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Inner ear hair cells convert the mechanical stimuli of sound, gravity, and head movement into electrical signals. This mechanotransduction process is initiated by opening of cation channels near the tips of hair cell stereocilia. Since the identity of these ion channels is unknown, and mutations in the gene encoding transmembrane channel–like 1 (TMC1) cause hearing loss without vestibular dysfunction in both mice and humans, we investigated the contribution of Tmc1 and the closely related Tmc2 to mechanotransduction in mice. We found that Tmc1 and Tmc2 were expressed in mouse vestibular and cochlear hair cells and that GFP-tagged TMC proteins localized near stereocilia tips. Tmc2 expression was transient in early postnatal mouse cochlear hair cells but persisted in vestibular hair cells. While mice with a targeted deletion of Tmc1 (Tmc1Δ mice) were deaf and those with a deletion of Tmc2 (Tmc2Δ mice) were phenotypically normal, Tmc1Δ Tmc2Δ mice had profound vestibular dysfunction, deafness, and structurally normal hair cells that lacked all mechanotransduction activity. Expression of either exogenous TMC1 or TMC2 rescued mechanotransduction in Tmc1Δ Tmc2Δ mutant hair cells. Our results indicate that TMC1 and TMC2 are necessary for hair cell mechanotransduction and may be integral components of the mechanotransduction complex. Our data also suggest that persistent TMC2 expression in vestibular hair cells may preserve vestibular function in humans with hearing loss caused by […]

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Inner ear hair cells convert the mechanical stimuli of sound, gravity, and head movement into electrical signals. This mechanotransduction process is initiated by opening of cation channels near the tips of hair cell stereocilia. Since the identity of these ion channels is unknown, and mutations in the gene encoding transmembrane channel–like 1 (TMC1) cause hearing loss without vestibular dysfunction in both mice and humans, we investigated the contribution of Tmc1 and the closely related Tmc2 to mechanotransduction in mice. We found that Tmc1 and Tmc2 were expressed in mouse vestibular and cochlear hair cells and that GFP-tagged TMC proteins localized near stereocilia tips. Tmc2 expression was transient in early postnatal mouse cochlear hair cells but persisted in vestibular hair cells. While mice with a targeted deletion of Tmc1 (Tmc1Δ mice) were deaf and those with a deletion of Tmc2 (Tmc2Δ mice) were phenotypically normal, Tmc1ΔTmc2Δ mice had profound vestibular dysfunction, deafness, and structurally normal hair cells that lacked all mechanotransduction activity. Expression of either exogenous TMC1 or TMC2 rescued mechanotransduction in Tmc1ΔTmc2Δ mutant hair cells. Our results indicate that TMC1 and TMC2 are necessary for hair cell mechanotransduction and may be integral components of the mechanotransduction complex. Our data also suggest that persistent TMC2 expression in vestibular hair cells may preserve vestibular function in humans with hearing loss caused by TMC1 mutations.

Introduction
Dominant and recessive mutations of human transmembrane channel-like 1 (TMC1) cause DFNA36 hearing loss and DFNB7/B11 deafness, respectively (1). Similarly, semidominant and recessive alleles of Tmc1 cause hearing loss in Beethoven (Bth) and deafness (dn) mutant mice (1, 2). TMC1 is a member of the novel TMC gene family that includes 7 other paralogs in mammals (3, 4). The function of TMC genes remains obscure, and the in silico translation products have no significant sequence similarity to proteins or domains of known function; however, all are predicted to encode integral membrane proteins with at least 6 membrane-spanning domains (3, 4). This 6-pass transmembrane topology has been experimentally confirmed for mouse TMC1 expressed in heterologous systems (5) and suggests it may function as a receptor, pump, transporter, or channel (3, 4).

In mouse, Tmc1 and the closely related Tmc2 are expressed at low levels in a restricted tissue distribution (1–3). Tmc1 mRNA is expressed in auditory and vestibular hair cells (1, 2), but there is no evidence of vestibular dysfunction associated with TMC1 or Tmc1 mutations (1, 2, 6–8). Tmc2 mRNA is also detectable in mouse inner ear (1), but neither TMC2 nor Tmc2 has been implicated in inner ear dysfunction. These observations could reflect incomplete penetrance of mutant phenotypes, functional redundancy of Tmc1 and Tmc2, differential requirements for Tmc1 versus Tmc2 in auditory and vestibular hair cells, or a combination of these mechanisms.

Marcotti et al. (9) investigated the function of Tmc1 in auditory hair cells excised from Tmc1ΔΔ and Tmc1+/Δ mice and suggested that Tmc1 is necessary for hair cell maturation but is not required for mechanotransduction, the process by which hair cells convert mechanical stimuli into electrical signals. Mechanotransduction is initiated near the tips of stereocilia (10–14) on the apical surface of hair cells and is mediated by an unidentified ion channel gated by mechanical force (15). Hair cell mechanotransduction is characterized by large single-channel conductances of approximately 100 pS (16–18), rapid activation with sub-millisecond kinetics (19, 20), and nonsynaptic permeability to cations (19) and other larger molecules (21) including the fluorescent dye FM1-43 (22, 23) and aminoglycoside antibiotics (24). There are several known pharmacologic agents that inhibit hair cell mechanotransduction, but a selective, high-affinity antagonist has not been identified (25). The difficulty in identifying the hair cell mechanotransduction channel at the molecular level could arise from functional pleiotropy, functional redundancy, or both (26). Here we provide evidence supporting the novel hypothesis that TMC1 and TMC2 are functionally redundant stereocilia components necessary for hair cell mechanotransduction.

Authorship note: Yoshiyuki Kawashima, Gwenaëlle S.G. Géléoc, and Kiyoto Kurima are co-first authors. Jeffrey R. Holt and Andrew J. Griffith are co-senior authors.

Conflict of interest: Charles C. Della Santina owns equity in Labyrinth Devices LLC. Kiyoto Kurima and Andrew J. Griffith hold the following U.S. patents: 7,166,433 (Transducin-2 and Applications to Hereditary Deafness), 7,192,705 (Transducin-1 and Applications to Hereditary Deafness), and 7,659,115 (Nucleic Acid Encoding Human Transducin-1 Polypeptide).

Citation for this article: J Clin Invest. 2011;121(12):4796–4809. doi:10.1172/JCI60405.
Results

Tmc1 and Tmc2 expression. Mouse utricular maculae and cochleae express two splice isoforms of Tmc1, Tmc1ex1 and Tmc1ex2, which exclude or include exon 2 and encode different translation initiation codons in exons 1 and 2, respectively (Supplemental Figure 1A; supplemental material available online with this article; doi:10.1172/JCI60405DS1). Levels of Tmc1ex1 and Tmc1ex2 RNA were similar to each other in either utricular maculae or cochleae excised at P0, P5, or P14 (data not shown).

We performed quantitative RT-PCR (qPCR) analysis of Tmc1 and Tmc2 RNA from mouse utricular maculae and cochleae harvested at different embryonic and postnatal time points (Figure 1A and B). The primer pair used for Tmc1 qPCR analysis amplified both Tmc1ex1 and Tmc1ex2. In utricular maculae, a rise in Tmc2 expression (Figure 1A) coincided with the onset of mechanotransduction between E16 and E17 (27). The rise in Tmc1 expression (Figure 1A) lagged mechanotransduction acquisition by several days. The levels of Tmc1 and Tmc2 mRNA remained elevated at P21, the last time point tested.

