IL-10–producing NKT10 cells are a distinct regulatory invariant NKT cell subset

Duygu Sag, … , Mitchell Kronenberg, Gerhard Wingender


Invariant natural killer T (iNKT) cells rapidly produce copious amounts of multiple cytokines after activation, thereby impacting a wide variety of different immune reactions. However, strong activation of iNKT cells with α-galactosylceramide (αGalCer) reportedly induces a hyporeactive state that resembles anergy. In contrast, we determined here that iNKT cells from mice pretreated with αGalCer retain cytotoxic activity and maintain the ability to respond to TCR-dependent as well as TCR-independent cytokine-mediated stimulation. Additionally, αGalCer-pretreated iNKT cells acquired characteristics of regulatory cells, including production and secretion of the immunomodulatory cytokine IL-10. Through the production of IL-10, αGalCer-pretreated iNKT cells impaired antitumor responses and reduced disease in experimental autoimmune encephalomyelitis, a mouse model of autoimmune disease. Furthermore, a subset of iNKT cells with a similar inhibitory phenotype and function were present in mice not exposed to αGalCer and were enriched in mouse adipose tissue and detectable in human PBMCs. These data demonstrate that IL-10–producing iNKT cells with regulatory potential (NKT10 cells) represent a distinct iNKT cell subset.

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IL-10–producing NKT10 cells are a distinct regulatory invariant NKT cell subset

Duygu Sag,1 Petra Krause,2 Catherine C. Hedrick,1 Mitchell Kronenberg,2 and Gerhard Wingender2

1Division of Inflammation Biology and 2Division of Developmental Immunology, La Jolla Institute for Allergy and Immunology, La Jolla, California, USA.

In contrast, we determined here that NKT cells from mice pretreated with αGalCer retain cytotoxic activity and maintain the ability to respond to TCR-dependent as well as TCR-independent cytokine-mediated stimulation. Additionally, αGalCer-pretreated NKT cells acquired characteristics of regulatory cells, including production and secretion of the immunomodulatory cytokine IL-10. Through the production of IL-10, αGalCer-pretreated NKT cells impaired antitumor responses and reduced disease in experimental autoimmune encephalomyelitis, a mouse model of autoimmune disease. Furthermore, a subset of NKT cells with a similar inhibitory phenotype and function were present in mice not exposed to αGalCer and were enriched in mouse adipose tissue and detectable in human PBMCs. These data demonstrate that IL-10–producing NKT cells with regulatory potential (NKT10 cells) represent a distinct NKT cell subset.

Results
αGalCer-pretreated NKT cells display an increased frequency of proliferating cells. αGalCer-pretreated NKT cells have been shown to have a long-term reduction in proliferation and proinflammatory cytokine production upon antigen rechallenge (9, 10, 12), which has been interpreted as a general loss of effector functions. However, when we analyzed the expression of Ki67 in NKT cells as a surrogate marker for proliferation on a single-cell level, the proportion of proliferating NKT cells at steady state, when analyzed ex vivo, was actually higher in animals pretreated with αGalCer 1 month earlier (Figure 1, A and B). Although Ki67 is expressed in all active phases of the cell cycle (G1, S, G2, and mitosis), it only marks cells with the potential to divide (13), and anergic T lymphocytes have been reported to be arrested in the G1 phase of the cell cycle (14). Therefore, we performed a detailed cell-cycle analysis and measured proliferation directly by BrdU incorporation. Congruent with the Ki67 data, significantly more αGalCer-pretreated NKT cells incorporated BrdU than did control NKT cells (Figure 1, A–C), and significantly more cells were detected in the S phase of the cell cycle (Figure 1, C and D). However, the ratio of cells in the early S phase compared with that in the late S phase was similar in control and αGalCer-pretreated NKT cells (Figure 1E), suggesting that the cell-cycle progression of αGalCer-pretreated NKT cells was undisturbed. We also performed a genome-wide gene expression analysis of untreated and αGalCer-pretreated NKT cells using microarrays. In agreement with the increased proliferation observed by flow cytometry, genes that promote transition through the cell cycle were upregulated in αGalCer-pretreated NKT cells, including genes that are required for cells to transit beyond the G1 phase, such as cyclin A, B, and E (Supplemental Figure 1A; supplemental material available online with this article; doi:10.1172/JCI72308DS1). Additionally, genes for cell-cycle inhibitors were not expressed or did not change consistently

Introduction
Invariant natural killer T (iNKT) cells are a unique subset of T lymphocytes characterized by an invariant Vα14 to Jα18 TCR rearrangement (Vα14i) in mice and an orthologous rearrangement (Vα24i) in humans. iNKT cells recognize glycolipids presented by CD1d, a nonpolymorphic MHC class I homolog. After activation, iNKT cells rapidly produce copious amounts of Th1, Th2, and Th17 cytokines, impacting a dazzling variety of different immune reactions, including responses to pathogens and tumors (1–5). The synthetic model antigen α-galactosylceramide (αGalCer), a potent agonist of mouse and human NKT cells, has been tested in clinical trials for the treatment of cancer patients, and it is under continuing development as a therapeutic agent (1–7). The influence of αGalCer-pretreated αGalCer is thought to lead to a general hyporesponsive state of NKT cells. Here, we report that the strong activation mediated by αGalCer does not lead to anergy or a complete unresponsiveness of iNKT cells, but rather induces a novel state characterized by IL-10 production. Importantly, a subset of iNKT cells with a similar phenotype and function were detected even in untreated mice and in PBMCs from healthy donors, indicating that iNKT cells with a similar function are present in vivo under steady-state conditions. This indicates that regulatory NKT cells occur naturally and that they represent a novel subset that we term NKT10 cells.

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in αGalCer-pretreated iNKT cells (Supplemental Figure 1B). Therefore, whereas anergic T cells show impaired proliferation (15), our data demonstrate that αGalCer-pretreated iNKT cells display increased steady-state proliferation.

αGalCer-pretreated iNKT cells retain antigen-specific cytotoxicity. Effector functions of iNKT cells are not limited to cytokine production, as activated iNKT cells display potent cytotoxic activity (16). To test whether αGalCer pretreatment would affect the cytotoxic potential of iNKT cells, we measured the in vivo cytotoxic activity of naive and αGalCer-pretreated mice. As shown in Figure 2A, the αGalCer-specific in vivo cytotoxicity tended to be lower in αGalCer-pretreated mice compared with that in control animals, but this decrease did not reach statistical significance in the majority (6 of 10) of experiments. iNKT cell thymic emigrants that reached the periphery after the injected αGalCer was cleared from the circulation could account for the cytotoxicity. To exclude this, we repeated the in vivo cytotoxicity assay with animals that had been thymectomized before the injection of αGalCer. However, we observed no significant difference between the wild-type and thymectomized groups (Figure 2A), indicating that recent thymic emigrants (RTEs) cannot account for the remaining cytotoxicity in the αGalCer-pretreated animals.

