Autoantibodies in systemic autoimmune diseases: specificity and pathogenicity

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Introduction

The first autoantibodies were discovered in the late 1940s, when both antinuclear antibodies (ANAs) and rheumatoid factors (RFs) were described as serum factors that could bind nuclear antigens and immunoglobulins, respectively (1, 2). ANA and RF were recognized as diagnostic features of systemic lupus erythematosus (SLE) or RA, respectively, and as contributors to disease pathogenesis. It is now becoming increasingly clear that autoantibodies play a pivotal role in the pathogenesis of many diseases and that autoantibodies mediate both systemic inflammation and tissue injury (3).

Here we focus mainly on the autoantibodies associated with the autoimmune diseases SLE and RA, and we discuss the recent developments on the generation of autoreactivity and the contribution of these antibodies to disease pathogenesis. We propose a model in which autoreactive B cells escape from tolerance because they bind TLR ligands and undergo clonal expansion because of continuous exposure to antigen. Furthermore, the generation of immune complexes containing TLR ligands leads to systemic inflammation through the activation of innate immune cells. This systemic inflammation is part of the disease process and further impairs tolerance mechanisms. While we cite data from studies of SLE and RA to support this paradigm, we believe the model has broader applications.

Central versus peripheral tolerance defects

During B cell development in the BM, autoreactivity is prevented through receptor editing, apoptosis, and induction of anergy in B cells expressing an autoreactive B cell receptor (BCR). When immature B cells express surface IgM, recognition of a self-antigen in the BM can induce these processes. Transitional B cells emerging from the BM continue to mature in the spleen, where additional tolerance mechanisms are in place. The exact mechanisms in this peripheral compartment are not fully understood, but they require ligand recognition by the BCR, similar to the tolerance checkpoints in the BM (4). After this stage, mature naive B cells can be activated upon antigen recognition, allowing them to enter the germinal center (GC). The GC is a site of rapid clonal expansion of B cells, affinity maturation, class switching, and differentiation to memory B cells or plasma cells. A unique feature of GC-matured B cells is that they have undergone extensive somatic mutation of the antibody genes. As somatic hypermutation can give rise to de novo autoreactivity as well as enhance affinity of existing autoreactive B cells, additional tolerance checkpoints in the post-GC compartment have been suggested to effectively prevent autoreactivity in memory B cells and plasma cells, through either apoptosis or receptor editing (5, 6).

The ability to sequence and clone Ig genes from individual B cells has opened the opportunity to study these tolerance checkpoints in health and disease. The results from such studies using B cells from healthy individuals suggest that a high number of autoreactive B cells is generated in the BM and that sequential tolerance checkpoints lead to a gradual decrease in autoreactivity as the B cells mature (7).

The presence of autoreactive B cells in healthy individuals indicates that central tolerance is not sufficient to remove all autoreactive B cells with self-targeting BCRs. Indeed, it is well-established that both the B1 subset and marginal zone B subset contain high numbers of autoreactive B cells, and in animal models B cells from these subsets can give rise to pathogenic autoreactivity (8–10). Although defects in early B cell tolerance have been found in patients with autoimmune disease (11, 12), current data suggest that most of their autoreactive B cells are derived from nonautoreactive precursors. Autoantibodies derived from memory B cells in systemic autoimmune disease are often class-switched and highly somatically mutated, which suggests that they have been involved in GC reactions (13–19). Memory B cells making anti-DNA antibodies and anticitrullinated protein antibodies (ACPAs) in patients with autoimmune disease are thought to be derived from nonautoreactive precursors because back-mutation of their somatic mutations.

Conflict of interest: Betty Diamond owns stock in Macrogenics and receives research support from Merck-Serono.

to germline sequences removes their autoreactivity (17, 18). Furthermore, although naïve polyreactive B cells can recognize citrullinated proteins or DNA and therefore could give rise to ACPAs and anti-DNA-producing plasma cells, the affinity of ACPAs and anti-DNA antibodies from naïve B cells is greatly reduced compared with antibodies in patient serum, suggesting that these naïve B cells have undergone affinity maturation before developing into plasma cells (11, 18). While sequence analyses of autoantibodies have thus far only been performed for a limited number of patients and access to plasma cells is limited by the unavailability of BM samples, there is consistent evidence that many autoreactive B cells in autoimmune disease mature in the GC.

Taken together, these data suggest that defects in central and early peripheral tolerance checkpoints are present in autoimmune disease patients, but the defects in early tolerance checkpoints do not necessarily lead to the generation of pathogenic autoantibodies. As studies point to an important role of somatic hypermutation during GC reactions as a source of autoreactivity, post-GC checkpoints are likely defective in patients with autoimmune disease, but our understanding of these checkpoints is limited.

