Engrafted human stem cell–derived hepatocytes establish an infectious HCV murine model

Arnaud Carpentier, …, Stephen M. Feinstone, T. Jake Liang

*J Clin Invest.* 2014;124(11):4953-4964. [https://doi.org/10.1172/JCI75456](https://doi.org/10.1172/JCI75456).

**Technical Advance**

The demonstrated ability to differentiate both human embryonic stem cells (hESCs) and patient-derived induced pluripotent stem cells (hiPSCs) into hepatocyte-like cells (HLCs) holds great promise for both regenerative medicine and liver disease research. Here, we determined that, despite an immature phenotype, differentiated HLCs are permissive to hepatitis C virus (HCV) infection and mount an interferon response to HCV infection in vitro. HLCs differentiated from hESCs and hiPSCs could be engrafted in the liver parenchyma of immune-deficient transgenic mice carrying the urokinase-type plasminogen activator gene driven by the major urinary protein promoter. The HLCs were maintained for more than 3 months in the livers of chimeric mice, in which they underwent further maturation and proliferation. These engrafted and expanded human HLCs were permissive to in vivo infection with HCV-positive sera and supported long-term infection of multiple HCV genotypes. Our study demonstrates efficient engraftment and in vivo HCV infection of human stem cell–derived hepatocytes and provides a model to study chronic HCV infection in patient-derived hepatocytes, action of antiviral therapies, and the biology of HCV infection.

Find the latest version:

[http://jci.me/75456/pdf](http://jci.me/75456/pdf)
Engrafted human stem cell–derived hepatocytes establish an infectious HCV murine model


1Liver Diseases Branch, National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK), NIH, Bethesda, Maryland, USA. 2Division of Viral Products, Center for Biologics Evaluation and Research, FDA, Bethesda, Maryland, USA. 3Laboratory of Experimental Carcinogenesis, Center for Cancer Research, National Cancer Institute, NIH, Bethesda, Maryland, USA.

The Journal of Clinical Investigation

Technical advance

Introduction

The technology to developmentally program human embryonic stem cells (hESCs) to various cell lineages offers great promise for both regenerative medicine and liver disease research. Here, we determined that, despite an immature phenotype, differentiated HLCs are permissive to hepatitis C virus (HCV) infection and mount an interferon response to HCV infection in vitro. HLCs differentiated from hESCs and hiPSCs could be engrafted in the liver parenchyma of immune-deficient transgenic mice carrying the urokinase-type plasminogen activator gene driven by the major urinary protein promoter. The HLCs were maintained for more than 3 months in the livers of chimeric mice, in which they underwent further maturation and proliferation. These engrafted and expanded human HLCs were permissive to in vivo infection with HCV-positive sera and supported long-term infection of multiple HCV genotypes. Our study demonstrates efficient engraftment and in vivo HCV infection of human stem cell–derived hepatocytes and provides a model to study chronic HCV infection in patient-derived hepatocytes, action of antiviral therapies, and the biology of HCV infection.

Conflict of interest: The authors have declared that no conflict of interest exists.
Figure 1. Hepatic differentiation of human pluripotent stem cells. (A–D) Protocol, phase-contrast microscopy, immunofluorescent assay, and FACS analysis of the cells at different stages. (A) hESCs and hiPSCs. (B) Stem cells were treated for 3 days with 100 ng/ml activin A and basic fibroblast growth factor (bFGF) to generate definitive endoderm. (C) Hepatic specification was induced by maintaining the cells in presence of 100 ng/ml hepatocyte growth factor (HGF) and 0.1% DMSO for 8 days. (D) Finally, hepatic maturation was achieved by treating the hepatoblasts for 3 days with 10^{-7} M dexamethasone. Differentiated HLCs were maintained for up to 2 weeks in culture in the presence of dexamethasone (DEX), hydrocortisone (HC), and insulin. Arrowheads indicate binucleate cells. Scale bars: 200 μm. (E) Expression of differentiation markers assessed by RTqPCR along the differentiation process (day 0: stem cells, day 3: definitive endoderm, day 11: hepatoblasts, days 14 and 18: differentiated hepatocytes). Results are expressed as relative expression. (F) Secretion of hepatic proteins AFP and albumin assessed by ELISA. Data represent mean ± SEM. (G–K) Functional characterization of HLCs at day 14 of differentiation. (G) Lipoprotein uptake assessed by incubation with Alexa 488-conjugated LDL. (H) Lipid storage demonstrated by Oil Red O staining of the lipid droplets. (I) Glycogen storage demonstrated by periodic acid–Schiff staining. (J) Cells were examined for uptake of indocyanine green. (K) Six hours later, internalized indocyanine green was released. Scale bars: 100 μm.
types for more than 3 months. This approach represents a unique and highly relevant model to study HCV infection both in vitro and in vivo, in the context of patient-specific genetic background.

