Neurofibromatosis type 1 (NF1) patients develop benign neurofibromas and malignant peripheral nerve sheath tumors (MPNST). These incurable peripheral nerve tumors result from loss of \textit{NF1} tumor suppressor gene function, causing hyperactive Ras signaling. Activated Ras controls numerous downstream effectors, but specific pathways mediating the effects of hyperactive Ras in NF1 tumors are unknown. We performed cross-species transcriptome analyses of mouse and human neurofibromas and MPNSTs and identified global negative feedback of genes that regulate Ras/Raf/MEK/ERK signaling in both species. Nonetheless, ERK activation was sustained in mouse and human neurofibromas and MPNST. We used a highly selective pharmacological inhibitor of MEK, PD0325901, to test whether sustained Ras/Raf/MEK/ERK signaling contributes to neurofibroma growth in a neurofibromatosis mouse model (\textit{Nf1}\textsuperscript{fl/fl};\textit{Dhh-Cre}) or in NF1 patient MPNST cell xenografts. PD0325901 treatment reduced aberrantly proliferating cells in neurofibroma and MPNST, prolonged survival of mice implanted with human MPNST cells, and shrank neurofibromas in more than 80% of mice tested. Our data demonstrate that deregulated Ras/ERK signaling is critical for the growth of NF1 peripheral nerve tumors and provide a strong rationale for testing MEK inhibitors in NF1 clinical trials.
MEK inhibition exhibits efficacy in human and mouse neurofibromatosis tumors

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Neurofibromatosis type 1 (NF1) patients develop benign neurofibromas and malignant peripheral nerve sheath tumors (MPNST). These incurable peripheral nerve tumors result from loss of NFI tumor suppressor gene function, causing hyperactive Ras signaling. Activated Ras controls numerous downstream effectors, but specific pathways mediating the effects of hyperactive Ras in NF1 tumors are unknown. We performed cross-species transcriptome analyses of mouse and human neurofibromas and MPNSTs and identified global negative feedback of genes that regulate Ras/Raf/MEK/ERK signaling in both species. Nonetheless, ERK activation was sustained in mouse and human neurofibromas and MPNSTs. We used a highly selective pharmacological inhibitor of MEK, PD0325901, to test whether sustained Ras/Raf/MEK/ERK signaling contributes to neurofibroma growth in a neurofibromatosis mouse model (Nf1fl/fl;Dhh-Cre) or in NF1 patient MPNST cell xenografts. PD0325901 treatment reduced aberrantly proliferating cells in neurofibroma and MPNST, prolonged survival of mice implanted with human MPNST cells, and shrank neurofibromas in more than 80% of mice tested. Our data demonstrate that deregulated Ras/ERK signaling is critical for the growth of NF1 peripheral nerve tumors and provide a strong rationale for testing MEK inhibitors in NF1 clinical trials.

Introduction
The hallmark manifestation of neurofibromatosis type 1 (NF1) is the development of peripheral nerve tumors (1). In benign and malignant peripheral nerve tumors, cells acquire somatic inactivation of the second NF1 allele and loss of NF1 (neurofibromin) protein function (2). With few exceptions, NF1 patients develop small benign dermal neurofibromas that can number into the thousands and be extremely disfiguring. At least a third of NF1 patients develop larger benign plexiform neurofibromas that cause disfigurement and morbidity when they compress vital structures. Surgical removal of neurofibromas is not always feasible due to tumor location, resulting in substantial morbidity for NF1 patients, and plexiform neurofibromas can transform to malignant peripheral nerve sheath tumors (MPNSTs), a leading cause of death in adults with NF1 (3–6). There is currently no chemotherapeutic regimen that will effectively treat NF1 tumors, warranting investigation into the development of novel molecular-targeted therapeutic strategies (7). Studies have begun to identify molecular alterations in MPNST tumors, yet the pathway of molecular events contributing to neurofibroma growth or progression to malignancy remains unclear (8).