**T cell help for autoantibody production**

Development of high-affinity antibodies requires cognate B cell/T cell interactions to initiate and sustain the GC. Somatically mutated autoantibodies in autoimmune disease are thought to be derived through T cell–dependent pathways, especially because the presence of autoantibodies is often correlated with the presence of specific HLA alleles (20, 21). However, at present little is known about the specificity of T helper cells that are able to stimulate autoantibody-producing B cells.

The most direct pathway for T cell helper function is for T cells to recognize the same antigen as the B cells. In RA, one of the genetic risk HLA alleles, HLA-DRB1*0401, has recently been associated with presentation of citrullinated peptides, allowing for activation of citrulline-specific T cells (22). These T cells may engage in direct antigen-specific interactions with ACPA-producing B cells when citrullinated antigens are internalized through BCR-dependent recognition. However, the T cell specificities involved in the activation of other autoreactive B cells, such as DNA-reactive B cells, are not known. Some studies suggest that the epitopes for B cells and T cells do not necessarily need to be the same. For example, B cells can internalize multimolecular complexes through BCR-mediated recognition and can then process peptides from different proteins present in the complex. This internalization of multimolecular complexes clearly enables processing of nucleoproteins capable of binding DNA in lupus-prone mice and SLE patients (23–26).

T cell help for autoantibody-producing B cells can also originate from T cell responses to foreign antigens through molecular mimicry between viral or bacterial antigens and self-antigens (27–30). Activation of nonautoreactive T cells by foreign antigens can lead to activation of B cells that recognize the foreign antigen but also cross-react with a self-antigen (31). It is hypothesized that such activation can lead to selection and maturation of high-affinity self-reactive B cells.

In addition to the typical GC response, class switching and somatic hypermutation can also occur outside of the classical GCs. In several autoimmune diseases, ectopic lymphoid structures can develop, usually in the target organ, such as the synovial tissue in RA or the kidney in SLE. These structures closely resemble the GC with the presence of B/T lymphoid clusters that include T follicular helper cells and follicular dendritic cells. Ectopic lymphoid organs are often associated with the production of high levels of autoantibodies, suggesting that local activation of these cells can contribute to T cell–dependent B cell differentiation, as in the GC (32, 33).

**Target antigens for autoantibodies**

Autoantibodies can be directed against a variety of molecules, such as nucleic acids, lipids, and proteins, and these target antigens can be located in the nucleus or cytoplasm, on the cell surface, or in the...
is substituted for arginine (34, 35). Most of these antigenic citrullinated proteins are extracellular proteins, although intracellular citrullinated proteins have also been identified as targets of ACPAs. To date, it is unknown whether intracellular or extracellular citrullinated antigens are the main driver of the ACPA response.

B cell propagation in response to self-antigen or foreign antigen

The intracellular location of many autoantibody targets raises the question of how these antigens can drive clonal expansion, a process that requires the recognition of antigens through the BCR, which occurs in the extracellular space.

Two main hypotheses exist that can explain how autoantibodies arise that recognize intracellular self-antigens (Figure 1A). The first theory proposes that apoptosis plays an important role as a source of nuclear and cytoplasmic self-antigens. During this process of programmed cell death, apoptotic blebs appear on the cell surface and microparticles can be released from the cell, both of which contain several cytoplasmic and nuclear components. These include several...
of the intracellular antigens recognized by autoantibodies, such as histones, dsDNA, RNA, ribonucleoproteins (RNPs), and myeloperoxidase and proteinase 3 in neutrophils (36–39). Furthermore, the process of apoptosis induces several modifications in the nucleus, including histone citrullination and acetylation, which can be recognized by autoantibodies in RA and SLE, respectively (40, 41). Apoptotic blebs are normally quickly engulfed by phagocytes to prevent immune activation (42); however, a failure to do this, due either to a genetic defect in one of the apoptosis clearance pathways (as often seen in autoimmune disease) or to excessive release of these blebs due to inflammation, allows for the sustained presence of these self-antigens in the extracellular environment (43).

Another pathway for propagation of autoantibody responses toward intracellular antigens is through cross-reactivity with extracellular antigens. An example of such cross-reactivity is anti-dsDNA antibodies in SLE, which have been shown to recognize antigens in the glomerular basement membrane, where these autoantibodies can deposit (44–47). While the binding of some anti-dsDNA antibodies to glomeruli depends on the presence of nucleosomes containing DNA on the glomerular membrane, the contribution of cross-reactivity to renal pathology has also been demonstrated (48, 49).