We also noted significantly lower frequencies of splenic and liver iNKT cells in the αGalCer-pretreated mice compared with those in control animals (Figure 2A and data not shown), suggesting that the small decrease in the in vivo cytotoxicity could be a consequence of lower numbers of effector iNKT cells rather than a reduction in their intrinsic functionality.

It could be argued that the iNKT cell population consists of a cytotoxic subset that is unaffected by previous αGalCer treatment and a cytokine-producing subset that can be anergized by αGalCer. To distinguish between these options, we used a single-cell assay by measuring CD107α and CD107β externalization as an indicator of iNKT cell degranulation (17). To this end, we stimulated splenocytes from control and αGalCer-pretreated mice with plate-bound αCD3ε antibody and analyzed the binding of αCD107α/b antibodies by iNKT cells. We observed no difference in the frequency or intensity of iNKT cells that bound CD107α and CD107β between the control and αGalCer-pretreated splenocytes (Figure 2, B and C), indicating that the degranulation of iNKT cells is comparable between wild-type and αGalCer-pretreated iNKT cells.

Stimulation of iNKT cells with αGalCer can protect mice against lung metastases resulting from challenge with B16 melanoma, and Parekh et al. (9) reported that this protection is lost in αGalCer-pretreated mice. However, B16 melanoma cells do not express CD1d (16), excluding the possibility that the tumor cells serve as direct targets of iNKT cells. Indeed, it is known that the cytotoxicity against B16 in this model is dependent on NK cell trans-activation, induced in part by iNKT cell–derived IFN-γ (18, 19). We previously showed, however, that B16 stably transfected with CD1d (B16-CD1d) and loaded with αGalCer can be a cognate target for iNKT cells, in which case the tumor cells are eliminated efficiently in vivo in an antigen-specific manner (16). We used the B16-CD1d cell line to directly address whether αGalCer-pretreated iNKT cells could protect mice against this melanoma. In line with previous data (9), we found that administration of αGalCer with B16-CD1d tumor challenge could protect mice from metastases (Figure 2D, compare bars 1 and 2), but iNKT cells were not effective in mice pretreated with αGalCer (Figure 2D, bars 4 and 5). This is consistent with the impaired IFN-γ release by iNKT cells previously exposed to αGalCer as reported before (9, 10) and shown below.

Figure 1. αGalCer-pretreated iNKT cells display an increased frequency of proliferating cells. (A and B) Steady-state expression of Ki67 and BrdU (3-day pulse) in splenic iNKT cells from C57BL/6 control (B6, tinted area) or C57BL/6 mice injected 1 month earlier with 4 μg αGalCer (B6/αGC, black line). Representative data (A) and a summary graph (B) are shown. Numbers in the histograms denote the percentage of positive cells within the depicted gate from the indicated mice. (C–E) Expression of DNA (labeled with 7-AAD) and BrdU (3-day pulse) in splenic iNKT cells from C57BL/6 control (B6) or C57BL/6 mice injected 1 month earlier with 4 μg αGalCer (B6/αGC). Representative data (C) and a summary graph (D) are shown. Numbers in C denote the percentages in the respective rectangles. Definition of the cell-cycle stages is given in the left dot plot. M, mitosis. (E) Graphic representation of the frequency of 7-AAD− (S early) and 7-AAD+ (S late) cells, considering BrdU+ cells rather than a reduction in their intrinsic functionality.
In contrast, when αGalCer was loaded directly onto the B16-CD1d cells, which will efficiently elicit cytotoxic function, we observed no difference in tumor protection between control and αGalCer-pretreated mice (Figure 2D, bar 3 versus bar 6). Therefore, we conclude that cytotoxicity induced by cognate antigen recognition is unimpaired in αGalCer-pretreated iNKT cells.

Effective TCR signaling of αGalCer-pretreated iNKT cells. To directly assess the response of αGalCer-pretreated iNKT cells to TCR triggering, we used Nur77GFP reporter mice. The Nur77 (Nur77) gene is an orphan nuclear receptor that is an immediate-early gene whose expression is increased by TCR stimulation. Therefore, in these mice, GFP expression is a sensitive and faithful-early gene whose expression is increased by TCR stimulation. The Nur77 GFP gene is an orphan nuclear receptor that is an immediate-early gene whose expression is increased by TCR stimulation.

Nur77 GFP reporter mice. The 3 hallmarks of anergic T cells are the lack of proliferation, the lack of effector functions, and the inhibition of the MAPK pathway. To this end, we injected LPS into control and previously thymectomized C57BL/6 mice (B6/ThX) (22, 23). We therefore tested whether αGalCer-pretreated iNKT cells would respond to cytokines from activated innate immune cells triggered by TLR ligands. To this end, we injected LPS into control and αGalCer-pretreated iNKT cells and analyzed GFP expression by iNKT cells 6 hours later. Following LPS injection, the increased expression of CD69 (Figure 3A) and the production of IFN-γ by αGalCer-pretreated iNKT cells were comparable to what was observed in wild-type controls (Figure 3B), indicating that the response of αGalCer-pretreated iNKT cells to TLR-stimulated APCs was unaltered. These data are in line with a previous report showing that αGalCer-pretreated iNKT cells could respond to IL-12/IL-18 stimulation in vitro with IFN-γ production, albeit with reduced intensity (12).
It has been reported that IL-2 provided in vitro can overcome the hyporesponsive phenotype of αGalCer-pretreated iNKT cells (9). Therefore, it could be argued that IL-2 or some other cytokine induced by LPS restored the response of αGalCer-pretreated iNKT cells to LPS. To determine whether LPS could overcome the hyporesponsiveness of αGalCer-pretreated iNKT cells, we injected LPS into mice that had been injected with αGalCer 4 weeks earlier.

Table 1. Summary of the changes observed on induced NKT10 cells

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<th>Downregulated (on induced NKT10 cells):</th>
<th>Upregulated (on induced NKT10 cells):</th>
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<td>CD25</td>
<td>CD185 (CXCR5)</td>
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Expression levels of the indicated markers on iNKT cells from C57BL/6 control and C57BL/6 mice i.v. injected 1 month earlier with 4 μg αGalCer were compared.

Figure 3. Effective TCR signaling of αGalCer-pretreated iNKT cells. (A and B) Control Nur77GFP mice and Nur77GFP mice injected 1 month earlier with 4 μg αGalCer (Nur77αGalCer) were challenged i.v. with 1 μg αGalCer, and GFP expression by splenic iNKT cells was analyzed 1 hour later. Representative data (A) and a summary graph of the GFP+ cells (B) are shown. αGalCer-pre, αGalCer pretreatment. Expression levels of unstimulated iNKT cells from C57BL/6 (B6) mice are shown in A as a negative control. (C) Enriched splenocytes from control congenic C57BL/6 mice (CD45.1+) and from αGalCer-pre, αGalCer-NKT cells from C57BL/6 (B6) mice are GalCer pretreatment. Expression levels of unstimulated iNKT cells was analyzed 1 hour later. Representative data (A) are compared.

