory distress syndrome (ARDS) is an important cause of acute respiratory failure that is often associated with multiple organ failure. Several clinical disorders can precipitate ARDS, including pneumonia, sepsis, aspiration of gastric contents, and major trauma. Physiologically, ARDS is characterized by increased permeability pulmonary edema, severe arterial hypoxemia, and impaired carbon dioxide excretion. Based on both experimental and clinical studies, progress has been made in understanding the mechanisms responsible for the pathogenesis and the resolution of lung injury, including the contribution of environmental and genetic factors. Improved survival has been achieved with the use of lung-protective ventilation. Future progress will depend on developing novel therapeutics that can facilitate and enhance lung repair.
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Introduction
Since the original description of the acute respiratory distress syndrome (ARDS) in 1967, considerable progress has been made in understanding the pathogenesis and pathophysiology of acute lung injury (ALI) (1–4). The likelihood of survival is determined by the severity of lung injury, the extent of nonpulmonary organ dysfunction, preexisting medical conditions, and the quality of supportive care. Because ARDS is a complex syndrome with a broad clinical phenotype, it has been challenging to translate the results of cell and animal studies to pharmacologic therapies that reduce mortality in humans. Nevertheless, laboratory-based investigations have produced valuable insights into the mechanisms responsible for the pathogenesis and resolution of lung injury, and preclinical studies paved the way for important improvements in supportive care. Two of these therapies, lung-protective ventilation and fluid-conservative management, have reduced mortality and morbidity, respectively. This review of ARDS will focus on some of these issues, including new insights into the molecular mechanisms of lung injury and repair.

Definitions, epidemiology, incidence, and mortality
Criteria for the diagnosis of ARDS have evolved. The original description emphasized rapidly progressive respiratory failure from noncardiogenic pulmonary edema, requiring mechanical ventilation because of severe arterial hypoxemia and difficulty breathing (5). In 1988, a 4-point scoring system provided a quantitative assessment of lung injury severity based on the degree of hypoxemia, the level of positive end-expiratory pressure (PEEP), static respiratory compliance, and the extent of radiographic infiltrates (6), and this scoring system has been useful for research and clinical trials. In 1994, a consensus conference recommended simplified criteria: arterial hypoxemia with \( \text{PaO}_2/\text{FiO}_2 \) ratio less than 300 mmHg and less than 200 mmHg to define ALI and ARDS, respectively, and bilateral radiographic opacities without evidence of left atrial hypertension (7). These criteria have been widely utilized, although some investigators believe that the definitions should specify the level of PEEP and/or the fraction of inspired oxygen. A recent report — what is now called the Berlin definitions — recommends use of three categories of ARDS, based on the degree of hypoxemia: mild (200 mmHg < \( \text{PaO}_2/\text{FiO}_2 \) ≤ 300 mmHg), moderate (100 mmHg < \( \text{PaO}_2/\text{FiO}_2 \) ≤ 200 mmHg), and severe (\( \text{PaO}_2/\text{FiO}_2 \) ≤ 100 mmHg) (8). Whether stratification of patients based on these descriptions will advance the efficacy of clinical detection and of charting the natural history of ARDS remains to be determined.

Most investigations have focused on ALI and/or ARDS patients who are already mechanically ventilated. Recently, progress has been made in diagnosing ALI in spontaneously breathing patients who have bilateral pulmonary infiltrates and arterial hypoxemia and in whom intravascular volume overload and congestive heart failure are excluded (9, 10). This approach facilitates patient identification and testing of new therapies prior to the need for mechanical ventilation. Figure 1 provides a clinical vignette describing early recognition of ALI.

Bacterial or viral pneumonia is the most common cause of ALI and ARDS (1). Sepsis due to nonpulmonary infections, aspiration of gastric contents, and major trauma with shock also commonly precipitate the injury. Less commonly, acute pancreatitis, transfusions, drug reactions, and fungal and parasitic lung infections are linked to ALI and ARDS. The coexistence of two or more of these risk factors can enhance the likelihood of developing ALI or ARDS (1).

A prospective epidemiologic study in 1999–2000 estimated an annual incidence of ALI and ARDS of 190,000 adult patients in the United States (11). There is a substantial incidence of ALI and ARDS in children as well (12, 13). Data from 2001–2008 indicate that the incidence of ALI and/or ARDS in hospitalized adults has declined, perhaps secondary to more widespread use of lung-protective ventilation, reductions in nosocomial infections, and more conservative use of blood products (14, 15).

