Tumor formation constitutes a major obstacle to the clinical application of embryonic stem cell–derived (ESC-derived) cells. In an attempt to find major extracellular signaling and intrinsic factors controlling tumorigenicity and therapeutic function of transplanted ESC-derived retinal progenitor cells (ESC-RPCs), we evaluated multiple kinds of ESC-RPCs in a mouse retinal degeneration model and conducted genome-wide gene expression profiling. We identified canonical WNT signaling as a critical determinant for the tumorigenicity and therapeutic function of ESC-RPCs. The function of WNT signaling is primarily mediated by TCF7, which directly induces expression of Sox2 and Nestin. Inhibition of WNT signaling, overexpression of dominant-negative Tcf7, and silencing Tcf7, Sox2, or Nestin all resulted in drastically reduced tumor formation and substantially improved retinal integration and visual preservation in mice. These results demonstrate that the WNT signaling cascade plays a critical role in modulating the tumorigenicity and functionality of ESC-derived progenitors.
WNT signaling determines tumorigenicity and function of ESC-derived retinal progenitors

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Tumor formation constitutes a major obstacle to the clinical application of embryonic stem cell–derived (ESC-derived) cells. In an attempt to find major extracellular signaling and intrinsic factors controlling tumorigenicity and therapeutic functionality of transplanted ESC-derived retinal progenitor cells (ESC-RPCs), we evaluated multiple kinds of ESC-RPCs in a mouse retinal degeneration model and conducted genome-wide gene expression profiling. We identified canonical WNT signaling as a critical determinant for the tumorigenicity and therapeutic function of ESC-RPCs. The function of WNT signaling is primarily mediated by TCF7, which directly induces expression of Sox2 and Nestin. Inhibition of WNT signaling, overexpression of dominant-negative Tcf7, and silencing Tcf7, Sox2, or Nestin all resulted in drastically reduced tumor formation and substantially improved retinal integration and visual preservation in mice. These results demonstrate that the WNT signaling cascade plays a critical role in modulating the tumorigenicity and functionality of ESC-derived progenitors.

Introduction

The unique properties of embryonic stem cells (ESCs), unlimited self-renewal and pluripotency, make them an attractive cell source for the treatment of various degenerative diseases. However, the same properties also present a major hurdle for their clinical application. Tumor formation has been reported in the transplantation of ESC derivatives despite predifferentiation or presorting (1–6), raising a safety concern for the therapeutic use of ESC-derived cell products in humans. On the other hand, transplantation of photoreceptor precursors in neonatal mice repaired retinal defect efficiently without development of any tumors (7, 8), which emphasizes the critical role of the developmental stage of donor cells in determining the cell fate following transplantation. Thus, steering ESCs to an appropriate state could be an important step for safe and effective cell therapies.

To date, ESCs from the mouse, monkey, and human have been successfully differentiated into retinal cells in vitro (9–13). Sasai’s group developed an efficient induction of retinal precursors by culturing mouse ESCs under a serum-free suspension condition (SFEB culture) and obtained high percentages of differentiated cells expressing key eye-field transcription factors (12–14). However, mechanisms governing efficient generation of various types of retinal cells and the optical cups from ESCs in vitro as well as their application potential in vivo, in regards to the functional integration and safety, are not clearly elucidated.