Neurofibromin is a Ras-GTPase activating protein (Ras-GAP), converting active Ras-GTP to inactive Ras-GDP (9, 10). Therefore, cells isolated from neurofibromas (11) and MPNSTs (12, 13) that lack neurofibromin have elevated levels of active Ras-GTP. Aberrant activation of Ras signaling generally leads to promotion of tumor cell proliferation and/or survival (14). However, in benign tumors, Ras signaling can alternatively be associated with oncogene-induced senescence (15, 16), and evidence supports a role for cellular senescence in NF1 dermal neurofibromas (17).

At least 11 effector pathways downstream of Ras-GTP have been described (18). Research focused on the biology of NF1 and pathogenesis of plexiform neurofibroma and MPNST has identified potential therapeutic targets including Ras itself, Ras effectors, growth factor receptors, and angiogenesis (8, 19). For example, Ral and PI3K/AKT/mTOR/S6K1, where mTOR indicates mammalian target of rapamycin, are known to regulate cell proliferation, survival, and cell death and have each been implicated in NF1 tumorigenesis (20, 21). S6K1 is activated in MPNST cells with NF1 mutations, and this response is attenuated by rapamycin in MPNST cell lines, MPNST xenografts, and in a genetic engineered mouse model with NFI and p53 mutations in cis (22, 23). On this basis, a phase II trial of rapamycin in plexiform neurofibromas is ongoing. However, no chemotherapeutic approach blocking any molecular target, including tyrosine kinases upstream of Ras, Ras effectors, or combination of effectors, has to date prevented or arrested neurofibroma formation or more than transiently delayed MPNST growth (7).

Genetically engineered mouse (GEM) NFI models have been developed using Cre/lox technology for ablation of NFI (24–28). We chose the Nf1fl/fl;Dhh-Cre model for preclinical testing, as the neurofibroma histology in this model replicates human neurofibroma histology (25, 26). While some mouse models of neurofibromatosis, and perhaps some human patients, require a heterozygous genetic background (29), the Nf1fl/fl;Dhh-Cre model does not, facilitating preclinical testing. We have used 7 Tesla small-animal MRI to assess tumor growth rate in the Nf1fl/fl;Dhh-Cre model.
mouse model using volumetric MRI analysis. However, treatment with the rapamycin analog RAD001 failed to block tumor growth, and the multikinase inhibitor sorafenib affected few mice (30). The same volumetric measurement technique is in use in ongoing clinical trials and has been proven to sensitively detect small changes in tumor size over time (31, 32). The reproducibility of this method is similar for tumors in mice and humans, and thus the response criteria used in human trials can be applied to the preclinical evaluation in mice. Here, we confirm the transcriptional similarities of human and mouse tumors using a bioinformatics approach and illustrate the use of our GEM model for preclinical evaluation of candidate molecular targets.

We compared the transcriptomes of human NF1 tumors and GEM Nf1 models to normal differentiated peripheral nerves of each species to identify molecular mechanisms contributing to tumorigenesis and shared potential therapeutic targets. Our results support the hypothesis that hyperactive Ras induces expression of genes that suppresses the canonical downstream pathway, Raf/MEK/ERK, in benign neurofibromas and MPNST. Although these transcriptional changes suggest that a negative feedback loop has been induced, ERK remains active in these tumors. Based on these data, we performed preclinical trials of the MEK inhibitor PD0325901 in mouse models of NF1-associated peripheral nerve tumors and observed remarkable efficacy. Based on these data, we performed preclinical trials of the MEK inhibitor PD0325901 in mouse models of NF1-associated peripheral nerve tumors and observed remarkable efficacy.