In RA, ACPAs can target both intracellular and extracellular citrullinated antigens that arise during an inflammatory response (Figure 1B and refs. 34, 35, 50). It has not been formally shown, but it is likely that the citrulline epitopes on different antigens form the basis for cross-reactivity between intracellular antigens and extracellular antigens. In cardiomyopathy, autoantibodies targeting cardiac myosin, an intracellular antigen, have been shown to cross-react with the β-adrenergic receptor, thereby inducing stimulation of cardiomyocytes. These findings suggest that cross-reactivity between intra- and extracellular antigens can drive clonal expansion.

Similarly, cross-reactivity between foreign antigens and self-antigens can drive the expansion of autoreactive B cells (Figure 1C). Several autoantibodies exhibit such cross-reactivity: dsDNA autoantibodies cross-react with pneumococcal bacteria and Epstein-Barr virus (29, 45) and anti-neutrophil cytoplasm antibodies (ANCAs) recognize the bacterial adhesin FimH (28). However, this cross-reactivity between foreign antigens and intracellular self-antigens still requires the release of intracellular antigens through apoptosis or cross-reactivity to surface antigens in order for autoantibodies to bind and exert their pathogenic activity.

We speculate that B cells with BCRs that recognize only intracellular antigens will propagate less well than those that cross-react with more ubiquitous cell surface or extracellular antigens. Therefore, the basis for clonal expansion of many autoreactive B cells can be found in their cross-reactivity.

**TLR recognition and its role in escape from tolerance**

Many antigens recognized by autoantibodies are able to trigger pattern recognition receptor (PRRs), including TLRs and Nod-like receptors (NLRs). These receptors are listed in Table 1 and include DNA, RNA, citrullinated fibrinogen, cardiac myosin, cardiolipin, and oxidized phospholipids (51–56).

B cells express several TLRs for nucleic acids, including TLR7 and TLR9. Engagement of these TLRs activates B cells. Notably, many antigens recognized by autoantibodies are ligands for B cell TLRs (Table 1), suggesting a role for TLR recognition and its role in escape from tolerance. Indeed, mice deficient in TLR7, TLR9, or the TLR-associated signaling adaptor MyD88 are partially protected against development of ANAs and lupus-like symptoms (57–60). This protection derives from the effects that TLR ligation has on reducing B cell tolerance.

Combined BCR and TLR ligation by nuclear antigens has been shown to induce a synergistic signaling response, mediated by colocalization of TLR9 and antigen internalized through BCR (61–63). Thus, self-antigens able to activate endosomal TLRs and BCRs will induce robust activation in B cells. Some studies suggest that TLR activation of immature B cells can dysregulate central tolerance pathways (64–66). We hypothesize that TLR ligation might also explain the failure of peripheral tolerance in autoimmune disease. Several studies show increased TLR expression and function in activated or memory B cells compared with immature B cells, allowing for robust activation through TLR in the GC (67–70). As tolerance in the GC is thought to be driven by a survival advantage through affinity-driven selection, combined BCR/TLR ligation in the GC may facilitate positive selection and escape from tolerance.
Pathogenicity of autoantibodies
The pathogenicity of autoantibodies has been extensively debated. Proof of pathogenicity is derived from the ability of autoantibodies to induce cellular damage or immune activation in vitro or to passively transfer autoimmune disease in animal models, as well as the effectiveness of B cell–targeting therapies. Although such evidence is not established in all autoimmune diseases, the most common systemic autoantibodies, such as ANAs and ACPAs, are considered pathogenic.

Pathogenicity has been most thoroughly studied in mouse models with lupus-like disease. Several studies have shown that passive transfer of anti-DNA antibodies or expression of autoreactive transgenic BCRs can cause inflammation in the kidneys, although not all antibodies have equal capacity to do so (78–80). In contrast, only limited evidence of the pathogenicity of ACPAs or RF is present in mouse models of arthritis. This may be due to the absence of these autoantibodies in most mouse models; however, when ACPAs are adoptively transferred into mice with low-level inflammation caused by anti-collagen antibodies, ACPAs enhances arthritis, directly implicating them in the inflammatory process in the joint (80, 81).

Autoantibody effector functions
The effector functions that are employed by autoantibodies resemble those of antibodies in general and are therefore largely dependent on their isotype. Specific autoantibody isotypes are present in different diseases, with IgA antibodies present in antiphospholipid syndrome and IgE autoantibodies present in SLE (82–84). IgM is a potent activator of the complement component 1, q subcomponent (C1q), which has recently been shown to inhibit TLR activation by acting as a ligand for the inhibitory leukocyte-associated immunoglobulin-like receptor 1 (LAIR1), suggesting that IgM might inhibit immune effector responses (86). In support of this model are experiments that showed that an inability to secrete IgM in B cells leads to IgG autoantibody production in mice (87).