In an attempt to find major extracellular signaling and intrinsic factors controlling tumorigenicity and therapeutic effects of ocular transplanted ESC-derived retinal progenitor cells (ESC-RPCs), we identified the canonical WNT signaling-activated TCF7-Sox2-Nestin cascade as a critical determinant for the consequence of ESC-RPC transplantation: whether tumors would form as well as whether successful integration into the host retina and prevention of the visual defect would be achieved. Canonical WNT signaling is known to play a key role in cell fate determination for various cell lineages, and its inappropriate activation is frequently associated with cancers (15–17). It has also been shown to promote proliferation of isolated retinal stem cells (18) and retina regeneration in adult mammals (19). However, the correlation between WNT signaling in ESC-derived donor cells and their therapeutic effect as well as tumorigenicity after transplantation in a disease model remains unexamined. In addition, factor(s) that mediate the function of WNT signaling in the control of cell fate commitment remain elusive. In this study, we link the activity of WNT signaling to the tumorigenic potential of ESC-RPCs and provide the experimental evidence for transcriptional factor TCF7 to regulate expression of Sox2 and Nestin, two important genes actively engaged in the neural development and tumor formation. The tumorigenic and therapeutic effect of transplanted ESC-RPCs is determined in a well-studied sodium iodate–induced (SI-induced) mouse retinal degeneration model (20, 21). We also show that the expression of TCF7, Sox2, and Nestin is closely associated in mouse neonatal retinae. These findings open new avenues to define and manipulate ESC-derived donor cells prior to transplantation for safe and effective cell therapies.

Results

ESC-RPCs but not primary retinal progenitor cells produce neural tumors in transplanted eyes. We began with differentiation of mouse ESCs (46C line containing a Sox1.EGFP–targeted gene) (22, 23) into retinal progenitor cells (RPCs) using a chemically defined SFE-
B-based protocol modified from a previously reported SFEB-based approach (ref. 12, Supplemental Figure 1, A–C, and Supplemental Methods; supplemental material available online with this article; doi:10.1172/JCI65048DS1). According to this protocol, ESC-RPCs were obtained by selection of SOX1.EGFP+ neural progenitors through FACS at day 7 of differentiation. Successfully differentiated cells were characterized by 3 independent approaches. Immunofluorescence staining showed that they expressed multiple neural progenitor and retinal markers, such as nestin, PAX6, SIX3, and RAX for RPCs at an early stage of differentiation (day 14), OTX2 and NRL for photoreceptors, and rhodopsin (OPsin) for rod photoreceptors at the end of differentiation (day 24) (Figure 1A). Hereafter, ESC-derived RPCs at day 24 were termed ESC-RPCs. Then, transcript levels of various markers among ESCs, ESC-RPCs, and primary RPCs (P-RPCs) isolated from neonatal mice at P1 were compared. Results of RT-PCR assays revealed that pluripotency-associated genes (OCT4, Nanog, and Rex1) were not detectable in ESC-RPCs and P-RPCs (data not show), whereas markers for various types of retinal cells were found in both ESC-RPCs and P-RPCs (Figure 1B; see Supplemental Table 1 for primers). Western blot analysis confirmed the absence of OCT4 and Nanog proteins in ESC-RPCs and P-RPCs (Supplemental Figure 1D). Moreover, the component of retinal cell types in ESC-RPCs was characterized by flow cytometry (FCM) analysis. The majority of ESC-RPCs were PAX6+. A fraction of ESC-RPCs was RAX+ or OTX2+; and 13.58% of cells expressed Nrl (Figure 1C and Supplemental Figure 1E). Thus, ESC-RPCs were composed of a mixture of neural retinal progenitors and committed photoreceptors.