Severe arterial hypoxemia (\( \text{PaO}_2/\text{FiO}_2 < 100 \)) and an increase in the pulmonary dead space fraction (>0.60) are associated with higher mortality (16), as are shock, liver dysfunction, acute kidney injury, age over 60 years, and higher severity of illness scores (17–19). Based on the NIH Heart, Lung and Blood Institute (NHLBI) ARDS Network trials, 60-day mortality has declined from 36% in 1996–1997 to 26% in 2004–2005 (20). The most recent ARDS Network clinical trials reported a 60-day mortality of 22% in adult patients despite higher APACHE III scores and a higher incidence of shock at enrollment compared with a prior trial in 2006 (Figure 2 and ref. 4).

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Environmental and genetic influences

Environmental and genetic factors that contribute to susceptibility and severity of ALI and ARDS have emerged as a major research focus. Chronic alcohol abuse increases the risk of ALI and ARDS (21) and multiple organ failure in septic shock (22). Both active and passive cigarette smoke exposure, as quantified by plasma levels of cotinine, have been independently associated with the development of ALI after severe blunt trauma (23). The mechanisms may include priming effects of cigarette smoke exposure on the lung endothelium, alveolar epithelium, or inflammatory cells.

Variants in more than 25 genes have been associated with developing ALI and/or ARDS and with clinical outcomes (24) including common variants of genes that regulate inflammation, coagulation, endothelial cell function, reactive oxygen radical generation, and apoptosis (25–29) — all processes that are important in lung injury and repair (2, 30, 31). For example, the Fas pathway modulates apoptosis, inflammation, and epithelial cell injury; in a candidate gene study, common genetic variants in Fas were associated with susceptibility to developing clinical lung injury (27). African Americans with ALI have a higher risk of death compared with white patients. A candidate gene study identified a functional T-46C polymorphism (rs2814778) in the promoter region of the Duffy antigen/receptor for chemokines (DARC) gene that was associated with a 17% increase in 60-day mortality in African-American patients enrolled in ARDS Network clinical trials. Plasma interleukin-8 levels were increased in those individuals with the DARC polymorphism, supporting one mechanism contributing to a worse clinical outcome (29).

Genetic polymorphisms that predispose individuals to the injurious effects of specific bacteria or viruses may influence the development of ALI and ARDS. Indeed, several polymorphisms are associated with more severe pneumococcal, Legionella, and viral lung infections (32). The genetic factors that regulate the virulence of infecting pathogens also require more research (33) to relate the severity of clinical lung injury to specific microbiologic variables that contribute to severe pneumonia and ALI. Genetic features of both the host and the microbe likely are important in determining the severity of lung injury.

In spite of the intriguing associations, putative causal genes require more validation in independent study populations, along with studies of gene and environmental interactions that may alter these associations. Genome-wide association studies for susceptibility and outcomes in multiple populations are in progress. One recent study of this nature identified PPFIA1 (which encodes liprin α, a protein involved in cell adhesion, integrin expression, and cell-matrix interactions) as a predictor for developing acute lung injury after major trauma (25).

Pathogenesis: dysregulated inflammation and alveolar barrier disruption

Dysregulated inflammation, inappropriate accumulation and activity of leukocytes and platelets, uncontrolled activation of coagulation pathways, and altered permeability of alveolar endothelial and epithelial barriers remain central pathophysiologic concepts in ALI and ARDS (1–3). Activation of the innate immune response by binding of microbial products or cell injury-associated endogenous molecules (danger-associated molecular patterns [DAMPs]) to pattern recognition receptors such as the Toll-like receptors on the lung epithelium and alveolar macrophages is now recognized as a potent driving force for acute lung inflammation (34). Newly reported innate immune effector mechanisms, such as formation of neutrophil extracellular traps — lattices of chromatin and antimicrobial factors that capture pathogens but also can cause endothelial injury — and histone release by neutrophils (35) may contribute to alveolar injury. Signaling between inflammatory and hemostatic effector cells, such
as platelet-neutrophil interaction, is important in some models, including acid-induced ALI, sepsis, and transfusion injury (36, 37). The delicate balance between protective and injurious innate and adaptive immune responses and hemostatic pathways may determine whether alveolar injury continues or is repaired and resolved. For example, in lung infection, acute inflammatory responses to pathogens and their toxins (38–40) cause ALI through leukocyte protease release, generation of reactive oxygen species, rampant synthesis of chemokines and cytokines, Toll-like receptor engagement, and actions of lipid mediators (33, 41, 42). Nevertheless, these same inflammatory mechanisms, when controlled rather than excessive, are requisite in pathogen containment and clearance. Recent research suggests that other pathways, such as the molecular events that govern the balance between angiotensin converting enzymes 1 and 2, may influence the degree of inflammatory lung injury consequent to viral infection and sepsis (43, 44). Similarly, newly recognized lipid modifications may contribute to resolution of lung inflammation (45).