Results

Hepatic differentiation of human pluripotent stem cells. hiPSCs were generated from skin fibroblasts using STEMCCA lentiviral vectors (ref. 30 and Supplemental Figure 1A; supplemental material available online with this article; doi:10.1172/JCI75456DS1). The hiPSCs share the same characteristics as the hESCs (Supplemental Figure 1B). hESCs remain the gold standard for in vitro differentiation. In this context, we used HSF-6 and H9 hESCs as positive controls for our experiments. Both hiPSCs and hESCs were subjected to a 3-step differentiation protocol (Figure 1). At day 0, hESC and hiPSC colonies were positive for OCT4, a pluripotency marker, and were negative for definitive endoderm and hepatic markers (Figure 1A, day 0). They were first treated for 3 days with 100 ng/ml activin A and basic fibroblast growth factor to generate definitive endoderm exhibiting a more spiky appearance. At day 3, more than 80% of the cells lost expression of OCT4 and became strongly positive for SOX17 and FOXA2 (also known as HNF3β), 2 markers of definitive endoderm (Figure 1B, day 3).

Confluent definitive endoderm cells were passaged at a ratio of 1:3 on growth factor-reduced Matrigel and cultured for 8 days in medium containing 1% DMSO and 100 ng/ml hepatocyte growth factor for hepatic specification. At day 11, around 80% of the cells were positive for AFP, a marker of hepatoblasts, but remained negative for albumin, a marker of mature hepatocytes. Hepatoblasts were also positive for the transcription factor HNF4α, a master regulator of hepatic differentiation (Figure 1C, day 11).

Finally, cells were maturated for 3 more days in medium containing 10⁻⁷ M dexamethasone. At the end of this protocol (day 14), differentiated cells exhibited characteristic hepatocyte morphology: cuboidal shape, distinctive round small nuclei, and compact cell-cell contacts (Figure 1D, day 14). On day 14 and under the best conditions, 80% of the cells were positive for albumin, consistent with hepatic maturation. Treatment with oncostatin M did not show an additional effect on hepatic differentiation and maturation (data not shown). The differentiated HLCs were also positive for α-1-antitrypsin (AAT) and the hepatic transcription factors HNF3β, also known as FOXA2, and HNF4α (Figure 1D and Supplemental Figure 2A). However, most of the cells remained AFP positive, suggesting that they maintained a more fetal phenotype than PHHs (Supplemental Figure 2A).

Staining of HLCs for different isoforms of CYP450, a marker for mature hepatocytes, remained mainly negative (CYP2D6) or weakly positive in few cells (CYP3A4), confirming that the ALB-positive cells differentiated in vitro did not fully mature (Supplemental Figure 2B), as seen in PHHs (Supplemental Figure 3A). Some cells maintained expression of CK7 (cholangiocyte marker) (Supplemental Figure 2C), suggesting that our cell population consists of maturated HLCs, AFP-positive hepatoblasts, and CK7-positive hepatic biprogenitor cells. The differentiated cells were then maintained in hepatocyte culture medium containing 1 μM insulin, 10 μM hydrocortisone, and 10⁻⁷ M dexamethasone for up to 1 week. Cell preparations consisting of more than 60% polygonal cells were considered good enough for further experimentation.

Quantitative RT-PCR (RTqPCR) (Figure 1E) and ELISA assays (Figure 1F) illustrated the transition from pluripotent NANOG- and OCT4-positive stem cells to SOX17- and FOXA2-positive definitive endoderm cells, followed by hepatic specification toward AFP-secreting hepatoblasts (day 11) and further maturation into albumin-secreting HLCs (day 14 and 18). HLCs exhibit a less mature phenotype than adult hepatocytes when compared with PHHs (Supplemental Figure 3B and C).

Finally, we demonstrated other hepatocyte-specific functions of these differentiated cells, including LDL uptake (Figure 1G), lipid storage using Oil Red O staining (Figure 1H), glycogen storage using periodic acid-Schiff staining (Figure 1I), and uptake and excretion of indocyanine green, an organic anion exclusively eliminated by mature hepatocytes through the liver-specific organic anion transporter 1 (Figure 1J and K). These results indicate that this protocol is highly efficient for generating differentiated hepatocytes from human pluripotent stem cells.
specific APOB100 was not expressed or secreted by stem cells and definitive endoderm cells. However, after hepatic specification and during hepatic maturation, APOB was highly overexpressed, and increasing levels of APOB100-containing lipoproteins were detected by ELISA in the supernatant of the hepatoblasts and differentiated hepatocytes (Figure 2D), but at a lower level than the supernatants of PHHs (Supplemental Figure 3, D and E). Taken together, these results show that the differentiated HLCs express several of the main cellular cofactors important for the various steps of the HCV life cycle.