Results

**PD0325901 inhibits MEK and moderately reduces MPNST cell survival in vitro.** To identify molecular similarities between mouse and human NF1 tumors, we performed cross-species transcriptome analysis of human NF1-derived peripheral nerve tumors, neurofibroma and MPNST (n = 32) (34), and mouse neurofibroma and MPNST (n = 33) compared with normal differentiated peripheral nerves of each species (Supplemental Table 1; supplemental material available online with this article; doi:10.1172/JCI60578DS1). Human data were publically available from our previous studies (34); mouse data have been deposited in the GEO database (GSE41747). Hierarchical clustering of orthologous transcripts defined expression clusters (C17–C20; Supplemental Figure 1A). To prioritize relevant genes, we queried transcripts from orthologous clusters (C17–C20; Supplemental Figure 1) representing significant transcriptional changes in mouse and human samples. Notably, both species upregulated expression of genes that negatively regulate the canonical Ras effector pathway Raf/MEK/ERK (Supplemental Table 2 and Supplemental Figure 1, B and C). Despite transcriptional upregulation of genes that repress Raf/MEK/ERK signaling, and consistent with neurofibroma (11) and MPNST (12, 13) cells having elevated levels of Ras-GTP, activation of the downstream effector ERK was maintained in neurofibromas and MPNSTs, as elevated levels of phosphorylated ERK (p-ERK) were detected in neurofibromas and MPNSTs (Supplemental Figure 2).

**PD0325901 is a MEK inhibitor currently in clinical cancer trials.** Of 70 tested kinases, PD0325901 blocked MEK1 at values of 1 μM in vitro, with the closely related MEK5 affected at 10-fold higher concentrations (37); no other kinases were affected. The unusual specificity of this inhibitor is due to the fact that
PD0325901 is not a direct kinase inhibitor, but rather an allosteric inhibitor. In comparison with its predecessor CI-1040, PD0325901 is more potent and has improved duration and bioavailability and increased metabolic stability (38). PD0325901 blocked Raf/MEK/ERK signaling based on pharmacodynamic measurement of p-ERK in MPNST xenografts at intervals after PD0325901 exposure (Figure 1, A–D). p-ERK disappeared by 30 minutes after dosing MPNST xenografts (Figure 1B); levels remained low at 6 hours (Figure 1C) and resembled pretreatment levels by 24 hours (Figure 1D). Pharmacodynamic assessment of PD0325901 was at the end of the 60-day treatment period, demonstrating that the inhibitor remained efficacious throughout the experiment. Dose-response analysis of PD0325901 was conducted on 5 MPNST cell lines in vitro. Four MPNST cell lines were derived from NF1 patients (S462TY, S462, ST8814, T265); 1 MPNST cell line was derived from a sporadic MPNST (STS26T). In all 60-day treatment period, demonstrating that the inhibitor remained efficacious throughout the experiment. Dose-response analysis of PD0325901 was conducted on 5 MPNST cell lines in vitro. Four MPNST cell lines were derived from NF1 patients (S462TY, S462, ST8814, T265); 1 MPNST cell line was derived from a sporadic MPNST (STS26T). After 4 days of treatment, effects on survival in the 4 NF1-derived cell lines were modest and variable (IC50 values 420–3100 nM (203–1495 ng/ml); (Figure 1E). Measurement of the same tumor from 2 individuals, and tumor size among mice, mimicking human neurofibromas, which are also monitored by serial MRI. There is less than 10% variability in measurement of the same tumor from 2 individuals, and tumor size measured by MRI has been verified by direct dissection (43). We did not observe complete suppression of tumor growth in vivo, supporting the moderate response to MEK inhibition in vitro.

**Efficacy of PD0325901 on neurofibroma growth in vivo.** In Nf110/−;Dhh-Cre mice, the Nf1 gene is deleted exclusively in Schwann cells. All Nf110/−;Dhh-Cre mice develop multiple neurofibromas with histological identity to human neurofibromas (25). Nf110/−;Dhh-Cre mice neurofibroma growth rates can be measured using serial volumetric MRI analysis (30). Neurofibroma growth rates vary among mice, mimicking human neurofibromas, which are also monitored by serial MRI. There is less than 10% variability in measurement of the same tumor from 2 individuals, and tumor size measured by MRI has been verified by direct dissection (43). We measured tumor volume at 5 (Figure 2, A and D) and 7 (Figure 2, B and E) months. Eighteen tumor-bearing mice were randomly assigned to PD0325901 treatment (10 mg/kg/day) (Figure 2F); 10 received vehicle (Figure 2C). All tumors in an individual mouse responded equally to treatment. Therefore, we compared the sum of tumor volumes in each individual mouse before and after treatment (Figure 2C). All tumors in an individual mouse responded equally to treatment. Therefore, we compared the sum of tumor volumes in each individual mouse before and after treatment.
ment with vehicle or PD0325901. For statistical analyses, we also studied 20 historical controls, untreated mice, and vehicle-treated control mice from the same genotype and strain background imaged previously.