These data indicate that αGalCer-pretreated iNKT cells retain the capacity to respond to stimuli provided by APC-derived cytokines. However, this type of stimulation does not overcome the phenotype of hyporesponsiveness to subsequent TCR activation, consistent with the view that the LPS-mediated activation is operating independently of the TCR (22).

The phenotype of αGalCer-pretreated iNKT cells is similar to that of Tregs. Having established that αGalCer-pretreated iNKT cells can respond to TCR- and cytokine-mediated activation, we performed a detailed phenotypic characterization of αGalCer-pretreated iNKT cells. A summary of the observed changes on αGalCer-pretreated iNKT cells is provided in Table 1. αGalCer-pretreated iNKT cells showed downregulated expression of NKL1 and CD69 and increased expression of CD185 (CXCR5), CD278 (ICOS), and CD279 (PD-1) (Figure 5, A and B and Supplemental Figure 3, A–D). The expression of additional NK cell markers, namely, CD94, NKG2D, and NKG2A/C/E, was reduced on αGalCer-pretreated iNKT cells as well (Supplemental Figure 3A). Other surface proteins that were expressed at lower levels on αGalCer-pretreated iNKT cells included CD25, CD122, CD127, CD186 (CXCR6), and CD199 (CCR9) (Supplemental Figure 3, B–D). In contrast, besides the increased expression of CD279 (PD-1), αGalCer-pretreated iNKT cells also displayed augmented expression of CD5, CD49d, CD150 (SLAM), CD200, CD244 (2B4), CD272 (BTLA), and SLAMF6/Ly108 (Figure 5, A and B, and Supplemental Figure 3, B–D). Of particular interest was the increased expression on αGalCer-pretreated iNKT cells of several markers, including CD152 (CTLA4), neuropilin-1 (NRPI), and FR4 (folate receptor 4), which have been associated with the inhibitory phenotype of Tregs (Figure 5, A and B). Whereas the relative percentage of CD4+ αGalCer-pretreated iNKT cells was largely unchanged, the expression levels of CD4 increased on the CD4+ αGalCer-pretreated iNKT cells and approached the expression levels observed on conventional CD4+ T cells (Figure 5C and Supplemental Figure 4A). Expression of the TCR/CD3 complex on αGalCer-pretreated iNKT cells was increased slightly but consistently (Figure 5D and Supplemental Figure 4B). Furthermore, in contrast to control cells, αGalCer-pretreated iNKT cells did not downregulate the TCR/CD3 complex following activation with αGalCer, and there-
fore, the αGalCer-pretreated iNKT cells could easily be detected 1 day after injection (Figure 5D and Supplemental Figure 4B). This indicates that the internalization and recycling of the TCR differs between αGalCer-pretreated and control iNKT cells. Whereas several of the above changes were reported previously (9, 12, 24–26), this is, to our knowledge, the most comprehensive investigation of αGalCer-pretreated iNKT cells to date.

The results of the genome-wide gene expression of untreated and αGalCer-pretreated iNKT cells were in good agreement with the flow cytometric data (Supplemental Figure 3). A set of 18 genes has been defined to be characteristically increased in expression by anergic T cells (27, 28). Importantly, from these 18 anergy-associated genes, only Egr2 was changed in the αGalCer-pretreated iNKT cells in a fashion similar to that seen in anergic T cells (Supplemental Figure 3E), supporting the conclusion that αGalCer-pretreated iNKT cells are not similar to anergic conventional T lymphocytes.

αGalCer-pretreated iNKT cells produce IL-10. Expression of CD152 (CTLA4), CD272 (BTLA), CD279 (PD-1), NRP1, and FR4 by T lymphocytes has been shown to be involved in regulatory and/or tolerogenic contexts (29). Therefore, we determined whether αGalCer-pretreated iNKT cells change their cytokine expression profile in a similar fashion from that of a pro- to an antiinflammatory pattern. As described previously (9, 10), αGalCer-pretreated iNKT cells produced reduced amounts of IL-4 and IFN-γ following a secondary stimulation with αGalCer (Figure 6A), and the response was biased toward Th1 cytokines (Supplemental Figure 5). Similarly, the production of TNF, IL-13, and GM-CSF was decreased in the αGalCer-pretreated iNKT cells (Figure 6, A and B). However, when we measured IL-10 by intracellular staining, we found that up to 10% of αGalCer-pretreated iNKT cells were IL-10+ after secondary antigen stimulation (Figure 6, A and B). To broadly detect cells with the capacity to produce IL-10, we analyzed IL-10 reporter mice, in which an internal ribosome entry site and the gene for the fluorochrome EGFP were inserted into the Il10 locus (Il10GFP, Vert-X) to faithfully label IL-10+ cells (30). Since GFP is a stable protein with a half-life of approximately 26 hours in cells (31) and since it is not secreted in this reporter strain, it accumulates over time and provides increased sensitivity for the detection of IL-10+ cells. However, because the kinetics of GFP induction is slower than that of the IL-10 protein (data not shown), we performed the analysis 16 hours after αGalCer injection. Approximately half of the αGalCer-pretreated iNKT cells were IL-10GFP+ following a second stimulation with αGalCer (50.03% ± 1.51%, Figure 6, C and D). All of the cells expressing IL-10 protein detected by intracellular cytokine staining were within the EGFP+ cell population (Supplemental Figure 6). However, the production of IL-10 was not required to induce the phenotype of αGalCer-pretreated iNKT cells, as αGalCer pretreatment in Il10−/− mice led to phenotypic and functional changes that were indistinguishable from those in wild-type mice (Supplemental Figure 7A and refs. 9, 32). In agreement with this, IL-10 could be detected by intracellular cytokine staining in αGalCer-pretreated iNKT cells from IL-10 receptor-deficient mice after restimulation in vitro (Supplemental Figure 7B). These data corroborate the conclusion that IL-10 is not required for the observed changes in iNKT cells previously exposed to αGalCer. To confirm secretion of IL-10, iNKT cells from control and αGalCer-pretreated mice were stimulated in vitro, and IL-10 in the supernatant was measured by ELISA. In line with the intracellular cytokine staining, we found that αGalCer-pretreated iNKT cells secreted large amounts of IL-10 (Figure 6E). Furthermore, the IL-10 cytokine staining results were corroborated by the results of cytokine mRNA from the microarray analysis (Supplemental Figure 3F). The microarray analysis also indicated increased amounts of Tgfβ3 mRNA (Supplemental Figure 3F), which was verified by RT-PCR (data not shown).

Taken together, these data demonstrate that αGalCer-pretreated iNKT cells acquire a novel surface phenotype and the potential to produce a different cytokine pattern, including IL-10, which mediates immune suppression. Based on this capability, we propose the name “induced NKT10 cells.”

