Interruption of progerin–lamin A/C binding ameliorates Hutchinson-Gilford progeria syndrome phenotype

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Hutchinson-Gilford progeria syndrome (HGPS) is a rare autosomal dominant genetic disease that is caused by a silent mutation of the LMNA gene encoding lamins A and C (lamin A/C). The G608G mutation generates a more accessible splicing donor site than does WT and produces an alternatively spliced product of LMNA called progerin, which is also expressed in normal aged cells. In this study, we determined that progerin binds directly to lamin A/C and induces profound nuclear aberrations. Given this observation, we performed a random screening of a chemical library and identified 3 compounds (JH1, JH4, and JH13) that efficiently block progerin–lamin A/C binding. These 3 chemicals, particularly JH4, alleviated nuclear deformation and reversed senescence markers characteristic of HGPS cells, including growth arrest and senescence-associated β-gal (SA–β-gal) activity. We then used microarray-based analysis to demonstrate that JH4 is able to rescue defects of cell-cycle progression in both HGPS and aged cells. Furthermore, administration of JH4 to LmnaG609G/G609G-mutant mice, which phenocopy human HGPS, resulted in a marked improvement of several progeria phenotypes and an extended lifespan. Together, these findings indicate that specific inhibitors with the ability to block pathological progerin–lamin A/C binding may represent a promising strategy for improving lifespan and health in both HGPS and normal aging.

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Introduction

Hutchinson-Gilford progeria syndrome (HGPS) is a rare autosomal dominant genetic disease that is caused by a silent mutation of the LMNA gene encoding lamins A and C (lamin A/C). The G608G mutation generates a more accessible splicing donor site than does WT and produces an alternatively spliced product of LMNA called progerin, which is also expressed in normal aged cells. In this study, we determined that progerin binds directly to lamin A/C and induces profound nuclear aberrations. Given this observation, we performed a random screening of a chemical library and identified 3 compounds (JH1, JH4, and JH13) that efficiently block progerin–lamin A/C binding. These 3 chemicals, particularly JH4, alleviated nuclear deformation and reversed senescence markers characteristic of HGPS cells, including growth arrest and senescence-associated β-gal (SA–β-gal) activity. We then used microarray-based analysis to demonstrate that JH4 is able to rescue defects of cell-cycle progression in both HGPS and aged cells. Furthermore, administration of JH4 to Lmna G609G/G609G mice, which phenocopy human HGPS, resulted in a marked improvement of several progeria phenotypes and an extended lifespan. Together, these findings indicate that specific inhibitors with the ability to block pathological progerin–lamin A/C binding may represent a promising strategy for improving lifespan and health in both HGPS and normal aging.

Despite several reports dealing with the role of this protein in cell-cycle regulation (11, 17, 18) and senescence (12, 19), how progerin induces various cellular defects and premature aging remains to be revealed. Remarkably, proteomic studies and 2-hybrid approaches have identified lamin A/C as a progerin-binding protein (20, 21). On this basis, we hypothesized that the interaction of progerin with lamin A/C contributes to the development of the senescence phenotype of HGPS and aged cells.

Results

Direct interaction between progerin and lamin A/C but not lamin B. To investigate our hypothesis, we performed a binding assay and found that C-terminal progerin and lamin A associated with lamins A and C but not with lamin B1 (Figure 1A and Supplementary Figure 1A; supplemental material available online with this article; doi:10.1172/JCI84164DS1). In addition, the binding affinity between progerin and lamin A/C was stronger than that between lamin A/C and lamin A/C (Figure 1B and Supplementary Figure 1B). To dissect the progerin domain involved in lamin A/C binding, we performed a glutathione-S-transferase pull-down (GST pull-down) assay using a His-tagged lamin A N-terminal region (LMNA-N; aa residues 1-300) and a lamin A middle region (LMNA-M; aa residues 301-564) with bead-coupled GST-C-terminal fragments of lamin A (GST-LMNA) or progerin (GST-progerin). We found that the middle region of lamin A was responsible for progerin binding (Figure 1, C and D). We could confirm the direct interaction of both proteins through Far Western blot
Figure 1. Identification of a lamin A–progerin–binding inhibitor. (A) Interaction of progerin and LMNA A/E but not lamin B. GST pull-down assay using lysates from HEK293 cells transfected with lamin A, B, C and progerin after incubation with the C-terminal 100-aa sequence of lamin A (564–664 aa; GST-LMNA) or 50-aa sequence of progerin (564–614 aa; GST-progerin). (B) Strong interaction between lamin A and progerin. Lamin A–progerin interaction was detected after a 1-hour incubation. However, progerin did not show evidence of self-association. Rx, treatment. (C) Direct interaction between lamin A and progerin. In vitro–binding assay using His-tagged N-terminal (1–300 aa; LMNA-N) and the middle region (LMNA-M) of lamin A after incubation with GST-LMNA or GST-progerin. Associated proteins were analyzed by Western blotting. N, LMNA-N; M, LMNA-M. (D) Scheme of the interaction between lamin A and progerin. (E) ELISA results. His-tagged LMNA-M was fixed in a 96-well plate and incubated with GST-LMNA or GST-progerin. Photograph of the 96-well plate (inset) and ELISA value. (F and G) Progerin did not induce nuclear deformation in neuron or embryonic neuron cells. Transfection of GFP-progerin or lamin A alone into the neuroblastoma cell line SH-SY-5Y (F) and the mouse neuroembryonic cell line N2A did not induce nuclear deformation. However, cotransfection could induce it. Cells were transfected with the indicated vectors for 24 hours. For visualization of the nuclear membrane, cells were stained with emerin (red) and DAPI for DNA (blue). Scale bar: 10 μm.