Because this dose of MEK inhibitor has been reported to cause toxicity in human trials after prolonged exposure (35, 36, 42), we also analyzed mice for effects at lower doses of PD0325901. Sixteen mice were treated with 5 mg/kg and 15 mice with 1.5 mg/kg PD0325901. The 1.5 mg/kg PD0325901 dose in mice provided a concentration similar to that which will be tested in upcoming human trials (8 mg BID). A striking reduction in neurofibroma volumes was achieved after 60 days of PD0325901 treatment (Figure 2G). Remarkably, tumor volumes were reduced in mice treated with 1.5 mg/kg (10/15 mice), 5 mg/kg (15/16 mice), or 10 mg/kg (14/18 mice) PD0325901 ($P < 0.001$ each dose; mixed models analysis; Supplemental Figure 4). PD0325901 was well tolerated with no apparent toxicity at any dose.

Reduced tumor volume suggested that cell proliferation or cell death might be altered in neurofibromas. Cell proliferation was decreased at the end point of PD0325901 treatment of Nf1fl/fl;Dhh-Cre neurofibromas (Figure 3A). Similar effects on cell proliferation were observed at all 3 doses ($P < 0.001$). Apoptosis was not observed at the end of the experiment (not shown), but neurofibromas did regress. MPNST xenografts treated with PD0325901 showed a modest and marginally significant ($P < 0.05$) reduction in proliferating Ki67+ cells (Figure 3B) relative to neurofibromas (Figure 3A). Similar effects on MPNST proliferation were observed with short-term (11 days) or long-term (28 days) exposure to PD0325901 (Figure 3B). Double labeling tumor sections with Ki67 and CNPase, a Schwann cell marker, indicated that many of the proliferating cells were Schwann cells (Figure 3, C and D).

In addition to effects on tumor cell proliferation, the number of MECAM+ blood vessels was significantly reduced in both neurofibromas (Figure 3E; $P = 0.003$) and MPNST (Figure 3F; $P = 0.008$) subsequent to PD0325901 treatment. This may have resulted indi-
rectly from effects of tumor cells on endothelial cells or directly through effects on blood vessels. In either case, changes in tumor vasculature likely contributed to changes in tumor volume.

Interestingly, we observed reduced p-S6K in PD0325901-treated Nf1fl/fl;Dhh-Cre neurofibromas, but not MPNSTs (Figure 3, G–J), suggesting that S6K is in part downstream of MEK in NF1 benign tumor cells. A reduction in p-S6K after MEK inhibition was also observed in melanoma cell lines (44). Ras/PI3K/Akt is a Ras pathway alternative to Ras/Raf/MEK/ERK, but PD0325901 did not affect p-AKT levels in neurofibromas or MPNSTs (Figure 3, K–N), validating PD0325901 specificity as a MEK inhibitor.

PD0325901 maintains MEK inhibition at low doses in Nf1fl/fl;Dhh-Cre neurofibromas. To determine whether the Raf/MEK/ERK negative feedback mechanism suggested by the microarray data (Supplemental Figure 1) is present in NF1 benign tumor cells, we observed reduced p-ERK after MEK inhibition. However, PD0325901 did not affect p-AKT levels in neurofibromas or MPNSTs (Figure 3, K–N), validating PD0325901 specificity as a MEK inhibitor.