Increased permeability of microvascular barriers, resulting in extravascular accumulation of protein-rich edema fluid, is a cardinal feature of acute inflammation and a central pathophysiologic mechanism in ALI and ARDS (Figure 3A and refs. 1, 3, 6).

**Figure 2**
Mortality in ALI and ARDS. Shown is the 60-day mortality reported over the last 11 years in randomized clinical trials from the ARDS Network. ARMA-12 refers to the mortality rate of 431 patients enrolled into the higher–tidal volume arm (12 ml tidal volume/kg predicted body weight), and ARMA-6 refers to the mortality of 430 patients enrolled in the lower–tidal volume arm (6 ml tidal volume/kg predicted body weight) of one study (97). FACTT fluid conservative refers to the mortality of the 500 patients enrolled into the fluid-conservative arm of the Fluid and Catheter Treatment Trial (120). ALTA and OMEGA refer to the combined mortalities of the 2 most recent trials (N = 517 in both trials combined), Albuterol for the Treatment of ALI (136) and Omega-3 Fatty Acid, Gamma-Linolenic Acid, and Antioxidant Supplementation in the Management of ALI or ARDS (138).

**Figure 3**
Molecular targets for new therapies that can lead to endothelial and epithelial barrier stabilization and reversal of increased permeability. (A) Disrupted alveolar barrier function, resulting in increased permeability to water, proteins, and other solutes, is a hallmark of clinical and experimental ALI. Intra-alveolar accumulation of neutrophils, other leukocytes, and erythrocytes is also associated with altered endothelial and epithelial barrier function. TNF-α, IL-1, thrombin, and microbes and their toxins — including LPS, noxious agents, and factors generated by neutrophils and platelet-leukocyte interactions — can destabilize and disrupt alveolar barrier function, leading to increased permeability. (B) Disruption of VE-cadherin bonds is a central mechanism of altered endothelial barrier function in experimental ALI and in models of sepsis and systemic vascular destabilization. VE-cadherin is an endothelial-specific adherens junction protein that mediates Ca2+-dependent homophilic interactions at the lateral cell membranes of adjacent endothelial cells. VE-cadherin is regulated by cytoplasmic associations with catenins and actin and by cytoskeletal organization, in addition to intracellular signaling by Rho and Rac. Disruption of VE-cadherin bonds also facilitates transendothelial migration of leukocytes and, in some studies, is associated with accumulation of leukocytes and platelets in microvessels. (C) Stabilizing agonists (i) or small-molecule mimetics bind to stabilizing receptors (ii) on endothelial cells in alveolar and systemic vessels, restoring barrier integrity. Stabilizing agonists include S1P, Slit2N, Ang1, atrial natriuretic peptide, APC, and ATP; multiple intracellular pathways and mechanisms are implicated (iii) (reviewed in refs. 58, 61, 64). These intracellular mechanisms favorably influence cytoskeletal architecture, preserve catenin–VE-cadherin cytoplasmic interactions, prevent VE-cadherin internalization, and/or promote adherens junction formation (iv and v).
Thus, repair may occur by endogenous stem cell proliferation, not just by epithelial cell migration and proliferation of existing differentiated cells. A central therapeutic paradigm involves prevention of dissociation of a phosphatase from VE-cadherin (57) or genetic manipulation of VE-cadherin–catenin interactions (56) or contrast, stabilization of VE-cadherin bonds (Figure 3C) through cAMP stimulation, the rate of alveolar fluid transport increases substantially, accomplished by increased expression and activity of ENaC, NaKATPase, and opening of the CFTR. For net fluid clearance to occur, however, there needs to be a reasonably intact alveolar epithelial barrier (see C). AQPS5, aquaporin 5. (B) The resolution of inflammation in ALI and ARDS requires the removal of neutrophils from the distal airspace of the lung. Neutrophils are normally taken up by alveolar macrophages, process termed efferocytosis. The rate of neutrophil clearance can be accelerated by regulatory T lymphocytes, in part by release of TGF-β. (C) Restoration of the alveolar epithelial barrier initially occurs by reepithelialization of the epithelial surface by alveolar type II cells. Although it was previously thought that this occurred via proliferation of resident type II cells, new work suggests there may be niches of progenitor cells that also contribute. An α6β4+ progenitor cell has been identified in the mouse lung that is responsible for restoration of the alveolar epithelial barrier after bleomycin-induced lung injury (88). Thus, repair may occur by endogenous stem cell proliferation, not just by epithelial cell migration and proliferation of existing differentiated cells.