NKT10 cells are a distinct iNKT cell subset. In recent years, functional subsets of Vα14i NKT cells have been defined that can be distinguished based on the expression of particular surface markers and/or transcription factors (33–35). We next investigated the
relation of NKT10 cells to published NKT cell subsets. The transcription factor FOXP3 is crucial for the function of Tregs (29), and its expression can be induced in NKT cells (36, 37). However, αGalCer-pretreated NKT cell subsets did not express FOXP3 (Figure 7A). Furthermore, it has been reported that early after αGalCer injection (day 6), NKT cells acquire a T follicular helper (Tfh) phenotype, characterized by the expression of the transcription factor BCL6 (25, 26, 38). In agreement with these results, on day 6 after αGalCer injection, we detected a population of BCL6+ NKT cells (Figure 7B and Supplemental Figure 9). Furthermore, after 1 month, the NKT10 cell population was largely BCL6+ (25, 26, 38). In agreement with these results, NKT cells (36, 37). However, αGalCer-pretreated NKT cells did not express FOXP3 (Figure 7A). Furthermore, it has been reported that early after αGalCer injection (day 6), NKT cells acquire a T follicular helper (Tfh) phenotype, characterized by the expression of the transcription factor BCL6 (25, 26, 38). In agreement with these results, on day 6 after αGalCer injection, we detected a population of BCL6+ NKT cells (Figure 7B and Supplemental Figure 9). Furthermore, after 1 month, the αGalCer-pretreated NKT cells expressed several cell-surface molecules characteristic of NKTfh cells, such as CD185 (CXCR5) and SLAMF6, and they had increased Il21 mRNA expression (Supplemental Figure 3, C, D, and F). However, other evidence indicated that NKT10 cells are not NKTfh cells. First and most important, BCL6 staining returned almost to background levels by 4 weeks (Figure 7B, Supplemental Figure 9, and Supplemental Figure 10B). These data indicate either that BCL6 expression by iNKT cells is transient after antigen activation or that the BCL6+ cell population of iNKT cells is short lived. Additionally, while the αGalCer-pretreated iNKT cells had reduced CD127 expression by day 6, similar to the reported phenotype for NKTfh cells (26), by day 33, expression of CD127 increased (Figure 7B, Supplemental Figure 9, and Supplemental Figure 10B). These data indicate that after 4 weeks, the αGalCer-pretreated iNKT cells were no longer NKTfh cells. Therefore, to exclude the involvement of NKTfh cells, experiments were performed at least 4 weeks after the injection of αGalCer.

NKT10 cells did express NRP1 and largely lacked expression of NK1.1 (Figure 5, A and B), which has also been reported for NKT17 cells (39–42). However, IL-10+ iNKT cells did not coexpress IL-17A (Figure 7C). Furthermore, the majority of NKT17 cells were CD4− and CD49d+, whereas IL-10+ iNKT cells were largely CD4+ and CD49d− (Figure 7, D and E). These data demonstrate that NKT10 and NKT17 cells represent 2 independent iNKT cell subsets, despite the shared expression of NRP1 and the lack of NK1.1.

It has been reported that 1 month after αGalCer, the proinflammatory cytokine response of the αGalCer-pretreated iNKT cells partially recovered (9), suggesting that the NKT10 cell phenotype might be short lived as well. To gain a better understanding of the development of the αGalCer-induced changes, we conducted time-course experiments for up to 3 months after αGalCer injection. Several markers associated with NKTfh cells, like BCL6 and CD127, changed rapidly but largely reverted 2–3 weeks af-
ter αGalCer injection (Supplemental Figure 10B). The phenotype of αGalCer-pretreated iNKT cells then remained stable for up to 3 months (Supplemental Figure 10), and data derived 2 or 3 months after αGalCer injection were largely indistinguishable from those from the 1-month time point. At later time points, the phenotype of iNKT cells appeared to slowly revert, but this was likely due to RTEs, as indicated by preliminary experiments with thymectomized mice (data not shown). These data suggest that αGalCer-pretreated iNKT cells might recover full activity, but that this takes at least 4 months and is probably overshadowed by the appearance of “fresh” RTE iNKT cells. As these RTE iNKT cells have not been exposed to αGalCer, their response should be comparable to that of iNKT cells in untreated control mice. Therefore, to exclude the involvement of RTE iNKT cells, we performed experiments within 2 months of αGalCer injection. Together, these data indicate that NKT10 cells are a defined iNKT cell subset that is stable for at least 3 months.

**IL-10–dependent immune regulation by induced NKT10 cells.** The phenotype of induced NKT10 cells, with some similarity to Tregs, and their ability to produce IL-10, suggested that they are capable of regulating immune responses. Two in vivo models were used to test this hypothesis.

First, we observed that αGalCer-pretreated animals challenged with B16 melanoma cells suffered from a significantly higher tumor burden when treated concomitantly with αGalCer (Figure 8A, bar 3 versus 4). This finding suggests that TCR-mediated activation of induced NKT10 cells was actively dampening the antitumor response, and therefore we reasoned that the

**Figure 6.** αGalCer-pretreated iNKT cells produce IL-10. (A and B) αGalCer (1 μg) was i.v. injected into wild-type (B6) or mice i.v. injected 1 month earlier with 4 μg αGalCer (B6/αGC) (3 mice/group). Splenic iNKT cells were analyzed 90 minutes later by intracellular staining for expression of the indicated cytokines. (A) Summary graph. Data in the 2 panels are derived from 2 independent experiments. (B) Representative flow cytometric data. ICS, intracellular cytokine staining. (C and D) Control Il10GFP mice or Il10GFP mice given 4 μg αGalCer 1 month earlier (Il10GFP/αGC, αGalCer-pre) were either left untreated or were i.v. injected with 1 μg αGalCer (Stimulated), and splenic iNKT cells were analyzed 16 hours later for EGFP and CD279 (PD1) expression. Representative data (C) and a summary graph of IL-10 expression (D) are shown. αGalCer-pre, unstimulated = mean: 3.92% ± 0.47%; median: 3.61%. αGalCer-pre, stimulated = mean: 50.03% ± 1.51%; median: 51.71%. Control, stimulated = mean: 13.76% ± 2.04%; median: 12.85%. Data shown in D are from at least 8 independent experiments with at least 16 mice per group. Background values for IL-10 detection by intracellular staining or EGFP are shown and discussed in Supplemental Figure 8. (E) Splenic iNKT cells from the indicated mice (B6 = C57BL/6; B6/αGC = C57BL/6 mice i.v. injected 1 month earlier with 4 μg αGalCer; Il10−/− = IL-10–deficient mice) were sorted and were either left untreated or were stimulated in vitro for 4 days on αTCRβ antibody–coated plates before supernatants were collected and IL-10 levels were determined by ELISA. Representative data from 3 independent experiments are shown.
αGalCer-induced IL-10 production by induced NKT10 cells may act to promote tumor growth. Indeed, αGalCer-pretreated Il10−/− mice displayed no difference in the amount of B16 tumor nodules following a secondary αGalCer challenge than did control mice (Figure 8A, bars 7 and 8 versus bars 3 and 4). These data are consistent with the hypothesis that induced NKT10 cells promote tumor growth in vivo via the production of IL-10. Interestingly, whereas the injection of αGalCer into αGalCer-pretreated animals had no effect on tumor growth when the mice were challenged with B16–CD1d (Figure 2D), it led to increased tumor burden when the mice were challenged with B16 (Figure 8A). These data suggest that the binding of αGalCer to the B16–CD1d cells in vivo led to cognate recognition and the direct cytotoxic elimination of some of the tumor cells by the αGalCer-pretreated iNKT cells, in line with previous observations (16).