PD0325901 maintains MEK inhibition at low doses in Nf1fl/fl;Dhh-Cre neurofibromas. To determine whether the Raf/MEK/ERK negative feedback mechanism suggested by the microarray data (Supplemental Figure 1, B and C) was valid, we analyzed p-ERK and expression of 2 candidate genes, SPRY4 and DUSP6, at the end of the 60-day treatment period of Nf1fl/fl;Dhh-Cre with low-dose (1.5 mg/kg) and high-dose (10 mg/kg) PD0325901. As observed with high-dose treatment of MPNST xenografts (Figure 1, A–D), p-ERK was reduced in Nf1fl/fl;Dhh-Cre neurofibromas relative to control (Figure 4A) by 30 minutes (Figure 4B), remained low at 6 hours (Figure 4C), and was robust at 24 hours after the last dose of drug (Figure 4D). Reduced p-ERK was observed whether neurofibromas shrank or did not shrink after exposure to PD0325901 (not shown). Similar results were observed with the low-dose treatment at 30 minutes and 6 hours (Figure 4, E–G). However, in sections from mice treated with 1.5 mg/kg MEK inhibitor, p-ERK remained suppressed at 24 hours (Figure 4H).

We confirmed overexpression of SPRY4 and DUSP6 in Nf1fl/fl;Dhh-Cre neurofibromas relative to WT mouse nerve (Figure 4I). Expression of SPRY4 and DUSP6 mRNA was each reduced relative to vehicle control–treated mice at 6 hours following the last dose of PD0325901 at 10 mg/kg (Figure 4J) or 1.5 mg/kg (Figure 4K) PD0325901. However, a significant increment in expression at 24 hours was only observed with the higher dose (Figure 4, J and K). These data suggest that different levels of MEK inhibition differentially affect accumulation of mRNAs encoding negative feedback regulators of ERK. Importantly, the data are consistent with the response of neurofibromas to single-agent MEK inhibition, even at low doses.

**Discussion**

A primary goal of this study was to identify and evaluate gene expression signatures shared between human NF1 tumors and GEM Nf1 models. We posited that shared signatures would reveal critical mechanisms of tumorigenesis and key therapeutic targets to facilitate preclinical studies and development of NF1 therapeutics. Functional enrichment analysis identified a prominent
theme, Raf/MEK/ERK suppression, shared between mouse and human. Despite this transcriptional signature, ERK signaling remained activated in NF1 tumors and inhibiting Raf/MEK/ERK signaling with a MEK inhibitor diminished tumor cell growth. Our data set provides a wealth of gene expression information and supports MEK signaling as an important clinical target in NF1.

Our results identify a transcriptional response to suppress Raf/MEK/ERK activity in neurofibromas and MPNST. In contrast to previous studies indicating overexpression of DUSP and Sprouty family members, feedback inhibitors of Raf/MEK/ERK signaling, Expression of DUSP family members was reported in response to NF1 ablation in human fibroblasts (17) and KRAS2-activating mutations in lung tumors (45), suggesting a common mechanism of suppression in Ras-driven tumors. Elevated SPRY4 expression was also reported in response to NF1 deficiency in fibroblasts, and exogenous expression of SPRY2 induced senescence in NF1-deficient fibroblasts (17). DUSP and Sprouty genes are members of a 52-gene transcriptional profile that is specifically downstream of Raf/MEK/ERK signaling; these genes are expressed at high levels in tumor cell lines with an activating Raf mutation (14) but not tumor cell lines with receptor tyrosine kinase (RTK) activation (46). Consistent with our results in NF1+/−;Dhh-Cre neurofibromas, PD0325901 MEK inhibition reduces expression of these genes and blocks cell proliferation in tumor cell lines with an activating Raf mutation (40).