Increased permeability is also linked to transfer of leukocytes and erythrocytes into the alveolar space in ARDS (46), as well as to inflammasome-regulated cytokines (47). A variety of mediators, pathways, and molecular systems contribute to altered alveolar endothelial and epithelial permeability (48–53). Vascular endothelial cadherin (VE-cadherin), an adherens junction protein, is critical for maintenance of endothelial barrier integrity in lung microvessels (54). Disruption of VE-cadherin homophilic bonds destabilizes lung microvascular barrier function (Figure 3B). Antibodies against VE-cadherin, destabilizing agonists such as TNF, thrombin, and VEGF, and leukocyte signals all interrupt VE-cadherin bonds and induce lung edema formation (54, 55). In contrast, stabilization of VE-cadherin bonds (Figure 3C) through genetic manipulation of VE-cadherin–catenin interactions (56) or prevention of dissociation of a phosphatase from VE-cadherin (57) reduces BAL protein and leukocytes in LPS-challenged mice. Thus, experimental manipulation of VE-cadherin alters alveolar and systemic endothelial barrier function and leukocyte transmigration, with pathogenic implications for clinical ALI and ARDS.

Molecular approaches to specifically reverse increased-permeability pulmonary edema have been long sought in ALI and ARDS research (1, 3). Candidate pathways for stabilization of lung and systemic endothelial barriers (reviewed in ref. 58) have recently been described. Systemic endothelium may be a critical therapeutic target in septic ALI (58) and in multiple organ failure associated with ALI and ARDS (1). A central therapeutic paradigm involves administration of stabilizing ligands that bind to receptors on endothelial cells and activate intracellular pathways, mediating cytoskeletal reorganization and catenin–VE-cadherin interactions that tighten VE-cadherin bonds (Figure 3C). Sphingosine-1-phosphate (S1P) is a lipid recognized by G protein–coupled receptors on endothelial cells (S1P1, S1P2, S1P3). S1P binding to S1P1 induces actin cytoskeletal reorganization, RAC activation, localization of α-, β-, and γ-catenin and VE-cadherin to regions of intercellular contact, and assembly of adherens junctions in cultured human endothelial cells (59, 60). S1P enhances pulmonary and systemic endothelial barrier integrity in vivo and in vitro (reviewed in ref. 61), and small-molecule agonists of endothelial S1P1 suppress cytokine storm and lung leukocyte recruitment in experimental influenza (62). S1P is present in high concentrations in plasma and in that compartment regulates basal and inflammation-triggered vascular leak in the lungs and systemic vessels of mice (63). Platelets may locally contribute S1P at sites of vascular injury (60) and may reduce alveolar hemorrhage — another complication of endothelial barrier disruptions (46) — under some conditions; this effect may in part be related to delivery of S1P. However, effects of S1P or synthetic S1P receptor agonists may depend on biologic context (60) and time/duration of administration (64), since S1P1 and S1P3 are barrier destabilizing and S1P1 undergoes time-dependent desensitization (60, 63). Similarly, other receptor-mediated signaling systems, such as those recognized by thrombin and other G protein–coupled protease-activated receptor agonists, may differentially trigger lung endothelial barrier disruption or stabilization, depending on time and context (65).

The Robo4/Slit signaling system also stabilizes the endothelial barrier (66, 67). In contrast to S1P receptors (60), Robo4 expression is restricted to endothelial cells (68). An active fragment of the Robo4 ligand Slit (Slit2N) inhibits tyrosine phosphorylation of VE-cadherin and preserves its association with P120 catenin, preventing VE-cadherin internalization and abnormal permeability of human microvascular endothelial cells induced by TNF-α, IL-1, or LPS (67). In mice, Slit2N reduces pulmonary and systemic vascular perme-
ability in LPS lung injury, cecal ligation and puncture, and influenza infection, increasing survival (67). Cytokine levels are not decreased, indicating that Robo4 signaling does not inhibit this component of inflammation and that endothelial barrier stabilization may be sufficient to improve outcomes in lethal infectious challenges (58, 67).