We then tested the therapeutic potential and risk of tumor formation by injecting 8 × 10⁴ of ESC-RPCs into the subretinal space of SF-E8-treated mice. All transplanted ESC-RPCs (except for D3.EGFP-derived RPCs) in this study were labeled by EGFP (ESC-RPC.EGFP) through infection with a lentiviral vector containing an EGFP sequence at day 24 of differentiation and were injected into mice 4 days later (day 28). The same number of P-RPCs carrying an EGFP transgene (P-RPCs.EGFP) (24) was used as a transplantation control. Three weeks later, tumors were detected in 63 eyes out of 104 ESC-RPC transplanted eyes, which maintained an intact external structure with the enlarged size (Supplemental Figure 1F). Most internal structures were entirely destroyed (Figure 1D). The cells within tumors seemed to have a homogenous morphology, without any distinct tissue structure (Figure 1E), and were EGFP positive (Figure 1F), indicating the origin of exogenous injection. The expression of Sox2, nestin, and PAX6 as well as PCNA was detected in tumor cells, hinting at the existence of immature RPCs (Figure 1G and Supplemental Figure 1G). Furthermore, FACS-isolated EGFP+ tumor cells had the capacity to form neurospheres and a high BrdU incorporation rate (Supplemental Figure 1, H and I). Immunostaining data showed that isolated tumor-derived cells were SOX1+, NESTIN+, and Sox2+ (Supplemental Figure 1J), implicating the neural progenitor nature of tumors. Nevertheless, about 33% of injected ESC-RPCs integrated well without forming tumors. In these eyes, the visual function was partially preserved (see below). This finding argues for the potential of ESC-RPCs to integrate and function in the host retina should tumor formation be prevented. In contrast to ESC-RPCs, strikingly, none of the eyes injected with P-RPCs developed tumors and efficient integration took place in 72 eyes out of 80 transplanted eyes (Table 1).

The visual function of eyes receiving P-RPCs and ESC-RPCs (for tumor-free eyes) was assessed by electroretinography (ERG) analysis, which reflects functional responses of photoreceptors and downstream neurons upon light stimulus (Figure 1H and Supplemental Figure 1K). In line with previous reports, SF-E8 treatment led to a complete loss of ERG response (the sham group; n = 10), as evidenced by the disappearance of scotopic a and b waves, and of oscillatory potentials, the latter representing the blood circulation in the retina and synaptic circuits in the inner nuclear layer. P-RPCs corrected both defects completely (n = 10). These results suggest that satisfactory cell transplantation would be achieved if we could steer ESC-RPCs into a state similar to that of P-RPCs.

**Inhibition of canonical WNT signaling enables ESC-RPCs to have P-RPC-like properties.** The distinct results obtained from the transplantation of ESC-RPCs and P-RPCs prompted us to search for molecular networks distinguishing these 2 types of donor cells. To this end, genome-wide transcript profiles were compared between ESC-RPCs and P-RPCs, with 3 replicates for each cell type. A total

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**Table 1**

Summary of transplantation results of 8 kinds of donor cells used in this study

<table>
<thead>
<tr>
<th>Donor cells</th>
<th>Tumor</th>
<th>Integration</th>
<th>Failure</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>No.</td>
<td>%</td>
<td>No.</td>
</tr>
<tr>
<td>P-RPC</td>
<td>63/104 60.58%</td>
<td>35/104 33.65%</td>
<td>6/104 5.77%</td>
</tr>
<tr>
<td>ESC-RPC</td>
<td>2/62 3.23%</td>
<td>58/62 93.54%</td>
<td>2/62 3.23%</td>
</tr>
<tr>
<td>DKK1-ESC-RPC</td>
<td>6/132 4.55%</td>
<td>102/132 77.27%</td>
<td>24/132 18.18%</td>
</tr>
<tr>
<td>Dkk1-ESC-RPC</td>
<td>6/122 4.92%</td>
<td>79/122 64.75%</td>
<td>37/122 30.33%</td>
</tr>
<tr>
<td>shSox2-ESC-RPC</td>
<td>12/92 13.04%</td>
<td>65/92 70.65%</td>
<td>15/92 16.31%</td>
</tr>
<tr>
<td>shNestin-ESC-RPC</td>
<td>2/36 5.56%</td>
<td>25/36 69.44%</td>
<td>9/36 25%</td>
</tr>
<tr>
<td>ShControl-ESC-RPC</td>
<td>45/60 75%</td>
<td>12/60 20%</td>
<td>3/60 5%</td>
</tr>
</tbody>
</table>