The above findings prompted us to search for putative progerin–lamin A–binding inhibitors by using an ELISA-based screening system. In brief, progerin, immobilized in 96-well plates, was incubated sequentially with a chemical library and His–lamin A (Supplemental Figure 1, A and B). This screening identified 5 chemicals that blocked this interaction (JH1, JH4, JH13, LG-02, and KEK-2-24, Supplemental Figure 2C). Because KEK-2-24 is a mixture of compounds from Panax ginseng and LG-02 requires complicated synthetic steps, we excluded them from further study. A detailed synthetic pathway for the selected chemicals is described in the Chemical Synthesis section of the Supplemental Methods. Interestingly, in spite of the blind screening, the selected chemicals were structurally similar (Supplemental Figure 2C) and showed very low cytotoxicity (Supplemental Figure 2D). Therefore, we decided to further investigate the activity of JH chemicals, particularly JH4, because JH1 and JH13, despite their similar effects, show very low solubility. We first confirmed their inhibitory effect of the JH chemicals on the binding between progerin and lamin A through in vitro binding assays (Supplemental Figure 2E). Indeed, less than 1 μM of JH4 (IC50 = 0.65 μM) could block 50% of the interaction between progerin and lamin A without disruption of lamin A self-association (Supplemental Figure 2E).

The effect of progerin–lamin A–binding inhibitors in HGPS model cells. To confirm the inhibitory effect of JH chemicals on binding between progerin and lamin A, we performed the GST pull-down assay and observed that JH4 blocked the interaction of GST-lamin A with GFP-progerin, but not with GFP–lamin A, in a GST pull-down assay using cell lysates (Figure 2A and Supplemental Figure 3A). However, the farnesyltransferase inhibitor FTI-277, known to ameliorate nuclear deformation (23, 24), did not show an obvious effect on progerin–lamin A binding (Figure 2A and Supplemental Figure 3B). Next, we checked their in vivo effect in progerin and GFP–lamin A–transfected HEK293 cells through IP assays with an anti-GFP Ab. The binding between progerin and lamin A was markedly reduced by the JH chemicals, while the binding between lamin A and lamin B was not affected by these compounds (Figure 2B). Additionally, a reduction of p16INK4A expression was detected in response to treatment with the JH chemicals (Figure 2B). We also observed the different localization of progerin from that of lamin A upon JH4 treatment (Figure 2C and Supplemental Figure 3C). Moreover, all these JH compounds, and in particular JH4, blocked nuclear deformation (Supplemental Figure 4, A and B). These results indicate that JH4 (and JH13 to some extent) blocked the interaction between progerin and lamin A and alleviated progerin-induced nuclear deformation. However, JH4 did not show any significant effect on the localization pattern of other lami-nopathy-related lamin A mutants (2) such as nuclear speckles of D192G (Supplemental Figure 4C). This result also supports the idea that JH4 possesses selective activity on progerin. Next, we examined the effect of JH chemicals on HGPS cells. Consistent with the above results, JH chemicals, in particular JH4, blocked the interaction between progerin and lamin A in these progeroid cells (Figure 2D and Supplemental Figure 4D), without alteration of progerin mRNA expression (Supplemental Figure 4E). In addition, JH chemicals, and especially JH4, ameliorated the frequency of nuclear deformation of HGPS cells (Figure 2, E and F, and Supplemental Figure 4, F and G).