Although numerous endothelial-stabilizing agonists and intracellular pathways have been identified (refs. 58, 61, and Figure 3C), use of these agents may have unintended consequences. Molecular mechanisms by which plasminogen activator inhibitor-1 mediates *Pseudomonas*-associated alveolar endothelial barrier disruption have been identified, but *Patl1*−/− mice have a defect in alveolar neutrophil recruitment and increased mortality compared with wild-type animals (69). This emphasizes the daedal relationships between barrier integrity and leukocyte transmigration and the precarious tension between injurious and protective inflammatory mechanisms that may operate in ALI and ARDS.

In contrast to endothelium, less is known about the potential mechanisms of alveolar epithelial stabilization, although epithelial permeability is critical in alveolar flooding (70, 71) and leukocyte accumulation (52) and potentially critical for intra-alveolar fibrin deposition and hyaline membrane formation (72). Epithelial barriers involve cadherin-mediated adherens junction bonds and tight junctions, although the topography differs from endothelium, and E-cadherin substitutes for VE-cadherin (54). Alveolar epithelial barriers are tighter than epithelial barriers (70), but the two have functional interactions (2). For example, under some (55), but not all (57), conditions, disruption of epithelial VE-cadherin bonds causes alveolar epithelial leak and epithelial injury. Mesenchymal stem cells (MSCs) (see Future directions) restore barrier integrity in cytokine-treated cultured human alveolar epithelial cells (73). The mechanism involves release of angiopoietin-1, which inhibits its actin stress fiber formation and redistribution of the tight junction protein claudin 18 in epithelial cells (73) and also induces S1P production and inhibits endothelial VE-cadherin internalization (58).

Resolution
Resolution of ALI requires effective and synchronous (a) resorption of alveolar edema, (b) repair of the epithelial and endothelial barriers, and (c) removal of inflammatory cells and exudate from the distal airspaces (Figure 4). Resolution of ALI