In cochleae, within 5 days after birth, Tmc1 and Tmc2 mRNA levels increased 20-fold and 40-fold, respectively (Figure 1B). The rise in Tmc2 expression (Figure 1B) coincided with the spatiotemporal onset of mechanotransduction in outer hair cells (OHCs) in the basal turn at P0 and the apical turn at P2 (28). The rise in Tmc1 expression (Figure 1B) lagged mechanotransduction acquisition by 2–3 days in all regions of the cochlea (28). Tmc2 mRNA was transiently expressed during the first postnatal week, while the Tmc1 transcript level increased more than 200-fold by P22. The transient Tmc2 expression profile was confirmed in a second independent sample of cochleae harvested from mice that ranged in age from E17 to P8 (Supplemental Figure 1B).

We also performed in situ hybridization analysis of Tmc2 to identify which specific cell type(s) expressed Tmc2 mRNA in P1 mouse vestibule and cochlea. Tmc2 expression was detected in wild-type crista ampullaris (CA), utricular macula (UM), hair cells of the basal cochlear turn, and, to a lesser extent, in the middle turn at P1. Tmc2 expression was not detected in hair cells of the apical turn. Adjacent sections probed for Myo15 transcripts are shown as a control for hair cell hybridization (52). Scale bar: 100 μm. See also Supplemental Figure 1.
**Tmc1^Δ** and **Tmc2^Δ** mice. To generate the Tmc1^Δ and Tmc2^Δ lines, we used homologous recombination in ES cells to delete exon(s) encoding the predicted first transmembrane domains of Tmc1 and Tmc2, respectively (Figure 2). The exons were replaced with a gene trap construct fused to a lacZ reporter gene to visualize mRNA expression from the endogenous Tmc1 or Tmc2 promoter (Figure 2, A and B). Both lines were backcrossed to C57BL/6J for 10 or more generations. X-gal staining of Tmc1^Δ/Δ P28 vestibular end organs detected β-galactosidase activity in both type I and type II hair cells of the saccular macula, utricular macula, and cristae ampullaris (Supplemental Figure 2, A and B). In Tmc2^Δ/Δ P28 inner ears, β-galactosidase activity was detected in hair cells of the cristae ampullaris (Supplemental Figure 2A) and was barely detectable in hair cells of the saccular and utricular maculae. To characterize transcripts encoded by the targeted Tmc2 locus, we performed RT-PCR analysis of Tmc2Δ RNA. We identified a Tmc2Δ transcript in which a portion of the gene trap cassette was spliced into the flanking exons (6 and 8) of Tmc2 (Supplemental Figure 2, C and D). This transcript lacked the lacZ reporter sequence, which may account for the low levels of β-galactosidase activity in Tmc2Δ hair cells.

Tmc1^Δ and Tmc2^Δ hearing and balance phenotypes. Tmc1^Δ mice developed normally, but Tmc1^Δ/Δ mice were deaf (Supplemental Figure 3) and exhibited normal motor behavior. Both Tmc2^Δ/Δ and Tmc2^Δ/Δ mice developed normally without hearing loss (Supplemental Figure 3) and showed normal motor behavior. To detect potential genetic interactions of Tmc1 with Tmc2, we crossed the Tmc1^Δ and Tmc2^Δ lines to generate mice with each of 8 different Tmc1/Tmc2 genotypes. All of the mutant mice were viable; however, Tmc1^Δ/ΔTmc2^Δ/Δ mice exhibited overly abnormal vestibular behaviors: head-bobbing and arching, ataxic gait, and circling (Supplemental Video 1). Moreover, early postnatal Tmc1^Δ/ΔTmc2^Δ/Δ pups lacked the ability to right from a supine position (Supplemental Video 1).

We employed vestibulo-ocular reflex (VOR) testing during high-acceleration, whole-body rotations in darkness to evaluate hair cell mechano-sensitivity in the crista ampullaris of the lateral semicircular canal. VOR testing of Tmc1^Δ/ΔTmc2^Δ/Δ mice at 2–3 months of age showed negligible acceleration gain values that were indistinguishable from those of dead mouse controls (Figure 3, A and B). In contrast, Tmc1^Δ/ΔTmc2^Δ/Δ mice had normal acceleration gain values. Tmc1 appears to modify the Tmc2Δ/Δ VOR phenotype, since Tmc1^Δ/ΔTmc2^Δ/Δ mice exhibited intermediate values that were significantly higher (P = 0.0059, Student’s t test) than those of Tmc1^Δ/ΔTmc2^Δ/Δ mice and lower (P = 0.0075) than those of wild-type mice. The mean acceleration gain of Tmc1^+/ΔTmc2^Δ/Δ mice was also higher (P = 0.0030) than that of Tmc1^Δ/ΔTmc2^Δ/Δ mice, but not significantly lower (P = 0.1416) than that of wild-type mice. This genetic interaction of Tmc1 with Tmc2 could reflect redundancy of their function(s) in the vestibular system.

Auditory brainstem response (ABR) thresholds measured in young adult mice showed that only Tmc1^Δ/Δ mice had hearing loss, with no effect of the Tmc2 genotype at any frequency tested (Figure 3C). The profound deafness in mice lacking functional Tmc1 was consistent with data from Tmc1^Δ/Δ/Δ and Tmc1^−/− mice (8, 29). This indicates that Tmc1, but not Tmc2, is necessary for normal auditory function in adult mice.