The PD0325901-induced reduction of neurofibroma volume in 39/49 (80%) of neurofibroma-bearing NF1+/−;Dhh-Cre mice represents the most dramatic result described to date for treatment of neurofibroma-bearing mice. In contrast, RAD001 did not decrease neurofibroma volume, and sorafenib, a multikinase inhibitor, decreased tumor volume in only 5 of 9 (56%) mice (30). Imatinib, a tyrosine kinase inhibitor, showed an un-quantified effect on tumor burden in another neurofibroma mouse model (29). Plexiform neurofibromas have a slow growth rate and a complex non-spherical shape. Therefore, standard response criteria for malignant solid tumors have limited applicability. A more sensitive and reproducible method of response evaluation for human NF1 plexiform neurofibromas was developed, evaluating response and progression using volumetric MRI analysis. Response is defined as 20% or greater decrease in tumor volume compared with baseline, and progression is defined as 20% or greater increase in tumor volume compared with baseline (32). Plexiform neurofibroma shrinkage has not been observed in clinical trials, with the exception of up to 22% in response to pegitinron (47) and rarely in response to imatinib (48). The method of volumetric MRI analysis used in our preclinical studies is identical to the method used in clinical trials, and we used identical response criteria.

Similar to our results, a previous study reported variable effects on MPNST cell survival in vitro with the MEK inhibitor from which PD0325901 was derived, PD184352 (CI-1040) (49). In contrast, the PD0325901 MEK inhibitor showed a robust, yet transient, in vivo effect on survival, possibly due to effects on tumor vasculature. We did not observe persistent apoptosis following PD0325901 treatment in vivo, in contrast with effects in vitro (49). Due to the multiplicity of Ras effectors and complexity of negative feedback regulation, therapeutic strategies against more aggressive Ras-related tumors are likely to include combinations of compounds that target multiple points in the Ras signaling network (40, 50–52). These studies support the investigation of combinatorial effects of PD0325901 with additional Ras pathway inhibitors in NF1 tumors.

Our results provide preclinical evidence implicating PD0325901 as a candidate NF1 therapeutic agent. This was unexpected, as the MEK inhibitor from which PD0325901 was derived failed to show efficacy in a GEM model of NF1 JMML (53). Recently, PD0325901 was found to be effective in reversing myeloproliferative disease in an activated KRas mouse model (54) and in NF1-driven JMML (33), likely due to the more persistent inhibition of MEK by PD0325901, a second-generation MEK inhibitor modified to improve efficacy for clinical cancer trials (55). Collectively, these studies suggest that Raf/MEK/ERK is a critical pathway in NF1.

Preliminary studies reported adverse effects after prolonged treatment of patients with advanced cancers with 10 mg or more BID PD0325901 (35, 36, 42). MPNST xenografts were only moderately sensitive to PD0325901 treatment at this dose, likely requiring combination therapies for MPNST treatment. However, the results of our experiments suggest that the lowest dose of 1.5 mg/kg in our preclinical neurofibroma mouse model, equivalent to 8 mg BID in humans, is as effective in inhibiting tumor growth as the higher dose of 10 mg/kg, supporting evaluation of new dosing schedules in clinical trials of PD0325901. Furthermore, the lower dose (1.5 mg/kg) appeared more effective in maintaining inhibition of MEK, with the higher dose activating the negative feedback response and elevating p-ERK to pretreatment levels. Fine-tuning the long-term maximal effective dose below the threshold of negative feedback may be relevant to monitoring PD0325901 in the clinic.

In summary, NF1 mutation causing neurofibromatosis results in hyperactive Ras, potentially activating numerous downstream signaling pathways. However, a successful targeted therapy in humans has not yet been developed. Combining mouse and human transcriptome data focused attention on increased expression of genes that suppress the Raf/MEK/ERK arm of Ras signaling. This transcriptional repression does not apparently compensate, as ERK phosphorylation was detected in neurofibromas and MPNST. Furthermore, inhibition of neurofibroma and MPNST growth with PD0325901 indicated dependence of both tumor types on sustained Raf/MEK/ERK signaling. The results of our preclinical tests of PD0325901 in MPNST and neurofibroma support investigation of MEK inhibitors as candidate therapeutics in the treatment of Ras-related diseases, including NF1 and other “RASopathies” (56). In addition, NF1 mutations are frequently found in tumor types other than neurofibroma and MPNST, including glioblastoma (57), lung adenocarcinoma (58), and ovarian cancer (59). Thus, PD0325901 may be a potential molecular-targeted treatment for a wide variety of disorders.