Table 1
Selected clinical trials of ALI and ARDS

<table>
<thead>
<tr>
<th>Intervention</th>
<th>Reference</th>
<th>Study phase</th>
<th>Study population a</th>
<th>Results</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lung-protective ventilation</td>
<td>96</td>
<td>Phase III</td>
<td>ARDS (<em>N = 53</em>)</td>
<td>Decrease in mortality</td>
</tr>
<tr>
<td>Lung-protective ventilation</td>
<td>97</td>
<td>Phase III</td>
<td>ARDS (<em>N = 861</em>)</td>
<td>Decrease in mortality</td>
</tr>
<tr>
<td>Lung-protective ventilation</td>
<td>98</td>
<td>Phase III</td>
<td>ARDS (<em>N = 103</em>)</td>
<td>Decrease in mortality</td>
</tr>
<tr>
<td>High PEEP</td>
<td>108</td>
<td>Phase III</td>
<td>ARDS (<em>N = 549</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>High PEEP</td>
<td>109</td>
<td>Phase III</td>
<td>ARDS (<em>N = 385</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>High PEEP</td>
<td>110</td>
<td>Phase III</td>
<td>ARDS (<em>N = 382</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>High-frequency ventilation</td>
<td>116</td>
<td>Phase II</td>
<td>ARDS (<em>N = 148</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>Prone position</td>
<td>111</td>
<td>Phase III</td>
<td>ARDS (<em>N = 102</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>Prone position</td>
<td>112</td>
<td>Phase III</td>
<td>ARDS (<em>N = 342</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>Neurouromuscular blockade</td>
<td>113</td>
<td>Phase III</td>
<td>ARDS (<em>N = 340</em>)</td>
<td>Decrease in mortality</td>
</tr>
<tr>
<td>Esophageal pressure to adjust PEEP</td>
<td>114</td>
<td>Phase II</td>
<td>ARDS (<em>N = 61</em>)</td>
<td>Improved oxygenation</td>
</tr>
<tr>
<td>Surfactant</td>
<td>125</td>
<td>Phase III</td>
<td>ARDS (<em>N = 448</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>Methylprednisolone</td>
<td>126</td>
<td>Phase III</td>
<td>ARDS (<em>N = 99</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>Methylprednisolone</td>
<td>127</td>
<td>Phase III</td>
<td>ARDS (<em>n = 24</em>)</td>
<td>Decrease in mortality, but small study</td>
</tr>
<tr>
<td>Methylprednisolone</td>
<td>128</td>
<td>Phase III</td>
<td>ARDS (<em>n = 180</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>Methylprednisolone</td>
<td>129</td>
<td>Phase III</td>
<td>ARDS (<em>N = 91</em>)</td>
<td>Reduction in duration of mechanical ventilation, but major limitations related to study design</td>
</tr>
<tr>
<td>Liposomal prostaglandin E1</td>
<td>130</td>
<td>Phase III</td>
<td>ARDS (<em>N = 350</em>)</td>
<td>No difference in mortality for results</td>
</tr>
<tr>
<td>Antioxidants</td>
<td>132</td>
<td>Phase II</td>
<td>ARDS (<em>N = 46</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>Nitric oxide</td>
<td>135</td>
<td>Phase III</td>
<td>ARDS (<em>N = 385</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>β2-Agonist (aerosolized)</td>
<td>136</td>
<td>Phase III</td>
<td>ARDS (<em>N = 282</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>β2-Agonist (intravenous)</td>
<td>137</td>
<td>Phase III</td>
<td>ARDS (<em>N = 330</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>ω-3 Fatty acid supplement</td>
<td>138</td>
<td>Phase III</td>
<td>ARDS (<em>N = 272</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>Pulmonary artery versus central venous catheter</td>
<td>121</td>
<td>Phase III</td>
<td>ARDS (<em>N = 1,000</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>Fluid-conservative versus fluid-liberal therapy</td>
<td>120</td>
<td>Phase III</td>
<td>ARDS (<em>N = 1,000</em>)</td>
<td>More ventilator-free days with fluid-conservative therapy</td>
</tr>
<tr>
<td>Extracorporeal membrane oxygenation</td>
<td>115</td>
<td>Phase III</td>
<td>ARDS (<em>N = 90</em>)</td>
<td>Decrease in mortality, but results not conclusive</td>
</tr>
<tr>
<td>APC</td>
<td>134</td>
<td>Phase III</td>
<td>Nonseptic ARDS (<em>N = 75</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>APC</td>
<td>133</td>
<td>Phase III</td>
<td>Sepsis (<em>N = 1,697</em>)</td>
<td>No difference in mortality</td>
</tr>
<tr>
<td>GM-CSF</td>
<td>131</td>
<td>Phase II</td>
<td>ARDS (<em>N = 130</em>)</td>
<td>No difference in mortality</td>
</tr>
</tbody>
</table>

aPhysiologic criteria for ALI and ARDS varied among these trials.
Mechanisms of ventilator-associated lung injury (VALI). (A) ALI leads to lung endothelial and epithelial injury, increased permeability of the alveolar-capillary barrier, flooding of the airspace with protein-rich pulmonary edema fluid, activation of alveolar macrophages with release of proinflammatory chemokines and cytokines, enhanced neutrophil migration and activation, and fibrin deposition (hyaline membranes). (B) If the injured lung is ventilated with high tidal volumes and high inflation pressures (high-stretch ventilation), then lung injury is exacerbated, with increased lung endothelial and epithelial injury and/or necrosis, enhanced neutrophil margination, release of injurious neutrophil products such as proteases and oxidants, increased release of proinflammatory cytokines from alveolar macrophages and the lung epithelium, increased fibrin deposition, and increased hyaline membrane formation. Injurious mechanical ventilation can also impair alveolar fluid clearance (AFC) mechanisms. (C) In contrast, a protective ventilatory strategy (low-stretch ventilation) can limit further lung endothelial and epithelial injury, reduce the release of proinflammatory cytokines, and enhance alveolar fluid clearance through the active transport of sodium and chloride across the alveolar epithelium (see Figure 4), thereby reducing the quantity of pulmonary edema and allowing endothelial and epithelial repair to occur. Epithelial repair occurs through migration, proliferation, and differentiation of alveolar epithelial type II cells to repopulate the denuded basement membrane. Acute inflammation resolves through apoptosis of neutrophils, which are phagocytosed by alveolar macrophages (see Figure 4).
and repair of alveolar structures may — like the initiation of alveolar damage — depend on a precise balance of inflammatory interactions and molecular signaling. For example, hyaluronan fragments found in the serum of ALI patients trigger release of chemokines by macrophages but also interact with Toll-like receptors to deliver signals that limit epithelial apoptosis and promote reestablishment of epithelial integrity in experimental lung injury (74).