**Tmc1^Δ** and **Tmc2^Δ** hair cell structure. We analyzed vestibular and cochlear hair bundle morphology using scanning electron microscopy (SEM). Both vestibular and cochlear Tmc1^Δ/ΔTmc2^Δ/Δ hair cells appeared to have intact hair bundles, tip links, and tenting of tips of shorter stereocilia that were indistinguishable from those of wild-type controls at P3 (Figure 4), when wild-type hair cells have acquired functional mechanotransduction (27, 28). Furthermore, vestibular hair bundles appeared normal through 4 months of age (Supplemental Figure 4A). In cochlear, degeneration of Tmc1^Δ/ΔTmc2^Δ/Δ OHC bundles was first apparent at P5 and P7 in the basal and apical cochlear turns, respectively, while IHC bundles appeared normal until approximately P9 (Supplemental Figure 4B). In contrast, Tmc1^Δ/ΔTmc2^−/− IHC and OHC bundles developed normally until P14 and then degenerated (Supplemental Figure 4C), consistent with reported observations of
Tmc1Δ/Δ and Tmc2Δ/Δ hair cell function. To examine mechanosensory function in vestibular hair cells of Tmc1;Tmc2 mutant mice, we recorded mechanotransduction currents from P1–P8 hair cells excised from the utricle (17, 30), an organ of balance sensitive to linear acceleration. To avoid using hair bundles damaged during dissection, we selected only hair cells with visibly intact stereocilia for analysis. Utricle hair cells that lacked Tmc1 had normal transduction currents (Figure 5, A and B), consistent with the lack of behavioral and VOR phenotypes in those mice. Hair cells that lacked Tmc2, however, had significantly attenuated transduction currents, reduced more than 80% relative to wild-type controls. In hair cells that lacked both Tmc1 and Tmc2, mechanotransduction currents were completely absent. Hair cells with intermediate genotypes, Tmc1Δ/Δ-Tmc2Δ/Δ and Tmc1Δ/+Tmc2Δ/Δ, had reduced current amplitudes relative to wild-type controls (P < 0.001, 1-way ANOVA) (Figure 5B). One or two wild-type alleles of Tmc2 yielded approximately 70% or 100% of wild-type mechanotransduction current amplitudes, respectively. However, one or two wild-type alleles of Tmc1 yielded no more than 15% of wild-type mechanotransduction current amplitudes. Data collected from 111 type II utricle hair cells are summarized in Figure 5B. In hair cells with intermediate genotypes, we analyzed the 10%–90% operating range of stimulus-response relationships, but found no significant difference relative to wild-type controls, indicating that the sensitivity of mechanotransduction was unaltered by deletion of Tmc1 or Tmc2. Marcotti et al. (9) reported that auditory hair cells of mice that carried either the dn or Bb mutation in Tmc1 had reduced voltage-dependent currents at adolescent and adult stages (<P14) but normal current amplitudes at earlier postnatal stages (<P7). We examined voltage-dependent currents from type II vestibular hair cells of Tmc1;Tmc2 mutant mice at postnatal stages (P5–P7). Consistent with the results of Marcotti et al. (9), we found no difference in current amplitudes, kinetics, or voltage dependence for any of the Tmc1;Tmc2 genotypes examined (Supplemental Figure 5). Thus, the only electrophysiological deficit we detected in Tmc1;Tmc2-deficient utricle hair cells was the absent or reduced mechanotransduction current.

In cochleae, we recorded whole-cell mechanotransduction currents from apical-turn OHCs at P5–P7 as previously described (28, 31). OHCs from mice with at least one functional allele of Tmc1 had mechanotransduction currents that were indistinguishable from those of wild-type controls, regardless of the Tmc2 genotype (Figure 6, A and B). In contrast to a previous report on Tmc1Δ/Δ-Tmc2Δ/Δ and Tmc1Δ/Δ-Tmc2+/+ hair cells (9), apical-turn OHCs that lacked both alleles of Tmc1 had transduction currents that were significantly reduced relative to those of wild-type hair cells (P < 0.001, 1-way ANOVA). Double homozygous mutant hair cells with neither Tmc1 nor Tmc2 lacked mechanotransduction currents entirely (Figure 6, A and B). The Tmc1Δ/Δ-Tmc2Δ/Δ apical-turn OHCs had normal voltage-dependent currents at P5–P7, consistent with the Marcotti et al. (9) data for the equivalent postnatal day. Therefore, the absence of mechanotransduction currents in Tmc1Δ/Δ-Tmc2Δ/Δ apical-turn OHCs was a specific physiologic deficit.

We also recorded mechanotransduction currents from OHCs excised from the basal end of P2–P4 cochleae. Interestingly, Tmc1Δ/Δ-Tmc2Δ/Δ basal-turn OHCs had amplitudes similar to those of wild-type OHCs (Figure 6, C and D). This result contrasted with data recorded from apical cochleae at P5–P7 that revealed a significant reduction in current amplitude in Tmc1Δ/Δ-Tmc2Δ/Δ cells. Together with the relatively high expression of Tmc2 and relatively low expression of Tmc1 in the wild-type cochlear base at P2, the data suggest that Tmc2 is necessary for mechanotransduction and can fully compensate for deletion of Tmc1 in basal-turn hair cells at this early postnatal stage. There was a significant reduction in transduction current amplitudes in Tmc1Δ/Δ-Tmc2Δ/Δ OHCs from P2–P4 basal cochleae, which may reflect haploinsufficiency of Tmc2. Consistent with our data from cochlear apex, we found that basal OHCs that lacked both Tmc1 and Tmc2 had no mechanotransduction currents (Figure 6, C and D).
Rescue of Tmc1Δ/Δ and Tmc2Δ/Δ hair cell mechanotransduction with Tmc1 or Tmc2. To investigate whether Tmc1 or Tmc2 is sufficient to restore mechanosensitivity in Tmc1Δ/ΔTmc2Δ/Δ hair cells, we generated adenoviral expression vectors encoding Tmc1ex1, Tmc1ex2, or Tmc2. A fragment of the human MYO7A promoter was initially used to produce relatively low levels of either Tmc1 or Tmc2 expression. The coding sequence for red fluorescent protein (RFP) with a separate promoter was included as a transfection marker. We prepared organotypic cultures from utricles harvested from Tmc1Δ/ΔTmc2Δ/Δ mice at P0. All tested hair cells (n = 11) that lacked RFP fluorescence also lacked mechanotransduction current (data not shown). Tmc1Δ/ΔTmc2Δ/Δ hair cells that were RFP positive, indicating transfection with Ad-Tmc1ex1, Ad-Tmc1ex2, or Ad-Tmc2, had mechanotransduction currents (Figure 7, A and B). This suggests that exogenous expression of either Tmc1 or Tmc2 is sufficient to rescue mechanotransduction in Tmc1Δ/ΔTmc2Δ/Δ utricles hair cells. We also generated a negative control adenoviral vector, Ad-Tmc1ex2-dn, encoding the deafness mouse allele of Tmc1ex2. Hair cells transfected with Ad-Tmc1ex2-dn did not recover mechanotransduction, indicating that rescue of mechanotransduction by TMC1 or TMC2 is a specific effect of expression of functional TMC protein.

We also tested the ability of exogenous TMC1 or TMC2 to rescue mechanotransduction in cochlear cultures harvested from Tmc1Δ/ΔTmc2Δ/Δ mice at P0. RFP-positive hair cells that expressed Ad-Tmc2 had mechanotransduction (Figure 7, C and D), while neither Ad-Tmc1ex1 nor Ad-Tmc1ex2 rescued mechanotransduction (data not shown). We also generated a Tmc1ex1 expression vector driven by a CMV promoter to produce relatively higher levels of Tmc1 expression. RFP-positive hair cells that expressed CMV promoter–driven Ad-Tmc1ex1 had unambiguous mechanotransduction currents (Figure 7, C and D), indicating that exogenous expression of either Tmc1 or Tmc2 was sufficient to restore mechanotransduction in Tmc1Δ/ΔTmc2Δ/Δ auditory hair cells. The rescue of hair cell transduction in organotypic cultures raises the possibility that Tmc gene replacement in neonatal mice may preserve or restore hearing and balance function in Tmc-deficient animals.