We had previously performed a genome-wide siRNA screen for host factors required for productive HCV infection (34), leading to the identification of 262 host factors involved in different steps of the HCV life cycle in Huh7-derived cell lines. We compared this list, updated with all other host factors described in the literature (a total of 323 genes) (35), with a list of 374 genes shown by microarray as significantly induced (i.e., >4-fold) during one

Expression of cellular factors important for HCV infection. We confirmed that these cells express the cellular factors necessary for productive HCV infection. Interestingly, the stem cells expressed detectable amounts of 4 HCV entry factors: CD81, SCARB1, CLDN1, and OCLN (Figure 2A). During the differentiation process, expression of CD81, SCARB1, and OCLN was maintained at a stable level, while CLDN1 mRNA expression was significantly increased at the hepatic maturation stage. This result was confirmed by visualization of these factors by immunofluorescence of pluripotent stem cells (day 0) and HLCs at day 14 (Figure 2B). Of note, the levels of expression of HCV entry factors were lower in HLCs than in PHHs (Supplemental Figure 3D).

HCV assembly and secretion have been associated with the lipoprotein secretion pathway and particularly with pathways mediated by APOB and APOE (31–33). Here, we show that the expression of APOB and APOE increased during hepatic differentiation, as demonstrated by RTqPCR (Figure 2C).
or several steps of in vitro hepatic differentiation of hiPSCs (36). From this comparison, we identified 8 genes that are important in HCV replication and significantly induced during hepatic differentiation. We confirmed the expression profile of these 8 genes in our differentiation protocol (Supplemental Table 1). Among these factors are the CLDN1 and APOB, as described above (Figure 2, A and C). In addition, microsomal triglyceride transfer protein (MTTP), another gene implicated in the lipoprotein secretion pathway and the assembly and maturation of the viral particles (31, 33), was also significantly induced during hepatic specification and maturation. The functions of 5 other identified genes (FER1L3, [also known as MYOF], FOXA2, SMAD6, TWIST1, HIST1H2BK) in the HCV infection cycle are unknown and under investigation.

\textit{In vitro infection of HLCs with HCV}. At day 14, differentiated cells were inoculated overnight with JFH1-HCVcc at a multiplicity of infection of 0.5. Intracellular HCV RNA was detected by RTqPCR on day 1, 3, and 6 postinfection (pi), indicative of HCV replication (Figure 3A). The levels of intracellular HCV RNA were similar among HCV-infected UCO6-, H9-, and hiPSC-derived HLCs, suggesting similar hepatic maturation among these cells suitable for HCV infection (data not shown). Treatment of the infected cells with 2′-C-methylcytidine, an HCV polymerase inhibitor, inhibited 90% of HCV replication after 2 days of treatment, confirming authentic HCV replication (Figure 3A, red lines). HCV RNA and core antigen were detected in the supernatant by RTqPCR and ELISA, respectively, indicating the production of progeny virus (Figure 3A). More importantly, infectivity assays, consisting of incubation of Huh7.5.1 cells with supernatant from infected HLCs, followed by immunostaining for HCV core, confirmed that these progeny virions were infectious (Figure 3B). The infectious titers in the medium were relatively low (~2 log focus-forming units/ml).

HCV protein expression in infected HLCs, like in PHHs, was not detectable by regular immunofluorescence, probably due to a low level of replication. To circumvent this limitation, we used a recently described approach to visualize low-level infection: the hepatitis C–dependent relocalization (HDFR) assay (37, 38). HLCs were transduced with EGFP-IPS or RFP-NLS-IPS reporters and then infected with HCV. Cells exhibiting relocalization of fluorescence (i.e., cells in which the viral protease NS3-4A cleaved the reporter) could be observed as early as day 2 pi, confirming the infection of these cells by HCVcc (Figure 3C). Three days after inoculation with JFH1-HCVcc at a multiplicity of 0.5, less than 1% of the HLCs showed relocalization events, suggesting that HLCs are much less permissive to HCVcc infection than Huh7 cell lines.

Immunostaining for ALB on infected cells expressing the HDFR vector confirmed that only differentiated cells (ALB positive) were permissive to HCV infection (Supplemental Figure 4). Relocal-
ization events were also detectable after in vitro inoculation with HCV-positive sera of different genotypes (1a, 1b, 2, 3, 4) (Figure 3D and Supplemental Table 2), confirming that HLCs are permissive to infection with authentic HCV particles of different genotypes. Moreover, we observed propagation of HCV infection from infected “relocalized” cells to adjacent cells (Figure 3E), suggesting possible direct cell-to-cell transmission of HCV, as has been described previously in Huh7 cells (37, 38).