All ESC-RPCs listed in this table were derived from the 46C cell line. No tumors were found in P-RPCs. Failure is defined by the absence of EGFP+ donor cells in injected eyes.
D3EGFP-derived RPCs (green) using antibodies against SOX2, N-Cad. Immunofluorescence staining for sections of eye tumors produced by Sox1 locus NCAM+ neural cells were selected by FACS, respectively (Supplemental population of EGFP+/NCAM+, OCT4–/NCAM+, and SOX1+/NCAM+ neural aggregates were digested to single cell suspension. Then, ESCs were induced to RPCs according to a recently published protocol of Fgf/Ras/Mek signaling and GSK3β to generate ESC-RPCs. These ESC-RPCs, as tumor cells expressed EGFP (Figure 2D). They also expressed SOX2, N-Cadherin, Pax6, Tcf7, and Nestin, revealing the neural nature of tumors (Figure 2E). Remarkably, DKK1 treatment reduced the tumor formation and enhanced integration drastically (Supplemental Table 3). These results are consistent with the transplantation result obtained with 46C ESC-RPCs. Therefore, our data indicate that ESC-RPC, regardless of the ESC source, induction protocol used, and lentiviral infection, possess the potential to generate tumors following ocular transplantation. Inhibition of canonical WNT signal before cell transplantation is an effective strategy to prevent tumor formation.

Moreover, successful integration and functional rescue of degenerative retina were achieved in DKK1-treated ESC-RPC–transplanted (DKK1-ESC-RPC–transplanted) eyes, as indicated by the following lines of experimental evidence. First, transplantation of DKK1-ESC-RPCs markedly rescued Si-induced reduction in the thickness of the whole retina and loss of the structure of the photoreceptor outer segment (OS) (Figure 3A), which are typical features of retinal degeneration. Second, injected DKK1-ESC-RPCs substantially preserved the scotopic ERG responses (n = 10). In comparison, untreated ESC-RPCs only partially preserved the ERG response in the tumor-free eyes (n = 10) (Figure 3D). Collectively, these data indicate that inhibition of WNT signaling enabled ESC-RPCs to have the therapeutic capacity closer to that of P-RPCs and that there was a correlation between higher activities of WNT signaling and tumorigenicity of ESC-RPCs.

We next attempted to find what molecular processes took place within DKK1-ESC-RPCs. QRT-PCR data showed that DKK1 repressed the expression of neural progenitor markers but upregulated committed retinal lineage markers, apart from downregulation of many key components of WNT signaling (Supplemental Figure 3A). Immunostaining also showed that percentages of cells positive to BrdU, PCNA, and SOX2 staining (Figure 3E) were reduced significantly by DKK1 treatment, whereas fractions of cells expressing Map2 or Opsin increased significantly (Supplemental Figure 3B). Clearly, Oct32- or Nrl+ cells and Nestin+ or Brdu+ cells were mutually exclusive, although expression of BrdU and Nestin was found in the same cells (Supplemental Figure 3C), indicating that these Nrl+/BrdU+/Nestin+ cells were fully differentiated postmitotic photoreceptor precursors. The specificity of the antibodies used here was further verified by retinal cross-section immunofluorescence staining of neonatal C57 mice (Supplemental Figure 3D). In addition, FCM analysis revealed reduced percentages of SOX2+ and Pax6+ cells but increased fractions of Rax+ and Crx+ cells (Figure 3F). Therefore, DKK1 treatment induced ESC-RPC into a more P-RPC-like state, which should underline its effect on the functional outcomes of ocular transplantation.