The inability to rescue mechanotransduction with MYO7A pro...
We used the styryl membrane dye ΔΔP. Although ΔΔP

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To investigate whether Tmc1 or Tmc2 was sufficient to rescue FM1-43 uptake in Tmc1Δ/ΔTmc2Δ/Δ hair cells, we excised utricles and cochleae from Tmc1Δ/ΔTmc2Δ/Δ mice at P0–P2 and exposed them to adenoviral Tmc1Δ/Δ or Tmc2Δ/Δ expression vectors with RFP as a transfection marker. We observed robust uptake of FM1-43 in utricles and cochleae exposed to vectors expressing Tmc2 (Supplemental Figure 6). In Tmc1Δ/ΔTmc2Δ/Δ utricles transfected with Tmc1Δ/Δ, fewer cells displayed robust FM1-43 uptake, consistent with the modest Tmc1Δ/Δ rescue of mechanotransduction (Figure 7) and the reduced transduction currents in Tmc1Δ/ΔTmc2Δ/Δ hair cells (Figure 5). As a control, we transfected Tmc1Δ/ΔTmc2Δ/Δ utricles with Ad-Tmc1Δ/Δ. Although there were many transfected cells, none took up FM1-43, indicating that Tmc1- and Tmc2-mediated rescue of FM1-43 uptake in Tmc1Δ/ΔTmc2Δ/Δ utricles is a specific effect of expression of functional Tmc1 and Tmc2.

Gentamicin uptake. To further investigate mechanotransduction in Tmc1;Tmc2 mutant hair cells, we evaluated uptake of gentamicin, a vestibulotoxic and cochleotoxic aminoglycoside antibiotic thought to enter hair cells mainly, if not entirely, through mechanotransduction channels (24). Utricular maculae were explanted from P2 mice, incubated overnight in culture medium, and exposed to 1 mM gentamicin for 48 hours. Wild-type hair bundles were almost completely eliminated (Figure 9, A and B), as previously reported (32). In contrast, hair bundles were intact in gentamicin-treated Tmc1Δ/ΔTmc2Δ/Δ hair cells as well as control explants not exposed to gentamicin (Figure 9, A and B). The number of Tmc1Δ/ΔTmc2Δ/Δ hair bundles was significantly reduced (P < 0.05, 1-way ANOVA) in comparison to non-exposed controls, whereas Tmc1Δ/ΔTmc2Δ/Δ hair bundles were eliminated almost completely (Figure 9, A and B). To confirm uptake of gentamicin in wild-type hair cells and lack of uptake in Tmc1Δ/ΔTmc2Δ/Δ hair cells, we stained gentamicin-exposed utricles with anti-gentamicin antibodies. After a 6-hour incubation with 1 mM gentamicin, robust gentamicin immunoreactivity was observed in wild-type hair cells, while no gentamicin immunoreactivity was detected in Tmc1Δ/ΔTmc2Δ/Δ hair cells (Figure 9C). The lack of gentamicin uptake in Tmc1;Tmc2 mutant hair cells provides another line of evidence supporting the hypothesis that Tmc1 or Tmc2 is required for vestibular hair cell mechanotransduction.

To examine the uptake of gentamicin in Tmc1;Tmc2 mutant cochlear hair cells, whole cochleae were explanted from P2 mice, incubated overnight in culture medium, exposed to 1 mM gentamicin for 48 hours, and then fixed immediately for visualization. In wild-type and Tmc1Δ/ΔTmc2Δ/Δ cochleae, both inner and outer hair bundles were eliminated completely, whereas Tmc1Δ/ΔTmc2Δ/Δ hair bundles remained (Supplemental Figure 7). Hair cells with functional Tmc1 but not Tmc2 (Tmc1Δ/ΔTmc2Δ/Δ) showed an intermediate phenotype; some hair bundles were eliminated, but only toward the basal end of the cochlea. To assess whether susceptibility to gentamicin was different at a later developmental stage, we repeated the same experiment on cultures explant-
We generated antibodies to Ex1-Ex2 and indicated the total number of cells examined. *notransduction current amplitudes (±SEM) from 45 basal OHCs. Upper numbers indicate the number of mice examined, and lower numbers of the cochlea from mice of the indicated genotypes. The stimulus protocol and scale bars in 1-way ANOVA, relative to wild-type values. (P<0.001, 1-way ANOVA, relative to wild-type values. See also Supplemental Figure 5.)

Figure 6

Tmc1Δ and Tmc2Δ cochlear hair cell mechanotransduction currents. (A) Representative mechanotransduction current families recorded from OHCs harvested at P5–P7 from the apical cochlea of mice of the indicated Tmc1−,Tmc2− genotypes. Hair bundles were deflected for 90-ms steps that ranged from 200 nm to 750 nm in 50-nm increments while cells were held at a physiologically relevant holding potential (~64 mV). Some traces have been removed for clarity. The envelope of the stimulus protocol is shown at the bottom right. Tmc1Δ,Tmc2Δ OHCs lack mechanotransduction currents entirely. Scale bars apply to all current families. (B) Summary bar graph of mean maximal mechanotransduction current amplitudes (±SEM) from 100 apical OHCs. For each bar, the upper number indicates the number of mice examined, and the lower number indicates the total number of hair cells examined. Tmc1Δ,Tmc2Δ and Tmc1Δ,Tmc2Δ hair cells had intermediate current amplitudes. *P<0.001, 1-way ANOVA, relative to wild-type values. (C) Representative transduction currents recorded from OHCs excised at P2–P4 from the basal end of the cochlea from mice of the indicated genotypes. The stimulus protocol and scale bars in A apply. (D) Summary bar graph of maximal mechanotransduction current amplitudes (±SEM) from 45 basal OHCs. Upper numbers indicate the numbers of mice examined, and lower numbers indicate the total numbers of cells examined. *P<0.001, 1-way ANOVA, relative to wild-type values. See also Supplemental Figure 5.

ed at P3 and incubated for 4 days. After a 48-hour incubation in 1 mM gentamicin, similar results were observed. Wild-type hair bundles were eliminated completely; some middle-turn and most basal-turn Tmc1Δ,Tmc2Δ hair bundles were eliminated, while Tmc1Δ,Tmc2Δ hair bundles remained intact (Supplemental Figure 8). To confirm uptake of gentamicin in wild-type hair cells and no uptake in Tmc1Δ,Tmc2Δ hair cells, we stained cochleae with anti-gentamicin antibodies. After a 6-hour incubation with 1 mM gentamicin, robust gentamicin immunoreactivity was detected in wild-type hair cells, while no gentamicin immunoreactivity was observed in Tmc1Δ,Tmc2Δ hair cells (Figure 9D). The lack of gentamicin uptake in Tmc1Δ,Tmc2Δ hair cells provides another line of functional evidence supporting the hypothesis that Tmc1 or Tmc2 is required for cochlear hair cell mechanotransduction. Moreover, the intermediate loss of hair cells in the basal region of cochleae lacking Tmc2 may reflect the base-to-apex developmental gradient of Tmc1 expression at this early postnatal stage (Figure 1B) and the developmental switch from Tmc2 to Tmc1 expression.

Hair cell localization of TMC proteins. We generated antibodies to TMC1 or TMC2 by in situ immunization of rabbits and in vitro screening of a recombinant human monoclonal antibody library with synthetic peptides and larger recombinant protein fragments corresponding to TMC1 and TMC2. We validated the specificity of antibodies on recombinant TMC1 or TMC2 expressed in heterologous cells, but did not detect specific labeling of hair cells. Our antibodies did not label the lateral membrane of hair cells (data not shown), where potential TMC2 immunoreactivity in the chicken basilar papilla has been observed (33).