We also demonstrated that the HLCs responded to HCV infection by mounting an interferon response, as demonstrated by an early induction of CXCL10 (also known as IP-10), followed by IL-28B (also known as IFNL3) (Figure 3F), as observed in PHHs upon HCV infection (24). However, the percentage of infected cells was probably too low to detect IFN-α, IFN-β, or IL-28 proteins by ELISA in the supernatant. Small amount of CXCL10 protein could be detected in the supernatant of infected cells (data not shown).

**Engraftment of HLCs in MUP-uPA/SCID/Bg mice.** Four million hiPSC-derived HLCs were injected into the spleens of MUP-uPA/SCID/Bg mice (26, 29). Human albumin (hALB) was then monitored by ELISA in the sera of 5 engrafted mice (black lines) and 1 control nonengrafted mouse (red line) for 2 weeks post-engraftment (pe) (Figure 4A). hALB was detectable in every injected mouse as early as day 4 pe and increased during the 2 weeks pe. A total of 25 more mice were engrafted with 4 million hESC- or hiPSC-derived HLCs. hALB was detectable in every injected mouse by day 10 pe, confirming engraftment of HLCs. It is noteworthy that the concentration at day 10 pe varied from 50 to 3,900 μg/ml, but no statistical difference could be seen between hESC- or hiPSC-derived cells (Figure 4B).

At day 14 pe, clusters of engrafted cells could be visualized by immunohistochemistry (IHC) for human Hep Par-1 (Figure 4C) and immunofluorescence assay (IFA) for hALB (Figure 4, D and E). At this time, the percentage of liver repopulation is between 1% and 7% of the liver parenchyma and correlates with serum hALB titers. Tissue morphology after hematoxylin staining (Figure 4D) and immunostaining for glutamine synthetase (Figure 4D) showed that engraftment mainly occurred around central veins and to a lesser extent around portal areas. Nuclear staining for the proliferation marker Ki67 in hALB-positive HLCs confirmed that the engrafted HLCs proliferate in the mouse parenchyma (Figure 4E). Importantly, costaining for human AAT (hAAT) and mouse ALB visualized in different cells confirmed the absence of fusion of the engrafted HLCs with mouse hepatocytes (Figure 4F).

**Maintenance of engrafted HLCs.** Importantly, a detectable level of hALB in the sera of engrafted mice was maintained for more than 100 days pe, confirming long-term engraftment of HLCs (Figure 5A). To better examine the kinetics of hALB production in these engrafted mice, we showed the time course of serum hALB in individual mice (Supplemental Figure 5A). hALB was never detectable in the sera of control nonengrafted mice.

One hundred days pe, H&E staining of sections of engrafted liver revealed large areas of hypoeosinophilic cells, believed to be engrafted human cells (Figure 5B). IHC assays with the Hep Par-1 antibody showed engrafted human HLCs in all engrafted mice (Figure 5C). The percentage of repopulation varied from mouse to mouse, from scattered single cells (<1%) to large areas of positive cells constituting up to 15% to 20% of the liver parenchyma (Figure 5C and Supplemental Figure 5B for pictures of whole
sections). Importantly, the percentage of engraftment correlated with the concentration of hALB in the sera of the mice 100 days pe (Figure 5, A and C). Hep Par-1–positive–engrafted human hepatocytes could also be detected with anti–cytokeratin 18 (anti-CAT18) (Figure 5D). Human cells could be visualized by IFA for hALB (Figure 5E), from small colonies to large areas of the liver parenchyma, consistent with IHC, mainly around central veins (Supplemental Figure 5C). Interestingly, hALB-positive areas often exhibited a distinct histological morphology, with smaller polygonal cells, smaller nuclei, and higher cell density (Figure 5F), similar to what was observed after H&E staining (Figure 5B). On day 100 pe, some of the hALB-positive cells could be stained for proliferation marker Ki67, indicating that HLCs continued to proliferate in the mouse liver parenchyma 100 days pe (Supplemental Figure 5D, arrows and inlay). Sections of nonengrafted control liver did not show areas of lighter-stained cells after H&E staining and were negative for human Hep Par-1, CK18, and hALB (Supplemental Figure 5E).

In situ maturation of engrafted HLCs. We performed coimmunostaining on sections of engrafted liver at day 14 and 100 pe and compared them to the HLCs used for the engraftment. HLCs at the time of engraftment were positive for human AFP (hAFP), hAAT, and hALB (Figure 1D and Supplemental Figure 2A). However, after engraftment, all engrafted hALB-positive cells were positive for hAAT expression (Figure 6A and Supplemental Figure 6A) but negative for hAFP expression (Figure 6B and Supplemental Figure 6A), similar to engrafted PHHs (Supplemental Figure 6B) (see Supplemental Figure 6C for positive control of AFP staining on section of human fetal liver). In situ IFA (Figure 6C) and hAFP ELISA assay (Figure 6D) during the first 2 weeks pe showed that hAFP expression was lost during the first week pe. Nonengrafted control liver tissues and sera were negative for hALB, hAAT, and hAFP (data not shown).