Methods

Gene expression microarray. Mouse RNA isolation, hybridization and data normalization, mouse-human ortholog mapping and cross-species data integration are described in Supplemental Methods. Mouse microarray data can be accessed in the GEO MIAME-compliant public database (GSE41747).

In vitro MPNST cytotoxicity assay. MPNST cell lines (STS26T, ST8814, S462, S462TY, T265p21) were obtained and maintained as described (41). All MPNST cell lines were derived from NF1 patients except STS26T. MPNST cells were plated in quadruplicate for each dose of PD0325901 MEK inhibitor (gift of Pfizer Inc.). 24 hours after plating, cells were treated with vehicle alone (0.1% DMSO) or inhibitor. Cell viability was quantified 96 hours after treatment by MTS assay as described (22).

Transient transfection of MPNST cells. Nucleofection and Western analyses are described in Supplemental Methods.

MPNST xenograft. The subcutaneous NF1+/− S462-TY human MPNST xenograft model has been previously described (41, 60). Administration
and dose of PD0325901 were determined by previous animal studies (61). Daily oral gavage of vehicle (0.5% [w/v] methylcellulose solution with 0.2% [v/v] polysorb 80 [TWEEN 80] or 10 mg/kg PD0325901 in vehicle) was given for 28 days beginning when tumors reached 200 mm³ ± 3 weeks after injection. We measured tumors and weighed mice twice weekly once tumors began enlarging. Tumor volume was calculated as follows: \( V = \frac{4}{3} \pi L W^2 \), where \( L \) is the longest diameter and \( W \) is the width. Mice were treated until tumors reached 2500 mm³ or a maximum of 92 days. 100% of control tumors reached 2500 mm³. Two of 18 PD0325901-treated mice did not reach 2500 mm³; one showed complete and sustained remission at 150 days. Survival was analyzed by log-rank test using GraphPad Prism.

**Neurofibroma MRI and tumor volume measurement.** MRI and tumor volume measurement of \( NF1^{fl/fl};Dhh-Cre \) mouse was conducted as described (30). Mice were administered vehicle control (0.5% [w/v] methylcellulose solution with 0.2% [v/v] polysorb 80 [TWEEN 80] or PD0325901 [1.5, 5.0, or 10 mg/kg/d]) by oral gavage for 60 days. Due to the high cost of performing serial MRI, littermate controls (\( n = 5 \)) were allocated to the vehicle treatment group. In addition, historical controls (\( n = 20 \)) were untreated or vehicle-treated control mice from the same genotype and strain background imaged over the 18 months preceding these experiments; these were included to increase statistical power (mixed effects model analysis).

**MPNST statistical analysis.** For MPNST, computing the mean and SEM based solely on surviving mice created downward bias, as mice with large tumors required sacrifice. We analyzed the longitudinally collected tumor volumes together across time points and conducted mixed effects analysis with autoregressive within-mouse dependent structure. Missing data from sacrificed mice with large tumors constituted a missing-at-completely-random (MACR) condition, under which the mixed effects analysis provided consistent estimates of mean tumor volume and SEM (62).

**Immunohistochemistry.** Paraffin sections were deparaffinized, hydrated and transferred to a 1 M citrate buffer (pH 6.0) for antigen retrieval. Slides were boiled for 10 minutes in citrate buffer, cooled at room temperature for 30 minutes, then rinsed in water twice and in PBS 3 times. Sections were quenched with 3% hydrogen peroxide for 10 minutes, rinsed in PBS, and blocked in 10% normal goat serum with 0.3% Triton X-100. Sections were incubated overnight in primary antibody diluted in block; rabbit pERK (#4370, 1:200, Cell Signaling; see Supplemental Methods), rabbit Ki67 (1:10,000, Novocastra), rat MECA 32 (1:5; Developmental Studies Hybridoma Bank), rabbit pS6Kinase (1:800; Cell Signaling), rabbit pAKT (1:50), and mouse CNPase (1:750; MAB226, Millipore). Sections were then incubated in appropriate biotinylated secondary antibodies; goat anti-rabbit or goat anti-rat (1:200; Vector). Some sections were counterstained with Harris hematoxylin. Numbers of MECA 32 blood vessels were counted in 10 ×20 fields per MPNST (\( n = 5 \)) and 5 ×20 fields per neurofibroma (\( n = 5 \)) for each condition. Statistical comparisons between control and tumor samples were conducted using Student’s t test. All microscopic images were acquired with Openlab software suites on a Zeiss Axiosver 200.