As an alternative to immunolocalization of endogenous TMC proteins, we used biolistic gene transfection to express TMC1Ex1::AcGFP and TMC1Ex2::AcGFP in organotypic cultures of hair cells. TMC proteins were fused at their carboxy termini to AcGFP for visualization. We used either CMV or MYO7A promoters to produce higher or lower levels of expression, respectively. AcGFP did not interfere with the ability of these TMC expression constructs to rescue FM1-43 uptake (data not shown). TMC1Ex1::AcGFP and TMC1Ex2::AcGFP expressed from the MYO7A promoter were not detected in stereocilia, but were detected in stereocilia in a few rare experiments using the CMV promoter (Supplemental Figure 9). In contrast, TMC2::AcGFP was consistently detected near the tips of stereocilia in both vestibular and cochlear hair cells when driven by the MYO7A promoter (Figure 10), but not the CMV promoter (data not shown), consistent with the results of the rescue experiments.
Our results indicate that TMC1 and TMC2 are necessary for mechanotransduction in early postnatal mouse inner ear hair cells. We suspect that the previously reported mechanotransduction currents in P8 cochlear hair cells from Tmc1<sup>−/−</sup> and Tmc2<sup>−/−</sup> mice (9) may have been the result of residual TMC2. Our observation of a developmental switch in the cochlea from Tmc2 to Tmc1 mRNA expression, nearly complete by P8, suggests that TMC2 is not available to compensate for loss of TMC1 in the mature auditory system. This failure to compensate appears to occur at the level of transcription or mRNA stability, since Tmc2 mRNA levels do not increase in Tmc1<sup>−/−</sup> cochleae (K. Kurima, unpublished observations). Steel and Bock (8) reported that young dn mice (P12–P20), presumably homozygous for the Tmc1<sup>−/−</sup> allele, had no cochlear potentials, a collective measure of hair cell receptor potentials, despite the presence of hair cells. We thus conclude that recessive alleles of TMC1/Tmc1 cause profound deafness via failure to maintain mechanotransduction in mature cochlear hair cells. The loss of transduction in TMC1-deficient mouse cochlear hair cells may ultimately lead to hair cell degeneration, which begins as early as P14, beyond which TMC2 is no longer available to compensate. We propose that the failure of Tmc1<sup>−/−</sup> and Tmc2<sup>−/−</sup> cochlear hair cells to develop mature voltage-dependent currents at P14 (9) may be an indirect consequence of the failure to maintain mechanotransduction.

Discussion

We observed that vestibular hair cells and early postnatal cochlear hair cells lacking TMC1 and TMC2 appear structurally normal in scanning electron micrographs (Figure 4) but have no detectable mechanotransduction (Figures 5 and 6), no FM1-43 uptake (Figure 8), and no gentamicin uptake (Figure 9) at time points when stereocilia and mechanotransduction are well developed in wild-type hair cells (27, 28). This structure-function phenotype has not been reported for any other mouse mutants (34) and provides strong genetic evidence for the direct contribution of these proteins to mechanotransduction in inner ear hair cells. A prominent role for TMC2 and TMC1 in inner ear hair cell transduction is further supported by the following experimental observations: First, the rise in Tmc2 mRNA expression in both vestibular and cochlear hair cells coincides with the developmental acquisition of mechanotransduction (27, 28), followed 2–3 days later by a rise in Tmc1 mRNA expression (Figure 1). Second, exogenous expression of either TMC1 or TMC2 can rescue mechanotransduction in either vestibular or cochlear hair cells of Tmc1<sup>−/−</sup>Tmc2<sup>−/−</sup> mice (Figure 7). Third, exogenously expressed TMC1 and TMC2 proteins can be localized at or near the tips of stereocilia (Figure 10 and Supplemental Figure 9), as predicted for proteins that directly contribute to mechanotransduction.

Although we did not consistently observe TMC1 protein localization in stereocilia, abundant unpublished data (not shown) suggest that this reflects the technical challenge of expressing sufficient quantities of tagged TMC proteins for detectable localization in stereocilia, without causing a stress response and cell degeneration due to overexpression and retention in the endoplasmic reticulum. These technical challenges associated with TMC localization are consistent with the expected properties of proteins constituting or directly associated with the hair cell mechanotransduction complex.

One hypothesis to explain our results is that TMC1 and TMC2 are required for trafficking, assembly, structure, or function of the mechanotransduction complex. However, since structurally visible components of the complex, such as tip links, remain intact (Figure 4), the missing component remains obscure. An alternate hypothesis that would explain our observations is that TMC1 and TMC2 may be components of the mechanotransduction channel itself (1, 3–5). The topology of TMC1 and TMC2 resembles that of a large superfAMILY of proteins, including Shaker K+ and TRP channels, with 6 membrane-spanning segments and cytoplasmic N and C termini (1, 5, 35, 36). Moreover, in silico hydropathy analyses imply the existence of two hydrophobic segments that are not predicted to span the membrane and may thus constitute a pore-loop structure (5). These segments are located within the most highly conserved region, the
TMC domain, of TMC proteins (3, 4). Part of this domain is deleted in the Tmc1\(^{dn}\) allele that fails to rescue mechanotransduction (Figure 7, A and B, and Supplemental Figure 6).

Formal proof that TMC1 or TMC2 can constitute the mechanotransduction channel requires data that can directly attribute channel properties to these proteins. Unfortunately, when exogenous TMC1 and TMC2 were expressed in a variety of heterologous systems, they were retained in intracellular compartments and never localized to the plasma membrane, where they could be directly tested for channel activity. We suspect these systems lack hair cell components required for proper trafficking of TMC1 and TMC2 to the plasma membrane of stereocilia.

Alteration of the intrinsic properties of the hair cell mechanotransduction channel with amino acid substitutions in TMC1 or TMC2 would be another approach to attribute pore-forming channel activity to these proteins. All of the known dominant mutations of TMC1/Tmc1 are missense amino acid substitutions (1, 2, 37, 38) that must act via a gain-of-function or dominant-negative mechanism (1). The aspartate 572 residue within the highly conserved TMC domain of TMC1 is substituted by two of three reported DFNA36 mutations (1, 37, 38). The other reported DFNA36 mutation affects the glycine 417 residue that is adjacent to methionine 418 (38), the orthologous residue for methionine 412 that is substituted in Tmc1\(^{Bth}\) (1, 2). This clustering of dominant mutations suggests...
Figure 9