In vitro HLCs were largely negative for isoforms of CYP450 (Supplemental Figure 2B). Here, we show that engrafted hALB-positive HLCs can be stained for different cytochrome P450 isoforms, which are markers of mature hepatocytes (Figure 6, E and F). The CYP antibodies recognized both human and mouse isoforms and were expressed only in the liver parenchyma cells. Interestingly, engrafted HLCs on day 14 pe were already positive for CYPs (Supplemental Figure 7A), like engrafted PHHs (Supplemental Figure 7B), confirming that the maturation of the human cells occurred within 2 weeks pe in the liver parenchyma. Together, these results demonstrate that engrafted HLCs underwent maturation in situ.

HCV infection of engrafted HLCs in vivo. We investigated the permissiveness of the engrafted HLCs to HCV infection in vivo. HLC-engrafted mice were inoculated by tail vein injection with 100 μl diluted plasma from chimpanzees chronically infected with HCV genotype 1a (strain HC-TN) (26). At 1, 2, and 3 months pi, 100 μl sera were harvested and assessed for HCV RNA titer by RTqPCR (Figure 7A). When inoculated with a low dose of HCV (30 chimpanzee infectious dose 50 [CID50]), only 1 of 6 mice exhibited detectable HCV RNA 1 month pi. On the other hand, inoculation with high titer of HCV (1,000 CID50, experiment M3, blue line) led to detectable HCV RNA 1 month pi in 75% of the inoculated mice (Figure 7A). Interestingly, HCV RNA could not be detected 2 weeks after infection, no matter what infectious dose was inoc-
Discussion

The natural history of HCV infection and response to antiviral treatment varies from patient to patient, and host genetic factors may play an important role in these differences. The most widely used cell culture model of HCV infection is based on subclones of the Huh7 cell line (19), which is derived from one individual and thereby is not useful for investigating the influence of patient-specific genetic background on the HCV infection. Productive in vitro HCV infection has also been achieved in PHHs (23), but several limitations (availability, invasive procedure, short window of time for cell culture) make them difficult to use for routine research purposes. It has been shown that human somatic cells can be reprogrammed into a stem cell–like state (hiPSC) and differentiated into mature HLCs (6–11). This approach may hold the key to more relevant models to investigate the link between patient genetic background and HCV infection, pathology, and treatment. Here, we demonstrate that both hESCs and fibroblast-derived hiPSCs can be efficiently differentiated into functional HLCs with many hepatocyte-specific functions. These HLCs do maintain some characteristics of immature fetal hepatocytes, such as expression of AFP and CYP3A7. However, despite these limitations, the differentiated cells express various CYP450 isoforms and reproduce key features of mature hepatocytes, such as glycogen and lipid storage, lipoprotein internalization, and indocyanine green metabolism. More importantly, the HLCs exhibit a level of hepatic differentiation high enough to be infected by HCV, a hepatotropic virus notoriously difficult to culture in vitro (40, 41). Consistent with previous publications (27, 28), we demonstrate that HLCs can be infected with HCVcc in vitro at a low level.
The Journal of Clinical Investigation

The reason for this low level of viral replication in HLCs may be several-fold. Initially, the virus most widely used for research (JFH1-HCVcc) (19) is produced and highly adapted to the Huh7 cell line, is limited to the genotype 2a (JFH1) replicating machinery, and poorly reproduces the characteristic of the natural virus found in the blood of HCV-infected patients in terms of specific infectivity and buoyant density (23, 42, 43). Using sera from HCV-infected patients, we demonstrated that the HLCs are also permissive to infection with authentic HCV of genotype 1a, 1b, 2, 3, and 4. HCV infection also triggers an antiviral innate immune response. In highly permissive Huh7 subclones, the high level of replication is a result of defects in the RIG-1 and TLR pathways, two key actors of the innate immunity pathways (44, 45). These pathways are expected to be functional in our HLCs. HCV-infected PHHs can mount an effective intrinsic innate immunity, with production of interferon and induction of antiviral genes, thereby limiting HCV replication and spread (24). Upon infection of HLCs with HCV, we detected an increased expression of CXCL10 and IL-28B. Thus, this model is a valuable tool to study the innate immune response to HCV infection.

Finally, the low replication of the virus in HLCs may be linked to the less matured phenotype of the HLCs, as compared with that of primary adult hepatocytes (PHHs). We confirmed that proviral host factors are expressed in our HLCs but at lower levels than those of PHHs. Interestingly, the infectious titers in the supernatant of infected HLCs were low compared with the intracellular HCV RNA level, suggesting a relative deficiency in assembly and/or secretion of infectious progeny viruses by the HLCs. Assembly and secretion of progeny virions involve the lipoproteins secretion pathway. While we confirmed that our HLCs expressed high levels of APOB and APOE, the secretion of APOB100-containing lipoprotein was lower than that described for PHHs.