As a second in vivo model, representing an autoimmune disease, we analyzed the impact of induced NKT10 cells on the development of experimental autoimmune encephalomyelitis (EAE). EAE is a mouse model of multiple sclerosis, whereby immunization of mice with myelin oligodendrocyte glycoprotein peptide (MOG33–55) and treatment of disease progression compared with IL-10+ iNKT cells (Figure 8B).

Taken together, these data from 2 experimental systems demonstrate that IL-10 production by induced NKT10 cells can significantly impact the immune response.

NKT10 cells are a naturally occurring subset. Although NKT10 cells can be induced by αGalCer treatment, this antigen provides an extremely strong stimulus that may not be typical. Therefore, we determined whether a similar iNKT cell subset could also be found in unchallenged mice. We injected αGalCer into naïve C57BL/6 mice and analyzed the production of IL-10 by intracellular staining 90 minutes later. We detected a small population of IL-10+ iNKT cells (0.45% ± 0.03%) in the spleen (Figure 9, A and B). To more inclusively capture iNKT cells capable of producing IL-10, we stimulated splenocytes with PMA and ionomycin for 4 hours in vitro. On average 1.77% ± 0.18% of iNKT cells were IL-10+ following this treatment (Figure 8A) and compared with mice without iNKT cells (Supplemental Figure 7). Importantly, Jα18−/− host mice that received wild-type iNKT cells displayed significantly ameliorated signs of disease progression compared with Jα18−/− host mice that received Il10−/− iNKT cells (Figure 8B).

To address the role of induced NKT10 cell-derived IL-10, we transferred iNKT cells from wild-type and Il10−/− mice into iNKT cell–deficient Jα18−/− hosts. As noted above, IL-10 is not required for the generation of induced NKT10 cells (Supplemental Figure 7). The recipient host mice were then immunized with myelin oligodendrocyte glycoprotein peptide (MOG33–55) and treated 3 times with αGalCer (see experimental outline in Supplemental Figure 11). In agreement with previous results, mice with iNKT cells treated multiple times with αGalCer were protected from EAE compared with mice without iNKT cells (Figure 8B).
that were expressed at least 3 times more frequently on the IL-10+ cells than on the IL-10- iNKT cells. Additionally, IL-10 expression correlated with the expression of CD49d, CD200, CD278, CD314, and β7-integrin, or IL-17A (data not shown). Importantly, nearly 50% of the scWAT NKT cells remained detectable 16 hours after injection, whereas IL-10GFP+ iNKT cells represented 13.5% ± 2.6% of the starting population of iNKT cells in the scWAT (Figure 10B). Therefore, NKT10 cells are enriched, or become enriched rapidly following antigenic exposure, in the WAT. Based on these data, we propose that such IL-10+ iNKT cells in unchallenged mice represent a naturally present iNKT cell subset. We refer to these IL-10+ iNKT cells as “natural NKT10 cells,” without claiming that they originate exclusively in the thymus or as a result of peripheral stimulation.

Given that we could detect natural NKT10 cells in unchallenged control mice, we tested whether NKT10 cells could also be found in human PBMCs. To this end, human PBMCs from healthy donors were stimulated with PMA and ionomycin for 4 hours. This stimulation yielded a clear IL-10 signal from Vα24i NKT cells by intracellular cytokine staining (Figure 11A). Similar to the findings with mouse splenocytes, the percentage of IL-10+ iNKT cells was mostly below 2% (mean: 0.42% ± 0.08%; median: 0.38%), but NKT10 cells could be detected in 19 of 20 donors tested (Figure 11B). Together, these data demonstrate that NKT10 cells are a distinct and novel iNKT cell subset in mice and humans that occurs naturally in the absence of αGalCer exposure.

The number of iNKT cells in the liver declined similarly (data not shown). This is likely due to downregulation of TCR expression (51, 52), although activation-induced cell death and migration of iNKT cells may also contribute to this decline. Importantly, 17.0% ± 1.3% of the remaining CD1d-αGalCer tetramer+ iNKT cells expressed IL-10GFP (Figure 10, A and B). This represented 0.53% ± 0.08% compared with the starting iNKT cell population in the spleen (Figure 10B), which was in agreement with the values derived from the intracellular IL-10 staining (Figure 9, A and B). For some other iNKT cell subsets, preferential homing to specific organs, such as the lymph nodes for NKT17 cells, has been reported (42). The recent suggestion of the presence of antiinflammatory iNKT cells in white adipose tissue (WAT) (53–55) and the regulatory phenotype of induced NKT10 cells we described above made the subcutaneous white adipose tissue (scWAT) a candidate tissue for preferential localization of NKT10 cells. Additionally, we observed that scWAT iNKT cells were enriched for the expression of markers also expressed by induced NKT10 cells (Figure 10, C and D). Therefore, we enumerated naturally occurring NKT10 cells in the scWAT of IL10GFP reporter mice, as we did for the spleen. Following primary stimulation with αGalCer, a significantly higher percentage of iNKT cells remained detectable in scWAT compared with that in the spleen (Figure 10B). Importantly, 31.6% ± 3.6% of the iNKT cells recovered from the scWAT-expressed IL-10GFP (Figure 10, A and B). Considering that nearly 50% of the scWAT iNKT cells were still detectable 16 hours after injection, these IL-10GFP+ iNKT cells represented 13.5% ± 2.6% of the starting population of iNKT cells in the scWAT (Figure 10B). Therefore, NKT10 cells are enriched, or become enriched rapidly following antigenic exposure, in the WAT.

Based on these data, we propose that such IL-10+ iNKT cells in unchallenged mice represent a naturally present iNKT cell subset. We refer to these IL-10-producing iNKT cells as “natural NKT10 cells,” without claiming that they originate exclusively in the thymus or as a result of peripheral stimulation.

Given that we could detect natural NKT10 cells in unchallenged control mice, we tested whether NKT10 cells could also be found in human PBMCs. To this end, human PBMCs from healthy donors were stimulated with PMA and ionomycin for 4 hours. This stimulation yielded a clear IL-10 signal from Vα24i NKT cells by intracellular cytokine staining (Figure 11A). Similar to the findings with mouse splenocytes, the percentage of IL-10+ iNKT cells was mostly below 2% (mean: 0.42% ± 0.08%; median: 0.38%), but NKT10 cells could be detected in 19 of 20 donors tested (Figure 11B). Together, these data demonstrate that NKT10 cells are a distinct and novel iNKT cell subset in mice and humans that occurs naturally in the absence of αGalCer exposure.
Discussion

Here, we describe a novel iNKT cell subset, which we have termed NK10 cells, that is characterized both by the expression of several proteins found on Tregs and by the capacity to produce IL-10 after antigenic stimulation. Importantly, NK10 cells occur naturally in mice and in humans, albeit at a low frequency, and they expand greatly after activation with the strong agonist αGalCer. Such αGalCer-induced NK10 cells were not anergic, as previously suggested, but retained their cytotoxic activity and their response to TCR-dependent and TCR-independent stimulation by cytokines from APCs. Furthermore, through the production of IL-10, induced NK10 cells were able to modulate the tumor load of B16 melanoma–challenged mice and the outcome of EAE. Therefore, our data reveal new aspects of iNKT cell functional diversity.

