Alteration of hypothalamic cellular dynamics in obesity

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The number of people who suffer from obesity and one or more of its adverse complications is rapidly increasing. It is becoming clear that diet, exercise, and other lifestyle modifications are insufficient strategies to combat this growing problem. Greater understanding of the mechanisms controlling our desire to feed and our ability to balance energy intake with energy expenditure are key to the development of pharmacological approaches for treating obesity. Although great strides have been made in our understanding of how the hypothalamus regulates feeding and energy balance, much less is known about how obesity affects the structure of the hypothalamus. The authors of two papers in this issue of the JCI have addressed this issue by examining the effects of obesity on neurons and glia in the hypothalamus. These studies reveal that obesity may be in part due to hypothalamic injury, which leads to inflammation and reduced neurogenesis. These findings support the notion that obesity is a disease that affects multiple organs, including the brain, and that disruption of normal brain function leads to abnormal regulation of peripheral metabolism.

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Introduction

As the number of people who are obese continues to rise so does the number of cases of obesity-associated morbidities, including type 2 diabetes, hypertension, heart disease, sleep apnea, and cancer. Diet, exercise, and other lifestyle modifications are important approaches to treating obesity. However, it is becoming clear that pharmacological approaches are also needed to combat this major healthcare problem. Thus far, pharmacological approaches to treating obesity have shown limited efficacy in the clinic and have been associated with serious side effects. This is largely because our understanding of the complex biology of obesity is incomplete. For example, while great strides have been made in our understanding of how the brain controls our desire to feed, as well as the processes underlying the balancing of energy intake and expenditure, little is known about how the structure and organization of the brain are altered by obesity.

Central regulation of peripheral metabolism

The role of the hypothalamus in the regulation of feeding and energy balance was first highlighted by lesion studies in rodents (1, 2). Although these classic studies proposing the existence of “feeding” and “satiety” centers in the hypothalamus lacked anatomic precision and were overly simplistic in their interpretation, the importance of the hypothalamus in the regulation of feeding and energy balance was subsequently highlighted by the discovery that hormone and peptide regulators of feeding and metabolism act on the hypothalamus as well as the brainstem and other areas of the brain (3). The adipose tissue–derived hormone leptin exerts its inhibitory effects on food intake by modulating the function of neurons in the arcuate nucleus (an aggregation of neurons in the medial basal region of the hypothalamus). Specifically, leptin suppresses neurons that release neuropeptide Y (NPY) and agouti-related peptide—neuropeptides that normally increase appetite—while stimulating neurons that release proopiomelanocortin (POMC) —the precursor of several neuropeptides, including some that suppress appetite (3). Leptin, ghrelin, and other hormones that control energy metabolism also affect synaptic plasticity—the structural connections of neurons in the hypothalamus and other areas of the brain (4–7).

A hallmark of obesity is the ability of adipose tissue to expand and undergo significant remodeling in order to fulfill its role as the major energy-storing organ. This involves coordinated responses among various cell types, including adipocyte precursors, blood vessels, and immune cells (8). Inflammation in obese adipose tissue is related to adipocyte death, accumulation of macrophages and other immune cells, and metabolic dysfunction (8). Recent stud-
Obesity and hypothalamic neurogenesis

Studies performed by Jeffrey Flier and colleagues have shown that treatment with ciliary neurotrophic factor (CNTF), a protein known to reduce body weight in obese animals and humans (14, 15), enhances neurogenesis (a process by which new neurons are formed) in the hypothalamus of adult mice (16, 17). Interestingly, the new neurons induced by CNTF express neuropeptides implicated in the regulation of feeding and energy balance (NPY and POMC) (16). In a follow-up study, in this issue of the *JCI*, McNay, Flier, and colleagues labeled neurons in the developing hypothalamus and studied their dynamics in lean mice compared with those in obese mice (Figure 1 and ref. 12). In normal mice fed regular (low-fat) chow, most neurons were “born” between embryonic days 10.5 and 12.5. Although the number of new neurons decreased dramatically (by 63%) after weaning, the total number of cells within the arcuate nucleus remained constant, indicating that adult neurogenesis was responsible for replacing a sizeable proportion of neurons in the arcuate nucleus. This dynamic remodeling of the arcuate nucleus included the turnover of both POMC and NPY neurons (Figure 1).

Diet-induced obesity (DIO) was associated with suppression of the number of actively proliferating cells; the affected cells were then determined to be progenitor-like cells rather than neural stem-like cells. The reduction in the number of actively proliferating progenitor-like cells was linked to an increased rate of apoptosis of newly divided cells. Of note, caloric restriction in mice with DIO restored the number of actively proliferating progenitor-like cells to that seen prior to the onset of DIO.