A major pathway for immune activation and tissue damage for systemic autoantibodies is through formation of immune complexes. In situ immune complexes form when antigen is present at specific sites within tissues. Different monoclonal anti-DNA antibodies have distinct patterns of renal deposition, depending on their specificity and cross-reactivity (45). Although RA is associated with the presence of circulating immune complexes, the disease is mainly limited to the joints. This is thought to arise in part from preferential binding of antibodies to citrullinated proteins present in the synovium (88).

Immune complexes formed in situ can activate the complement system and Fc receptors, leading to tissue inflammation and damage. Complement activation by autoantibodies has been shown to occur through both classical and alternative pathways (89, 90) and can lead to direct cell lysis and damage as well as recruitment of leukocytes to further enhance inflammatory responses. Complement-mediated cell lysis has also been shown to serve as a source of antigen, such as DNA, RNP, and citrullinated proteins (91).
Activating Fc receptors are expressed by a variety of cells, particularly by immune cells of the myeloid lineage. Autoantibodies have been shown to activate these immune cells through FcγR-dependent pathways (92–94). In vivo, FcγR-deficient mice often show decreased autoimmunity (95, 96). The capacity of IgG to activate Fc receptors contributes to their increased pathogenicity compared with IgM autoantibodies. However, IgM-RF was recently shown to enhance FcγR-dependent activation by ACPAs, demonstrating that isotypes other than IgG can contribute to pathogenesis (97).

Clinical evidence for autoantibody pathogenesis

Most autoantibodies are present years before disease onset and thus are not merely a consequence of autoimmunity (98–100). Furthermore, for ACPAs, there is evidence for epitope spreading, affinity maturation, isotype switching, and glycosylation changes before clinical symptoms start. Each of these changes is associated with greater pathogenic potential (101–105). In addition to the changes in the intrinsic properties of autoantibodies before disease onset, the target organ can also become susceptible to autoantibody deposition or binding. This is in accordance with the multiple-hit theory of autoimmunity, in which genetic and environmental factors induce an initial break in tolerance leading to production of autoantibodies. A “second hit” is needed for the antibody to exert pathogenic effects and to sustain the inflammatory cascade of the autoimmune response. This phenomenon is perhaps most clearly illustrated in myositis, in which the autoantibodies bind regenerating myocytes and thus sustain a cycle of injury and regeneration (106).

Chronic inflammation

As stated above, many autoantibodies target ubiquitously expressed intracellular molecules. For antibodies to bind these targets, these antigens need to be released to the extracellular environment, where antibodies or immune complexes are deposited. As the self-antigens are ubiquitously present, autoantibody effector mechanisms fail to eliminate the antigen, causing inflammatory injury to tissues that lead to additional release of these intracellular antigens. This process causes a vicious cycle wherein each disease flare can initiate further autoantibody production and tissue injury (107–109). Furthermore, combined TLR- and Fc receptor-mediated activation of myeloid cells often leads to an enhanced inflammatory response that is thought to contribute to the chronicity of inflammation (51, 94, 110, 111). Therefore, those self-antigens that can most potently activate B cells because they are TLR ligands may also potently enhance pathogenic inflammatory responses (Figure 3).

Conclusions

A break in tolerance against self-antigens or modified self-antigens can lead to the production of autoantibodies. Such a breach in tolerance can originate from a lack of clearance of apoptotic debris, inflammation-mediated modification of self-antigen, or cross-reactivity between foreign and self-antigens. Some autoantibodies cross-react between an intracellular antigen that is a TLR ligand and cell surface or extracellular antigen, allowing for continued B cell activation and autoantibody-mediated injury. After initiation of autoantibody production, inflammation may cause release of intracellular or modified self-antigens, leading to antibody effector function and instigation of clinical symptoms as well as to the propagation of the autoreactive B cell clones. These pathogenic processes are mainly mediated by IgG autoantibodies, although other isotypes may contribute. TLRs play an important role during both initiation and pathogenesis of autoantibody responses, as they can recognize several self-antigens such as alarmins. TLR activation can enhance the initiation of autoantibody responses and can synergize with Fc-medi- ated effector functions. We speculate that those autoreactive B cells that can escape tolerance and expand and differentiate in response to antigen have BCRs that bind antigens that associate with or are TLR ligands. Reactivity with abundant cell surface or extracellular antigens permits the clonal expansion and survival of these B cells once inflammation has subsided. Furthermore, this cross-reactivity may contribute to their pathogenicity.

Acknowledgments

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