For double labeling, paraffin sections were processed for antigen retrieval as above and incubated overnight in a cocktail of 2 of the following primary antibodies: rabbit pERK (1:200; 4370, Cell Signaling), mouse CNPase (1:750; MAB226, Millipore), rabbit Ki67 (1:1000, Novocastra), and rat MECA 32 (1:5; Developmental Studies Hybridoma Bank). Sections were then incubated with a cocktail of 2 appropriate secondary antibodies: Goat anti-rabbit Alexa Fluor 488 (1.800; Invitrogen), goat anti-mouse Alexa Fluor 594 (1.1200; Invitrogen), and goat anti-rat Alexa Fluor 594 (1.1200; Invitrogen). Sections were stained with DAPI (1:10,000) to visualize cell nuclei and mounted with Fluoromount G. Pictures were taken under a fluorescent microscope, and cells were counted using ImageJ software.

**qRT-PCR.** mRNAs were extracted from WT sciatic nerves, \( NF1^{fl/fl};Dhh-Cre \) mouse neurofibromas, and PD0325901- or vehicle-treated \( NF1^{fl/fl};Dhh-Cre \) mouse neurofibromas using the Mini-mRNA isolation kit (QIAGEN). The mRNA was reverse-transcribed using the Superscript Preamplification System (Gibco; Invitrogen). Superscript II reverse transcriptase was used in the reaction, as per the manufacturer’s protocol. Duplicate samples lacking reverse transcriptase were conducted to control for genomic DNA contamination. Primers were designed and synthesized by Integrated DNA Technologies (IDT) for quantitative RT-PCR (qRT-PCR) of SPRY4 and DUSP6 (SPRY4F: 5′-TGTGATCCCGCTGCTAGTG-3′; SPRY4R: 5′-ATTTCACCAGTGCTGTCCTCAT-3′; DUSP6F: 5′-TGCCCAATCTTGTTGAAATGCGG-3′; DUSP6R: 5′-CAATGCAACAGCACCA-CAGTGT-3′). Mouse tubulin primers (IDT) were included in each reaction as a positive control for cDNA. Triplicate reactions were performed in an ABI Prism 7500 as described (63). For statistical analyses, 3 mice were included for each treatment group. Relative gene expression was calculated to tulin expression. Fold change of SPRY4 or DUSP6 in untreated \( NF1^{fl/fl};Dhh-Cre \) mouse neurofibromas compared with WT levels or in PD0325901-treated \( NF1^{fl/fl};Dhh-Cre \) mouse neurofibromas compared with vehicle-treated \( NF1^{fl/fl};Dhh-Cre \) mouse neurofibromas was calculated using the ΔΔCt method.

**Statistics.** For gene expression microarray data analyses, ANOVA tests were used to identify transcripts differentially expressed; all tests were corrected for multiple testing effects by applying the Benjamini and Hochberg false discovery rate correction. Significance was accepted at \( P < 0.05 \) for all biological experiments. MPNST survival analyses used Kaplan Meier analysis followed by a Gehan-Breslow-Wilcox log-rank test. Neurofibromatosis growth was modeled by Mixed Effects Model Analysis; we generated \( P \) values with a random effects model analysis on log transformed tumor volume data using the SAS mixed procedure. Proliferation, cell death, and blood vessel quantification used 2-tailed Student’s t test.

**Study approval.** Human paraffin-embedded tissues were collected under Cincinnati Children’s Hospital Medical Center (CCHMC) IRB approval. Informed consent was not required, as we used archival samples from a tissue bank that remained anonymous. The CCHMC Animal Use and Care Committee approved all animal use.

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