The authors then investigated whether hypothalamic neurogenesis was similarly altered in leptin-deficient ob/ob mice, which display overeating and early-onset obesity (4). They found that the number of stem cells, the small pool of cells that gives rise to actively proliferating progenitor-like cells, was drastically reduced in the hypothalamus of ob/ob mice (12). Treating normal mice with leptin also seemed to increase the number of hypothalamic stem cells. This latter finding is consistent with the known stimulatory effects of leptin on neurogenesis in the hippocampus and the development of feeding circuits (5, 18). The results generated by McNay et al. using ob/ob mice further demonstrate that obesity affects the capacity for renewal of neurons in the arcuate nucleus (12); however, it remains unclear whether hypothalamic neurogenesis is directly regulated by leptin or whether indirect effects via alterations in the levels of glucocorticoids or insulin or various other metabolic defects impact hypothalamic neurogenesis in ob/ob mice (4).

Feeding the flames of inflammation in obesity

Obesity results in chronic low-grade inflammation of several organs, including the liver, muscle, and adipose tissue, as well as the hypothalamus (8, 10). Inflammation is thought to exacerbate metabolic abnormalities by disrupting homeostatic pathways within target tissues (8, 10). For example, infiltration of adipose tissue by macrophages has been linked to the development of insulin resistance (8, 10). Inflammation is also thought to be increased in the hypothalamus of obese individuals.
rodents and may promote food intake and hypertension (9–11, 19). Although inflammation in the hypothalamus is thought to be harmful to neuronal function, little is known about the effects of a high-fat diet (HFD) on glia—nonneuronal cells that are more abundant in the central nervous system than neurons, provide support and nutrition to neurons (astrocytes), and mediate inflammation in the brain (microglia). Moreover, it is not clear how closely hypothalamic inflammation mirrors peripheral inflammation. In their study in this issue of the JCI, Thaler et al. found some interesting differences (Figure 1 and ref. 13). In liver and adipose tissue, inflammatory markers increase in number and magnitude slowly over the course of weeks to months in rats fed HFD (8, 10). In contrast, Thaler et al. found that inflammatory gene expression increased in the arcuate nucleus of rats within one day of initiating a HFD (13). This initial spike in inflammation in the hypothalamus occurred in the absence of weight gain and coincided with increased gliosis, the process whereby astrocytes and microglia are activated and proliferate in response to a brain insult, as measured by the expression of microglial and astrocytic genes (Figure 1). The gene expression patterns were corroborated by immunohistochemistry, which showed an increase in the number and size of reactive glia. However, the rise in inflammatory gene expression and gliosis subsided within a week and reappeared only after continued prolonged exposure to HFD. This biphasic inflammatory response to a HFD was unique to the arcuate nucleus; it was not observed in the liver, adipose tissue, and other regions of the brain. Similar patterns of inflammation and gliosis were also observed in mice fed a HFD.

What are the consequences of arcuate nucleus inflammation and gliosis in rodents fed a HFD? Thaler et al. looked for evidence of neuronal injury, particularly in POMC neurons within the arcuate nucleus (13). At seven days after a HFD was initiated, they found that many POMC neurons displayed an increase in expression of Hsp70, a chaperone protein that is induced in response to various stressors and neuronal injury. There was also evidence of increased autophagy within POMC neurons after 20 weeks on a HFD. After 8 months on a HFD, the number of POMC neurons within the arcuate nucleus was reduced by 25%.

The rodent studies performed by Thaler et al. (13) provide a time line of injury to the arcuate nucleus in response to a HFD. In particular, the early induction of inflammation and reactive gliosis, together with evidence of neuronal stress within one week, suggest that the arcuate nucleus is injured acutely and that perhaps this early insult may be linked to the development of an abnormal energy balance and obesity. Thaler et al. then extended these findings by studying brain images in patients (13). They found that body mass index, a measure of fat content, was proportional to T2 signal in the hypothalamus by MRI. T2-weighted MRI is particularly sensitive for identifying brain lesions, including inflammation, gliosis, and edema. This finding raises the possibility that obesity increases hypothalamic gliosis (20), but further studies in humans are required to ascertain whether this is indeed the case and whether hypothalamic inflammation and injury occur in the arcuate nucleus, as in rodents.

A long way to go
What are the mechanisms underlying these two interesting results showing that obesity is associated with suppression of hypothalamic neurogenesis (12) and increased hypothalamic injury and gliosis (13)? With regards to the effects of obesity on hypothalamic neurogenesis (12), it appears that two distinct pools of cells, actively proliferating progenitor-like and stem-like cells, are inhibited in the hypothalamus of mice with DIO and ob/ob mice, respectively. The extent to which leptin or other metabolic signals regulate neurogenesis in mice with DIO or ob/ob mice is unknown. With regards to the hypothalamic inflammation (13), the data demonstrate that an initial rise in inflammatory markers is triggered by the HFD and not obesity. Rodents respond acutely to a HFD by becoming hyperphagic and consuming more calories than animals fed regular chow. It is possible that any number of factors that are altered by excessive consumption of a HFD (e.g., lipids, leptin, insulin, and ghrelin) contribute to hypothalamic inflammation, neuronal injury, and gliosis. These factors could directly impact neurons and/or glia or indirectly affect the structures and functions of these cells by changing the transport of circulating factors across the blood-brain barrier.