Reabsorption of alveolar edema (Figure 4A) occurs through vectorial transport of sodium and chloride across alveolar epithelial type I and II cells to create a mini-osmotic gradient to reabsorb water (75–77). This process is impaired in ALI and ARDS because of apoptosis and necrosis of alveolar epithelium (31, 46) and defects in transcellular ion transport induced by proinflammatory cytokines, oxidants, and hypoxia (75, 78–81). Effective reabsorption of edema fluid from the air spaces requires reestablishment of an effective epithelial barrier (70, 82). Reepithelialization occurs initially by proliferation of type II cells (46), and the traditional view is that type II cells are the main source of new alveolar epithelial cells (83). Nevertheless, there is new evidence that progenitor cells may exist in strategic niches in bronchoalveolar junctions and that the alveolar epithelium can be activated to regenerate the epithelial and endothelial barriers (84–87). For example, α6β4-expressing progenitor cells have been identified that account for a substantial fraction of the type II cells that reepithelialize the injured alveolar barrier (Figure 4C and ref. 88). There may also be a human c-kit+ lung stem cell capable of renewing all lung cells (89), although this hypothesis is controversial and requires validation. There has been progress in understanding how inflammation is resolved through clearance of alveolar neutrophils, monocytes, and necrotic debris including the contributions of lipid mediators such as lipoxin A4, resolvin E1, and other antiinflammatory pathways (Figure 4B and refs. 45, 90). Macrophages remove apoptotic neutrophils and monocytes via molecular mechanisms that have recently been more clearly identified (91–93). A deficiency of alveolar macrophages worsens influenza-related pneumonia and lung injury in mice, leading to an increase in neutrophils and neutrophil extracellular traps (94). Lymphocytes also contribute to resolution. In a mouse model of endotoxin-induced lung injury, CD4+CD25+ regulatory T cells that were recruited to the air spaces played an essential role in resolving inflammation by enhancing neutrophil apoptosis and suppressing cytokine secretion, in part by release of TGF-β (95).

New insights based on clinical trials

Many clinical trials have assessed pharmacologic interventions, innovative strategies for positive-pressure ventilation, and other supportive approaches to ALI and ARDS treatment, advancing our understanding of the mechanism of the disease (Table 1).

Lung-protective ventilation. Three randomized clinical trials demonstrated that lung-protective ventilation with lower tidal volumes and airway pressures reduces mortality in ALI and ARDS (96–98). Why did this simple change in how we ventilate patients have such an impressive impact?

Lung-protective ventilation decreases accumulation of pulmonary edema by preserving barrier properties of the alveolar endothelium and alveolar epithelium (99–102). In rats, the rate of resolution of alveolar edema was 3-fold higher with 6 versus 12 ml/kg tidal volume, in part because of reduced epithelial cell injury (103). Reductions in markers of lung epithelial injury have also been observed in clinical studies of reduced ventilation tidal volume (104). Lung-protective ventilation also downregulates mechanosensitive proinflammatory pathways, resulting in reduced neutrophil accumulation in the alveoli and lower plasma levels of IL-6, IL-8, and soluble TNF receptor 1 (refs. 105, 106, and Figure 5).

Lung-protective ventilation also benefits nonpulmonary organ function. For example, ventilation with high tidal volume in rabbits with acid-induced ALI causes apoptosis of renal tubular cells, an effect that is attenuated with lower tidal volume (107), and lower tidal volume ventilation was associated with an increase in renal failure–free days in patients with ARDS (97).

The effects of prone positioning, optimal levels of PEEP, high-frequency ventilation, neuromuscular blockade, measurement of esophageal pressure, and extracorporeal circuits to enhance carbon dioxide excretion have also been tested in clinical trials (refs. 108–116 and Table 1). These approaches have value as rescue therapies but have not achieved sufficient efficacy to recommend them as primary treatments (117).