Specific interaction of JH4 and progerin. To determine the direct target of JH chemicals, we monitored their effect on the previously observed progerin-p14ARF binding (14). In this assay, we found that JH chemicals blocked the interaction between progerin and p14ARF (Supplemental Figure 5A), but not between p53 and p14ARF (ref. 25 and Supplemental Figure 5B). In addition, JH4 did not block the interaction between progerin and MEL18/BMI1 that is achieved by the middle region of lamin A (ref. 26 and Supplemental Figure 5, C–E). Considering these facts, the C-terminal region of progerin may be the direct target of JH4 (Supplemental Figure 5F). To verify this hypothesis, we generated biotinylated JH4 (Supplemental Figure 5G; B33: precursor for biotinylation; 07: biotinylated JH4; 08: biotin plus linker). Modified chemicals showed inhibitory activity similar to that of JH4 on binding between lamin A and progerin (Supplemental Figure 5H) as well as on nuclear deformation (Fig-
Figure 2. Ameliorating effect of JH chemicals on nuclear deformation. (A) Specific effect of JH chemicals on the binding of lamin A–progerin but not on that of lamin A–lamin A. GST pull-down assay using lysates from HEK293 cells transfected with GFP-LMNA after incubation with GST-LMNA or GST-progerin under treatment with the indicated chemicals. (B) Dissociation of lamin A and progerin by JH4 in human cells. Dissociation of progerin from lamin A/C was confirmed by IP in HEK293 cells. For this, GFP-progerin–transfected HEK293 cells were incubated with the indicated chemicals (5 μM) for 24 hours. A reduction of p16ink4a was also detected in whole-cell lysate. Actin was used as a loading control. SUP, supernatant. (C) JH4 blocked the colocalization of lamin A and progerin. For this, HEK293 cells were cotransfected with GFP-LMNA and nontagged progerin (NT progerin) for 24 hours and incubated with JH4 for an additional 24 hours. JH4 could abolish the colocalization of lamin A and progerin (red) in the nuclear membrane or in nuclear speckles. DAPI was used for DNA staining. Scale bar: 10 μm. (D) JH4 blocks the interaction of lamin A and progerin in HGPS cells. Treatment with 5 μM JH4, but not FTI-277, for 24 hours blocked the interaction of lamin A and progerin in HGPS cells. Cell lysates were immunoprecipitated with a lamin A–specific Ab (H-102, sc-20880) and immunoblotted with a progerin–specific Ab. Normal cells (9 yr; N9) were used as a negative control for progerin. (E) Effect of JH chemicals on nuclear deformation of HGPS cells. Three kinds of HGPS cells were incubated with the indicated chemicals for 24 hours and fixed for lamin A/C immunofluorescence analysis (green). The same effect was observed in other HGPS cells (Supplemental Figure 4F). Scale bars: 10 μm. (F) Quantification of nuclear abnormalities in HGPS cells. JH chemicals, in particular JH4, ameliorated nuclear deformation of HGPS cells. P values were determined by Student’s t test. PPT, protein pellet.

Figure 3A and Supplemental Figure 5I). Then, we checked the location of biotinylated JH4 using avidin-HRP and found that JH4 was located in the nuclear membrane of progerin-transfected cells (Figure 3B) and HGPS, but not in the nuclear membrane of normal cells (Figure 3C and Supplemental Figure 5I). In addition, a biotin-avidin bead pull-down assay showed specific interaction of JH4–biotin with progerin (Figure 3D) but not with lamin A, lamin B, p53, or emerin (Figure 3, D and E), indicating that progerin is the primary and direct target of JH4.