Treatment of iNKT cells with αGalCer in vivo has been proposed to lead to a general hyporesponsive state similar to that of energy, based on reduced antigen-induced proliferation and production of IFN-γ and IL-4 following restimulation. However, the original data in support of this hypothesis analyzed the splenic iNKT cell response at the cell population level after in vitro culture (9, 10). Given that the relative percentage of αGalCer-pretreated iNKT cells is significantly lower in the spleen of αGalCer-pretreated mice (Figure 2A and Supplemental Figure 10A), such an approach could underestimate the per-cell response of iNKT cells. Furthermore, the in vitro response of αGalCer-pretreated iNKT cells does not necessarily reflect their in vivo response. Therefore, to gain insight into the per-cell response of iNKT cells from αGalCer-pretreated mice, we used single-cell flow cytometric assays to directly measure iNKT cell activity ex vivo. This is not typically done, although a few reports have analyzed the iNKT cell cytokine response on a single-cell level in vitro (12) or in vivo (12, 56). Using our approach, we confirmed the reduced production of proinflammatory cytokines by αGalCer-pretreated iNKT cells (Figure 6, A and B). In contrast, while it was reported that the expansion of αGalCer-pretreated iNKT cells after a secondary stimulation with αGalCer was reduced (12, 56), their steady-state proliferation in vivo, as measured by Ki67 and BrdU staining, was actually increased (Figure 1), suggesting an increased response to homeostatic signals from cytokines or self-antigen.

The induction of GFP in αGalCer-pretreated iNKT cells from Nur77GFP reporter mice, following antigen reexposure, showed productive TCR signaling, albeit reduced from controls (Figure 3, A and B). The induction of Nur77 is downstream of Ras activation, which is defective in anergic T cells (27, 28). Staining for p-ERK1/2 following TCR stimulation in primary iNKT cells (Figure 3C) directly demonstrated unimpaired TCR-induced MAPK signaling.
of αGalCer-pretreated iNKT cells. Furthermore, the results from the microarray analysis indicated that expression of most of the genes associated with the anergic state (27, 28) was not increased in αGalCer-pretreated iNKT cells (Supplemental Figure 3E). Together, our data clearly demonstrate that αGalCer-pretreated iNKT cells are not anergic.

We therefore tested the responsiveness of αGalCer-pretreated iNKT cells by 3 additional readouts. First, when we analyzed αGalCer-specific in vivo cytotoxicity (16), hypersensitivity was not observed (Figure 2A). We obtained similar results in thymectomized animals, indicating that RTEs could not account for the cytotoxicity (Figure 2A). Second, we activated αGalCer-pretreated iNKT cells via TLR ligand–induced cytokines from APCs, and their response was comparable to that of control cells in this context as well (Figure 4, A and B). Third, αGalCer-pretreated iNKT cells displayed an increased production of IL-10 (Figure 6). Whereas the term anergy implies the absence of an iNKT cell response, our data demonstrate that αGalCer pretreatment leads instead to modulation of the iNKT cell response.

In recent years, functional subsets of iNKT cells have been defined. Some, such as NKT1 and NKT2 (33–35), NKT17 (39–42), and IL17RB+ iNKT (57) cells, originate predominantly in the thymus, while others, such as NKTh (25, 26, 38) and FOXP3+ iNKT (36, 37) cells, arise after immunization. The definition of iNKT cell subsets is largely based on their expression of transcription factors and their function, especially significant biases in cytokine production of the respective iNKT cell types. NKT1, NKT2, and NKT17 cells are defined as the iNKT cell subset biased toward Th1, Th2, or Th17 cytokines, respectively. Like their Th subset counterparts, these iNKT cell subsets also produce several cytokines in addition to the signature one(s) that define the subsets. In line with the existing iNKT cell nomenclature, here we define NKT10 cells as the iNKT cell subset capable of producing IL-10 as one of its predominant cytokines and demonstrate that these NKT10 cells are distinct from the other known iNKT cell subsets (Figure 7). Dynamic changes occur in the iNKT cell population after αGalCer treatment, including the transient appearance of a Thh-like phenotype of iNKT cells (NKTh cells) in the spleen that is characterized by expression of the transcription factor BCL6 (25, 26). Consistent with this, while BCL6 is expressed in iNKT cells on day 6 after αGalCer injection, it declines thereafter (Figure 7B, Supplemental Figure 9, and Supplemental Figure 10B). However, NKT10 cells did not express FOXP3, BCL6, or IL-17A after stimulation, and thus by choosing the 1-month time point for most of our experiments, we were able to exclude the involvement of NKTh cells as well as limit the contribution of RTE iNKT cells. The relationship of NKTh to NKT10 cells remains to be determined. Our data indicated that NKT10 cells expand after αGalCer treatment in vivo. This is not specific to αGalCer stimulation, however, since repeated injection (3 times) of a weaker antigen also caused an increase in this cell population (data not shown). Additionally, αGalCer stimulation does not always lead to the generation of induced NKT10 cells, as we did not observe this, in line with previous reports (9, 10), when we injected bone marrow–derived DCs loaded with αGalCer (data not shown). However, whether the expansion of NKT10 cells after αGalCer is the result of selective expansion of a preexisting population of NKT10 cells or of conversion to the NKT10 cell phenotype cannot be answered definitively at this point. When we sorted NRP1+ iNKT cells, transferred them into Jα18−/− mice, and injected αGalCer, we could detect a rapid increase in NKT10 cells (data not shown), consistent with induction of these cells. However, even though sorting NRP1+ iNKT cells likely excluded naturally occurring NKT10 cells, the presence of a small amount of contaminating cells cannot be ruled out. Nonetheless, we favor the idea that some non-NKT10 cells can convert into NKT10 cells following αGalCer stimulation. Furthermore, it is not known whether those iNKT cells capable of producing IL-10 in the absence of αGalCer pretreatment are natural, in the sense that they differentiated in the thymus, or whether they are induced in the periphery, possibly by chronic antigenic exposure. A role of NKL1+ T-cell–derived IL-10, in the absence of previous αGalCer exposure, has so far only been reported in the systemic

Figure 10. NKT10 cells are enriched in WAT. (A and B) Il10−/− mice were left untreated or were i.v. injected with 1 μg αGalCer, and iNKT cells in spleen and scWAT were analyzed 16 hours later. (A) Expression of EGFP and CD49d from 1 representative experiment. Numbers in the dot plots denote the percentage of cells in the respective quadrants. (B) Percentage of EGFP (light gray) and EGFP– (dark gray) iNKT cells from spleen and scWAT was calculated as the percentage of iNKT cells in untreated control mice. P < 0.001 for spleen versus scWAT for all comparisons. Graph summarizes data from 4 independent experiments with 13 mice per group. (C and D) iNKT cells from spleen and scWAT of C57BL/6 mice were analyzed for expression of the indicated markers. Representative data (C) and a summary graph (D) are shown. Numbers in the histograms in C denote the geometric mean values for the depicted antigens on iNKT cells. Representative data from 1 of at least 3 independent experiments are shown.
Agreement with the percentage of IL-10+ NKT10 cells observed in scWAT suggests that natural NKT10 cells present in scWAT may help to maintain healthy adipose tissue.