Tcf7 mediates the function of canonical WNT signaling in ESC-RPCs. We then asked which factor(s) mediated the functions of canonical WNT signaling in ESC-RPCs. A high level of full-length Tcf7 (Tcf7) was noticed in ESC-RPCs (Figure 4A) and in eye tumors from ESC-RPC injection (Figure 4B and Supplemental Figure 4A). Moreover, DKK1 treatment reduced the fractions of tcf7+ and Nestin+...
cells in ESC-RPCs significantly (Supplemental Figure 4B). Furthermore, we found that, of TCF/LEF factors, only TCF7 was highly expressed in the developing mouse retina (Supplemental Figure 4, C and D). Based on these observations and considering that β-catenin usually binds TCF/LEF factors to execute functions of canonical WNT signaling, we anticipated that TCF7 might play a critical role in the control of ESC-RPC differentiation and proliferation, although the implication of TCF7 in neural development and retinal lineage commitment was previously not reported. To test this, a naturally truncated form of TCF7 (ΔNTCF7), lacking the N terminus, was used. It produces an inhibitory effect on WNT signaling, as the transcriptional activation activity of TCF/LEF factors is dependent on their association with β-catenin through the N terminus (29). The dominant-negative effect of ΔNTCF7 on canonical WNT signaling was validated by a TOPFlash luciferase assay (Supplemental Figure 4E). At molecular levels, overexpressing ΔNTcf7 in ESC-RPCs (ΔNTcf7-ESC-RPCs) brought about changes resembling those following DKK1 treatment (Figure 4, C and D). At functional levels, engraftment of ΔNTcf7-ESC-RPCs not only reduced the incidence of tumors to 6 out of 132 eyes but also increased the donor cell integration to 102 out of 132 eyes (Table 1). Transplanted ΔNTcf7-ESC-RPCs survived and migrated to multiple layers in the host retina. Typical integration and differentiation into rod photoreceptors were indicated by the observation of OS and IS of rod photoreceptors with EGFP expression inside the host retina (Supplemental Figure 4F). In addition, transplantation also preserved ERG in a manner similar to that of DKK1-ESC-RPCs (Figure 4E). Altogether, antagonizing TCF7’s activity almost phenocopied the effect of DKK1 treatment both at molecular and visual functional levels.

To further define the role of TCF7 in WNT signaling–mediated tumor formation following ESC-RPC transplantation, a specific Tcf7 RNA interference vector (shTcf7) was introduced into ESC-RPCs (Supplemental Figure 4G). Results from FCM analysis indicated that silencing Tcf7 evidently decreased the fraction of SOX2+ and PAX6+ cells but increased the fraction of RAX+; OTX2+; and CRX+ cells, respectively (Figure 5A). Hence, silencing Tcf7 rendered ESC-RPCs more retinal lineage-committed and matured. This conclusion was further supported by transplantation results. Similar to ΔNTcf7-ESC-RPCs, transplantation of shTcf7-treated ESC-RPCs gave rise to 69.44% of successful integration, with tumor formation in 5.56% of injected eyes (Table 1). This result further strengthened the notion that TCF7 could indeed be the primary mediator of WNT-controlled tumorigenicity of ESC-RPCs.

Last, we tested whether overexpression of ΔTcf7 could alter the differentiation state of P-RPCs and induce tumor formation in the mouse retina. Exogenous expression of ΔTcf7 in P-RPCs together with WNT3A treatment enhanced percentages of cells expressing neural progenitor cell (NPC) markers (PCNA, SOX2, and PAX6) substantially, whereas percentages of cells expressing RPC markers (RAX, OTX2, and CRX) decreased markedly (Figure 5B), suggesting that overexpression of ΔTcf7 switched P-RPCs to a state more like that of NPCs. We subsequently injected lentiviral vectors containing either ΔTcf7 or EGFP coding sequence together with WNT3A proteins into the retina of P1 mice. Three weeks after the injection, antibodies against glutamine synthetase (a Müller cell marker) and BrdU were used to carry out immunofluorescence staining of retina sections. Disorganized neural tumors were observed in 90% of eyes injected with ΔTcf7 lentiviruses and WNT3A, whereas there were no neural tumors detected in the eyes injected with EGFP lentiviruses and WNT3A proteins (Figure 5, C and D, and Supplemental Table 4). Therefore, WNT/TCF7 signaling could induce tumor formation in vivo.