Antisenescence effect of JH chemicals on HGPS cells. We next examined the effect of JH chemicals on senescence. Since an important feature of senescence is an irreversible cell proliferation arrest (27), we checked the effect of JH chemicals on cell proliferation and found that these compounds (and in particular JH4) promoted the proliferation of HGPS cells (Figure 4A). In addition, JH4 promoted the proliferation of other kinds of HGPS cells (Figure 4B). Indeed, JH4 could induce the expression of Ki-67, a cell proliferation marker, in HGPS cells (Figure 4C). SA–β-gal staining, a standard senescence assay (28, 29), also showed antisenescent activity of JH4 in HGPS cells, which was similar to that induced by FTI treatment (Figure 4D and Supplemental Figure 6, A and B). Decoy Receptor 2 (DcR2) expression (29) was also reduced in HGPS cells at both the RNA and protein levels after treatment of these cells with JH chemicals (Supplemental Figure 6, C and D). Since progerin levels were reduced by JH4 (Supplemental Figure 6D) without a reduction of the mRNA levels (Supplemental Figure 4F), we speculated that progerin, dissociated from lamin A, may lose its stability. Indeed, JH4 could reduce the half-life of progerin (Figure 4E), which was restored by incubation with the protesome inhibitor N-acetyl-leucyl-leucyl-norleucinal (ALLN) (Figure 4F). This result is consistent with our previous finding that JH4 reduces progerin expression in immunofluorescence staining (Figure 2C). To confirm the antisenescence effect, we examined the expression of H3K9Me3 and DNA-dependent protein kinase (DNA-PK), whose reduced levels have been proposed as markers of senescence (9, 15). Interestingly, JH4 and JH13 increased the expression of H3K9Me3 (Figure 4G and Supplemental Figure 6, E and F) and the DNA-PK catalytic subunit (DNA-PKcs) in HGPS cells (Supplemental Figure 6, G and H).

Antisenescence effect of JH chemicals on normal aged human fibroblasts. Since progerin-induced nuclear deformation is also observed in aged cells, we hypothesized that the JH chemicals may also decelerate the normal aging process. To address this, we checked the expression of progerin in normal fibroblasts obtained from an 81-year-old donor (N81). These cells expressed progerin at the transcriptional level (Supplemental Figure 7A) and showed abnormal nuclear morphology, similar to what was observed in HGPS cells (Supplemental Figure 7B; arrowheads). These results, together with previous reports showing that progerin accumulates in aged cells and causes nuclear deformation (8, 9), prompted us to hypothesize that the JH chemicals could prevent senescence of aged cells. Indeed, the nuclear morphology of the 81-year-old cells was clearly improved by JH chemicals and FTI-277 (Figure 5A and Supplemental Figure 7C). Moreover, the slow proliferation of normal 81-year-old fibroblasts was completely rescued by JH4 (Figure 5B). To check the possible long-term toxicity of this treatment, we incubated fibroblasts from young, old, and HGPS human tissues with JH4 for 10 days and found that this compound could promote cell growth and colony formation without obvious toxicity (Supplemental Figure 7, D and E). In SA–β-gal assays, we also observed that JH4 could block cellular senescence (Figure 5C). Consistently, expression of Ki-67 (30), a cell proliferation marker, was increased after JH4 treatment (Supplemental Figure 7F). Additionally, increased H3K9Me3 (Figure 5D) and reduced DcR2 and p16ink4a levels were detected by JH4 treatment in normal aged fibroblasts (Figure 5E). Thus, we next checked the effect of JH4 on oncogene-induced or growth factor–induced senescence (31, 32). Senescence induced by IGF-1 treatment or transfection of oncogenic Ras into WI-38 (normal foreskin fibroblast) was not blocked by JH4 (Supplemental Figure 7, G and H), because oncogene-induced senescence did not induce progerin (Supplemental Figure 7I), nor was induction of p16 blocked by JH4 treatment (Supplemental Figure 7I). In addition, JH4 did not promote cell viability in human cancer cell lines, despite a 72-hour treatment (Supplemental Figure 8A), suggesting that the antisenescence effects of JH4 do not result in the promotion of cancer progression. Instead, JH4 could ameliorate the nuclear deformation in renal cell carcinoma induced by progerin expression (Supplemental Figure 8B), with slight growth suppression at a high concentration (Supplemental Figure 8C). These results suggest that JH chemicals, in particular JH4, selectively block progerin-mediated senescence, without growth promotion of progerin-negative cells.
present in the array, 1,921 showed differential expression (cutoff of 2-fold; Figure 5A and Supplemental Figure 9A). After elimination of nonsignificant changes resulting from the comparison between HGPS and young cells (Supplemental Figure 9A and Supplemental Table 1), we classified differential genes in an HGPS unique gene set (HUG), an aging unique gene set (AUG), and an HGPS/aging-related gene set (HARG) (Figure 6A, Supplemental Figure 9B, and Supplemental Tables 2 and 3). Next, we defined a set of genes whose expression is restored by JH4 treatment (HARG/JR) (298 genes; Supplemental Figure 9B and Supplemental Table 3).