EAE is a mouse model of multiple sclerosis, in which iNKT cells influence the disease course (44–46, 61). It has been shown that repetitive injection of αGalCer can protect mice against EAE (44–47). Importantly, a single injection of αGalCer did not protect mice against EAE according to most (45, 48–50), albeit not all (45, 50), reports. Differences were attributed to the genetic background, the route and timing of application, and the dosage of αGalCer (45, 46, 50). The protection was originally thought to be due to a Th2 bias in the secondary response of αGalCer-pretreated iNKT cells (8, 9, 62, 63). However, this conclusion was based on the analysis of the αGalCer response on a single-cell level, either no bias or a Th1 cytokine bias was detected (Supplemental Figure 5 and refs. 12, 56, 63).

The apparent discrepancy of the results is likely due to the lack of NK cell trans-activation following a secondary stimulation of αGalCer-pretreated iNKT cells (63). In regard to the αGalCer-induced EAE protection, however, is not entirely clear which iNKT cell-derived cytokine is important. IL-4 and/or IL-10 have been suggested (44, 45, 48, 64–66), but some data contradict this (46, 61). Similarly, divergent results have been reported for IFN-γ (pro: refs. 45, 46, 66, 67; con: ref. 64).

However, based on our data, we propose that the protection against EAE by repetitive αGalCer challenge depends on the requirement to first induce sufficient numbers of NKT10 cells and then to subsequently stimulate IL-10 production from them. The generation of cells with an NKT10 cell phenotype in this model does not require their ability to produce IL-10, but we noticed a worsening of disease in mice that received IL-10-deficient iNKT cells compared with the control Jα18+ mice (Figure 8B). Therefore, it is possible that in the absence of the ability to produce IL-10, the proinflammatory cytokines still produced by αGalCer-induced EAE protection, however, is not entirely clear which iNKT cell-derived cytokine is important. IL-4 and/or IL-10 have been suggested (44, 45, 48, 64–66), but some data contradict this (46, 61). Similarly, divergent results have been reported for IFN-γ (pro: refs. 45, 46, 66, 67; con: ref. 64).

The recent appreciation that iNKT cells are heterogeneous suggests that some of the controversies regarding the function of iNKT cells could depend on which subset is activated. Our data indicate that αGalCer-pretreated NKT10 cells regulate concomitant adaptive immune responses. The fact that up to 13.5% of iNKT cells in control mice and healthy human donors resembled αGalCer-pretreated iNKT cells suggests that they represent a novel iNKT cell subset. The characterization of suppressive NKT10 cells could help to resolve current controversies about the dichotomous nature of iNKT cells observed in various studies, namely their ability to exert either pro- or antiinflammatory effects. Such knowledge also could be important for understanding the functional consequences of αGalCer or other...
glycolipid antigens in therapy and how to deliberately tailor iNKT cell responses for therapeutic applications.

**Methods**

**Mice and cell lines.** All mice were housed under specific pathogen-free conditions at the animal facilities of the La Jolla Institute for Allergy and Immunology (La Jolla, California, USA) in accordance with IACUC guidelines. II-10^-/- (B6.129S2-Il10rb^tm1Agt/J), II-10 receptor-deficient mice (B6.129S2-Il10rb^tm1Agt/J), C57BL/6j control mice, and C57BL/6 mice thymectomized at 7 weeks of age were purchased from The Jackson Laboratory. B6.129-Tcrα^tm1Tgi (Ja18^-/-) mice, Nr4a1 (Nur77^GFP+ (20) and II-10^-/- (II-10^--/--)) mice, or conjugated with Pacific Blue, eFluor 450, V450, Brilliant Violet 421, PE-Cy5, APC, Alexa Fluor 647, eFluor 660, Alexa Fluor 700, APC-Cy7, or APC-eFluor 780. Anti-mouse CD16/32 antibody (2.4G2) used for Fc receptor blocking was isolated in our laboratory. FcR inhibitor for human cells was obtained from eBioscience. Unconjugated mouse and rat IgG antibodies were purchased from Jackson ImmunoResearch. Dead cells were labeled with a Blue, Aqua, or Yellow Dead Cell Stain Kit (Invitrogen).

**In vivo challenge and cell preparation.** In vivo cytotoxicity assays were performed as reported previously (16). In brief, splenic CD19^-B cells were either pulsed with αGalCer (250 ng/ml, 1 hour) or were mock treated, then differentially labeled with CFDA-SE (Invitrogen), and equal numbers (5 x 10^6) of both cell populations were i.v. injected into the experimental mice and into the Ja18^-/- mice as a negative control. Four hours later, the presence of target cells in spleen was determined by flow cytometry, and the specific lysis was calculated as described (16). iNKT cells were pretreated with αGalCer by i.v. injection of 4 μg αGalCer and analyzed 4–6 weeks later or as otherwise indicated. Acute activation in vivo was induced by i.v. injection of 1 μg αGalCer followed by analysis 90 minutes later or as otherwise indicated. For in vivo labeling with BrdU (Sigma-Aldrich), the indicated mice were injected i.p. with 1 mg BrdU (in PBS), and at the same time, BrdU (1 mg/ml) was placed in the drinking water supplemented with 5% glucose (w/v). Three days later, splenic iNKT cells were analyzed for BrdU incorporation using a BrdU flow kit (BD Biosciences) according to the manufacturer’s instructions.

**Human PBMCs were isolated by use of Ficoll-Paque (GE Healthcare) density gradient centrifugation.** Single-cell suspensions from mouse liver, spleen, and thymus were prepared as described (16). Mouse scWAT was minced, digested (1 mg/ml collagenase type II and 8 μg/ml DNase I; both from Sigma-Aldrich) for 40 minutes (37°C, 200 rpm) and filtered through a 70-μm cell strainer (BD Biosciences). In some experiments intended for intracellular staining of IL-10, splenocytes were purified by use of Lymphoprep (Axis-Shield) density gradient centrifugation and by depletion of B cells with anti-CD45R- coated magnetic beads (Invitrogen). For enrichment of Vα14i NKT cells from splenocytes, cells were incubated with biotin-conjugated antibodies for CD8α, CD11b, CD19, CD24, CD45RA, F4/80, Ly6C/G, and TER119, followed by negative selection with streptavidin magnetic beads (STEMCELL Technologies), resulting in cell suspensions containing 8%–15% iNKT cells.