Different strategies have been used to improve the level of differentiation of the HLCs in vitro, such as the use of special matrix, 3D culture, spheroids formation, or coculture with stromal cells (46–52). However, none of them so far allowed maturation of the HLCs at a level similar to that of PHHs. We therefore hypothesized that engraftment of HLCs in the native liver niche, a complex 3D multicellular environment, may allow further maturation of the cells.

Several articles reported engraftment of stem cell–derived HLCs in the livers of various models of transgenic mice (9, 12–16). While one study did not report any engraftment using stem cell–derived hepatocytes in Alb-uPA Rag2γc–/– mice (12), other studies showed low levels of repopulation in induced hepatotoxicity models (9, 10, 13) and in Alb-uPA SCID mice (9). In contrast, 2 studies showed 8%–15% (14) and up to 24% repopulation (15) after transplantation of hESCs or hiPSC-derived HLCs in 2 mouse models of induced hepatotoxicity. Recently, coculture of hiPSC-derived hepatocytes with human umbilical vein endothelial cells and human mesenchymal stem cells has been described to result in formation of 3D “liver buds” in vitro that could be efficiently engrafted and vascularized (53). Engraftment of spheroid-forming HLCs into CCl4–injured mice has also been described (54). Finally, 2 studies showed that human induced hepatocytes, transdifferentiated from human fibroblasts, could be engrafted in the liver parenchyma of transgenic mice and repopulate up to 0.3% to 4.2% of the liver parenchyma of Fah–/– Rag2–/– mice (55) and up to 30% in Tet-uPA Rag2γc–/– mice (56).

Here, we describe engraftment of hESC- and hiPSC-derived HLCs in the liver parenchyma of the MUP-uPA/SCID/Bg mouse model. The MUP-uPA/SCID/Bg model is relatively easy to use for human hepatocyte engraftment (26, 29). Healthy mice are used in this model, which allows a longer window for engraftment compared with the more commonly used ALB–uPA/SCID model. In this model, the differentiated cells were maintained for more than 100 days in the liver parenchyma. The efficiency of repopulation at day 100 pe varied from about 1% to a maximum of 15% to 20% of the liver parenchyma. Concentrations of hALB in the sera of engrafted mice also varied from 100 to 6,415 μg/ml but correlated with percentages of repopulation. The percentage of repopulation is similar to that in some of the published results using human stem cell–derived HLCs (14, 15); however, the hALB titers in our MUP–uPA/SCID/Bg mice are surprisingly high. Only 2 studies, using hiPSC-derived HLCs in a chemically induced hepatotoxicity model (13) or transdifferentiated human induced hepatocytes in a transgene-induced toxicity model (56), reported concentrations of hALB at levels close to ours. The reasons for these differences are not clear and may be explained by different protocols and models used.

One difference in our protocol is the use of the MUP–uPA/SCID/Bg mouse model. Previous studies showed that this model allows high repopulation by PHHs—an optimal number of 4 million PHHs could lead to an average of 40% repopulation and to an average hALB titer of 1,750 μg/ml after intrasplenic injection in mice aged 5–8 months (26). In comparison, HLCs are somewhat less efficient than PHHs for engraftment in the MUP–uPA/SCID/Bg model.

Importantly, our HLC population at day 14 of differentiation was heterogeneous, mainly composed of hALB−hAAT+hAFP− hepatocytes, but also contained hALB−hAAT−hAFP− hepatoblasts and hAFT−hAAT−hCK7− biprogenitor cells. It is not clear whether these heterogeneous cells have a different ability to engraft or be maintained in the liver parenchyma, but this approach differs from that of previous studies engrafting purified, homogeneous HLC populations (9, 15). Interestingly, a previous study demonstrated that hiPSC-derived cells at a different stage of hepatic differentiation have different abilities to repopulate the liver parenchyma of NSG mice, with more mature cells repopulating less efficiently (14). In our experiments, we did not see a correlation between global hepatic maturation (assessed by ELISA for hAFP and hALB at day 14 of differentiation) and efficiency of engraftment (assessed by ELISA for hALB 10 days pe) or long-term maintenance (hALB ELISA at day 100 pe) in the mouse liver parenchyma (data not shown). At this point, we can only hypothesize that the level or nature of maturation of our cells plays a role in the efficiency of engraftment or maintenance in vivo. The absence of quantitative and defined criteria to assess hepatic maturation and cellular homogeneity makes it difficult to compare quality of differentiated cells from study to study.

Increasing hALB concentration during the 14 days pi, higher level of albumin in some mice 100 days pe, and immunostaining for Ki67 at day 14 and 100 suggest that HLCs are capable of proliferation once engrafted in the liver parenchyma, but this ability varies from experiment to experiment.