Figure 3. Progerin is a direct target of the JH4 chemical. (A) Effect of biotinylated JH4 on nuclear deformation of HGPS cells. Cells were incubated with the indicated chemicals for 24 hours and fixed for lamin A/C immunofluorescence analysis (green). Scale bars: 10 μm. (B) Localization of JH4 in progerin-transfected HEK293 cells. JH4-biotin (07), but not biotin alone (08), was stained in the nuclear membrane by streptavidin-HRP in progerin-transfected cells. Pro, progerin. Original magnification, ×200. (C) JH4 localizes in the nuclear membrane. After treatment with biotinylated JH4 for 48 hours, cells were fixed and stained with streptavidin-FITC. Green signal was detected only in JH4-treated HGPS. Scale bars: 10 μm. (D) Specific interaction of JH4 and progerin. Streptavidin-biotin-binding assay using lysates from HGPS cells incubated with biotinylated JH4 or biotin. After incubation with streptavidin-coated magnetic beads, the biotinylated JH4 with the protein complex was isolated. (E) Direct and specific interaction between JH4-biotin and progerin. Cells incubated with the indicated chemicals were lysed with RIPA and incubated with streptavidin-coated beads. Material precipitated by the biotin-streptavidin-bead complex was analyzed by Western blot analysis. GFP-progerin was precipitated by the biotin-streptavidin complex in JH4-biotin–treated cells. However, other nuclear membrane proteins such as lamin B (LMNB), emerin and GFP–lamin A, and p53 were not associated with JH4-biotin. WCL, whole-cell lysate.
To verify these transcriptomic results, we confirmed that the expression of several genes such as *IL33*, breast cancer 1 (*BRCA1*), bloom syndrome RecQ-like helicase (*BLM*), *RAD51* recombinase, *IL6*, *IL8*, and tumor necrosis factor superfamily member 18 (*TNFSF18*) was altered at both the protein and mRNA levels after JH4 treatment (Figure 6, B and C). By contrast, farnesyltransferase inhibitor (FTI) treatment did not revert the expression changes of the HARG/JR genes (Supplemental Figure 9C), indicating that the...
effects of JH4 and FTI are markedly different. Since the alternation of gene expression is due to a different genetic background but not to aging, we examined the expression of several genes induced or suppressed in aged fibroblasts and HGPS cells in other kinds of human aged fibroblasts and different HGPS cells. Expression levels of IL33, RAD51, and BRCA1 were reduced in 3 kinds of aged fibroblasts (N81; N72, 72-year-old; and N94, 94-year-old) and HGPS-2 cells compared with the levels detected in fibroblasts from N9 (Supplemental Figure 9D). In contrast, IL6 and IL8 were overexpressed in these cells (Supplemental Figure 9D). This result suggested that alternation of these expression levels would result from aging. To investigate the biological significance of these transcriptional alter
Figure 6. Aging-related gene expression is recovered by JH4 treatment. (A) Heatmap of gene expression profiles showing a difference of at least 2-fold in both cell lines. Colors represent values normalized to the range of intensity for each gene in all conditions. (B) Reduced expression of IL33, BRCA1, BLM, and RAD51 in HGPS cells was reverted by JH4. Conversely, IL6, IL8, and TNFSF18 overexpression was suppressed by JH4 treatment. (C) JH4 could alter HGPS-specific gene expression. RAD51, BRCA1, cyclin B1, and CDC25C protein expression levels were increased by treatment with JH4. (D) Network analysis using cBioportal. Genes in the HARG/JH group were clustered into chromosome separation, DNA replication, DNA repair, and cell-cycle groups. (E) JH4 overcame progerin-induced cell-cycle arrest. Cell-cycle arrest at the G2/M phase by progerin transfection was released by JH4 treatment (5 μM, for 24 hours).
LmnaG609G/G609G mice following JH4 injection. P values were determined by Student’s t test. (C) Increase in muscle strength following JH4 injection. Grip strength was measured for Lmna+/G609G, Lmna+/−, and Lmna−/− male mice by pulling the spring balance after treatment with JH4 or vehicle alone. P value was determined by Student’s t test. (D) Growth morphology of thymus and spleen from 9-week-old LmnaG609G/G609G mice treated with vehicle or JH4. An obvious enlargement of both organs after JH4 treatment was observed. (E) JH4 injection time–dependent organ size increase. Compared with vehicle-injected mice, the size of several organs was gradually increased by treatment time. Numbers above each bar indicate the fold increase at each time point (see also Supplemental Figure 14). (F) JH4 injection promoted the cell cycle of splenocytes obtained from 7-week-old mice (injected 6 times). Cell-cycle promotion was observed in JH4-treated cells. (G) Rescue of cell and tissue defects. Foot pad skin was thickened and dermal connective tissue was enriched in JH4-treated animals (upper panels). Kidney tissue was well organized after JH4 treatment. Tissues were obtained from 18-week-old (vehicle) and 22-week-old (JH4) mice. Scale bar: 80 μm, original magnification ×10. (H) Increase in cell density and reduction of deformed cell nuclei in JH4-treated heart tissue obtained from the same mice above. On the basis of the histology, nuclear morphology (right) and cell density (left) were counted for 3 different mice and 5 different regions per mouse by 3 researchers. P values were determined by Student’s t test. (I) Representative images of heart muscle. Arrows indicate deformed nuclei. Scale bar: 10 μm. (J) JH4 suppressed progerin and induced RAD51 and phosphorylated CDK2 (p-CDK2). Western blot analysis was performed using extracts from liver and kidney tissue obtained from 18-week-old sibling mice.