**Sox2 and Nestin, direct transcriptional targets of TCF7, control the tumorigenicity and visual repair capability of transplanted ESC-RPCs.**

To investigate the role of SOX2 in controlling the property of ESC-RPCs, its expression was stably silenced by a Sox2-specific RNA interference construct (shSox2) (Supplemental Figure 5E). Immunofluorescence staining and qRT-PCR results showed that silencing of Sox2 significantly reduced the expression of neural progenitor-associated genes but increased the transcription of genes associated with neural retinal lineages (Figure 6C and Supplemental Figure 5, F and G). Notably, the expression of WNT signaling components, such as Ctnnb1, Tcf7, and Axin2, was also repressed to various degrees, implying a possible feedback loop between SOX2 and WNT signaling.
Figure 4

TCF7 mediates functions of WNT signaling in transplanted ESC-RPCs. (A) The comparison of Tcf7 mRNA levels among ESCs, ESC-RPCs, and P-RPCs by qRT-PCR analysis. Values in ESCs were set at 1.0. (B) Protein levels of TCF7 in ESC-RPCEGFP-derived tumors and normal adult retinae were determined by Western blot analysis. (C) The relative expression levels of neural progenitor markers and WNT signaling in ESC-RPCs, ΔNTcf7-ESC-RPCs, and P-RPCs were determined by qRT-PCR. Values in empty vector–infected ESC-RPCs were set at 1.0. (D) Effects of ΔNTcf7 on the expression of neural progenitor and proliferation markers in ESC-RPCs were analyzed. Bar charts show the quantitative analysis. Scale bar: 50 μm. (E) ERG results for eyes grafted with empty vector–infected ESC-RPCs, ΔNTcf7-ESC-RPCs, and P-RPCs. n = 10 for each group. Data are shown as mean ± SD; ANOVA (A, C, and E), t test (D); *P < 0.05, **P < 0.01.
Figure 5
The expression level of TCF7 is directly linked to the state of ESC-RPCs and eye tumor formation in vivo. (A) FCM analysis to compare the percentage of ESC-RPCs expressing SOX2, PAX6, RAX, OTX2, and CRX, respectively, between shControl and shTcf7-ESC-RPCs. (B) FCM analysis to compare the percentage of P-RPCs expressing PCNA, SOX2, PAX6, RAX, OTX2, and CRX, respectively, between vector-infected and fTcf7-infected P-RPCs in the presence of WNT3A protein. (C and D) Immunofluorescence staining with antibodies against glutamine synthetase (GS) and BrdU in retinæ injected with lentiviral fTCF7 and WNT3A protein. Retinæ injected with lentiviral EGFP plus recombination WNT3A protein and the medium were used as controls. Scale bar: 50 μm (C); 250 μm (D).
SOX2 and NESTIN act as direct targets of TCF7, determining functional consequences of ESC-RPC transplantation. (A) ChiP assays were performed using rabbit IgG or the specific antibody against TCF7 in ESC-RPCs. The locus of DHFR was used as a negative control. (B) TCF7 and β-catenin cooperatively transactivated the SOX2 promoter. (C) Effects of shSOX2 on immunostaining for proliferation and neural progenitor markers in ESC-RPCs. DAPI is shown in gray. Bar charts show the quantitative analysis of the corresponding staining. (D) ChiP assay results are shown. (E) The effect and interaction of TCF7, SOX2, and β-catenin on the activation of the Nestin promoter was examined using luciferase reporter assays. (F) ERG analysis of Si-treated mice after transplantation with shSOX2- and shNESTIN-ESC-RPCsEGFP, n = 10 for each group. Data are shown as mean ± SD; t test (A, C, and D), ANOVA (B, E, and F). **P < 0.01. Scale bar: 50 μm (C).