*Tmc1*Δ and *Tmc2*Δ hair cell uptake of gentamicin. (A) After a 48-hour incubation in culture medium with 1 mM gentamicin, wild-type utricular hair bundles were almost completely eliminated, while *Tmc1*Δ/*Tmc2*Δ utricular hair cells retained an intact hair bundle. Scale bar: 10 μm. (B) The mean density of intact hair bundles (±SD) within the central area of more than 6 utricular maculae of each genotype is shown for each genotype. *Tmc1*Δ/*Tmc2*Δ and *Tmc1*Δ/*Tmc2*Δ utricular maculae showed no significant difference between cultures grown with gentamicin in comparison with control cultures grown without gentamicin. *Tmc1*Δ/*Tmc2*Δ utricular hair cells showed intermediate sensitivity to gentamicin. One-way ANOVA results for differences from normal control values are indicated: *P* < 0.05; **P** < 0.001; NS, *P* > 0.05. (C) After a 6-hour incubation in culture medium with 1 mM gentamicin, robust immunoreactivity for gentamicin was detected in wild-type utricular hair cells, whereas no immunoreactivity for gentamicin was detected in *Tmc1*Δ/*Tmc2*Δ hair cells. Red, anti–myosin VI antibodies; green, anti-gentamicin antibodies. Scale bar: 10 μm. (D) After a 6-hour incubation in 1 mM gentamicin, robust immunoreactivity for gentamicin was detected in wild-type IHCs and OHCs, whereas no immunoreactivity for gentamicin was detected in *Tmc1*Δ/*Tmc2*Δ hair cells. Red, anti–myosin VI antibodies; green, anti-gentamicin antibodies. Filled arrowheads, IHCs; open arrowheads, OHCs. Scale bar: 10 μm. See also Supplemental Figures 7 and 8.
there is a critical function(s) associated with the wild-type or mutant amino acids at these positions. These mutations are thus excellent candidates to explore potential effects on the intrinsic properties of mechanotransduction. The data could provide insight into the pathogenesis of DFNA36 hearing loss and further evidence supporting the role of TMC proteins in hair cell mechanotransduction.

**Methods**

**qPCR analysis.** Twenty mouse utricular maculae were excised from 10 wild-type mice at each stage between E15 and P21. Ten cochleae were excised from 5 wild-type mice at each age between P0 and P22. RNA was extracted and quality was confirmed using an Agilent Bioanalyzer (Agilent Technologies), and it was reverse transcribed into cDNA for qPCR analysis with efficient primer sets specific to either Tmc1 or Tmc2 with SYBR GreenER qPCR reagent (Invitrogen) as previously described (39). To amplify a fragment of Tmc1 common to both Tmc1Δ/+ and Tmc1Δ/Δ, we used primers 5′-CATCTGGCAGCCAACATTGGTGTTG-3′ and 5′-AGAGGTAACGGAAATTACGCTACTGATC-3′. To amplify a fragment of Tmc2, we used primers 5′-AGATCTTTGCGTCCCTTGCAAC-3′ and 5′-GATCTTCTTGGCACTGGCCATT-3′. Expression levels were normalized to those of Actb (encoding β-actin) amplified with 5′-TGAGGCGAAGTACTCTGTGATG-3′ and 5′-ACTCATCGTACTCTGGTTGGCTGA-3′. All primers were designed to span introns and validated using melt curve analysis and negative controls.

**In situ hybridization analysis.** Cryopreserved half-heads between P0 and P5 were sectioned at 12-μm thickness and processed for in situ hybridization as described previously (40). Tmc2 transcripts were detected using both 5′ and 3′ RNA probes generated from Tmc2 cDNA. The 5′ RNA probe corresponds to exons 2–6, and the 3′ RNA probe corresponds to exons 17–20 of Tmc2.

**Tmc1 and Tmc2 mutant mice.** Tmc1Δ/+ and Tmc2Δ/+ targeting constructs were designed to replace exons encoding the highly conserved first transmembrane domains (5) (Figure 2, A and B). We generated homology arms from lambda bacteriophage clones encoding Tmc1 exons 8 and 9 or Tmc2 exon 7 isolated from a 129X1/SvJ mouse genomic library (Agilent Technologies). Each targeting vector comprised a pPNT plasmid (41) containing Tmc1 or Tmc2 homology arms, a gene trap cassette (42), an internal ribosomal entry site (IRES) followed by a lacZ reporter gene, and a neomycin resistance gene (PGK-Neo) flanked by LoxP sites. R1 mouse ES cells (43) were electroporated with NotI-linearized targeting vector and grown in the presence of G418 and ganciclovir, using standard protocols, at the University of Michigan Transgenic Animal Model Core. ES cell clones with proper homologous recombination events were identified by Southern blot analysis, expanded, and injected into C57BL/6J blastocysts. Male chimeras were bred with C57BL/6J females to generate germline Tmc1Δ/+ or Tmc2Δ/+ heterozygotes. Germline transmission and Cre recombinase-mediated removal of the PGK-Neo cassette were confirmed by Southern blot (Figure 2, C and D) and PCR analyses (data not shown). Primer information for genotyping is shown in Supplemental Table 1. We ruled out clonal effects by comparison of founder lines derived from two different ES cell clones for both Tmc1Δ/+ and Tmc2Δ/+.

**lacZ reporter analysis.** Otic capsules were dissected from C57BL/6J wild-type, Tmc1Δ/+; Tmc2Δ/+; Tmc1Δ/+; Tmc2Δ/Δ; or Tmc1Δ/+; Tmc2Δ/Δ mice ranging from P1 to P28, and β-galactosidase activity was detected using X-gal staining (44). Results were confirmed with at least 3 mice of each genotype and age.

**Vestibulo-ocular reflex testing.** In the absence of CNS or oculomotor abnormalities, the first 100 ms of the VOR response to high-acceleration whole-body rotations in darkness about an axis through the stereotactic center of the skull and perpendicular to the planes of the horizontal semicircular canals is a sensitive and specific measure of lateral semicircular canal function. Vestibulo-ocular reflex responses to transient yaw rotations were measured (45) in at least 3 adult mice of each genotype at 2–3 months of age (C57BL/6J wild-type, n = 3; Tmc1Δ/+; Tmc2Δ/+; Tmc1Δ/+; Tmc2Δ/Δ, n = 3; Tmc1Δ/+; Tmc2Δ/Δ, n = 3; Tmc1Δ/+; Tmc2Δ/Δ, n = 3). An array of three fluorescent non-collinear markers was affixed to the anesthetized cornea using cyanoacrylate. Movement of the array was measured using 180-frame/s, 3-dimensional video-oculography while the mouse’s body was rotated about the stereotactic center and it was reverse transcribed into cDNA for qPCR analysis with efficient primer sets specific to either Tmc1 or Tmc2 homology arms, a gene trap cassette (42), an internal ribosomal entry site (IRES) followed by a lacZ reporter gene, and a neomycin resistance gene (PGK-Neo) flanked by LoxP sites. R1 mouse ES cells (43) were electroporated with NotI-linearized targeting vector and grown in the presence of G418 and ganciclovir, using standard protocols, at the University of Michigan Transgenic Animal Model Core. ES cell clones with proper homologous recombination events were identified by Southern blot analysis, expanded, and injected into C57BL/6J blastocysts. Male chimeras were bred with C57BL/6J females to generate germline Tmc1Δ/+ or Tmc2Δ/+ heterozygotes. Germline transmission and Cre recombinase-mediated removal of the PGK-Neo cassette were confirmed by Southern blot (Figure 2, C and D) and PCR analyses (data not shown). Primer information for genotyping is shown in Supplemental Table 1. We ruled out clonal effects by comparison of founder lines derived from two different ES cell clones for both Tmc1Δ/+ and Tmc2Δ/+. Tmc1Δ/+ and Tmc2Δ/+ lines were backcrossed to C57BL/6J for at least 10 generations to generate congenic lines for all experiments.