JH4 blocks progerin-induced lamin A aggregation in ER during the G2/M phase. Because many of the JH4-responsive genes shown in Supplemental Table 4 and Figure 4B are involved in cell-cycle and chromosome regulation and progerin induces cell-cycle arrest at the G2/M checkpoint (Figure 6E), we examined the effect of JH4 on the behavior of lamin A at the G2/M phase. Dispersal of lamin A upon treatment with Taxol (paclitaxel) was observed in young, but not aged, cells. Rather, lamin A surrounded chromosomal or nuclear DNA (Supplemental Figure 10A, white arrows). However, treatment with JH4 could promote lamin A elimination in aged cells (Supplemental Figure 10A, yellow arrows). We also observed that progerin-induced lamin A aggregation in nocodazol- or Taxol-treated cells was abolished by JH4, but not by FTI-277 (Supplemental Figure 10, B and C). To gain further insight into this observation, we analyzed the localization of progerin in nocodazol- or Taxol-treated cells. Cytoplasmic aggregates in nocodazol- or Taxol-treated cells were undetectable upon JH4 treatment, but not after incubation with FTI-277 (Supplemental Figure 11A). Given the role of the ER in nuclear lamina remodeling during the G2/M phase (23), we investigated the subcellular localization of lamin A and an ER-specific marker (35) in JH4-treated cells and found that aggregates of lamin A in the ER disappeared after JH4 treatment (Supplemental Figure 11B). Next, we checked the location of progerin after arrest at the M phase by Taxol treatment and found that JH4 also blocked ER-associated progerin during the M phase (Supplemental Figure 11C). To confirm whether M-phase aggregates contain lamin A–progerin complexes, we costained lamin A–GFP and progerin in Taxol-treated cells. Colocalized lamin A and progerin in the non-nuclear region was abrogated by JH4 (Supplemental Figure 11D). This result indicates that progerin disrupts lamin A disassembly and reduction during the G2/M phase by direct binding to this nuclear envelope protein. Indeed, it has been recently reported that HGPS cells show downregulated and defective poly(ADP-ribose) polymerase 1 (PARP1) in G2/M-phase progression (36).

Therapeutic effect of JH4 on the HGPS mouse model. To evaluate the therapeutic potential of JH4 in vivo, we injected i.p. 10 mg/kg of this compound twice per week into 4-week-old Lmna+/G609G/HGPS mice, a suitable murine model of human HGPS (37). After 4 weeks of treatment, we observed a significant increase in BW (Figure 7, A and B), which continuously increased out to 12 weeks (Supplemental Figure 12A). By contrast, vehicle-treated mice showed a significant decrease in BW after the eighth week (Figure 7B and Supplemental Figure 12A). At 11 to 12 weeks, the difference in BW between vehicle- and JH4-treated mice reached 4 g (about 25%; Supplemental Figure 12A). In addition, injection of JH4 increased grab strength (Figure 7C) and suppressed hair loss at week 15 (Supplemental Figure 12B). Consistently with the increase in BW, the size of different organs was increased by JH4 administration (Figure 7D). Moreover, organ size recovery was fully dependent on the number of injections given (Figure 7E and Supplemental Figure 13), such that 5 weeks after the start of treatment (10 injections), thymus and spleen volumes were enlarged by approximately 2-fold when compared with tissues from vehicle-treated mice (Figure 7E). Since progerin induces a cell-cycle arrest that seems to be critical for growth retardation (17, 18, 37), we also analyzed cell-cycle dynamics and found that it was also promoted by JH4 (Figure 7F and Supplemental Figure 12C) in a number of injection-dependent ways (Supplemental Figure 13, A7 and B7). Histological analysis revealed that JH4 increased skin thickness, reduced kidney destruction, and alleviated nuclear deformation (Figure 7G and Supplemental Figure 12D). To obtain more detailed evidence, we measured the cell density and frequency of deformed nuclei in heart (Figure 7, H and I), liver (Supplemental Figure 12E), lung (Supplemental Figure 12F), and kidney (Supplemental Figure 12G). In all examined tissues, JH4 treatment increased cell density and reduced the number of deformed nuclei. For example, the reduction in cell numbers in the same region or in tubes and...
Discussion