In addition, we found 2 evolutionarily conserved TCF/LEF binding sites in the second intron of the Nestin gene (Supplemental Figure 6A). NESTIN was previously reported as a direct target of SOX2 (31, 32) and has been recently suggested as a potential marker for cancer stem cells, particularly for cancers of neuroectodermal origin (33). Data from ChiP, EMSA, and reporter assays all demonstrated that TCF7 positively regulated Nestin expression through direct association with Nestin’s second intron (Figure 6D and Supplemental Figure 6, B and C). Further reporter assays showed that WNT3A and LiCl also activated Nestin expression (Supplemental Figure 6D). A stronger activation of the Nestin reporter was detected when SOX2 was coexpressed with β-catenin or TCF7. In contrast, the effect of SOX2 on activation of the Nestin reporter was blocked by ΔNTCF7, and the activation effect of TCF7 was abolished by shSOX2 expression (Figure 6E), revealing a functional link between SOX2 and TCF7/β-catenin complexes. To verify the interaction among TCF7, β-catenin, and SOX2 proteins, coimmunoprecipitation and coimmunofluorescence staining assays were carried out. Results from both assays supported the notion that TCF7, β-catenin, and SOX2 form protein complexes to collaboratively regulate Nestin expression (Supplemental Figure 6, E and F).

The above results established the transcriptional regulation links among TCF7/β-catenin, SOX2, and NESTIN. We next determined whether SOX2 and NESTIN play a role in the control of tumor formation and retina repair. To this end, ESC-RPCs transfected with a Sox or Nestin-specific RNA interference construct (shNestin) were transplanted into the same mouse model, respectively, as described above. Like in the case of ΔNTcf7-ESC-RPCs, markedly higher rates of donor-cell integration (65% and 70% for shSox2 and shNestin, respectively) and lower incidence of tumor formation (5% and 13% for shSox2 and shNestin, respectively) were obtained as compared with control ESC-RPCs transfected with a control RNA interference construct (shControl) (Table 1). Functionally, ERG analysis revealed improved b wave amplitudes in the eyes in which shSox2 or shNestin-ESC-RPCs integrated successfully, while no measurable response was detected in mice of the sham group (Figure 6F).

Discussion

In this study, through functional evaluation of multiple kinds of ESC-RPCs in comparison with P-RPCs in a retinal degeneration model (Table 1) and genome-wide transcriptome analyses, we report that the sustained activation of the canonical WNT signaling-coupled TCF7/β-catenin-SOX2-NESTIN cascade contributed to tumorigenesis in the ocular transplantation of ESC-RPCs. Inhibition of the cascade in ESC-RPCs prior to transplantation gave rise to a drastic reduction in tumor formation and substantial improvement in visual preservation after transplantation in the mouse retinal degeneration model (Figure 7C). This finding is of particular significance, given that tumor formation is one of the major obstacles for clinical application of ESC-derived cells, and emphasizes the importance of controlling and monitoring donor cell status for successful transplantation.

Another impact of this study is the uncovering of a crucial role for TCF7 in neural and eye development. TCF7 is best known for its function in T cell development (35). Our data discovered its function in neural and eye development. TCF7 is best known for its function in T cell development (35). Our data discovered its function in neural and eye development. TCF7 is best known for its function in T cell development (35). Our data discovered its function in neural and eye development. TCF7 is best known for its function in T cell development (35). Our data discovered its function in neural and eye development. TCF7 is best known for its function in T cell development (35). Our data discovered its function in neural and eye development. TCF7 is best known for its function in T cell development (35). Our data discovered its function in neural and eye development. TCF7 is best known for its function in T cell development (35). Our data discovered its function in neural and eye development. TCF7 is best known for its function in T cell development (35). Our data discovered its function in neural and eye development. TCF7 is best known for its function in T cell development (35). Our data discovered its function in neural and eye development. TCF7 is best known for its function in T cell development (35). Our data discovered its function in neural and eye development. TCF7 is best known for its function in T cell development (35). Our data discovered its function in neural and eye development. TCF7 is best known for its function in T cell development (35). Our data discovered its function in neural and eye development. TCF7 is best known for its function in T cell development (35). Our data discovered its function in neural and eye development.
Figure 7
Canonical WNT signaling does not only define the cell fate of transplanted ESC-derived retinal progenitors but is also closely associated with early development of the mouse retina. (A) Immunofluorescence staining for the expression of TCF7 and SOX2 in the retina of a Nestin-EGFP transgenic mouse from P1 to P30. DAPI is shown in gray. The asterisks indicate the nonspecific staining. ONBL, outer nuclear layer. Scale bar: 50 μm. (B) Representative Western blots of β-catenin, TCF7, TCF7L2, SOX2, NESTIN, and PCNA proteins in mouse retina from E14 to adult. (C) The proposed model explains how canonical WNT signaling determines the proliferation and differentiation state of ESC-derived progenitors via TCF7 to regulate the expression of Sox2 and Nestin in vitro, controlling tumorigenesis and functional visual preservation in a disease model. The potential targets to block the pathway are indicated: β-cat, β-catenin.