**lacZ reporter analysis.** Otic capsules were dissected from C57BL/6J wild-type, Tmc1Δ/+; Tmc2Δ/+; Tmc1Δ/+; Tmc2Δ/Δ, or Tmc1Δ/+; Tmc2Δ/Δ mice ranging from P1 to P28, and β-galactosidase activity was detected using X-gal staining (44). Results were confirmed with at least 3 mice of each genotype and age.

**Vestibulo-ocular reflex testing.** In the absence of CNS or oculomotor abnormalities, the first 100 ms of the VOR response to high-acceleration whole-body rotations in darkness about an axis through the stereotactic center of the skull and perpendicular to the planes of the horizontal semicircular canals is a sensitive and specific measure of lateral semicircular canal function. Vestibulo-ocular reflex responses to transient yaw rotations were measured (45) in at least 3 adult mice of each genotype at 2–3 months of age (C57BL/6J wild-type, n = 3; Tmc1Δ/+; Tmc2Δ/+; Tmc1Δ/+; Tmc2Δ/Δ, n = 3; Tmc1Δ/+; Tmc2Δ/Δ, n = 3; Tmc1Δ/+; Tmc2Δ/Δ, n = 3). An array of three fluorescent non-collinear markers was affixed to the anesthetized cornea using cyanoacrylate. Movement of the array was measured using 180-frame/s, 3-dimensional video-oculography while the mouse’s body was rotated about
an Earth-vertical axis passing through the animal’s head with its vision occluded. The stimulus was a constant acceleration of 30.00°/s² for 100 ms and a subsequent cruise at 300°/s for 400 ms. Responses were quantified as acceleration gain, the ratio of slow-phase eye velocity to head velocity averaged over the constant acceleration phase of the stimulation. Nystagmus quick phases with high peak velocity (>300°/s) and peak acceleration (>2,000°/s²) indicate normal function of the extraocular motor system.

**ABR testing.** ABR thresholds were measured at 5 or 12 weeks of age in at least 4 mice of each genotype (Tmc1Δ/ΔTmc2+/+, n = 5; Tmc1+/ΔTmc2+/+, n = 5; Tmc1Δ/ΔTmc2+/+, n = 5; Tmc1+/ΔTmc2+/+, n = 5; Tmc1+/ΔTmc2+/+, n = 6; Tmc1+/ΔTmc2+/+, n = 6; Tmc1Δ/ΔTmc2+/+, n = 9; Tmc1Δ/ΔTmc2+/+, n = 7; Tmc1Δ/ΔTmc2+/+, n = 7; Tmc1Δ/ΔTmc2+/+, n = 4). We used alternating polarity click and tone-burst stimuli of 5 ms duration (29). Stimulus intensities were initiated at suprathreshold values and initially decreased by 10 dB steps, which were followed by 5 dB steps to determine the ABR threshold. When no ABR waveform was detectable at the highest stimulus level of 100 dB sound pressure level (SPL), the threshold was considered to be 105 dB SPL for subsequent analyses.

**SEM.** Samples were prepared from C57BL/6j wild-type, Tmc1Δ/ΔTmc2+, Tmc1Δ/ΔTmc2+, or Tmc1+/ΔTmc2+ mice ranging in age from P0 to P28 using the OTOTO method (46) with some modifications (47). Results were confirmed with at least 3 mice of each genotype and age. Otic capsules were fixed in 2.5% glutaraldehyde buffered with 0.1 M sodium cacodylate containing 0.1 M sodium cacodylate containing 2 mM CaCl₂ for 1–1.5 hours at 4°C, rinsed in 0.1 M sodium cacodylate buffer containing 2 mM CaCl₂ and postfixed with 1% osmium tetroxide (OsO₄) with 0.1 M sodium cacodylate containing 2 mM CaCl₂ for 1 hour at 4°C. Cochlear and vestibular sensory epithelial tissues were dissected, and the tectorial membrane and otolithic membranes with otoconia were removed in 70% ethanol. Tissues were gradually hydrated to distilled water, and the tectorial membrane and otolithic membranes with otoconia were removed in 70% ethanol. Tissues were gradually hydrated to distilled water, rinsed with 0.1 M sodium cacodylate buffer, the TCH and OsO₄ treatments were repeated twice. The tissues were then gradually dehydrated to ethanol, critical point dried, and imaged with a Hitachi S-4800 field emission electron microscope at 1- to 10-kV acceleration voltages.

**Hair cell electrophysiology.** Utricles and cochleae were excised, mounted on glass coverslips, and viewed on an Axioskop FS upright microscope (Zeiss) equipped with a 63× water-immersion objective and differential interference contrast optics. Electrophysiological recordings were performed at room temperature in solutions containing (in mM): 137 NaCl, 5.8 KCl, 10 HEPS, 0.7 NaH₂PO₄, 1.3 CaCl₂, 0.9 MgCl₂, and 5.6 d-glucose, vitamins (1:100) and amino acids (1:50) as in MEM (Invitrogen), pH 7.4 (311 mOsm/kg). Recording electrodes (2–4 MΩ) were pulled from R-6 glass (King Precision Glass) and filled with (in mM): 135 KCl, 5 EGTA-KOH, 5 MgCl₂, and 1 CaCl₂, pH 7.4 (284 mOsm/kg). The whole-cell, tight-seal technique was used to record mechanotransduction currents using an Axopatch 200B (Molecular Devices) for cochleae. Cells were filled with 0.1 M sodium cacodylate containing 2 mM CaCl₂ and prepared for SEM as described above.

**Gentamicin uptake.** One cochlea or utricle from each experimental mouse was exposed to gentamicin, and the contralateral cochlea or utricle was used as a nonexposed control. Utricular maculae were excised from P2 mice of each genotype (C57BL/6j wild-type, n = 6; Tmc1Δ/ΔTmc2+/+, n = 7; Tmc1+/ΔTmc2+/+, n = 8; Tmc1Δ/ΔTmc2+/+, n = 6; Tmc1Δ/ΔTmc2+/+, n = 6) in L-15 medium and explanted onto a glass-bottom dish (MatTek) previously coated with type I collagen (Millipore). Explants were bathed in DMEM supplemented with 7% FBS and incubated overnight at 37°C with 5% CO₂. Whole cochleae were excised from P2 or P3 mice of each genotype and incubated overnight or 4 days, respectively. Numbers of P2 mice were as follows: Tmc1Δ/ΔTmc2+/+, n = 6; Tmc1Δ/ΔTmc2+/+, n = 6; Tmc1+/ΔTmc2+/+, n = 6; Tmc1Δ/ΔTmc2+/+, n = 4; Tmc1Δ/ΔTmc2+/+, n = 3. Numbers of P3 mice were: Tmc1Δ/ΔTmc2+/+, n = 3; Tmc1Δ/ΔTmc2+/+, n = 3; Tmc1Δ/ΔTmc2+/+, n = 4. The culture medium was replaced with fresh gentamicin-free medium for controls or with medium containing 1 mM gentamicin and incubated for 48 hours at 37°C with 5% CO₂. The explants were then fixed with 4% PFA for 30 minutes at room temperature. F-actin was visualized with rhodamine phalloidin (Invitrogen).