The aging process is complicated, and a full understanding of the molecular mechanisms controlling it is not likely to be realized in the near future. However, some basic clues are known from studying human progeria syndromes such as HGPS (39–41). Although HGPS is caused by progerin, a product of a point mutation in the LMNA gene, progerin is also expressed in normal aged cells as well as in cancers (14). Thus, understanding the pathological function of progerin is useful for the development of a therapeutic strategy to treat HGPS as well as for the prevention of aging.

In fact, over the past decade, the farnesylation of progerin has been targeted for drug development to treat HGPS. (24, 42). Because the farnesylated domain of progerin rigidly anchors it to the nuclear membrane, it may induce nuclear deformation. Blocking farnesylation by FTIs in HGPS cells restores nuclear morphology, cell proliferation, and heterochromatin organization. Moreover, treatment with FTIs in HGPS-like mouse models improves lifespan, growth, and BW and relieves premature aging symptoms such as bone defects (23). In particular, FTIs can extend the lifespan of Zmpste24+/− mice (24). However, a clinical trial of FTI did not produce promising results (43), indicating that other targets should be explored.

In this study, we found that the most important target of progerin is lamin A/C. Indeed, progerin shows strong binding affinity for lamin A/C but not lamin B (Figure 1E). On this basis, we found new chemicals that can block the interaction between progerin and lamin A/C through direct interaction with progerin (Figure 2B and Figure 3E). Since the chemical JH4 can bind selectively to progerin, there would be very minimal, nonspecific side effects. In our study, JH4 did not show an effect on the progerin-independent progeria model (Zmpste24+/− mice; Supplemental Figure 14C) or on mouse and monkey cell lines (data not shown). These results indicate that JH4 has potential use as a progerin-inhibitory drug.

Although JH4 did not fully overcome the short lifespan of the Lmna+/G609G/G609G mouse model, given the low dose and frequent injections of the compound, an optimized treatment strategy using JH4 could offer an effective therapy. Indeed, the same scheduled injection produced more obvious effects in Lmna+/G609G/G609G mice (Figure 8), and JH4 appeared to be excreted within 24 hours.

In summary, in this study, we have identified 3 compounds, JH1, JH4, and JH13, that efficiently block progerin–lamin A/C
binding and reverse or alleviate different senescence features characteristic of HGPS cells, including growth arrest, SA-β-gal accumulation, and nuclear aberrations. Moreover, we observed similarly favorable effects of JH chemicals on normal aged fibroblasts, which also express progerin. Indeed, JH4 suppressed SA-β-gal activity and increased the proliferation of these cells. Moreover, many genes altered by JH4 treatment were common between HGPS and aging cells. The fact that this compound also rescued several progeroid features of Lmna<sup>G609G/G609G</sup> mice and extended the lifespan of this HGPS mouse model represents a proof of principle for the in vivo relevance of blocking progerin–lamin A/C interaction as a new treatment strategy for HGPS and, eventually, for age-associated alterations involving nuclear envelope abnormalities.

**Methods**

**Animal experiments.** Progerin-heterozygous Lmna<sup>−/G609G</sup> mice were provided by Carlos López-Otín (Universidad de Oviedo, Asturias, Oviedo, Spain). Timed matings of heterozygous Lmna<sup>−/−</sup> and homozygous Lmna<sup>G609G/G609G</sup> males and females were conducted, and chemical was intraperitoneally injected for indicated weeks or months (Figure 8, E–G). All mice were maintained on outbred C57BL6/J backgrounds.

**Histology.** Following sacrifice at the end of the injection regimen, the indicated organs (foot pad, kidney, and heart) were isolated in PBS and fixed in 10% formaldehyde in PBS at 4°C for 6 to 12 hours. Tissue was dehydrated using ethanol (50%, 70%, 90%, 3 × 100%; 5 minutes each) and paraffin embedded for sectioning. Sections (5-μm-thick) were used for H&E staining.