The individual profiles of secretion of hALB show variable engraftment kinetics—some mice exhibited a high level of serum hALB at early time points followed by gradual decrease. Other
mice, exhibiting a lower concentration of hALB on day 10 pe, showed a gradual increase during the next 3 months, similar to the kinetics of PHH-engrafted mice. However, all the mice but one, engrafted with hESC- or hiPSC-derived HLCs, exhibited detectable hALB titer for more than 100 days pe, confirming that long-term engraftment of HLCs is feasible. The reason for this variability is not totally clear. Decrease of some of the hALB titers suggests death of engrafted HLCs. Analyzing the environmental and cellular factors influencing the survival or death of engrafted cells would be of particular interest to understand and achieve long-term engraftment of transplanted human hepatocytes in the liver parenchyma.

Our data suggest that the quality of the cell preparation may be important for the engraftment kinetics. Four mice (mice 1-1 to 1-4; see Supplemental Figure 5A) were engrafted with the same lot of HLCs and showed decreasing hALB titers at late time points. However, cell quality was not the only reason, as cells from a same preparation achieved different fates after engraftment in mice of a same litter (for example, M101/102 compared with M103/105; see Supplemental Figure 5A). To be noted, we didn’t see correlation between repopulation efficiency and sex or age of the mice at time of the engraftment (data not shown).

More importantly, we demonstrated that engrafted stem cell-derived HLCs undergo further maturation once in the hepatic niche, as described recently with transdifferentiated hepatocytes (56) and iMPC-Heps (57). While in vitro HLCs always expressed high levels of both fetal (hAFP) and mature (hALB, hAAT) hepatocyte markers, all engrafted cells lost expression of hAAT. ELISA assay at early time points revealed loss of AFP within the first week pe. After engraftment HLCs became positive for CYP450 isoforms, markers of mature hepatocytes, confirming that the cells acquired a more mature phenotype in the liver parenchyma within 2 weeks pe. Importantly, the in situ maturation of the cells could be associated with a higher expression and production of hALB by the cells, which could be partly accountable for the rapid increase of serum hALB titers after engraftment.

Because mice were injected with a heterogeneous cell population, we looked for appearance of teratomas at the time of sacrifice. No teratomas were detected in mice sacrificed 100 days pe. Recently, a study showed that hepatocytes generated from human fibroblasts, functionally similar to HLCs, could be safely engrafted in the livers of FAG mice (<1 cm) was performed in the upper left quadrant of the abdomen of mice between 5 and 8 months old, as this has been described with PHHs. Here, the engrafted HLCs maintained a high level of maturation and detectable level of HCV RNA for more than 3 months. We believe this novel in vivo system constitutes a unique model for studying long-term HCV infection of HLCs.

Genome-wide association studies have demonstrated that genetic polymorphisms near the IL28B gene are highly associated with spontaneous virus clearance (60) and response to antiviral treatment (61) in patients infected with HCV. However, in vitro studies of these mechanisms are limited by the lack of a relevant in vitro model to study HCV in the context of the patient’s genetic background. Here, for the first time, we demonstrate that hepatocytes differentiated from patient-specific hiPSCs can be infected both in vitro and in vivo by HCV. Our approach represents a pertinent and valuable model to study HCV infection and subsequent innate immune response in the context of patient-specific genetic background.

Methods

Hepatic differentiation, characterization of differentiated cells, and in vitro HCV infection of HLCs. Detailed protocols for hepatic differentiation of hESCs and hiPSCs, characterization of HLCs, and in vitro HCV infection and monitoring are provided in the Supplemental Methods and Supplemental Tables 3-5.

Engraftment of HLCs in MUP-uPA/SCID/Bg mice. HLCs were incubated for 10 minutes at 37°C in a solution of 0.25% Trypsin and 2.21 mM EDTA in HBSS (Cellgro), further dissociated by pipetting, and washed twice in PBS (centrifugation 50 g) for 10 minutes at 4°C. Resuspended cell pellet was filtered through a 40-μm nylon mesh to remove non-dissociated cells. Trypan blue exclusion test was performed: only cell preparations with viability over 60% were used for engraftment. After one more wash, cells were resuspended in HBSS (Vedco) at a final concentration of 20 million living cells per ml.

Engraftment was performed following the procedure based on our experience with PHHs (26). Only heterozygotes MUP-uPA/SCID/Bg mice between 5 and 8 months old were used, as this has been described as the best age to ensure the most optimal engraftment with PHHs. Briefly, after anesthesia with isoflurane inhalation, a short incision (<1 cm) was performed in the upper left quadrant of the abdomen of the recipient mouse. The dark spleen could be visualized through the intact peritoneum. 4 million HLCs (the optimal number of cells when injecting PHHs; ref. 26), in a maximum volume of 200 μl HBSS, were injected with a 27-gauge needle directly into the spleen. Control non-engrafted mice were injected with 200 μl HBSS. The skin was finally sutured. Details about mice, cell type, and the differentiation protocol used for the engraftments are provided in Supplemental Table 5.