Fluid-conservative therapy. Approximately 30 years ago, it was shown experimentally that reducing lung vascular hydrostatic pressures decreases lung edema in the setting of increased lung vascular permeability (118, 119). The clinical importance of this observation was confirmed when the ARDS Network reported in a 1,000-patient randomized clinical trial that a fluid-conservative strategy significantly reduced the average duration of mechanical ventilation by 2.5 days (120), a difference that was not affected by use of the pulmonary artery catheter (121). The primary beneficial mechanism can be explained by a favorable effect on Starling forces: lower vascular pressure reduces transvascular fluid filtration, particularly in the presence of increased lung vascular permeability. Also, animal studies indicate that reduced lung vascular pressure can attenuate lung endothelial translocation of P-selectin and accumulation of intravascular neutrophils (122). Plasma levels of angiopoietin-2 were lower in patients who were treated with a fluid-conservative versus a fluid-liberal strategy, which supports the hypothesis that an antiinflammatory mechanism explains this effect (123).

Other trials. Abnormalities of surfactant in ALI and ARDS include decreased production, alterations in phospholipid composition, and inhibition of surfactant function by exuded plasma proteins, oxygen radicals, and proteases (124). All these abnormalities promote atelectasis. Nevertheless, unlike the success of surfactant-replacement therapy for infant respiratory distress syndrome, clinical trials of surfactant replacement have not improved clinical outcomes in ALI and ARDS (ref. 125 and Table 1).

Although some animal studies support the potential efficacy of antiinflammatory therapies for decreasing lung injury, clinical trials have not demonstrated a convincing reduction in mortality using GM-CSF or glucocorticoids, antioxidants, or anticytokine therapies that were tested in patients with sepsis (126–132). Explanations for these outcomes are likely multifactorial, and these therapeutic strategies may merit reexamination as our understanding of subpopulations of patients with ALI and ARDS grows.

There has been considerable interest in the possibility that anticoagulant therapy may be effective in ALI and ARDS because of the close link between procoagulant and proinflammatory pathways (133, 134). Nevertheless, activated protein C (APC) did not reduce mortality in a small trial in nonseptic ALI and ARDS (134). Furthermore, a recent trial of APC has provided evidence that this anticoagulant and antiinflammatory agent does not have efficacy
for patients with severe sepsis (133), the most lethal cause of ALI and ARDS, in spite of previous clinical and experimental support for its use (3). Strategies and rationale for anticoagulants for ALI and ARDS will now need to be reevaluated. Although pulmonary hypertension and lung vascular injury are important features of ALI and ARDS, vasodilator therapies including prostaglandin E1 and nitric oxide have not reduced mortality (refs. 130, 135, and Table 1). Treatment to accelerate the resolution of pulmonary edema with aerosolized or intravenous β-adrenergic agonists also failed to improve survival (136, 137). Nutritional supplement with ω-3 fatty acid may be harmful (138).

Future directions

Cell-based therapy with allogeneic human MSCs has emerged as a promising approach to therapy for ALI and ARDS (139–144). MSCs secrete multiple effector molecules, including antiinflammatory cytokines, growth factors, and antimicrobial peptides. These can reverse the major abnormalities of lung injury, including altered lung endothelial and epithelial permeability, impaired alveolar edema fluid clearance, dysregulated inflammation, and infection. In addition, in an experimental model, MSCs attached to the alveolar wall by connexin-43–based gap junctional channels and transferred mitochondria to endothotin-injured alveolar epithelium, restoring alveolar ATP production and normalizing surfactant production and epithelial barrier properties (145). Also, the discovery that short-term function that can be achieved in repopulated decellularrized lungs raises the prospect that basic understanding of lung regeneration (146), including signals from the lung matrix, may lead to novel strategies to enhance repair in the severely injured lung.

With declining mortality, clinical trials of new therapies will need to enroll patients with the highest predicted mortality (4) and include approaches that combine clinical and biological predictors (147, 148) to select the patients at highest risk. Clinical trials are in progress to test whether statin therapy can favorably modulate inflammation and restore barrier integrity in ALI and ARDS (Figure 3C). In animals, aspirin was effective in preventing acid-induced and transfusion-associated lung injury (37). Consequently, a clinical trial is planned to test the efficacy of aspirin in patients at high risk for developing ALI. There is also progress on developing more effective extracorporeal systems to support gas exchange for patients with very severe lung injury (149). Further research into the complex factors that influence the functional status of patients after recovery from ALI or ARDS (150) may also stimulate new approaches to attenuate long-term muscle and neurologic impairments that cause disabilities in ALI and ARDS survivors.

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