**Lung metastases with B16 melanoma cells.** B16 and B16-CD11d melanoma cells were either loaded with 250 ng/ml αGalCer (37°C, 90 minutes) or mock treated, washed twice with PBS, and 1 x 10^6 tumor cells were i.v. injected into C57BL/6 mice as indicated in Figure 2D and Figure 8A. Fourteen days after challenge, the number of metastatic nodules on the lung surface was counted. Five hundred tumor nodules were established as the upper limit for counting, since at higher densities, discrete tumor nodules could not be accurately separated.

**EAE.** EAE was induced by immunization with MOG_35-55 (MEVGYWRSPFSRRVHLRNGK; Synthetic Biomolecules). MOG_35-55 (200 μg) was s.c. injected with a 25-gauge needle in 0.1 ml CFA (Sigma-Aldrich) into 3 to 4 different sites on the flank. Concomitant with the peptide and 2 days later, 200 ng pertussis toxin (Sigma-Aldrich) was injected i.v. on day 0 and i.p. on day 2. Disease was monitored daily.
from day 8 onward. Freshly isolated lymphocytes enriched for \textit{iNKT} cells (containing $5 \times 10^6$ iNKT cells) were i.v. injected into mice shortly before the immunization. Disease was monitored visually using a 0–5 ranking (0 = normal, 0.5 = tail weakness, 1 = hind limb weakness, 2 = mild hind limb paralysis, 3 = moderate hind limb paralysis, 4 = complete hind limb paralysis, 5 = moribund or dead). Gel packs were added to all cages as soon as mice reached a score above 2. Animals with a score of 4 were euthanized once they remained in this state for more than 2 consecutive days and were then categorized as score 5.

\textit{In vitro cultures.} For ex vivo experiments intended for intracellular staining of IL-10 or IL-17A, lymphocytes were cultured for 2 hours in medium consisting of RPMI 1640 (Invitrogen) supplemented with 10% (v/v) FCS and 1% (v/v) Pen-Strep-Glutamin (10,000 U/ml penicillin, 10,000 g/ml streptomycin, 29.2 mg/ml L-glutamine; Invitrogen) in the presence of GolgiPlug and GolgiStop (BD Biosciences) at 37°C. Ex vivo stimulation was performed with PMA and ionomycin (both from Sigma-Aldrich) for 4 hours at 37°C in the presence of GolgiPlug and GolgiStop. For detection of degranulation, splenocytes from congenic control (CD45.1) and \textit{αGalCer}-pretreated animals (CD45.2) were incubated for 5 hours at 37°C at a 1:1 ratio in \textit{αCD3$:}$ antibody-coated (5 \mu g/ml, 145.2C11) plates in the presence of 0.5 \mu g/ml \textit{αCD107a$-$(1D4B) and \textit{αCD107b$}$ (ABL93) Alexa Fluor 488-labeled antibodies (Santa Cruz Biotechnology Inc.). For the measurement of p-ERK1/2, enriched splenocytes from congenic control (CD45.1) and \textit{αGalCer}-pretreated animals (CD45.2) were mixed at a 1:2 ratio, incubated for 10 minutes on ice with 10 \mu g/ml \textit{αCD3$:}$ antibody (145.2C11, BD Biosciences), then washed and stimulated with 40 \mu g/ml cross-linking anti-hamster IgG antibodies (G70-204 and G94-56; BD Biosciences) together with \textit{CD1d$-αGalCer$} tetramers at 37°C. After 2 minutes, cells were fixed and stained for p-ERK1/2. Staining of primary \textit{iNKT} cells in this assay required the use of \textit{CD1d$-αGalCer$} tetramer labeled with the fluorochrome BV421 (BD Biosciences), together with the stimulation and rapid addition of the fixation reagent after the 2 minutes incubation.

\textit{Flow cytometry and ELISA.} Flow cytometry and preparation of fluorochrome-conjugated \textit{αGalCer}-loaded CD1d tetramers were performed as previously described (71). Cell sorting was performed with a FACSAria cell sorter (BD Biosciences). \textit{iNKT} cells were defined throughout as live \textit{CD8$^+$}$ \textit{TCR}^+$ (clone 6B11) cells (human). IL-10 levels in culture supernatants were determined by ELISA using BioLegend reagents, according to the manufacturer’s recommendations.

\textit{RNA microarray.} Enriched \textit{V$\alpha$t4i$\times$} \textit{iNKT} cells (CD1d$-αGalCer$ tetramer$^-TCR^+)$ were isolated from spleen using a FACS$\alpha亚$ cell sorter (BD Biosciences) and stored as cell pellets at $-80°C$. RNA was isolated from the frozen cell pellets using the RNeasy Mini Kit (QIAGEN), and transcripts were amplified with the Ovation RNA amplification system V2 (NuGEN) and purified with the QIAquick PCR purification kit (QIAGEN) according to the manufacturer’s instructions. In vitro transcription (IVT) probe generation and hybridization to Affymetrix Mouse Genome 430 2.0 arrays (Platform GPL1261) were performed at the Veterans Medical Research Foundation GeneChip Microarray facility (UCSD). Signal intensity data and lists of differentially expressed genes were compiled using VAMPIRE (Subramanian Laboratory, UCSD) and Genespring (Agilent) software. Microarray data were deposited in the NCBI’s Gene Expression Omnibus (GEO GSE47959; http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE47959) (72).

\textit{Statistics.} Results are expressed as the mean $\pm$ SEM. Statistical comparisons were drawn using a 2-tailed Student’s t test (Excel; GraphPad Prism, GraphPad Software) for all paired samples or otherwise using an ANOVA test (GraphPad Prism). Normal distribution was validated by the D’Agostino-Pearson omnibus normality test (GraphPad Prism), either directly (n $\geq$ 10) or after combining values from repetitive experiments. P values less than 0.05 were considered statistically significant and are indicated as $^*P < 0.05$, $^{**}P < 0.01$, and $^{***}P < 0.001$ in the figures. Each experiment was repeated at least twice, and background values were subtracted. Graphs were generated with GraphPad Prism.

\textit{Study approval.} Human PBMCs were obtained from healthy donors in accordance with the La Jolla Institute for Allergy and Immunology Normal Blood Donor Umbrella Program (VD-057). All mouse experiments were performed in an AAALAC-accredited facility with prior approval of the IACUC of the La Jolla Institute for Allergy and Immunology and in accordance with the US Public Health Service (PHS) Policy on Humane Care and Use of Laboratory Animals.

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Address correspondence to: Mitchell Kronenberg or Gerhard Wingender, La Jolla Institute for Allergy and Immunology, 9420 Athena Circle, La Jolla, California 92037, USA. Phone: 858.752.6540; E-mail: mitch@liai.org (M. Kronenberg); Phone: 858.752.6738; E-mail: gwing@liai.org (G. Wingender).
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