**Chemical screening and chemical synthesis.** For chemical screening, an ELISA was established by modifying a previously described platform (44). Details on the synthetic methods used are provided in the Supplemental material.

**Cell culture and reagents.** Human fibroblast cells were obtained from the Coriell Cell Repositories. Human cancer cell lines (HCT116, A549, and HEK293), obtained from ATCC, were maintained in liquid medium containing 10% FBS and 1% antibiotics at 37°C. Nonhuman cell lines such as Vero (monkey), HICIC7 (mouse), MDCK (canine), and Mv1Lu (mink) were obtained from the Korean Cell Line Bank (KCLB, Seoul, Korea).

**Plasmids and transfection.** GFP-progerin and GFP-fused lamin A expression vectors were provided by T. Misteli (National Cancer Institute [NCI], Bethesda, Maryland, USA). GFP-lamin B1 and lamin C were provided by J. Lammerding (Brigham and Women’s Hospital/ Harvard Medical School, Boston, Massachusetts, USA). Mutant forms of lamin A were gifts of F. Tesson (University of Ottawa, Ottawa, Ontario, Canada). The Myc-ARF vector and OST-LAMINA A50 vector were purchased from Addgene. Transfection was performed using JetPEI (Polyplus Transfection) following the manufacturer’s protocol.

**Recombinant proteins and GST pull-down assay.** For investigation of protein-protein interactions, recombinant proteins were generated. Recombinant lamin A N-terminal region (His-LMNA-N; aa residues 1-300) and middle region (His-LMNA-M; 301-564 aa) were cloned into His-tagged pPROEXHT. GST-fused recombinant lamin A-C terminal region (GST-LMNA) and progerin C-terminal region (GST-progerin) were produced by PCR cloning 100 aa upstream of the termination codon through PCR. Full-length p14<sup>ARF</sup> was produced by PCR, cloned into the pGEX vector, and confirmed by DNA sequencing. These recombinant proteins were purified using a nickel column. For the binding assay, GST-bead-lamin A/C or progerin-C was incubated with lysate from 293 cells that had been transfected with recombinant lamin A N-terminal region, lamin A middle region, GFP-lamin A, lamin B or lamin C for 2 hours at room temperature. After washing twice with PBS and once with RIPA buffer (50 mM Tris, 150 mM NaCl, 1% NP-40, 0.5% deoxycholate, and 0.5% SDS), precipitated materials were collected and subjected to SDS-PAGE and Western blotting using anti-GFP, His, and GST Abs.

**Streptavidin-biotin-binding assay.** Biotin-conjugated JH4 was produced by chemical synthesis (see the Chemical Synthesis section in the Supplemental materials). To determine the location of JH4–biotin, cells were incubated with biotin or biotin-JH4 for the indicated durations (Figure 3) and fixed with 2% PFA. After permeabilization, blocking with BSA, and prereaction with H<sub>2</sub>O<sub>2</sub> to eliminate endogenous HRP activity, cells were incubated with streptavidin-HRP (Sigma–Aldrich) and DAB solution (Vector Laboratories). To investigate the direct binding of JH4 and progerin, cells lysates, precleared by streptavidin-bead, were incubated with biotin-JH4 or biotin and streptavidin-bead, sequentially. The precipitated materials were subjected to SDS-PAGE and Western blot analysis.

**Microarray data.** The full data set is available in the NCBI’s Gene Expression Omnibus (GEO) database (GEO GSE84147; www.ncbi.nlm.gov/geo/query/acc.cgi?acc=GSE84147).

**Statistics.** All data are expressed as the mean ± SEM. Statistical significance was determined using a 2-tailed Student’s t test. For all experiments, a P value of less than 0.05 was considered statistically significant.

**Study approval.** All in vivo studies were performed at an Association for Assessment and Accreditation of Laboratory Animal Care–certified facility, in compliance with animal policies approved by Pusan National University (Busan, Korea) and the Universidad de Oviedo (Asturias, Oviedo, Spain).

**Author contributions**

SJL, YSJ, MHY, SK, AYO, JHL, SYJ, TGW, HYC, SKK, KJC, HYL, KL, GK, MKN, NCH, CB, JMF, and BJF performed the experiments. SJL, YSJ, and BJP conceived the experimental designs. CLO, GYS, and BJF wrote the manuscript. SJL, GYS, and BJP analyzed the data.

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