Three months pi, all the inoculated HLC-engrafted mice showed high serum titers of HCV RNA (4-6 log [copies HCV RNA/ml]). The reason for this delayed appearance of detectable HCV RNA in some of the HLC-engrafted mice is not clear but may be linked to an expansion of the engrafted HLCs or to a higher level of maturation. However, at day 100 pe, HCV RNA did not correlate with the concentration of hALB, suggesting that the level of repopulation is not a limiting factor to HCV spreading and that a higher level of repopulation is not enough to attain high viral titer.

This in vivo model of HCV infection shows long-term infection. The viral titers after in vitro infection tend to drop after 2 weeks (data not shown), associated with a dedifferentiation of the HLCs with long-term cell culture. It is similar to what has been described with PHHs. Here, the engrafted HLCs maintained a high level of maturation and detectable level of HCV RNA for more than 3 months. We believe this novel in vivo system constitutes a unique model for studying long-term HCV infection of HLCs.
At different time points after engraftment, mice were bled from the tail veins, and sera was collected after centrifugation. 30 μl sera were used to measure the concentration of hALB (Bethyl Laboratories) and hAFP (Calbiotech) by ELISA according to the manufacturers instructions.

Mice were sacrificed at day 14 or 100 pe, and liver tissues were collected. Liver tissues were fixed with 4% phosphate-buffered formalin overnight, processed, and embedded in paraffin. Sections were routinely stained with Eosin and Hematoxylin (both from Sigma-Aldrich) to monitor the liver parenchyma architecture and visualize the human cells (appearing white after eosin staining). Detailed protocols and antibody information for in situ IHC and IFAs are provided in the Supplemental Methods.

**HCV in vivo infection of chimeric mice.** The chimeric mice were inoculated intravenously with 100 μl diluted plasma from chimpanzees infected with genotype 1a (strain H), 1b (strain CGB1b) (62), and 3a (strain S52) (30 or 1,000 CID₅₀ per ml of genotype 1a, 100 CID₅₀ per ml for genotype 1b and 3a). The infectivity of genotype 1a and 3a has been previously demonstrated in MUP-uPA/SCID/Bg mice engrafted with PHHs (26).

Serum samples were obtained by tail bleeding at day 30, 60, and 90 after inoculation. Viral RNA was isolated using the QIAamp UltraSens Virus Kit (QIAGEN) and assessed for HCV RNA by RTqPCR as described previously (21, 63, 64). HCV core antigen was assessed using the QuickTiter HCV Core Antigen ELISA Kit (Cell Biolabs) according to the manufacturer’s instructions.

**Statistics.** In vitro data are expressed as average of representative experiment, and 2-tailed Student’s t test was performed to assess statistical significance. The results from in vivo experiments are presented for each individual animal. A P value of less than 0.05 was considered significant.

**Study approval.** All engraftment and animal procedures were performed according to the NIH guidelines for animal care and were approved by the Office of Animal Care and Use, Bethesda, Maryland, USA (protocol NIDDK-K090-LDB-13) and the Center for Biologies Evaluation and Research/FDA Institutional Animal Care and Use Committee, Bethesda, Maryland, USA (animal study protocol no. 2001-42).

**Acknowledgments.** We wish to thank Ronda Sapp, Frank Qisheng Li, Zhensheng Zhang, Zongyi Hu, Emmanuel Thomas, and Chen Du (all from NIDDK); Marian E. Major (FDA); Stephen Strom (Karolinska Institute); Hengli Tang (University of Florida); and the Flow Cytometry Core of National Heart, Lung, and Blood Institute for technical assistance and helpful discussions. This work was supported by the Intramural Research Program of the NIDDK (Z01 DK54504 and DK54505) and the NIH Center for Regenerative Medicine. PHHs were provided by the NIH-funded Liver Tissue Procurement and Cell Distribution System (N01-DK-7-0004/HHSN26700700004C).

Address correspondence to: T. Jake Liang, Liver Diseases Branch (LDB), NIDDK, NIH, 9000 Rockville Pike, Bldg. 10, Room 9B16, Bethesda, Maryland 20892, USA. Phone: 301.496.1721; E-mail: jakei@bdg10.nih.gov. Or to: Arnaud Carpentier, Liver Diseases Branch (LDB), NIDDK, NIH, 9000 Rockville Pike, Bldg. 10, Room 8D46, Bethesda, Maryland 20892, USA. Phone: 301.451.1674; E-mail: arnaud.carpentier@nih.gov.