To test the effects of 5 mM 1,2-bis(2-aminophenoxy)ethane-tetraacetic acid (BAPTA), HHBS-Ba was replaced with 5 mM BAPTA in HHBS-Ba in every step. FM1-43 fluorescence from whole cochlea was captured with a 10x objective lens and magnified images from middle turns were captured with a 40x objective at 5 and 10 minutes after FM1-43 application, respectively. All images were obtained with identical settings on an LSM 710 confocal microscope equipped with ZEN 2009 software (Zeiss). Gamma settings for the green channel were adjusted equally throughout entire images and for all images using Adobe Photoshop CS5 software. Immediately after imaging, samples were fixed with 2.5% glutaraldehyde buffered with 0.1 M sodium cacodylate containing 2 mM CaCl₂ and prepared for SEM as described above. Gentamicin uptake. One cochlea or utricle from each experimental mouse was exposed to gentamicin, and the contralateral cochlea or utricle was used as a nonexposed control. Utricular maculae were excised from P2 mice of each genotype (C57BL/6j wild-type, n = 6; Tmc1Δ/ΔTmc2+/+, n = 7; Tmc1+/ΔTmc2+/+, n = 8; Tmc1Δ/ΔTmc2+/+, n = 6; Tmc1Δ/ΔTmc2+/+, n = 6) in L-15 medium and explanted onto a glass-bottom dish (MatTek) previously coated with type I collagen (Millipore). Explants were bathed in DMEM supplemented with 7% FBS and incubated overnight at 37°C with 5% CO₂. Whole cochleae were excised from P2 or P3 mice of each genotype and incubated overnight or 4 days, respectively. Numbers of P2 mice were as follows: Tmc1Δ/ΔTmc2+/+, n = 6; Tmc1Δ/ΔTmc2+/+, n = 6; Tmc1+/ΔTmc2+/+, n = 6; Tmc1Δ/ΔTmc2+/+, n = 4; Tmc1Δ/ΔTmc2+/+, n = 3. Numbers of P3 mice were: Tmc1Δ/ΔTmc2+/+, n = 3; Tmc1Δ/ΔTmc2+/+, n = 3; Tmc1Δ/ΔTmc2+/+, n = 4. The culture medium was replaced with fresh gentamicin-free medium for controls or with medium containing 1 mM gentamicin and incubated for 48 hours at 37°C with 5% CO₂. The explants were then fixed with 4% PFA for 30 minutes at room temperature. F-actin was visualized with rhodamine phalloidin (Invitrogen). We counted intact hair bundles within an arbitrarily selected region of 100 μm × 100 μm within the central area of each utricular macula. To more directly evaluate gentamicin uptake into hair cells, additional tissues were explanted and incubated in culture medium containing 1 mM gentamicin for 6 hours at 37°C with 5% CO₂ fixed, and stained with mouse monoclonal anti-gentamicin antibodies (QED Biosciences), followed by incubation with goat anti-mouse Alexa Fluor 488 secondary antibodies (Invitrogen). Hair cells were labeled with rabbit polyclonal anti-myosin VI antibodies (Protein Biosciences) and donkey anti-rabbit Alexa Fluor 568 antibodies (Invitrogen). Images were captured with an LSM 710 confocal microscope equipped with ZEN 2009 software.

**DNA expression vectors.** Mouse Tmc1 and Tmc2 cDNAs were generated from RNA extracted from inner ear. Tmc1Δ<sup>ex2</sup> encodes a transcript lacking exon 2, and its translation initiation codon is predicted to be in exon 1 (Supplemental Figure 1A). Tmc1Δ<sup>ex2</sup> encodes a transcript including exon 2, and the translation initiation codon is predicted to be in exon 2 (Supplemental Figure 1A). cDNAs were cloned into AcGFP-N2 vector (Clontech). Tmc1Δ<sup>ex2</sup>-AcGFP, Tmc1Δ<sup>ex2</sup>-AcGFP, and Tmc2::AcGFP cDNA fragments were subcloned into a pAdTrack shuttle plasmid (48) in which the CMV promoter was replaced with a fragment (c. –46 to –3321) of the human MYO7A promoter.
Bioinformatic analysis. Samples were prepared as described previously (51) for use with the Helios Gene Gun System (Bio-Rad). Gold particles (1.0-μm, Bio-Rad) were coated with plasmid DNA at a ratio of 1 μg of gold particles to 2 μg plasmid DNA encoding Tmc1ex1⋯AcGFP, Tmc1ex2⋯AcGFP, or Tmc2:AcGFP with CMV or MYO7A promoters (see DNA expression vectors above). DNA-coated particles were precipitated onto the inner wall of Tefzel tubing (Bio-Rad), which was cut into individual cartridges containing approximately 1 μg of plasmid DNA. Inner-ear sensory epithelial cultures were explanted from P0 or P3 rats. Explants were bathed in DMEM supplemented with 7% FBS and incubated overnight at 37°C with 5% CO₂. Explants were then transfected with a Helios Gene Gun, returned to 37°C for 24–72 hours, and fixed with 3% paraformaldehyde in PBS (pH 7.4). F-actin was visualized with either rhodamine phalloidin (Invitrogen) or Alexa Fluor 647 phalloidin (Invitrogen). Some samples transfected with CMV-Tmc1ex2⋯AcGFP were stained with Alexa Fluor 488-conjugated anti-GFP antibodies (Invitrogen) to facilitate detection of faint GFP expression. Adjacent untransfected hair bundles served as negative controls. Images were captured with an LSM 710 confocal microscope equipped with ZEN 2009 software (Zeiss). Gamma settings for red and green channels were adjusted equally throughout entire images using Adobe Photoshop CS5.

Statistics. Two-tailed Student’s t test, with a significance criterion of $P < 0.05$, was used to compare acceleration gains of the VOR. One-way ANOVA was used to compare ABR thresholds and maximal mechanotransduction current amplitudes (significance criterion, $P < 0.001$) and hair bundle densities after gentamicin exposure (significance criterion, $P < 0.05$).

Acknowledgments
We thank Ken Kitamura for support; Charles Askew, Laura Digilio, Geoffrey Horwitz, and Michael Mulheisen for technical assistance; Inna Belyantseva, Jonathan Bird, Felipe Salles, Takashi Sato, and Shin-ichiro Kitajiri for technical advice; Rechaa Kachar for helpful discussions; and NIDCD colleagues for critical comments. This work was supported by NIH intramural research funds Z01-DC00060-10 (A.J. Griffith) and Z01-DC00021-18 (D.K. Wu); NIH grants R01-DC05439 (J.R. Holt), R01-DC008853 (G.S.G. Gélecö), R01-DC009255 and R01-DC002390 (C.C. Della Santina); and a grant from the Hazel Thorpe Carman & George Gay Carman Trust for Scientific Research (J.R. Holt). Y. Kawashima was supported in part by Tokyo Medical and Dental University.

Received for publication August 10, 2011, and accepted in revised form October 19, 2011.

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