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survival and differentiation into photoreceptors of donor cells, as indicated by the expression of rod photoreceptor markers in EGFP donor cells, although the integrated cell number was small. Previous studies reported that even a relatively small number of transplanted retinal cells can be sufficient to rescue visual function and that some vision is retained in patients having few photoreceptors (37). On the other hand, the impact of transplanted cells on the remaining host retinal cells should play a role as well, since we observed significant recovery in the thickness of different layers of host retinae, even for the layer in which few donor cells could be detected. Secreted soluble factors, which have previously been implicated in the mechanism for visual rescue by human NPCs (38), could be partially responsible for the improvement of retinal functions. Meanwhile, excessive proliferation or dedifferentiation of immature donor cells due to their response to local growth signals might also contribute to tumor formation after transplantation. The precise mechanisms underlying the morphological and functional recovery as well as the development of neural tumors following ocular transplantation of ESC-RPCs warrant further investigation. Overall, this study provides the proof of principle for optimizing the activity of extracellular signaling and related intrinsic factors prior to donor cell transplantation to prevent tumor formation and improve therapeutic effects.

Methods
SI-induced retinal degeneration model and subretinal transplant surgery. Experimental C57BL/6 mice at 8 weeks of age were used to induce a retinal degeneration model by a single intravenous injection of 30 mg/kg SI (Sigma-Aldrich) (20). Injected cells were suspended in the GMEM at the density of 8 x 10^6/μl (Gibco). Then, cell suspensions or enriched lentivirus (1 μl) were transferred into subretinal space in a manner similar to the one previously described (7).

ERG examination. Corneal ERG recordings from both eyes of mice were obtained 3 weeks after cell transplantation. ERG recordings were performed with an AVES system (Kanghua Rui Ming Technology Co. Ltd.) following the procedures described previously (39).

Microarray analyses. RNA samples from 3 independent experiments were hybridized to a whole mouse gene expression microarray (Affymetrix Mouse 430 2.0) following the manufacturer’s instruction. For each sample, the background was removed, and data were normalized in accordance with MAS 5.0. A hierarchical clustering of samples was performed using Cluster 3.0 software (Michael Eisen, Stanford University). Heat maps were generated using Java Treeview software (http://jtreeview.sourceforge.net; under GPL).

Accession number. Microarray data are accessible at the GEO database under accession number GSE34002.

Statistics. Data were pooled from at least 3 independent sets of experiments, unless otherwise indicated, and are presented as the mean ± SD. All statistical analyses were carried out using SPSS 11 (SPSS). Comparisons of mean values were analyzed by Student’s t test (2 sided) or 1-way ANOVA followed by the post-hoc Dunnett’s test for experiments with more than 2 groups (Levene’s tests for equal variance). Dunnett’s T3 test was used as post-hoc test comparison for the analysis of unequal variances (Welch’s and Brown-Forsythe’s test). Differences were considered statistically significant at P < 0.05. Graphing was performed using SigmaPlot software.

Study approval. All animal procedures and experiments were approved by the Shanghai Jiao Tong University School of Medicine Animal Care Committees. Animals were cared for in accordance with the Association for Research for Vision and Ophthalmology statement for the use of Animals in Ophthalmic and Vision Research.

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