In summary, the studies by McNay et al. (12) and Thaler et al. (13) demonstrate that obesity is associated with profound changes in the structure of neurons and glia in the hypothalamus. Unlike the earlier hypothalamic lesion studies that were shown to cause extensive damage to neurons within and outside the hypothalamus (1, 2), the work of McNay et al. and Thaler et al. suggests that the consumption of a HFD results in acute hypothalamic injury, followed by chronic changes in the structure and organization of neurons and glia. However, there is no evidence that these changes in hypothalamic structure are responsible for the onset and progression of obesity and associated metabolic disorders. Understanding how the intake of a HFD leads to injury and remodeling of the hypothalamus and whether this affects the development of obesity may provide novel insights into the pathogenesis of obesity. Furthermore, it is crucial to investigate whether hypothalamic neurogenesis, inflammation, and gliosis are reversible after weight loss in rodents and humans; whether the dynamics of neurons and glia are causally related to the failure to sustain weight reduction after dieting; and whether the structural changes in the hypothalamus could be targeted in the treatment of obesity and related diseases.

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5. Bouter SG, Draper SJ, Simerly RB. Trophic action of leptin on hypothalamic neurons that regulate
Human adenoviral vectors are being developed for use in candidate vaccines for HIV-1 and other pathogens. However, this approach suffered a setback when an HIV-1 vaccine using an adenovirus type 5 (Ad5) vector failed to reduce, and might even have increased, the rate of HIV infection in men who were uncircumcised and who had preexisting antibodies specific for Ad5. This increased interest in the evaluation of serologically distinct adenoviral vectors. In this issue of the JCI, Frahm and coworkers report evidence that preexisting cellular immune responses directed toward Ad5 reduce the immunogenicity of antigens expressed in Ad5 vectored vaccines and have cross-reacting potential with non-Ad5 adenoviral vectors. The implications of this observation need to be carefully evaluated in future clinical trials of all serotypes of adenovirus-vectored vaccines.

The joint United Nations Program on HIV/AIDS (UNAIDS) estimates that more than 33 million people were living with HIV at the end of 2009 (1). As witnessed by a series of watershed results in the HIV prevention field of late, some progress has been made toward tackling this pandemic. Adult male circumcision (2–4), antiretroviral-based vaginal microbicides (5), preexposure prophylaxis (6), and antiretroviral therapy as a means of prevention (7) have shown varying means of prevention (7) have shown varying means of prevention. For the prevention of HIV infection: subunit vaccines, recombinant virus–vector vaccines, and DNA vaccines. Three of the most significant clinical trials conducted thus far tested the efficacy of different combinations of the first two types of vaccine (9–11). When the first vaccine type was tested in two phase III clinical trials, it was disappointingly found that vaccination with repeated doses of gp120 protein subunit vaccines (AIDSVAX B/B’ and B/E) failed to protect either men who have sex with men (MSM) (10) or injection drug users (11) from HIV infection. Further disappointment came in the form of the results of the phase III Step study (9), which showed that three injections of a Merck-developed adenovirus serotype 5 (Ad5) vector containing gag/pol/nf/HIV-1 gene inserts (referred to as the MRKAd5/HIV-1 vaccine) also failed to protect MSM. In contrast to these results, the RV144 trial demonstrated that an HIV vaccine might be possible (12). The approach tested in this trial was to first prime participants with four injections of a canarypox vector containing gag/pro/gp120/gp41 HIV-1 gene inserts (ALVAC-HIV) and then boost with two injections of a gp120 protein subunit vaccine (AIDSVAX B/E), and it reduced the rate of HIV infection in a low-incidence population of Thais with heterosexual HIV transmission risk.

The Step study raised the added concern that, following vaccination, uncircumcised MSM with naturally acquired immunity to Ad5, in the guise of preexisting Ad5-specific neutralizing antibodies (nAbs), appeared to experience a transient period of increased risk of infection with HIV (11). These concerns led to the cancellation of a planned phase IIB clinical trial of a related Ad5-vectored gag/pol/env vaccine, which was to be used together with a DNA vaccine prime, in diverse risk groups and the generation of a new, smaller efficacy study focusing on circumcised MSM with no serologic evidence of previous Ad5 exposure (14). Additional research has suggested that the immunoge-